Processing ddRAD for population history inference

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01-06-2016

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- Lots of data returned
- Stable software pipelines for using these data

A Quick Note

Slides that contain ddRAD specific info will be noted. Some steps can be used with multiple data sources.

The Edwards Plateau

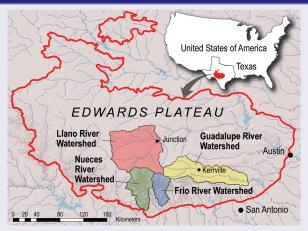
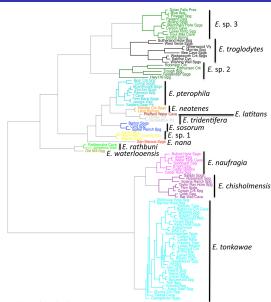


Figure 1: Image: AGU



13 putative species of *Eurycea*

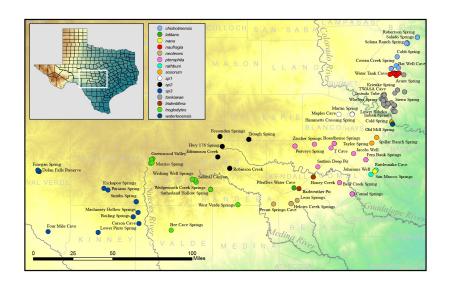
13 putative species of *Eurycea* All of which are fairly threatened by development



- 100 nucleotide changes

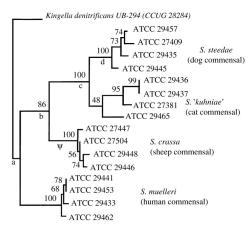
How many species of ${\it Eurycea}$ are there, really?

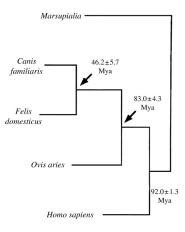
How many species of *Eurycea* are there, really? And is there introgression between them?



Simonsiella phylogeny

vertebrate phylogeny



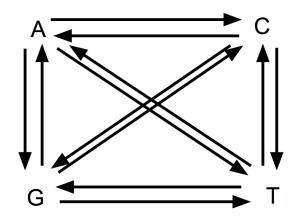


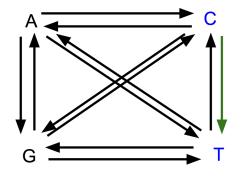
Maximum likelihood

Maximum likelihood is a framework for estimating phylogeny by modeling the process of evolution that generated our sequence data

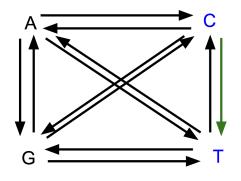
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Probability of C to T change Equilibrium frequency of C 0 * .25 = 0



Probability of C to T change Equilibrium frequency of C .75 * .25 = .1875

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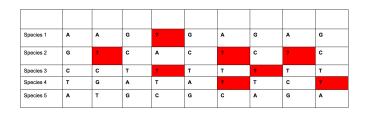
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Problems

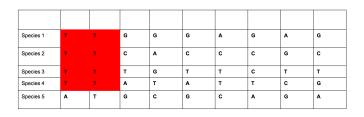
- Problems
- Missing data

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- Biased Missing data

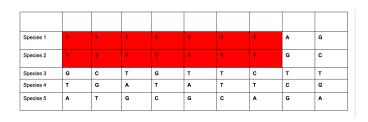


- Problems
- Biased Missing data

Missing data concentrated in specific individuals



- Missing data concentrated in specific individuals
- Missing data concentrated in certain loci in your data matrix



- Problems
- Model misspecification

Problems

 Model misspecification: when your data are not adequately described by your model

Today, we'll be visualizing our data at every step to try and minimize a bias in which individuals have missing data

Phylogenetics

We'll also look at ways to make sure we aren't overly-conservative in our choosing of SNPs (i.e., biasing our collection towards sites that exhibit little change)

One of the things that makes RADseq, and especially ddRADseq, so cheap is the pooling of samples

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The way we recover individual samples is via demultiplexing

This allows for the cost-saving properties of batching, without the cost-increasing properties of synthesizing oligonucleotides.

The STACKS step for this is called Process RAD Tags

Output

• FASTQ files

Let's look at the output

- FASTQ files
- Reads, grouped by individual

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- Reads, grouped by individual
- We haven't done any SNP calling. This is just the step that gets our data ready to do that

For this step, we will use ustacks

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Key Parameters

- -m: Minimum stack depth
- -M: Maximum mismatches allowed between reads in a stack

Other Parameters

• -i: ID for this sample

Exercise

One of the issues we discussed was biased missing data

Catalog Building

Once we have our within-individual stacks, we build a catalog of loci across individual catalogs (cstacks)

Catalog Building

Key Parameters

• -n: number of mismatches to allow between a putative tag, and a tag in the catalog

Outputting Data for Phylogenetics

We use populations for this.

Outputting Data for Phylogenetics

A new file is needed, here: the population map

Outputting Data for Phylogenetics

Key Parameters

- -r: Percentage of individuals in a population that must have a locus to output it
- -m: Minimum stack depth at a locus

Exercise

Looking at this output is easy.

Exercise

Looking at this output is easy. But we can also look in a more complex way: countPhyloMissing.sh and plotPhyloMissing.py

RAxML approximate likelihoods

```
323320
        -288336.664115
323340
         -84377.460743
323360
         -27407.770692
323380
         -10281.525371
323390
          -1699.210794
```

Lastly, let's build the tree

 $RAxML\ Approximate\ final\ L\ scores$

Lastly, let's build the tree

Garli

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```
323320 -287898.7874
323340 -84289.0263
Garli 323360 -27384.6012
323380 -10273.63021
323390 -1697.6284
```