

INCOMPLETE DRAFT: Smashing single cells into k -mer sketches

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Abstract

We introduce `kmermaid`, a lightweight computational method for quantifying similarity between RNA-seq transcriptomes across species. As `kmermaid` skips both alignment and gene orthology assignment, it can be applied to transcriptomes from organisms with no or poorly annotated genomes, and can perform the task of quantifying similarities much faster than current methods without loss of sensitivity. By enabling analyses across divergent species' transcriptomes in an orthology-, genome- and gene annotation-agnostic manner, `kmermaid` illustrates the potential of non-model organisms in building the cell type evolutionary tree of life.

Single-cell RNA-sequencing is a powerful technology for identifying cell types in a variety of species. However, the task of identifying even known cell types in species with poorly annotated genomes is nontrivial, as orthologous gene mapping remains an unsolved problem [1,2] and most of the 8.7 million Eukaryotic species on Earth have no sequenced genome [3]. Thus, there is an unmet need to quantitatively compare single-cell transcriptomes across species, without the need for orthologous gene mapping, gene annotations, or a reference genome. Short, k -long sequence substrings, or k -mers, have been proposed for comparing single cells [4] as they are a fast, simple way to create cell-cell similarities. Combining k -mers from putatively translated RNA-seq reads with reduced amino acid alphabets [5,5,6,7], we developed a method to identify known cell types in a non-model organism using a reference atlas from another organism, without the need for a genome or gene annotation from the non-model organism. This method relies solely on divergence time between species, which we show can be estimated from RNA-seq nucleotide k -mers (Supplemental Figure [??]). We benchmark the genome-agnostic method on the Quest for Orthologs Opisthokonta dataset, showing that k -mers from reduced amino acid alphabets are sufficient to estimate orthology. Using human amino acid sequences, we show that one can extract putative protein-coding reads from 239 Opisthokonta species in ENSEMBL, and present the best k -mer size and alphabet for different

divergence times. We apply this method on a bulk comparative transcriptomic dataset consisting of nine species and six tissues [10], showing that we can recapitulate the results as using only reads mapping to 1:1 orthologs of protein-coding genes, and we are able to resolve ... which can only be seen by using the reference-free k -mer method.

We then show that reduced amino acid protein k -mers can be used for comparing single-cell transcriptomes across mouse, human, and zebrafish hematopoiesis. We find that using k -mers has the advantage of resolving ... in comparison to using read counts from 1:1 gene orthologs. Using k -mers, we were able to resolve cell types ... , which was hidden using read counts alone. Thus, we have shown the reference-free method using the k -mers from single cells is a novel, annotation-agnostic method for comparing cells across species that is capable of identifying cell states unique to a particular organism, helping to build the cell type evolutionary tree of life.

Figure 1 – Overview of kmermaid pipeline

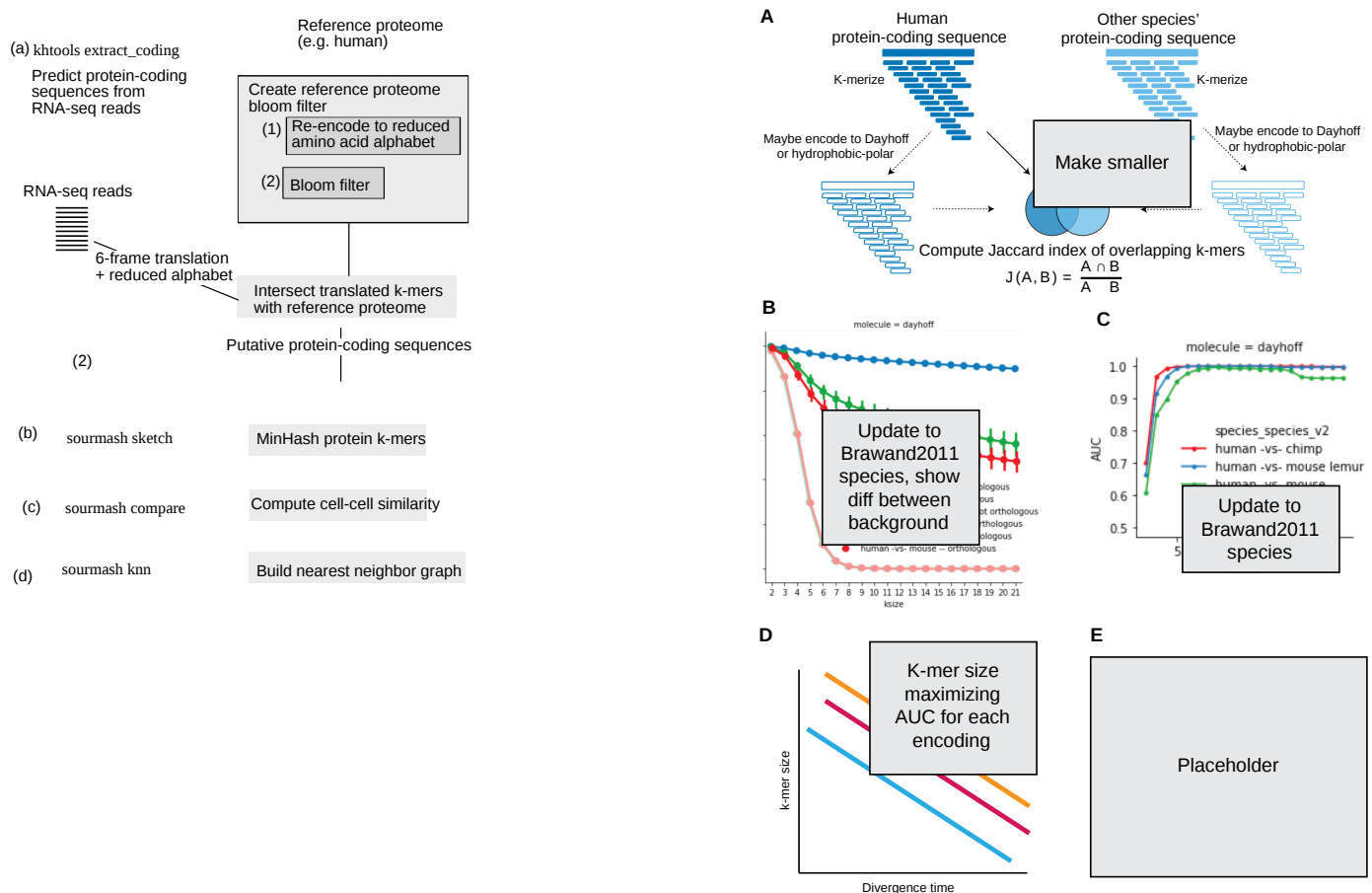


Figure 1: A. Overview of the **kmermaid** pipeline. **(a, b, c)** **kmermaid** consists of a protein-coding prediction phase **(a)** that is invoked by the command `khtools extract_coding`, a k -mer sketch computation phase **(b)** invoked by the command `sourmash sketch`, a signature similarity comparison phase **(c)** invoked by the command `sourmash compare`, and an optional database-creation phase **(d)** invoked by the command `sourmash index`. The coding prediction phase has three components: (1) six-frame translation, removal of stop-codon frames, and subsequent k -merization of RNA-sequencing reads; (2) a degenerate protein alphabet which allows for protein-coding detection from a wide variety of species; (3) a bloom filter containing known protein-coding sequences from a well annotated organism; and (4) computation of the Jaccard index of translated RNA-seq reading frames. The sketch computation phase involves randomly subsetting the degenerate peptide k -mers using a MinHash algorithm. The sketch comparison phase consists of computing the Jaccard intersection of MinHashed degenerate peptide k -mers between all pairs of samples.

To determine whether short segments of sequences could detect gene orthologues, we k -merized orthologous genes derived from the ENSEMBL version 97 [11] COMPARA database [12] (Figure [1]). We compared human protein sequences to orthologous chimpanzee, mouse, (orangutan, bonobo, gorilla, macaque, opossum, platypus, chicken) protein sequences, as these are species used in [10]. As

a background, we randomly chose 10 non-orthologous genes relative to the human gene. In addition to k -merizing the protein-coding sequence, we also re-encoded the protein-coding sequence into a six-letter Dayhoff alphabet [13], a nine-letter encoding [8], and a two-letter hydrophobic-polar encodings [14,15], show in Table [1].

We found that, consistent with previous knowledge, that 1:1 orthologues had higher k -mer similarities as determined by the Jaccard Index. This approach is similar to SwiftOrtho [8], a k -mer based orthology relationship finder.

Additionally, more recently diverged genes had higher k -mer similarity as well.

Outline

- Kmers can approximate orthologies
 - Jaccard similarity of orthologues is higher than non-orthologues
 - Benchmarking using <https://orthology.benchmarkservice.org/cgi-bin/gateway.pl>
 - Finding orthologues
 - Gold standard
 - ENSEMBL COMPARA
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 - Orthologous groups/Conserved Domain Database [17]

Figure 2 – k -mers from lossily-encoded putative protein-coding reads faithfully pull out reads from protein-coding genes within amniotes

A

B

C

Use all
Brawand2011
species

Figure 2: Figure 2.

- Overview of kmermaid pipeline
 - Comparison of tissue across species
 - Partition reads to coding/noncoding bins
 - MinHash the Dayhoff-encoded coding sequences
 - Jaccard similarity on the MinHashes
- Which reads are found to have coding features but didn't map to the genome?
- Do these features map to novel genes or gene fusions?
- Kmers can find correct reading from of RNA-seq reads

- Human peptides → human, chimp, bonobo, orangutan, gorilla, macaque, mouse, opossum, platypus, chicken RNAseq from Brawand2011 data
- Comparison to other methods: RNASamba [[18](#)]

Figure 3 – k -mers can pull out only reads from transcription factors and Amniotes can be compared on the MinHashes of their protein-coding sequences

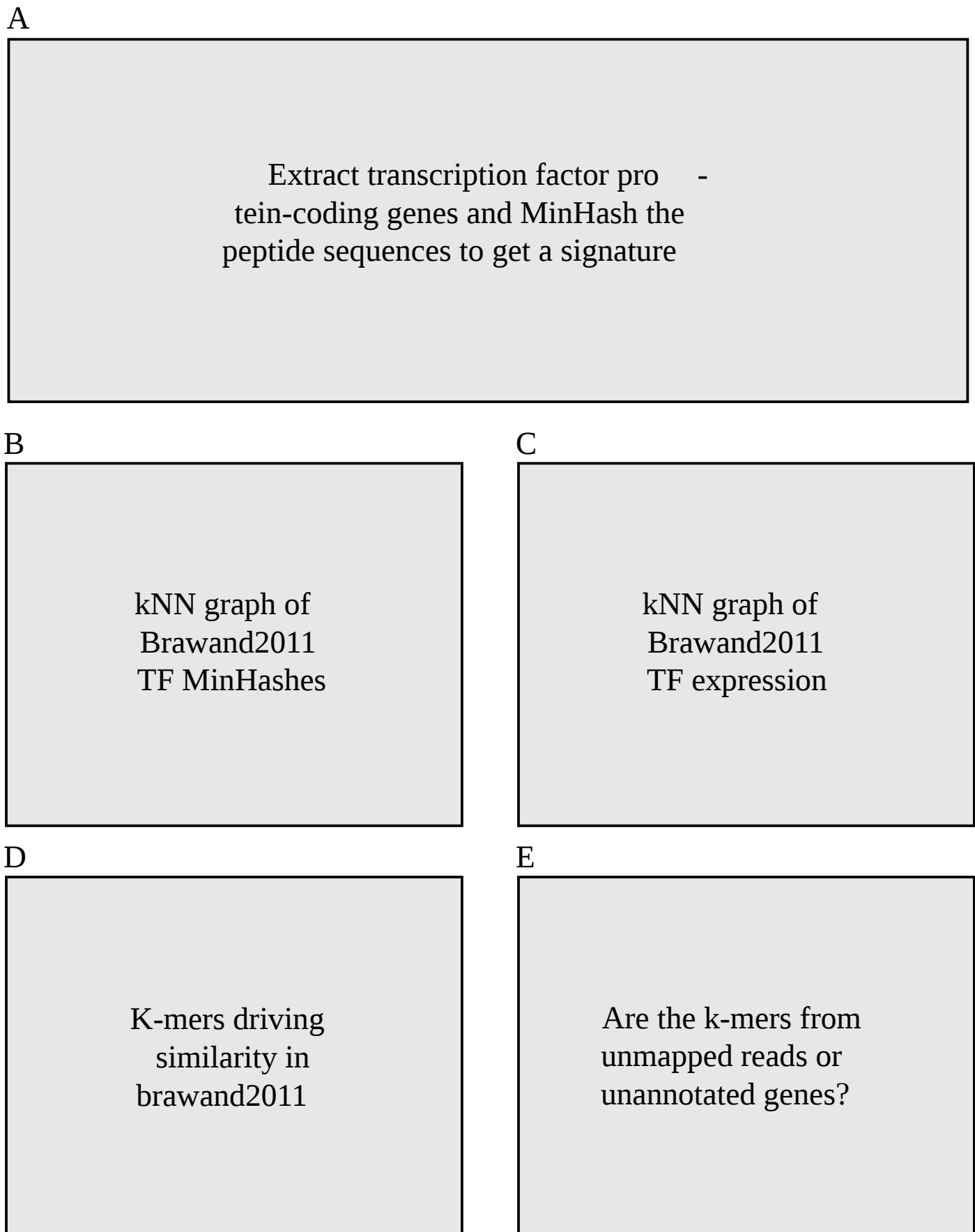


Figure 3: Figure 3.

- Kmers can find only transcription factor reads of TFs from RNA-seq reads
 - Human peptides → human, chimp, bonobo, orangutan, gorilla, macaque, mouse, opossum, platypus, chicken RNAseq from Brawand2011 data

`kmermaid` implements the concept of lightweight orthology assignment using k-mers to the problem of cross-species RNA-seq analyses and achieves unprecedented speed of analysis. By removing the orthology inference step, `kmermaid` opens up the possibility of finding shared and divergent tissue and cell types across a broad range of species, paving the way for evolutionary analyses of cell types across species. `kmermaid` can be used in *de novo* setting for non-model organisms, finding similar cell types within an organism, or finding similar cell types relative to a reference organism, without the need for a reference genome or transcriptome. The memory usage of `kmermaid` is quite low, using only 50MB for extracting coding sequences and 50MB for assigning protein k-mer signatures. As the number of RNA-seq datasets, especially single-cell RNA-seq datasets continues to grow, we expect `kmermaid` to be widely used for identifying cell types in non-model organisms.

`kmermaid` is free and open-source software and is available as Supplementary Data and at <http://github.com/czbiohub/kmermaid> and as a scalable Nextflow workflow at <http://github.com/nf-core/nf-kmermaid>.

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Methods

Methods go here.

Experimental

Primate brain organoid protocols

We did things. One sentence per line. Prefer DOI for references, but for Biorxiv use the URL. DOI example: [16]. Biorxiv example: [27]. Multiple citations per line example: [16,27].

Single-cell capture of primate brain organoids

We did things. One sentence per line. Prefer DOI for references, but for Biorxiv use the URL. DOI example: [16]. Biorxiv example: [27]. Multiple citations per line example: [16,27].

Long read library prep

We did things. One sentence per line. Prefer DOI for references, but for Biorxiv use the URL. DOI example: [16]. Biorxiv example: [27]. Multiple citations per line example: [16,27].

Short read library prep

We did things. One sentence per line. Prefer DOI for references, but for Biorxiv use the URL. DOI example: [16]. Biorxiv example: [27]. Multiple citations per line example: [16,27].

Sequencing

We did things. One sentence per line. Prefer DOI for references, but for Biorxiv use the URL. DOI example: [16]. Biorxiv example: [27]. Multiple citations per line example: [16,27].

Computational

Caption for Supplemental figure 1

k-mer comparison of orthologous genes

We used ENSEMBL version 97. We did things. One sentence per line. Prefer DOI for references, but for Biorxiv use the URL. DOI example: [16]. Biorxiv example: [27]. Multiple citations per line example: [16,27].

Extraction of putative coding reads from RNA-seq

We did things. One sentence per line. Prefer DOI for references, but for Biorxiv use the URL. DOI example: [16]. Biorxiv example: [27]. Multiple citations per line example: [16,27].

Supplemental Methods

Table 1: Dayhoff and hydrophobic-polar encodings are a reduced amino acid alphabet allowing for permissive cross-species sequence comparisons. For example, the amino acid sequence SASHAFIERCE would be Dayhoff-encoded to bbbdbfecdac , and HP-encoded to phpphhpppp .

Amino acid	Property	Dayhoff	Hydrophobic-polar (HP)
C	Sulfur polymerization	a	p
A, G, P, S, T	Small	b	A, G, P: h S,T: p
D, E, N, Q	Acid and amide	c	p
H, K, R	Basic	d	p
I, L, M, V	Hydrophobic	e	h
F, W, Y	Aromatic	f	h

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