Mini Project #3

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Contribution of each group member: 100% for Junmei Fan, 100% for Xi Cui

Question1

```
a) The full logistic model is fitted as below:
```

 $> fit.full <- gl(chd \sim sbp + tobacco + Idl + adiposity + famhist + typea + obesity + alcohol + age, family = binomial, data = SAheart)$

> summary.glm(fit.full)

Call:

glm(formula = chd ~ sbp + tobacco + ldl + adiposity + famhist + typea + obesity + alcohol + age, family = binomial, data = SAheart)

Deviance Residuals:

Min 1Q Median 3Q Max -1.7781 -0.8213 -0.4387 0.8889 2.5435

Coefficients:

Estimate Std. Error z value Pr(>|z|) (Intercept) -6.1507209 1.3082600 -4.701 2.58e-06 *** sbp 0.0065040 0.0057304 1.135 0.256374 0.0793764 0.0266028 2.984 0.002847 ** tobacco ldl 0.1739239 0.0596617 2.915 0.003555 ** 0.0185866 0.0292894 0.635 0.525700 adiposity famhistPresent 0.9253704 0.2278940 4.061 4.90e-05 *** 0.0395950 0.0123202 3.214 0.001310 ** typea obesity -0.0629099 0.0442477 -1.422 0.155095 alcohol 0.0001217 0.0044832 0.027 0.978350 age 0.0452253 0.0121298 3.728 0.000193 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1

(Dispersion parameter for binomial family taken to be 1) Null deviance: 596.11 on 461 degrees of freedom Residual deviance: 472.14 on 452 degrees of freedom

AIC: 492.14

Number of Fisher Scoring iterations: 5

The p values for the following predictors: sbp, adiposity, obesity and alcohol are large. So there is no clear evidence of a real association between chd and sbp(or adiposity, obesity or alcohol).

The hypothesis test is as below:

```
> fit.re<-glm(chd~tobacco+ldl+famhist+typea+age,family=binomial,data=SAheart)
> anova(fit.re,fit.full,test = "Chisq") #Note: The dropped predictors are not significant
Analysis of Deviance Table
```

```
Model 1: chd ~ tobacco + IdI + famhist + typea + age

Model 2: chd ~ sbp + tobacco + IdI + adiposity + famhist + typea + obesity +
alcohol + age
Resid. Df Resid. Dev Df Deviance Pr(>Chi)

1 456 475.69

2 452 472.14 4 3.5455 0.471
```

Hypothesis test:

H0: The coefficients for sbp, adiposity, obesity and alcohol are all zero.

Ha: at least one of the coefficients for sbp, adiposity, obesity and alcohol is not zero at significance level of 5%.

<u>Test statistics</u>: $F^* = \frac{3.5455/4}{472.14/452} \sim F(0.05, 4, 452)$, if $F^* > F(0.05, 4, 452)$ or p-value<0.05, then reject H0, otherwise fail to reject null hypothesis.

Conclusions: Since p-value 0.471>0.05, so we fail to reject H0 and conclude that the following 4 predictors: sbp, adiposity, obesity, and alcohol are not associated with chd.

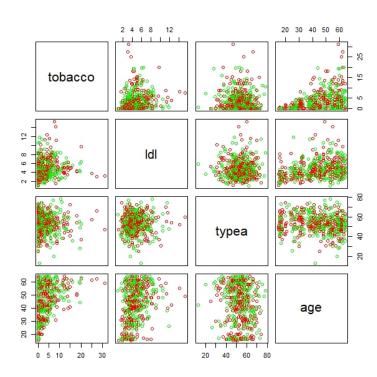
> fit.re\$coefficients

(Intercept) tobacco Idl famhistPresent typea age -6.44644451 0.08037533 0.16199164 0.90817526 0.03711521 0.05046038

The logistic regression coefficients give the change in the log odds of the outcome for a one unit increase in the predictor variable.

- For every one unit change in tobacco while keeping all other predictors constant, the log odds of chd (presence of coronary heart disease versus absence of coronary heart disease) increases by 0.08037533.
- For additional one unit increase in ldl, the log odds of presence of coronary heart disease increases by 0.16199164.
- For additional one unit increase in typea, the log odds of presence of coronary heart disease increases by 0.03711521.
- The indicator variables for famhistPresent have a slightly different interpretation. Having family history of coronary heart disease, versus no family history of coronary heart disease, changes the log odds of chd by 0.90817526.

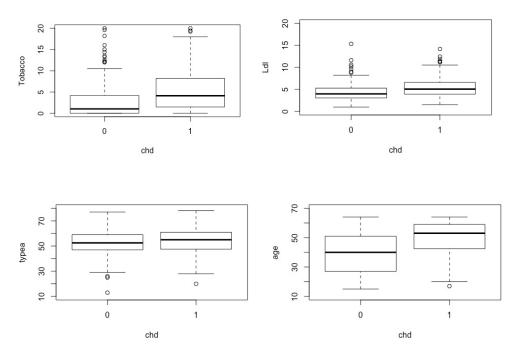
b)



From the scatter plot, points belong to different class of "chd" are overlapped, it seems that neither of these four predictors can strongly predict the class of "chd".

```
> cor(SAheart2)
```

```
tobacco ldl typea age chd tobacco 1.00000000 0.15890546 -0.01460788 0.4503302 0.2997175 ldl 0.15890546 1.00000000 0.04404758 0.3117992 0.2630527 typea -0.01460788 0.04404758 1.00000000 -0.1026063 0.1031558 age 0.45033016 0.31179923 -0.10260632 1.0000000 0.3729733 chd 0.29971754 0.26305268 0.10315583 0.3729733 1.0000000
```



From the correlation coefficients and boxplots, we know that all four of the predictors have some positive correlations with chd. Probably age has the most correlation with chd, tobacco and ldl are the second and typea has the least correlation with chd.

```
c) Fit the logistic regression model:
```

```
glm(formula = chd \sim tobacco + ldl + typea + age, family = binomial, data = SAheart)
```

Deviance Residuals:

```
Min 1Q Median 3Q Max -2.0444 -0.8718 -0.4645 0.9741 2.5118
```

Coefficients:

```
Estimate Std. Error z value Pr(>|z|) (Intercept) -6.334452 0.897809 -7.055 1.72e-12 *** tobacco 0.075031 0.025699 2.920 0.00350 ** typea 0.037914 0.011885 3.190 0.00142 ** age 0.055040 0.009948 5.533 3.15e-08 ***
```

(1) Logistic regression decision boundary equation:

```
\{x: \hat{p}(x) = 0.5\} or beta0 + beta1 *tobacco + beta2 * ldl+beta3 * typea+beta4 * age = 0 => -6.334452+0.075031*tobacco + 0.179891* ldl+0.037914 * typea+0.055040 * age = 0
```

(2) Confusion matrix:

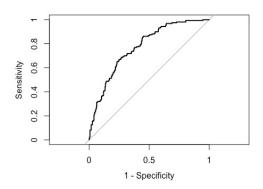
True chd

lr.pred 0 1

0 260 85

1 42 75

- (3) Sensitivity: $\underline{0.4687500}$; Specificity: $\underline{0.8609272}$. So this logistic model set the high threshold to detect chd, although specificity is relatively high, this model might miss a lot of positive chd cases.
- (4) Overall misclassification rate: 0.2748918
- (5) ROC plot:



(6) Cross-validation error rate:

> cv.est [1] 0.2835338

The test error rate using 10-fold cross-validation is 0.2835338.

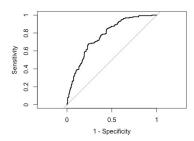
- d) Fit the LDA model
- (1) Decision boundary equation:
- -0.0893448*tobacco -0.1879042* ldl-0.03371824* typea-0.04818174* age = 5.872758
- (2) Confusion matrix:

Ture chd

pred 0 1

0 257 87 1 45 73

- (3) Sensitivity: 0.4562500; Specificity: 0.8509934, sensitivity and specificity are both worth than logistic model.
- (4) Overall misclassification rate: 0.2857143
- (5) ROC plot:



(6) Cross-validation error rate:

> cv.err.lda [1] 0.2943571

The test error rate using 10-fold cross-validation is 0.2943571.

- e) Fit the QDA model:
- (1) Decision boundary equation:

$$-\frac{1}{2}x^{T}\begin{bmatrix}0.0630 & -0.0256 & -0.0039 & -0.0048\\ -0.0256 & 0.1134 & -0.0003 & -0.0044\\ -0.0039 & -0.00027 & 0.0012 & -0.0008\\ -0.0048 & -0.0044 & -0.00084 & -0.0041\end{bmatrix}x + \begin{bmatrix}-0.3824\\ 0.0572\\ -0.0284\\ -0.3373\end{bmatrix}x = 13.1587$$

(2) Confusion matrix:

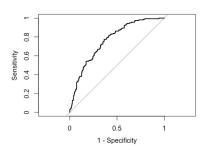
Ture chd

pred 0 1

 $0\ 261\ 88$

1 41 72

- (3) Sensitivity: 0.4500000; Specificity: 0.8642384, comparing to logistic and LDA model, specificity increased a tiny bit.
- (4) Overall misclassification rate: 0.2792208
- (5) ROC plot:



(6) Cross-validation error rate:

> cv.err.qda [1] 0.2898242

The test error rate using 10-fold cross-validation is 0.2898242.

f) Fit the KNN model:

(1) Find the optimal K=18.

> result[err.knn == min(result\$err.knn),]

ks err.knn

18 18 0.2834875

(2) Confusion matrix:

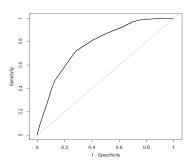
Ture chd

knn.fit 0 1

0 267 93

1 35 67

- (3) Sensitivity: 0.418750; Specificity: 0.884106 specificity increased but sensitivity decreased even more.
- (4) Overall misclassification rate: 0.2770563
- (5) ROC plot:



(6) Cross-validation error rate: The test error rate using 10-fold cross-validation is 0.2834875.

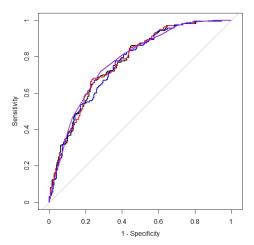
g)

Comparation of four models:

ROC curve:

AUC:

Test error rate by 10folds cross-validation:



	AUC
Logistic Regression	0.7719
LDA	0.7721
QDA	0.7641
KNN	0.779

Logistic Regression	0.2791859
LDA	0.2943571
QDA	0.2898242
KNN	0.2834875

The difference between these four models is small for this "SAheart" dataset. ROC curve and AUC shows the specificity and sensitivity for models, the performances of these four models are similar. However, the cross-validation error rate shows the logistic regression has the lowest test error rate. The response value we aimed to predict is a binary class, the logistic regression perform better is reasonable, so we recommend the logistic regression to do classification for this dataset.

Question2

Fit the QDA by using 10 folds cross-validation:

Test error rate for Model1: "Derection~Lag1+Lag2": 0.4824

Test error rate for Model2: "Derection~Lag1+Lag2+Lag3+Lag4+Lag5+Volume": 0.5072

The test error rate is higher after adding "Lag3, Lag4, Lag5, Volume" to the model, which means the additional predictors could not improve the predictive power of QDA.

T-test of determining the difference of the 10 folds error rate:

H0: the error rate of Model 1 and Model 2 is equal.

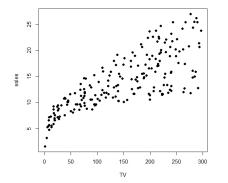
Ha: the error rate of Model 1 and Model 2 is not equal. (True difference in means is not equal to 0)

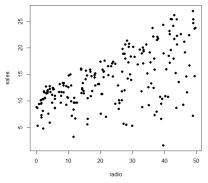
data: cv.err.qda1.10 and cv.err.qda2.10 Test statistics: t = 0.7102 p-value = 0.4881

Conclusion: P-value >0.05, failed to reject H0, and we conclude that the error rate of these two models are the same, the additional predictors don't give any help for QDA.

Question3

(1) Scatter plot:





The scatter plot shows a linear relationship between sales and TV, also sales and radio. The slop shows their positive relationship, and the linear relationship is more obvious for sale and TV.

(2) Point Estimate of Population correlation:
Sales and TV => Rho1: 0.7822244
Sales and radio=> Rho2: 0.5762226
(3) Bootstrap Estimates
Sales and TV: Original= 0.7822244; Bias=-0.0001563068;
Standard error of point estimate=0.02689515
Sales and radio: Original = 0.5762226; Bias=0.0002223246;
Standard error of point estimate=0.05243057
(4) 95% confidence interval:
Sales and TV: (0.7283, 0.8305)

Sales and radio: (0.4734, 0.6748)

The original point estimates of Rho1 and Rho2 are included in the 95% confidence interval from bootstrap, so the bootstrap method here could reliably predict point estimation and its distribution in the population.

R-Code:

```
install.packages("ElemStatLearn")
library(ElemStatLearn)
#data prepared
attach(SAheart)
summary(SAheart)
SAheart$chd<-as.factor(SAheart$chd)
#1.a
fit.full<-glm(chd~sbp+tobacco+ldl+adiposity+famhist+typea+obesity+alcohol+age, family = binomial, data = SAheart)
summary.glm(fit.full)
fit.re<-glm(chd~tobacco+ldl+famhist+typea+age,family=binomial,data=SAheart)
#hypothesis test
anova(fit.re,fit.full,test = "Chisq") #Note: The dropped predictors are not significant
#coefficients
summary.glm(fit.re)
#b
#scatter plot
train<-SAheart[,c(2,3,6,9)]
plot(train,col=(c("red","green")))
SAheart2 = SAheart[,c(2,3,6,9,10)]
str(SAheart2)
#pair correlation
cor(chd)
#plot correlation
pairs(SAheart2)
boxplot(SAheart2[, "tobacco"] ~ SAheart2$chd, ylab = "Tobacco",xlab = "chd", ylim = c(0, 20))
boxplot(SAheart$ldl \sim SAheart2$chd, ylab = "Ldl", xlab = "chd", ylim = c(0, 20))
boxplot(SAheart\$typea ~ SAheart\$chd, ylab = "typea", xlab = "chd",ylim = c(10, 80))
boxplot(SAheart$age ~ SAheart2$chd, ylab = "age", xlab = "chd",ylim = c(10, 70))
par(mfrow = c(1, 1))
#c logistic regression model
fit<-glm(chd~tobacco+ldl+typea+age,family=binomial,data=SAheart)
lr.prob <- predict(fit,SAheart, type = "response")</pre>
#classification
lr.pred <- ifelse(lr.prob >= 0.5, "1", "0")
#confusion matrix
```

```
table(lr.pred, SAheart[, "chd"])
#sensitivity and specificity
cm<-matrix(table(lr.pred, SAheart[, "chd"]),nrow=2,ncol = 2,byrow=FALSE)
c(cm[2,2]/sum(cm[,2]),cm[1,1]/sum(cm[,1]))
#overall misclassification rate
1 - mean(lr.pred == SAheart[, "chd"])
#ROC curve
install.packages('pROC')
library(pROC)
roc.lr < -roc(SAheart[, "chd"], lr.prob, levels = c("0", "1"))
plot(roc.lr, legacy.axes = T)
#10 fold cross validation (Method2)
#Create 10 equally size folds
SAheart<-SAheart[sample(nrow(SAheart)),]
folds <- cut(seq(1,nrow(SAheart)),breaks=10,labels=FALSE)
K<-10
cv.err.lr.10<-sapply(1:K, FUN=function(i){
 #Segement your data by fold using the which() function
 testIndexes <- which(folds==i,arr.ind=TRUE)
 testData <- SAheart[testIndexes, ]
 trainData <- SAheart[-testIndexes, ]</pre>
 train.X<-trainData[,c(2,3,6,9)]
 train.y<-trainData$chd
 test.X < -testData[,c(2,3,6,9)]
 test.y < -testData[,c(10)]
 lr.fit<-glm(chd~tobacco+ldl+typea+age, family=binomial, data=trainData)
 # Estimated probabilities for test data
 lr.prob <- predict(lr.fit, testData, type = "response")</pre>
 # Predicted classes (using 0.5 cutoff)
 lr.pred <- ifelse(lr.prob >= 0.5, "1", "0")
 cv.test <-1-sum(lr.pred == test.y)/length(test.y)
 return(cv.test)}
cv.err.lr=mean(cv.err.lr.10)
cv.err.lr
#d,e
train.X < -SAheart[,c(2,3,6,9)]
train.y<-SAheart[,c(10)]
#LDA
#find decision boundary
s6340.lda \leftarrow function(v, X) {
 \# y = training data response vector (a factor)
 \# X = training data predictor matrix
 N \le - length(y) # no of observations
 K <- nlevels(y) # no of classes
 p \le -ncol(X) \# no of predictors
 n <- as.numeric(table(y)) # class frequencies
 names(n) \le levels(y)
 pi <- n/N # class proportions
 # mean vector
 mu \le matrix(unlist(by(X, y, colMeans)), byrow = T, ncol = p)
 rownames(mu) <- levels(y)
 colnames(mu) <- colnames(X)
 # pooled covariance matrix
 S \leq by(X, y, cov)
 Sigma \leftarrow Reduce("+", lapply(1:K, FUN = function(k) \{(n[k] - 1) * S[[k]]\}))/(N - K)
 # its inverse
 Sigma.inv <- solve(Sigma)
 # delta functions
```

```
delta \leftarrow t(sapply(1:K, FUN = function(k))
  c(-(1/2) * drop(t(mu[k, ]) %*% Sigma.inv %*% mu[k, ]) +
     log(pi[k]), t(mu[k, ]) %*% Sigma.inv)}))
 rownames(delta) <- levels(y)
 colnames(delta) <- c("(Intercept)", colnames(X))</pre>
 # pairwise difference of delta functions
 idx.pair <- combn(K, 2)
 delta.diff <- t(apply(idx.pair, MAR = 2, FUN = function(pair) {
  delta[pair[1], ] - delta[pair[2], ]}))
 rownames(delta.diff) <- apply(idx.pair, MAR = 2, FUN = function(pair) {
  paste0(levels(y)[pair[1]], "-", levels(y)[pair[2]])})
 # multiply intecept difference by 1 to get the cutoff c
 delta.diff[, 1] <- -delta.diff[, 1]
 colnames(delta.diff)[1] <- "Cutoff"
 # result
 result \leftarrow list(N = N, n = n, pi = pi, mu = mu, Sigma = Sigma,
          delta = delta, disc = delta.diff)
 return(result)
our.lda.fit <- s6340.lda(train.y,train.X)
our.lda.fit$disc
#fit by package
library(MASS)
lda.fit <- lda(chd~tobacco+ldl+typea+age,data=SAheart)
lda.pred<- predict(lda.fit, SAheart)</pre>
#fit by function
train.X<-as.matrix(as.data.frame(train.X))</pre>
coeff <- our.lda.fit$disc[1, -1]
coeff
cutoff <- our.lda.fit$disc[1, 1]
score.test <- train.X %*% coeff
#predict class
pred.test <- ifelse(score.test >= cutoff, "0", "1")
#confusion matrix
table(pred.test, train.y)
#sensitivity and specificity
cm<-matrix(table(pred.test, train.y),nrow=2,ncol = 2,byrow=FALSE)
c(cm[2,2]/sum(cm[,2]),cm[1,1]/sum(cm[,1]))
#overall misclassification rate
1 - mean(pred.test == SAheart[, "chd"])
#ROC curve
lda.prob<-lda.pred$posterior[,2]
roc.lda < -roc(SAheart[, "chd"], lda.prob, levels = c("0", "1"))
plot(roc.lda, legacy.axes = T)
#k=10 folds cross-validation
SAheart<-SAheart[sample(nrow(SAheart)),]
cv.err.lda.10<-sapply(1:K, FUN=function(i){
 testIndexes <- which(folds==i,arr.ind=TRUE)
 testData <- SAheart[testIndexes, ]
 trainData <- SAheart[-testIndexes, ]
 train.X < -trainData[,c(2,3,6,9)]
 train.y<-trainData$chd
 test.X < -testData[,c(2,3,6,9)]
 test.y<-testData$chd
 lda.fit<-lda(chd~tobacco+ldl+typea+age, data=trainData)
 lda.pred<-predict(lda.fit,test.X)
 cv.est<-mean(lda.pred$class==test.y)
 return(cv.est)}
cv.err.lda=1-mean(cv.err.lda.10)
cv.err.lda
```

```
#ODA
#find decision boundary
s6340.qda \leftarrow function(y, X) {
 \# y = training data response vector (a factor)
 \# X = training data predictor matrix
 N \le - length(y) # no of observations
 K <- nlevels(y) # no of classes
 p \le -ncol(X) \# no of predictors
 n <- as.numeric(table(y)) # class frequencies
 names(n) \le levels(y)
 pi <- n/N # class proportions
 # mean vector
 mu \le matrix(unlist(by(X, y, colMeans)), byrow = T, ncol = p)
 rownames(mu) <- levels(y)
 colnames(mu) <- colnames(X)
 #covariance matrix
 S \leq by(X, y, cov)
 # its inverse
 Sigma.inv <- lapply(1:K, FUN=function(k){solve(S[[k]])})
 # delta functions
 delta <-t(sapply(1:K, FUN = function(k) \{c(-(1/2) * drop(t(mu[k, ]) %*% Sigma.inv[[k]] %*% mu[k, ]) +
     \log(\operatorname{pi}[k])-(1/2)*\log(\operatorname{drop}(\det(S[[k]])),\operatorname{drop}(t(\operatorname{mu}[k,])\%*\% \operatorname{Sigma.inv}[[k]]),\operatorname{drop}(\operatorname{Sigma.inv}[[k]]))))
 rownames(delta) <- levels(y)
 idx.pair <- combn(K, 2)
 delta.diff <- t(apply(idx.pair, MAR = 2, FUN = function(pair) {
  delta[pair[1], ] - delta[pair[2], ]}))
 #rownames(delta.diff) <- apply(idx.pair, MAR = 2, FUN = function(pair) {
 # paste0(levels(y)[pair[1]], "-", levels(y)[pair[2]])})
 # multiply intecept difference by 1 to get the cutoff c
 delta.diff[, 1] <- -delta.diff[, 1]
 colnames(delta.diff)[1] <- "say"
 result \leftarrow list(N = N, n = n, pi = pi, mu = mu, Sigma = S,
           delta = delta.disc = delta.diff)
 return(result)
our.qda.fit <- s6340.qda(train.y, train.X)
our.qda.fit$disc[,c(1:5)]
sigma1 sigma2<-matrix(our.qda.fit$disc[,c(6:21)],nrow=4,ncol=4,byrow = TRUE)
sigma1 sigma2
#fit qda model and predict class
qda.fit <- qda(chd~tobacco+ldl+typea+age,data=SAheart)
qda.pred<- predict(qda.fit, SAheart)</pre>
#confusion matrix
table(qda.pred$class, SAheart[, "chd"])
#sensitivity and specificity
cm<-matrix(table(qda.pred$class, SAheart[, "chd"]),nrow=2,ncol = 2,byrow=FALSE)
c(cm[2,2]/sum(cm[,2]),cm[1,1]/sum(cm[,1]))
#overall misclassification rate
1 - mean(qda.pred$class == SAheart[, "chd"])
#ROC curve
qda.prob<-qda.pred$posterior[,2]
roc.qda < -roc(SAheart[, "chd"], qda.prob, levels = c("0", "1"))
plot(roc.qda, legacy.axes = T)
#10fold cross validation for qda
SAheart<-SAheart[sample(nrow(SAheart)),]
#Create 10 equally size folds
folds <- cut(seq(1,nrow(SAheart)),breaks=10,labels=FALSE)
cv.err.qda.10<-sapply(1:K, FUN=function(i){
```

```
#Segement your data by fold using the which() function
 testIndexes <- which(folds==i,arr.ind=TRUE)
 testData <- SAheart[testIndexes, ]
 trainData <- SAheart[-testIndexes, ]</pre>
 #Use the test and train data partitions however you desire...
 train.X < -trainData[,c(2,3,6,9)]
 train.y<-trainData$chd
 test.X < -testData[,c(2,3,6,9)]
 test.y < -testData[,c(10)]
 qda.fit<-qda(chd~tobacco+ldl+typea+age, data=trainData)
 qda.pred<-predict(qda.fit,test.X)
 cv.est<-mean(qda.pred$class == test.y)
 return(cv.est)}
cv.err.qda=1-mean(cv.err.qda.10)
cv.err.qda
#f
#10fold cross validation
library(boot)
library(class)
#SAheart<-SAheart[sample(nrow(SAheart)),]
#Create 10 equally size folds
folds <- cut(seq(1,nrow(SAheart)),breaks=10,labels=FALSE)
#Perform 10 fold cross validation
 # Fit KNN for several values of K
 ks < -c(seq(1, 50, by = 1), seq(40, 200, by = 10))
 nks <- length(ks)
 err.rate.train <- numeric(length = nks)
 err.rate.test <- numeric(length = nks)
 err.knn<-numeric(length = nks)
 names(err.rate.train) <- names(err.rate.test) <- ks
 for (j in seq(along = ks)) \{
  cv.err.knn.10<-sapply(1:10, FUN=function(i){
    #Segement your data by fold using the which() function
    testIndexes <- which(folds==i,arr.ind=TRUE)
    testData <- SAheart[testIndexes, ]
    trainData <- SAheart[-testIndexes, ]
    #Use the test and train data partitions however you desire...
    train.X<-trainData[,c(2,3,6,9)]
    train.y<-trainData$chd
    test.X < -testData[,c(2,3,6,9)]
    test.y<-testData$chd
  set.seed(1)
  mod.train <- knn(train.X, train.X, train.y, k = ks[i])
  set.seed(1)
  mod.test <- knn(train.X, test.X, train.y, k = ks[i])
  err.rate.train[j] <- 1 - sum(mod.train == train.y)/length(train.y)
  err.rate.test[j] <- 1 - sum(mod.test == test.y)/length(test.y)
  return (err.rate.test[j])
 })
err.knn[j]<-(mean(cv.err.knn.10))
 #result <- data.frame(ks, err.rate.train, err.rate.test)</pre>
 result <- data.frame(ks, err.knn)
```

```
result[err.knn == min(result$err.knn), ]
# Optimal KNN (K = 18) --- from 10 fold cross validation
train.X < -SAheart[,c(2,3,6,9)]
train.y<-SAheart$chd
library(class)
set.seed(1)
knn.fit < -knn(train.X, train.X, train.y, k = 18, prob = T)
knn.prob <- attr(knn.fit, "prob") # prob is voting fraction for winning class
knn.prob <- ifelse(knn.fit == "1", knn.prob, 1 - knn.prob) # now it is voting fraction for Direction == "1"
#confusion matrix
table(knn.fit, SAheart[, "chd"])
#sensitivity and specificity
cm<-matrix(table(knn.fit, SAheart[, "chd"]),nrow=2,ncol = 2,byrow=FALSE)
c(cm[2,2]/sum(cm[,2]),cm[1,1]/sum(cm[,1]))
#misclassification rate
1 - mean(knn.fit == train.y)
#roc plot for knn
roc.knn < -roc(train.v, knn.prob, levels = c("0", "1"))
plot(roc.knn,add = T, legacy.axes = T)
#g
plot(roc.lr, legacy.axes = T)
plot(roc.lda, add = T, col="red")
plot(roc.qda, add = T, col = "blue")
plot(roc.knn,add = T,col = "purple")
auc(roc.lr)
auc(roc.lda)
auc(roc.qda)
auc(roc.knn)
#Q2
library(ISLR)
attach(Smarket)
#QDA for Derection~Lag1+Lag2
qda1.fit <- qda(Direction ~ Lag1 + Lag2, data = Smarket)
qda1.pred<-predict(qda1.fit, Smarket)
1 - mean(qda1.pred$class == Smarket[, "Direction"])
qda2.fit <- qda(Direction ~ Lag1 + Lag2 + Lag3 + Lag4 + Lag5 + Volume, data=Smarket)
qda2.pred<-predict(qda2.fit, Smarket)
1 - mean(gda2.pred$class == Smarket[, "Direction"])
#10 fold cross-validation
Smarket<-Smarket[sample(nrow(Smarket)),]
folds <- cut(seq(1,nrow(Smarket)),breaks=10,labels=FALSE)
K<-10
#QDA for Derection~Lag1+Lag2
cv.err.qda1.10<-sapply(1:K, FUN=function(i){
 testIndexes <- which(folds==i,arr.ind=TRUE)
 testData <- Smarket[testIndexes, ]
 trainData <- Smarket[-testIndexes, ]
 train.X<-trainData[,c("Lag1","Lag2")]
 train.y<-trainData$Direction
 test.X<-testData[,c("Lag1","Lag2")]
 test.y<-testData$Direction
 qda.fit <- qda(Direction ~ Lag1 + Lag2, data=trainData)
 qda.pred<-predict(qda.fit,test.X)
 cv.est<-mean(qda.pred$class==test.y)
 return(cv.est)}
```

```
1-mean(cv.err.qda1.10)
#QDA for Derection~Lag1+Lag2+Lag3+Lag4+Lag5+Volume
cv.err.qda2.10<-sapply(1:K, FUN=function(i){
 testIndexes <- which(folds==i,arr.ind=TRUE)
 testData <- Smarket[testIndexes, ]</pre>
 trainData <- Smarket[-testIndexes, ]</pre>
 train.X<-trainData[,c("Lag1","Lag2","Lag3","Lag4","Lag5","Volume")]
 train.y<-trainData$Direction
 test.X<-testData[,c("Lag1","Lag2","Lag3","Lag4","Lag5","Volume")]
 test.y<-testData$Direction
 qda.fit <- qda(Direction ~ Lag1 + Lag2 + Lag3+Lag4+Lag5+Volume, data=trainData)
 qda.pred<-predict(qda.fit,test.X)
 cv.est<-mean(qda.pred$class==test.y)
 return(cv.est)}
1-mean(cv.err.qda2.10)
#t test
t.test(cv.err.qda1.10,cv.err.qda2.10)
#O3
install.packages('boot')
library(boot)
#Data preparing
ad<-read.table("C:/Users/xicui/Desktop/Advertising.csv", header = TRUE, sep=",")
plot(sales \sim TV, data = ad, pch=19)
plot(sales \sim radio, data = ad, pch=19)
#point estimate
rho1<-cor(ad$sales,ad$TV)
rho2<-cor(ad$sales,ad$radio)
c(rho1,rho2)
#bootstrap estimate
fit1.fn <- function(data, index) {
 data=data[index,]
 result<-cor(data$sales,data$TV)
 return(result)}
n <- nrow(ad)
fit1.fn(ad, 1:n)
reg1.boot \leftarrow boot(ad, fit1.fn, R = 1000)
reg1.boot
fit2.fn <- function(data, index) {
 data=data[index,]
 result<-cor(data$sales,data$radio)
 return(result)}
fit2.fn(ad, 1:n)
reg2.boot \leftarrow boot(ad, fit2.fn, R = 1000)
reg2.boot
#95% confidence interval
boot.ci(reg1.boot,type="perc")
boot.ci(reg2.boot,type="perc")
```