

## Band Transects

### **Purpose**

To determine abundance and distribution of rare and clumped organisms not adequately sampled by quadrats

### **Materials**

- 2 dive slates with pencils
- 2 1.5 meter PVC rods
- 2 15 m tapes w/carabiners
- 2 underwater band transect data sheets (Appendix K)

### **Personnel**

- 2 SCUBA equipped observers experienced in the identification and search image needed for species listed in Table 6.

### **Methods**

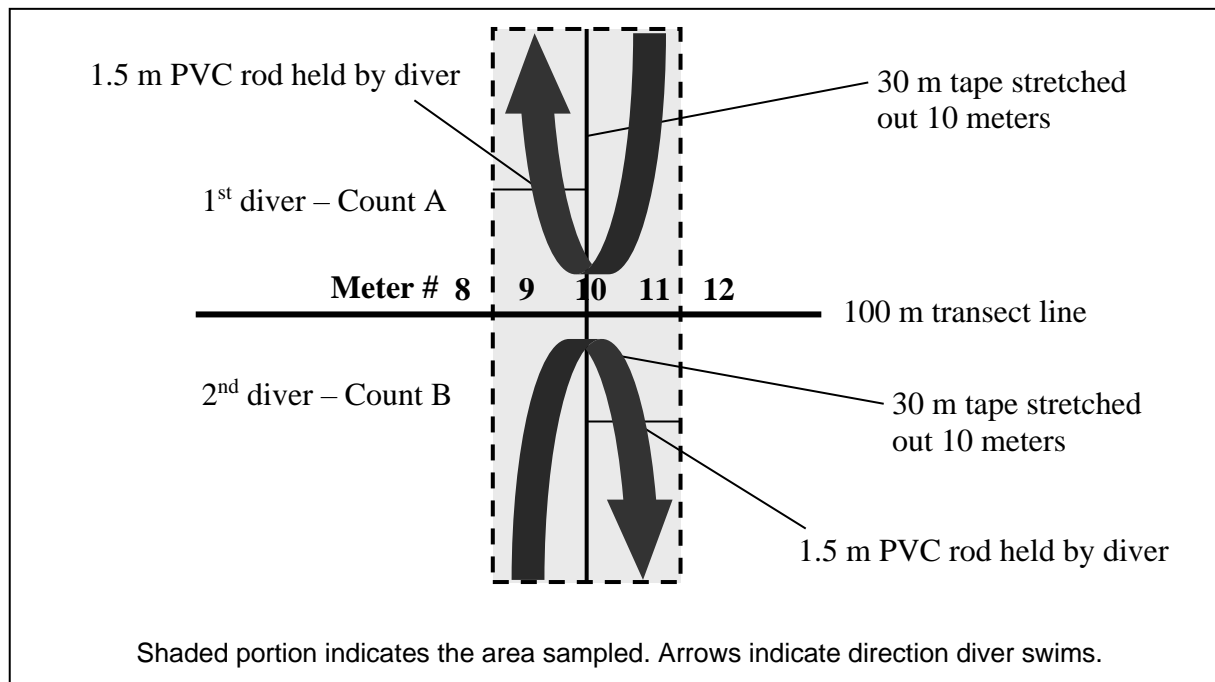
Divers will sample twelve points along the transect line. The points are systematic with a random starting point. At the start of each field season new sampling points must be randomly selected. The initial sampling point (a number between 0-7) is chosen using a randomized selection method. That number will be the meter mark for the first quadrat placement. The subsequent sampling points along the line are at 8.33 m intervals rounded to the nearest whole number (8 m, 17 m, 25m, etc). Each sampling point is recorded consecutively on the top of the data sheets corresponding to quadrat 1,2,3 etc. This systematic set of numbers with a random start will be the same for all sites sampled throughout a field season.

Divers will attach a 15 m tape to the lead line near each sampling point with a carabiner. The divers swim out 10 m in opposite directions as your dive partner perpendicular to the 100 m transect (Figure 5). Next, drop and secure the tape reels to the bottom and swim back towards the transect line along one side of the 10 m tape while using the 1.5 m PVC rod to determine the width of the transect and count the organisms listed on the data sheet that occur within that area. Once the lead line is reached, work back towards the reel along the opposite side of the tape, counting target organisms within 1.5 m of the tape. Once the reel is reached, wind up the tape and move to the next subsequent meter number along the 100 m transect line. Be sure to search the habitat thoroughly, including cracks, and crevices. However, do not conduct any invasive sampling (i.e. do not turn over rocks). At the beginning, midpoint and end of each transect you should make sure you have good contact with your dive partner.

After returning to the surface, check your data sheet and your dive partner's data sheet for readability and outliers. Rinse the data sheets in fresh water, allow them to air dry, and store them in the completed data sheet notebook.

On each transect, each diver covers an area of 3 m x 10 m. Add adjacent segments (Count A and Count B) from both divers to produce a sampled area of 3 m x 20 m at each of the 12 points.

At the bottom of Table 6 is a list of “write-in” species. These species are very rare and are not listed on the datasheet since there is not enough room to include them. If one is observed on band transects it can be written in to the blank row at the bottom of the datasheet. In general, all abalone species should be counted on band transects if observed.



**Figure 5.** Band transect sampling procedure.

### **Time Required**

Typically 100 to 150 minutes is required for sampling. Sampling time is decreased for flat, low relief habitats. Sampling time will increase at high relief, high cover areas. In addition, areas with large densities of *Lytechinus anamesus*, white sea urchin, will significantly slow sampling and may be best sampled using quadrats (in addition to band transects) at the discretion of the chief scientist on each cruise.

**Table 6.** Organisms sampled on band transects.

Species Name	Common Name
<i>Algae</i>	
<i>Sargassum horneri</i>	sargassum (adult = greater than 0.5 m tall or reproductive receptacles present)
<i>Invertebrates</i>	
<i>Tethya aurantia</i>	orange puffball sponge
<i>Stylaster (Allopora) californicus</i>	California hydrocoral
<i>Urticina (Tealia) lofotensis</i>	white-spotted rose anemone
<i>Lophogorgia chilensis</i>	red gorgonian
<i>Muricea fruticosa</i>	brown gorgonian
<i>Muricea californica</i>	California golden gorgonian
<i>Megathura crenulata</i>	giant keyhole limpet
<i>Haliotis rufescens</i>	red abalone
<i>Haliotis corrugata</i>	pink abalone
<i>Haliotis fulgens</i>	green abalone
<i>Kelletia kelletii</i>	Kellet's whelk
<i>Crassadoma gigantea</i>	rock scallop

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<i>Lytechinus anamesus</i>	white sea urchin
<i>Pycnopodia helianthoides</i>	sunflower sea star
<i>Aplysia californica</i>	California brown sea hare
<i>Panulirus interruptus</i>	California spiny lobster
<b>Additional “Write In” species:</b>	
<i>Haliotis sorenseni</i>	white abalone
<i>Haliotis assimilis</i>	threaded abalone
<i>Cryptochiton stelleri</i>	gumboot chiton
<i>Undaria pinnatifida</i>	wakame (juvenile = less than 0.5 m tall, reproductive sporophyll absent)
<i>Undaria pinnatifida</i>	wakame (subadult = greater than 0.5 m tall and not reproductive [can have frills on either side of blade above the holdfast, but lacks mature, dark brown sporophyll])
<i>Undaria pinnatifida</i>	wakame (adult = greater than 0.5 m tall with well developed/reproductive sporophyll)

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