

# Detection of specific biomolecules following immobilization: interaction and report

## 1) Nucleic acid hybridization (base pairing)

- a) Southern blots: DNA-DNA hybridization (Methods for labeling “probe” DNA)
- b) Northern blots: DNA-RNA hybridization

## 2) Antibody-antigen interactions

- a) Western blots (detection of proteins with specific antibodies)

## Guide to readings: Specific Biomolecule Detection

- 1) *9 MC4 Southern blots*. Technique for detecting specific **DNA** fragments by nucleic acid base hybridization
- 2) *10 MC4 Northern blots*. Technique for detecting specific **RNAs** by nucleic acid hybridization
- 3) *11 MC4 Western blots*. Technique for detecting specific **proteins** by antibody recognition
- 4) *12 MC4 Specific detection*. Detailed discussion of the various reagents available for probe detection.

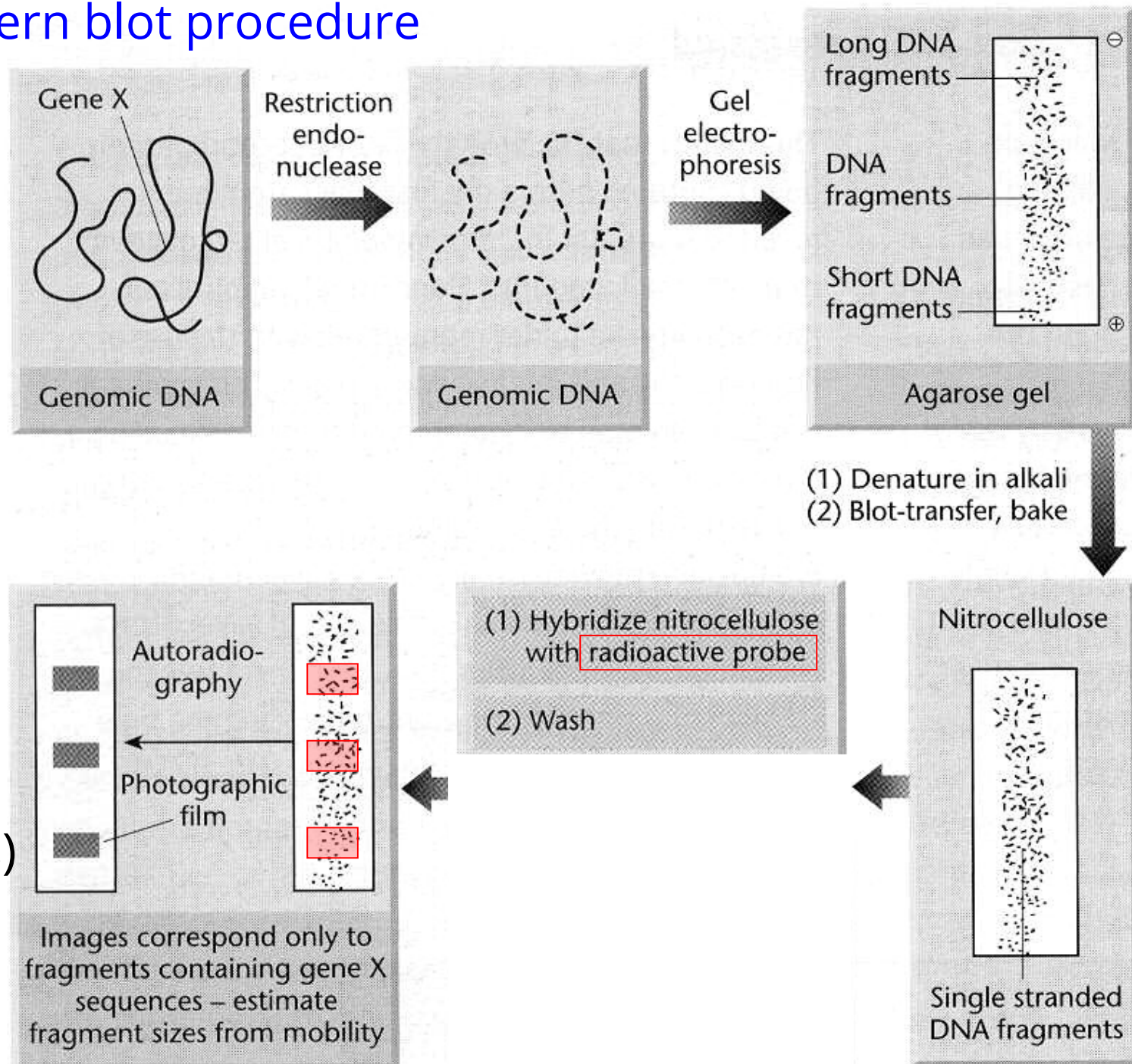
# Visualizing DNA, RNA and Protein: detecting specific sequences or proteins

- Detect **specific** DNA, RNA, or protein in a large, mixed population: cell extracts, genomic DNA preparations, etc.
- For DNA and RNA:
  - specific sequence detection
  - based on DNA and RNA complementarity/base-pairing/hybridization
- For proteins
  - Specific shape/chemistry of the protein
  - Antibodies recognize the protein of interest
  - a specific assay for activity of the protein

## **Southern blot: the original method for detecting presence of a specific DNA sequence**

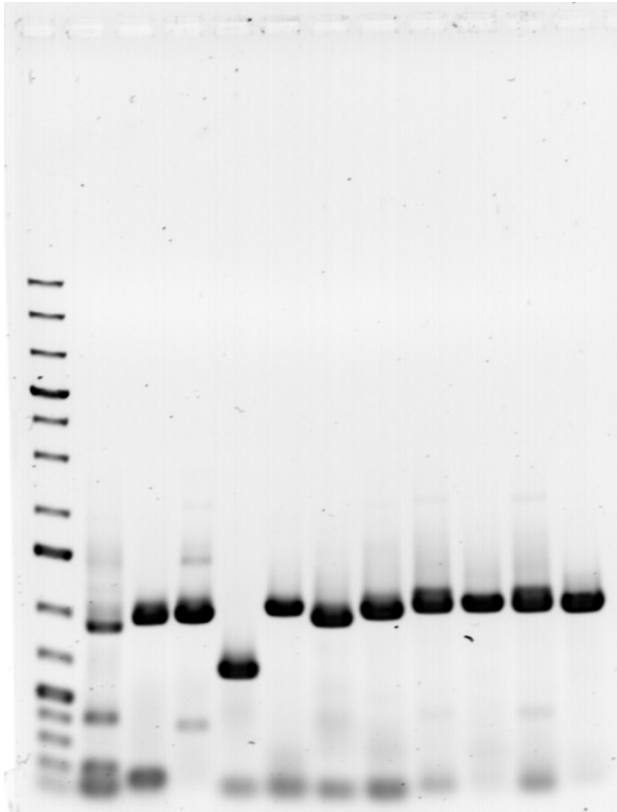
- 1) Prepare genomic DNA
- 2) Digest sufficient amount of DNA to completion with restriction enzyme
- 3) Run gel to separate DNA fragments according to size
- 4) Transfer, fix DNA to a membrane
- 5) Prepare probe DNA
- 6) Wash membrane with probe DNA
- 7) Visualize probe on membrane (appearing as bands where probe binds)

# Southern blot procedure

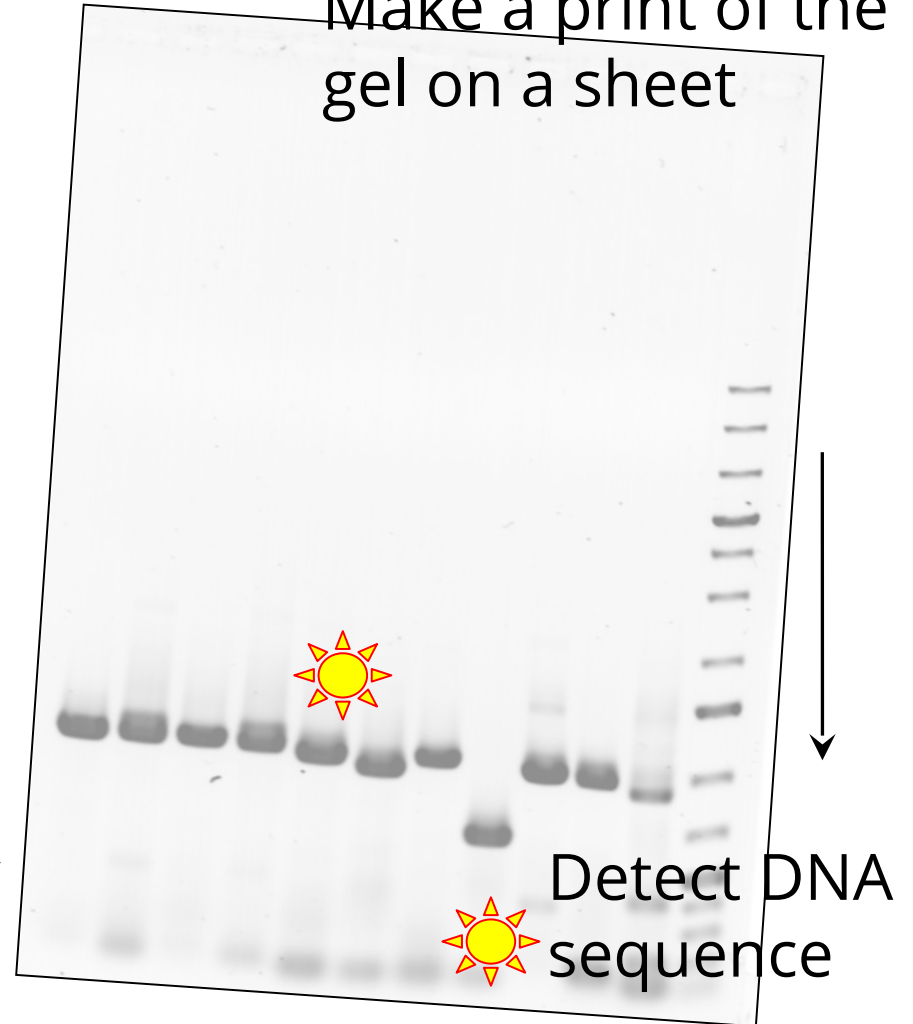


# DNA transfer: making a print from a gel

Run the gel



Make a print of the gel on a sheet



Detect DNA  
sequence

# ***Probe to detect sequence of interest: base-pairing (hybridization)***

## – Probe DNA

- synthetic oligonucleotide  
or
- cloned gene (single stranded)

## – The probe has to be easy to detect

- Radioactivity
- Fluorescence
- Enzyme dependent color change
- Enzyme dependent luminescence

# Hybridize (base pair) probes to target DNA

- blocking agents (e.g. milk, SDS) **prevent non-specific interactions between probes and membrane**
- Volume exclusion agents (eg. dextran sulfate) **increase rate and level of hybridization**
- Wash blot with increasing stringency...
  - Low stringency: high salt, low temperature, probe base pairs with sequences with mismatches
  - High stringency: low salt, higher temp., probe will base pair only to fully complementary sequences



# How to make a nucleic acid probe: order online

(Example: IDTdna.com)

Paste DNA sequence of the oligonucleotide in online order form

Define a modification:

- Fluorescence
- Attachment chemistry, e.g. biotin, digoxigenin
- Modified bases
- Randomized bases

Place order (company synthesizes oligo by automated phosphoramidite chemistry, sends oligo to you) (

<https://www.sigmaaldrich.com/technical-documents/articles/biology/dna-oligonucleotide-synthesis.html> )

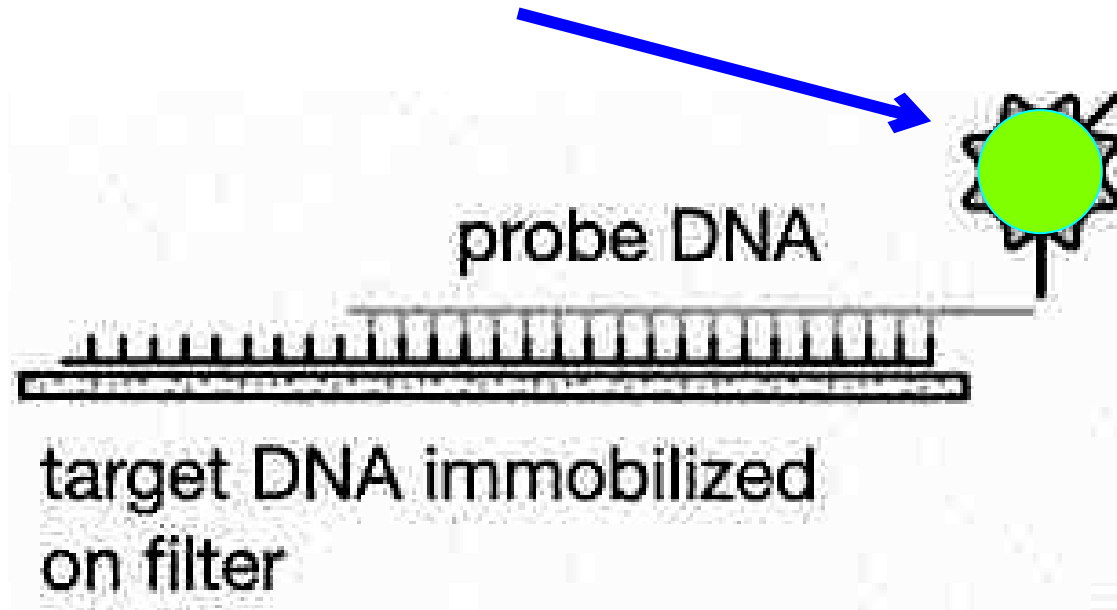
Do experiment

# Radioactive probes

- Example:  $^{32}\text{P}$  label
  - Add  $^{32}\text{P}$  ATP to the 5' end of probe DNA by kinase reaction
  - Probe DNA base pairs with target DNA
  - $^{32}\text{P}$  radioactive decay produces detectable signal
- Detect radiolabel with
  - autoradiography: X ray film
  - phosphorimager: phosphor coated plates store the energy of the radioactive decay

# Fluorescence for detection

Fluorophores: Cy3, Cy5, etc.



Induction and detection of fluorescence (example: Cy3):  
excitation wavelength: 547 nm  
emission wavelength: 563 nm

(<http://www.bdbiosciences.com/us/s/spectrumviewer> )

# How to amplify the DNA probe signal: add enzyme

Peroxidase, alkaline phosphatase enzyme activity leads to easily detected color change or emitted light

1.1) Covalently attach the **enzyme** to the DNA

Or

1.2) Attach a tag to the DNA:

- Digoxigenin (DIG)
- Biotin

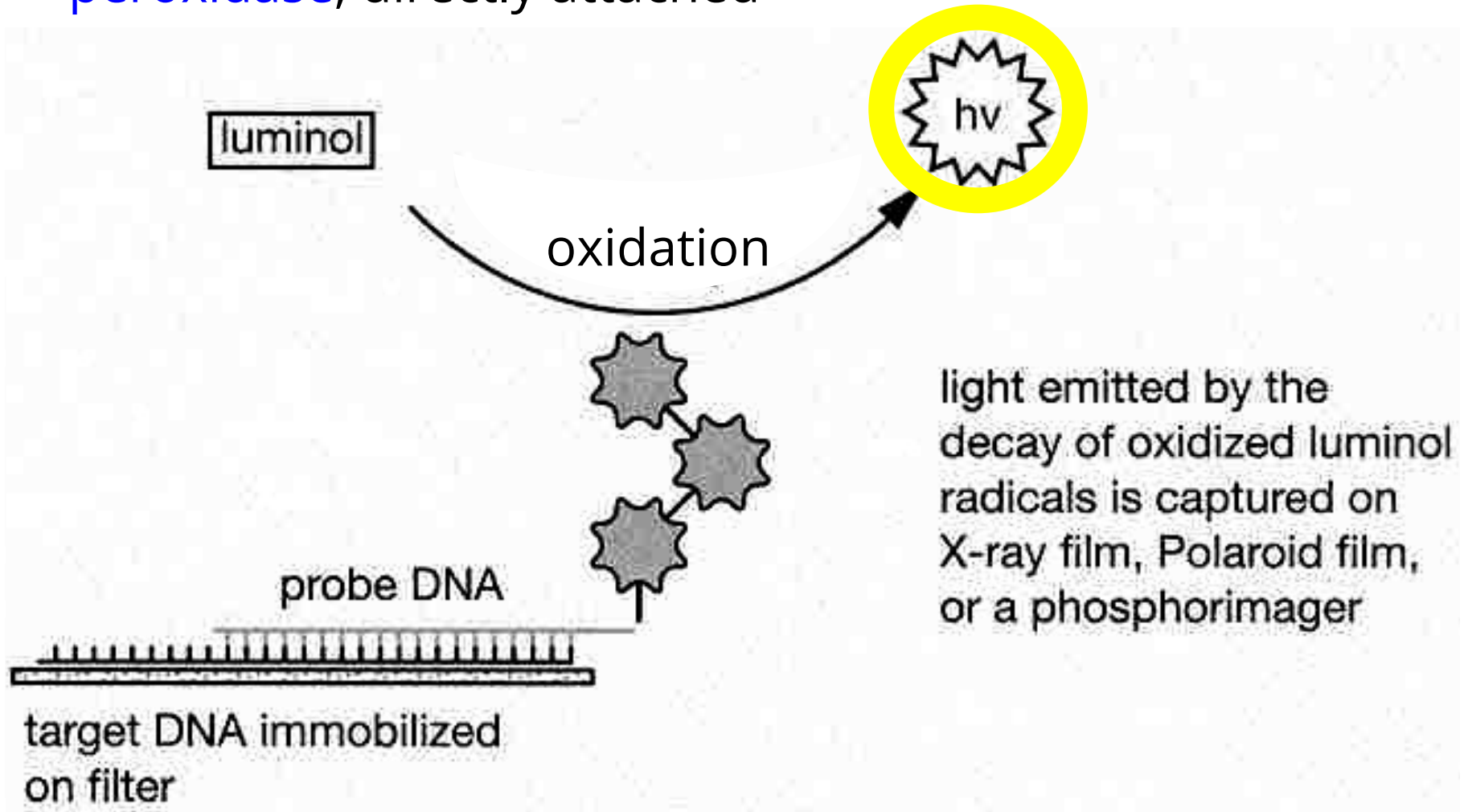
Then bring **enzyme** to the DNA tag:

- Conjugate it to antibody that recognizes DIG
- Conjugate it to streptavidin that binds to biotin

2) Detect **enzyme** through its activity

# Enzyme-linked probes: covalent linkage

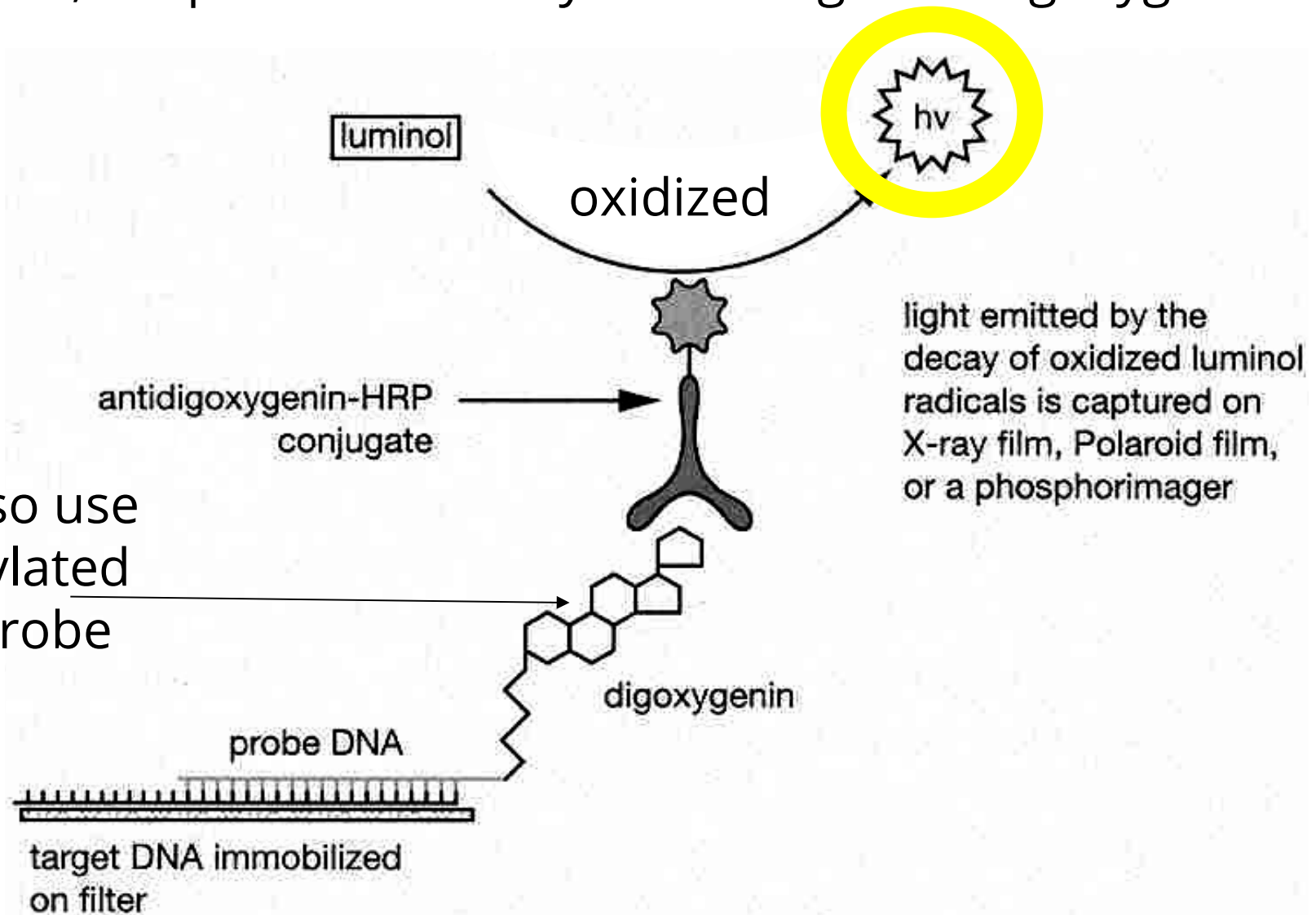
peroxidase, directly attached



# Enzyme-linked probes: epitope recognition

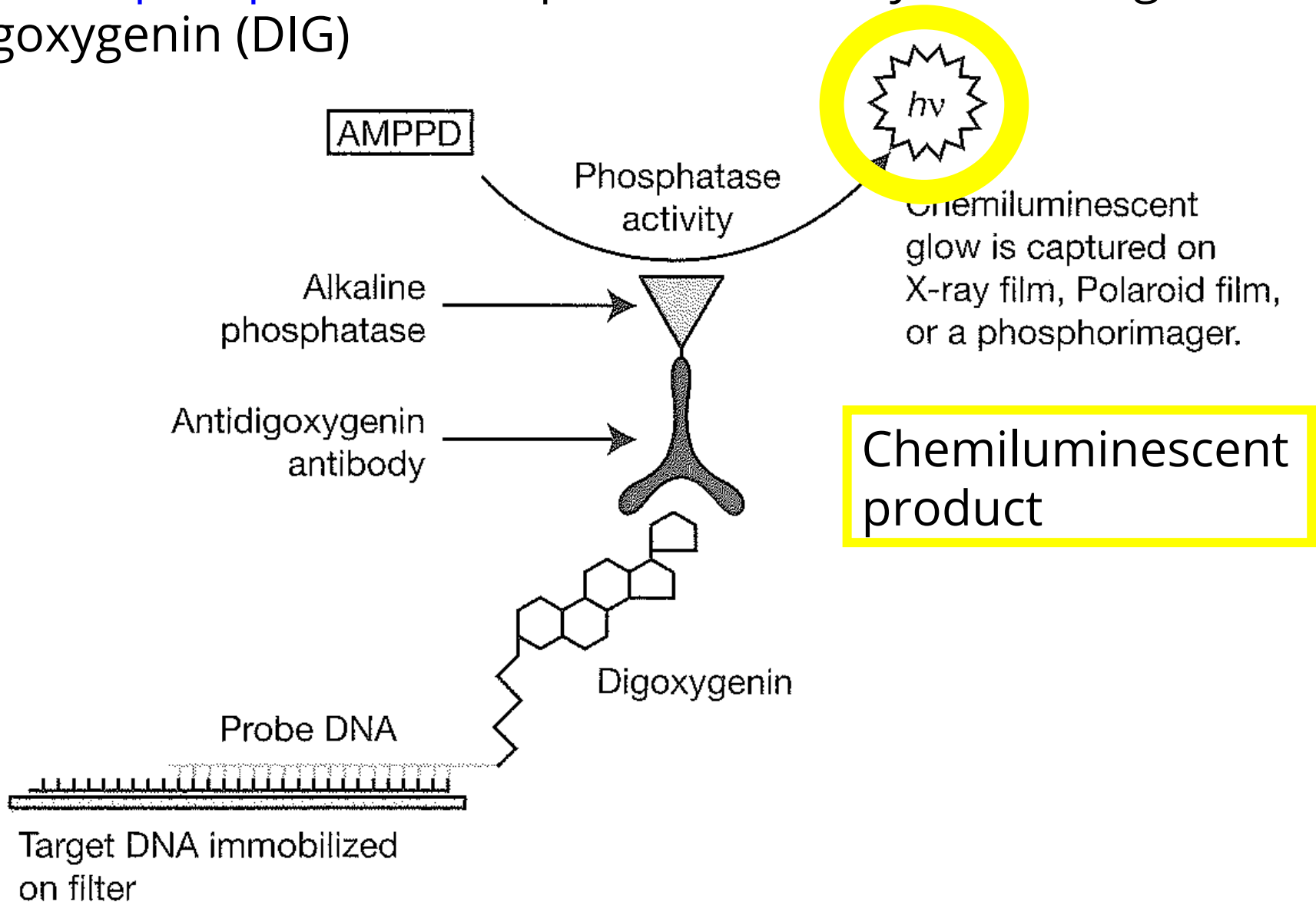
peroxidase, coupled to antibody that recognizes digoxigenin (DIG)

can also use  
biotinylated  
DNA probe



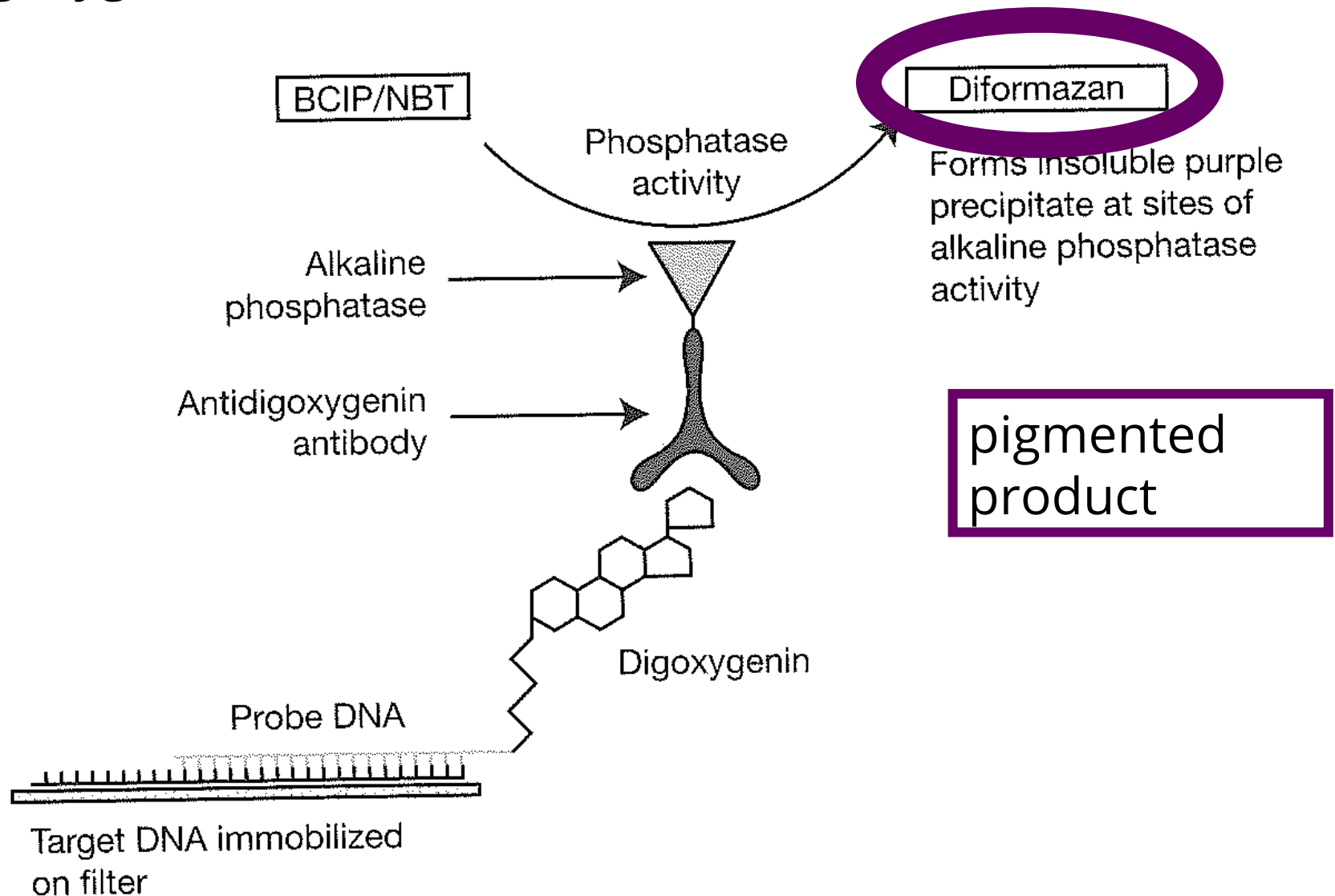
# Enzyme-linked probes: epitope recognition

alkaline phosphatase, coupled to antibody that recognizes digoxigenin (DIG)



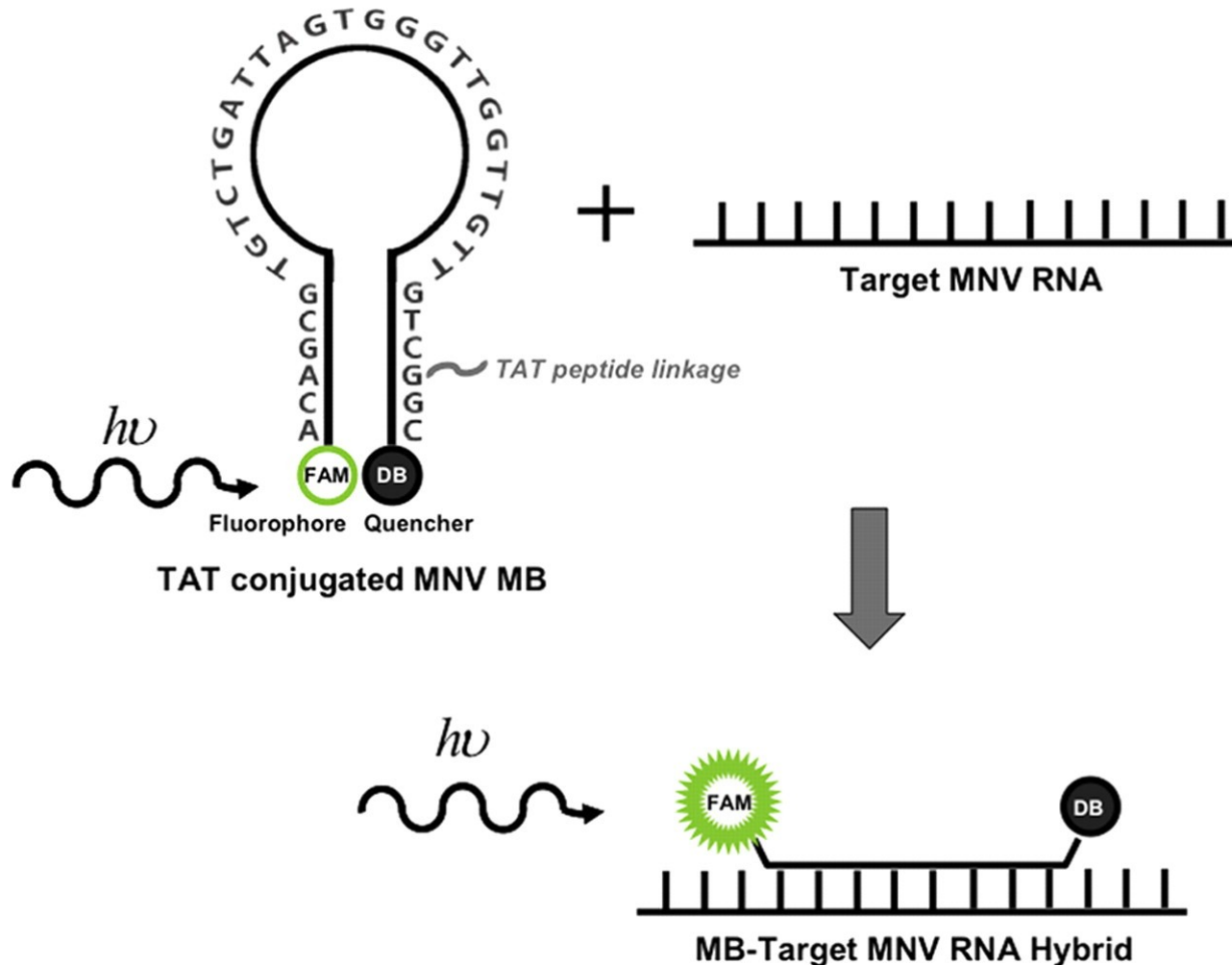
# Enzyme-linked probes: epitope recognition

alkaline phosphatase, coupled to antibody that recognizes digoxigenin (DIG)





## Detection of specific nucleic acids: in solution



The 'quencher' suppresses fluorescence in the hairpin structure

Target  
recognition  
disrupts hairpin,  
allows  
fluorescence

Probe DNA fluorescent only after target is detected

Detection of murine norovirus-1 by using TAT peptide-delivered molecular beacons. Ganguli PS, Chen W, Yates MV. *Appl Environ Microbiol.* 2011 Aug;77(15):5517-20. doi: 10.1128/AEM.03048-10.

# Northern blots: RNA

Same basic technique as Southern blots, but **RNA** is run on the initial gel and is transferred to the membrane.

This method was used to measure levels of gene transcription *in vivo* (detecting changes in the levels of RNA transcript under differing conditions)

**Microarrays** for measuring mRNA abundance are based on this principle, but many probes are immobilized in a regular array -- reverse transcribed (and fluorescently labelled) RNA “lights up” the probes on the microarray

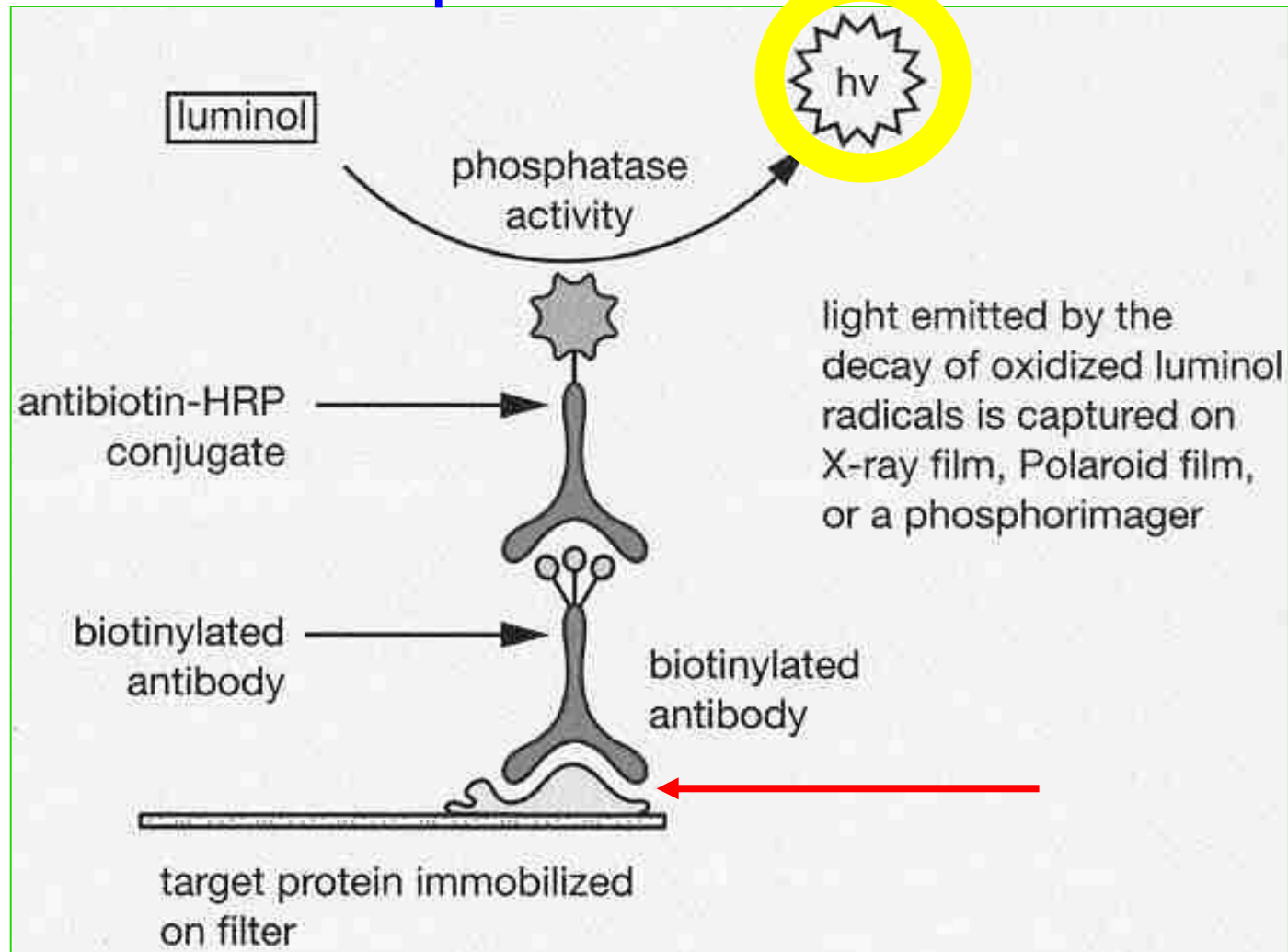
Current alternative for single genes is RT-PCR (reverse transcriptase to convert RNA to DNA, then PCR

# Protein detection in samples

Is a specific protein being made in a cell? How much is there?  
When is the protein present?

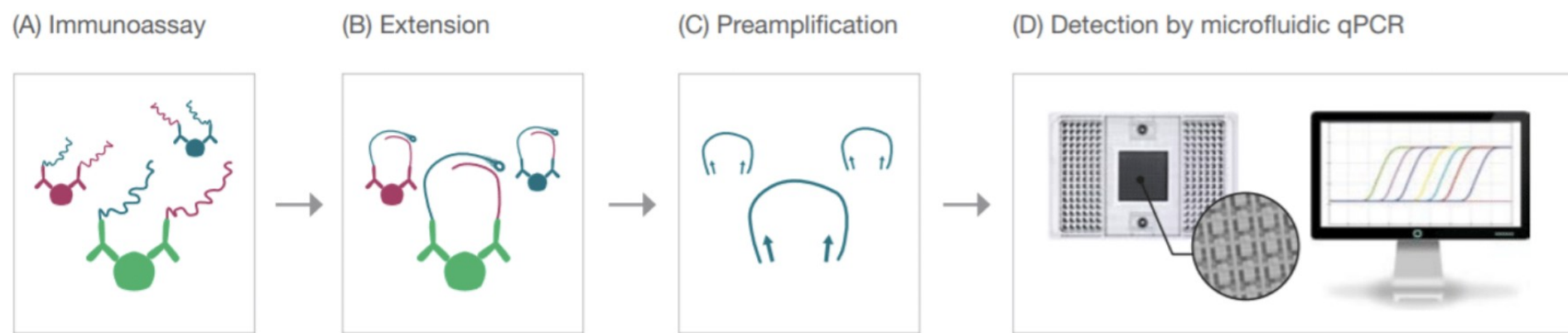
- Many proteins are made by cells. You need a way to detect a specific protein. Hybridization won't work!
  - Purify the protein
  - Raise antibodies to the protein (rabbits, goats, chickens, llamas)
  - Isolate the antibodies from animal blood
  - Test the antibodies for specificity
- Proteins separated by SDS PAGE transferred to membranes using the same principle as Southern blots
- Specific proteins detected by probing blot with antibodies to protein of interest

# Western blots: proteins



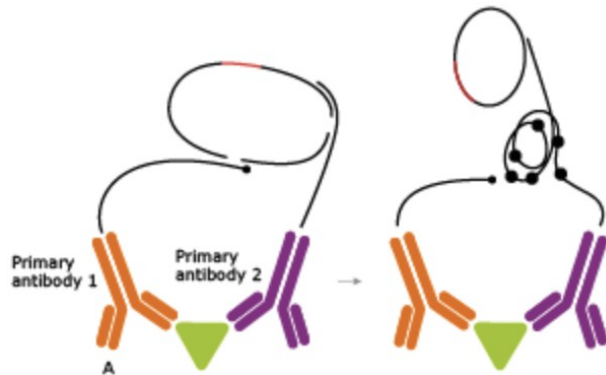
First antibody binding is detected by 'secondary' antibody that has enzyme (horseradish peroxidase, alkaline phosphatase) or radioactivity ( $^{125}\text{I}$ ) conjugated to it

Using antibodies to detect proteins in cells is not always straightforward – there can be ‘cross-reactivity’  
One solution is the “Proximity Extension Assay” to increase specificity



a) PCR detects primer extension product

b) Primer extension creates DNA hybridization sites



<http://www.proteinatlas.org/learn/method/proximity+ligation+assay#proximityligationassay>

<https://www.olk.com/data-you-can-trust/technology/>

# Methods for detecting specific biomolecules

- 1) (If necessary, separate DNA, RNA, or proteins on the basis of size, by gel electrophoresis)
- 2) Immobilize (blot) the DNA, RNA, or protein
- 3) “Probe” the blot with something that will specifically interact with a target
  - a) DNA and RNA: interacts with a complementary nucleic acid
  - b) Protein: interacts with an antibody that specifically recognizes the protein

Types of blots:    Southern, DNA (named for [E.M. Southern](#))  
                         Northern, RNA  
                         Western, protein