# **Conditional Diffusion Models for Uncertainty Estimation in Super Resolution Microscopy**

# **Anonymous Author(s)**

Affiliation Address email

# **Abstract**

Single-molecule localization microscopy (SMLM) techniques are a mainstay of fluorescence microscopy and can be used to produce a pointillist representation of living cells at diffraction-unlimited precision. Classical SMLM approaches leverage the deactivation of fluorescent tags, followed by spontaneous or photoinduced reactivation, which can be used to estimate of the density of a tagged biomolecule in cellular compartments. Standard SMLM localization algorithms based on maximum likelihood estimators or least squares optimization require tight control of activation and reactivation to maintain sparse emitters, presenting a tradeoff between imaging speed and labeling density. Recently, deep models have generalized SMLM to densely labeled structures by predicting high-resolution kernel density estimates (KDEs) from low resolution images with convolutional networks. However, estimated KDEs may contain irregularities due to finite sample sizes and limited model capacity. Denoising diffusion probabilistic models (DDPMs) are well suited conditional super resolution tasks, demonstrating promising results in detail reconstruction, while directly providing uncertainties in model predictions. Here, we adapt DDPM to the task of single molecule localization, and demonstrate that combining traditional CNNs with a DDPM permits uncertainty quantification of KDEs and improves localization precision over a wide range of experimental conditions.

### 20 1 Introduction

2

3

5

6

7

8

10

11

12

13

14

15

16

17

18

19

Single molecule localization microscopy (SMLM) relies on the temporal resolution of fluorophores 21 whose spatially overlapping point spread functions would otherwise render them unresolvable at the detector. Common strategies for the temporal separation of molecules involve molecular 23 photoswitching from dark to fluorescent states, permitting resolution of flurophores beyond the 24 diffraction limit. Estimation of molecular coordinates is typically carried out by modeling the optical 25 impulse response of the imaging system and fitting model functions to the data. However, such models are only well-suited to isolated molecules, reducing the number of molecules in the field of 27 view and limiting temporal resolution in super resolution microscopy. This issue has incited a series 28 of efforts to increase the density of fluorescent molecules imaged in a single frame while developing 29 appropriate models for dense localization. 30

In previous approaches to this issue, predicted super-resolution images can be recovered from a sparse set of localizations with conditional generative adversarial networks (Ouyang 2018) or direct prediction of molecular coordinates using neural networks (Nehme 2020; Speiser 2021). Here, we focus on the latter class of models which perform single molecule localization using neural networks. In this approach, one estimates molecular coordinates by predicting kernel density estimates (KDEs) y, which are latent in the raw data x, using a convolutional neural network. Importantly, inferences

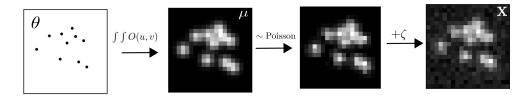


Figure 1: Generative model of single molecule localization microscopy images

in SMLM are often necessarily made on a single measurement, thus common measures of model performance are based on localization errors computed over ensembles of simulated images. However, this choice precludes computation of aleatoric uncertainty at test time under a fixed model, and may result in the application of models to out of distribution datasets. Therefore, we propose a deep generative approach which is complimentary to existing methods, to model the posterior distribution on reconstructions. Our approach could be readily integrated with existing localization performance measures to address both model accuracy on training data and precision on datasets produced by experiments.

# 45 2 Background

### 46 2.1 Image Formation and Origins of Uncertainty

The central objective of single molecule localization microscopy is to infer a set of molecular coordinates from measured low resolution images  $\mathbf{x}$ . We begin by defining the likelihood on measured low-resolution images  $p(\mathbf{x}|\theta)$ . In fluorescence microscopy, each pixel is treated as a Poisson random variable (Smith 2010; Nehme 2020; Chao 2016), with expected value

$$\omega = i_0 \int O(u) du \int O(v) dv \tag{1}$$

where  $i_0 = \eta N_0 \Delta$ . The scalar parameters  $\eta, \Delta$  are the photon detection probability of the sensor and the exposure time, respectively. Without loss of generality, we assume  $\eta = \Delta = 1$ . Most importantly,  $N_0$  represents the signal amplitude, which we assume maintains a fixed value. The optical impulse response O(u,v) is often approximated as a 2D isotropic Gaussian with standard deviation  $\sigma$  (Zhang 2007). This approximation has the convenient property, that the effects of pixelation can be expressed in terms of error functions. For example, given a fluorescent emitter located at  $\theta = (u_0, v_0)$ , we have that

$$\int O(u)du = \frac{1}{2} \left( \operatorname{erf} \left( \frac{u_k + \frac{1}{2} - u_0}{\sqrt{2}\sigma} \right) - \operatorname{erf} \left( \frac{u_k - \frac{1}{2} - u_0}{\sqrt{2}\sigma} \right) \right) \tag{2}$$

where we have used the common definition  $\operatorname{erf}(z) = \frac{2}{\sqrt{\pi}} \int_0^t e^{-t^2} dt$ . Our generative model also incorporates a normally distributed white noise per pixel  $\zeta$  with offset o and variance  $\sigma^2$ . Ultimately, we have a Poisson component of the signal, which scales with  $N_0$  and a Gaussian component, which does not. Therefore, in a single exposure, we measure:

$$\mathbf{x} = \mathbf{s} + \zeta \tag{3}$$

The distribution of x is the convolution of the distributions of s and  $\zeta$ ,

$$p(\mathbf{x}_k|\theta) = A \sum_{q=0}^{\infty} \frac{1}{q!} e^{-\omega_k} \omega_k^q \frac{1}{\sqrt{2\pi}\sigma_k} e^{-\frac{(\mathbf{x}_k - g_k q - \sigma_k)}{2\sigma_k^2}}$$
(4)

where  $p(\zeta_k) = \mathcal{N}(o_k, \sigma_k^2)$  and  $p(s_k) = \operatorname{Poisson}(\omega_k)$ , A is some normalization constant. In practice, (4) is difficult to work with, so we look for an approximation. We will use a Poisson-Normal approximation for simplification. Consider,

$$\zeta_k - o_k + \sigma_k^2 \sim \mathcal{N}(\sigma_k^2, \sigma_k^2) \approx \text{Poisson}(\sigma_k^2)$$
 (5)

Since  $\mathbf{x}_k = \mathbf{s}_k + \zeta_k$ , we transform  $\mathbf{x}_k' = \mathbf{x}_k - o_k + \sigma_k^2$ , which is distributed according to

$$\mathbf{x}_k' \sim \text{Poisson}(\omega_k')$$
 (6)

where  $\omega_k' = \omega_k + \sigma_k^2$ . This result can be seen from the fact the convolution of two Poisson distributions is also Poisson. We then arrive at the following log likelihood

$$\ell(\mathbf{x}|\theta) = -\log \prod_{k} \frac{e^{-\left(\mu_{k}'\right)} \left(\mu_{k}'\right)^{n_{k}}}{n_{k}!} \approx \sum_{k} n_{k} \log n_{k} + \mu_{k}' - n_{k} \log \left(\mu_{k}'\right) \tag{7}$$

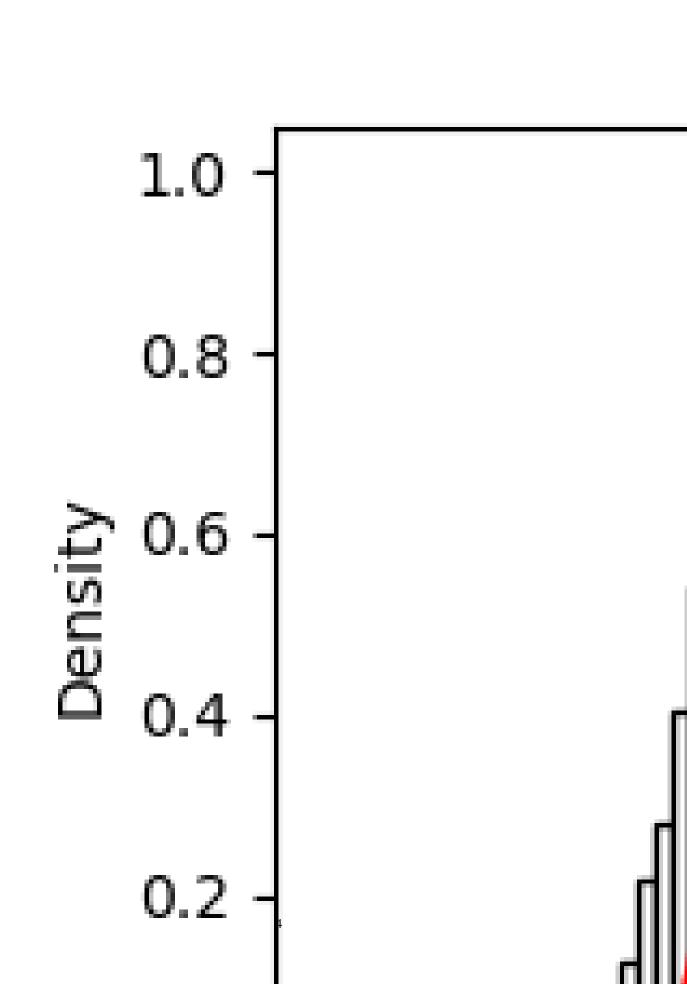
### 9 2.2 Localization Error

We use the Fisher information as an information theoretic criteria to assess the quality of the proposed algorithms, with respect to the root mean squared error (RMSE) of our predictions of  $\theta$ . The generative model  $\ell(\mathbf{x}|\theta)$  is also convenient for computing the Fisher information matrix (Smith 2010) and thus the Cramer-Rao lower bound, which bounds the variance of a statistical estimator of  $\theta$ , from below i.e.,  $\operatorname{var}(\hat{\theta}) \geq I^{-1}(\theta)$ . It is shown in the appendix, that the Fisher information is straightforward to compute under the Poisson likelihood (7)

$$\mathcal{I}_{ij}(\theta) = \mathbb{E}\left(\frac{\partial \ell}{\partial \theta_i} \frac{\partial \ell}{\partial \theta_j}\right) = \sum_k \frac{1}{\omega_k'} \frac{\partial \omega_k'}{\partial \theta_i} \frac{\partial \omega_k'}{\partial \theta_j}$$
(8)

# 6 2.3 Kernel density estimation with deep networks

Direct optimization of the log-likelihood in (7) from observations x alone is challenging when 77 fluorescent emitters are dense within the field of view and fluorescent signals significantly overlap. 78 Convolutional neural networks (CNN) have recently been used in fluorescence microscopy to extract 79 parameters describing fluorescent emitters such as color, emitter orientation, z coordinate, background 80 signal. For localization tasks, CNNs typically employ upsampling layers to reconstruct Bernoulli 81 probabilities of emitter occupancy or kernel density estimates with higher resolution than experimental 82 measurements. Kernel density estimates are the most common data structure used in SMLM, and 83 84 can be easily generated from molecular coordinates using the optical impulse response (2). While ground-truth data from experiments to train the neural network are typically not available, synthetic 85 86 training data can also be generated by (2) and simulation from the Poisson likelihood (7). The DeepSTORM CNN, initially proposed in [1] for 3D localization, can be viewed as a deep kernel 87 density estimator, reconstructing kernel density estimates y from low-resolution inputs x. We utilize 88 a simplified form of the original architecture for 2D localization, which we denote  $\phi$  hereafter, which 89 consists of three main modules: a multi-scale context aggregation module, an upsampling module, and 90 a prediction module. For context aggregation, the architecture utilizes dilated convolutions to increase 91 the receptive field of each layer. The upsampling module is then composed of two consecutive 2x 92 resize-convolutions, computed by nearest-neighbor interpolation, to increase the lateral resolution 93 by a factor of 4. For a common sCMOS camera, each pixel has a lateral size of approximately 108 94 nanometers, giving approximately 27 nanometer pixels in the KDE. The terminal prediction module 95 contains three additional convolutional blocks for refinement of the upsampled image, followed by an element-wise HardTanh.



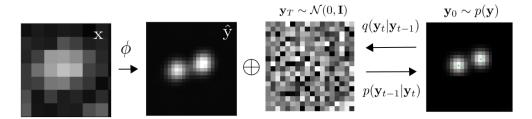


Figure 3: Conditional diffusion model for sampling kernel density estimates

# 98 3 Denoising Diffusion Probabilistic Model for SMLM

We consider datasets  $(\mathbf{x}_i, \mathbf{y}_i, \hat{\mathbf{y}}_i)_{i=1}^N$  of observed images  $\mathbf{x}_i$  true kernel density estimate (KDE) images  $\mathbf{y}_i$ , and KDE estimates  $\hat{\mathbf{y}}_i = \phi(\mathbf{x}_i)$ . Observations  $\mathbf{x}_i$  are generated under the image degradation model. We aim to develop a framework for sampling from  $p(\hat{\mathbf{y}}|\mathbf{x}, \mathbf{y})$ .

# 102 4 Conditional Denoising Diffusion Model

Point estimates  $\hat{\mathbf{y}}_i$  produced by the DeepSTORM architecture lack uncertainty quantification. To address this, we propose a DDPM to model the conditional distribution  $p(\hat{\mathbf{y}}|\mathbf{x},\mathbf{y})$ . Consider the factorization  $p(\hat{\mathbf{y}}|\mathbf{x},\mathbf{y})p(\mathbf{x}|\mathbf{y})p(\mathbf{y}) = p(\mathbf{x}|\mathbf{y},\hat{\mathbf{y}})p(\mathbf{y}|\hat{\mathbf{y}})p(\hat{\mathbf{y}})$ . Given that  $\mathbf{x}$  is conditionally independent of  $\hat{\mathbf{y}}$ , we find

$$p_{\Psi}(\hat{\mathbf{y}}|\mathbf{x},\mathbf{y}) = p(\mathbf{y}|\hat{\mathbf{y}})$$

Evidently, the DDPM  $\Psi$  can be trained on pairs  $(\mathbf{y}_i, \hat{\mathbf{y}}_i)_{i=1}^N$ . The conditional DDPM generates a target KDE  $\mathbf{y}_0$  in T refinement steps. Starting with a pure noise image  $\mathbf{y}_T \sim \mathcal{N}(0, \mathbf{I})$ , the model iteratively refines the KDE through successive iterations according to learned conditional transition distributions  $p(\mathbf{y}_{t-1}|\mathbf{y}_t)$  such that  $\mathbf{y}_0 \sim p(\mathbf{y}|\hat{\mathbf{y}})$ 

# 4.1 Gaussian Diffusion

111

Diffusion models (Sohl-Dickstein 2015; Ho 2020) are a class of generative models inspired by nonequilibrium statistical physics, which slowly destroy structure in a data distribution  $p(\mathbf{y}_0|\mathbf{x})$  via a fixed Markov chain referred to as the *forward process*. In the present context, the forward process gradually adds Gaussian noise to the KDE  $\mathbf{y}$  according to a variance schedule  $\beta_{0:T}$ 

$$q(\mathbf{y}_t|\mathbf{y}_0) = \prod_{t=1}^{T} q(\mathbf{y}_t|\mathbf{y}_{t-1}) \quad q(\mathbf{y}_t|\mathbf{y}_{t-1}) = \mathcal{N}\left(\sqrt{1-\beta_t}\mathbf{y}_{t-1}, \beta_t I\right)$$
(9)

An important property of the forward process is that it admits sampling  $y_t$  at an arbitrary timestep t in closed form (Ho 2020). Using the notation  $\alpha_t := 1 - \beta_t$  and  $\gamma_t := \prod_{s=1}^t \alpha_s$ , we have

$$q(\mathbf{y}_t|\mathbf{y}_0) = \mathcal{N}\left(\sqrt{\gamma_t}\mathbf{y}_0, (1-\gamma_t)I\right) \tag{10}$$

The usual procedure is then to learn a parametric representation of the *reverse process*, and therefore generate samples from  $p(\mathbf{y}_0)$ , starting from noise. Formally,  $p_{\theta}(\mathbf{y}_0|\hat{\mathbf{y}}) = \int p_{\theta}(\mathbf{y}_{0:T}|\hat{\mathbf{y}})d\hat{\mathbf{y}}_{1:T}$  where  $\mathbf{y}_t$  is a latent representation with the same dimensionality of the data.  $p_{\theta}(\mathbf{y}_{0:T}|\hat{\mathbf{y}})$  is a Markov process, starting from a noise sample  $p_{\theta}(\mathbf{y}_T) = \mathcal{N}(0, \mathbf{I})$ .

$$p_{\theta}(\mathbf{y}_{0:T}) = p_{\theta}(\mathbf{y}_T) \prod_{t=1}^{T} p_{\theta}(\mathbf{y}_{t-1}|\mathbf{y}_t) \quad p_{\theta}(\mathbf{y}_{t-1}|\mathbf{y}_t) = \mathcal{N}\left(\mu_{\theta}(\mathbf{y}_t), \beta_t I\right)$$
(11)

where we reuse the variance schedule of the forward process (Ho 2020). We seek to learn a denoising model  $\mu_{\theta}$  which computes the mean of the Gaussian transition density at each time step t. For all 123 t > 0, the mean of the transition density is computed as 124

$$\mu_{\theta}(\mathbf{y}_{t}, \hat{\mathbf{y}}, \gamma_{t}) = \frac{1}{\sqrt{\alpha_{t}}} \left( \mathbf{y}_{t} - \frac{(1 - \alpha_{t})}{\sqrt{1 - \gamma_{t}}} f_{\theta}(\mathbf{y}, \hat{\mathbf{y}}, \gamma_{t}) \right)$$
(12)

where  $f_{\theta}$  is a neural network. Only at t=0 is this mean directly a function of  $\mathbf{x}$ . 125

#### 4.2 Optimization of the Denoising Model 126

To reverse the diffusion process, we optimize a neural denoising model  $f_{\theta}$  that takes as input  $\hat{y}$  and a 127 noisy target image  $y_t \sim q(y_t|y_0)$ . That is, this noisy target image  $y_t$  is drawn from the marginal 128 distribution of noisy images at a time step t of the forward diffusion process. 129

$$\mathbf{y}_t = \sqrt{\gamma} \mathbf{y}_0 + \sqrt{1 - \gamma} \epsilon, \quad \epsilon \sim \mathcal{N}(0, \mathbf{I})$$
 (13)

In addition to a source image  $y_0$  and a noisy target image  $y_t$ , the denoising model  $f_\theta$  takes as input the sufficient statistics for the variance of the noise  $\gamma$ , and is trained to predict the noise vector  $\epsilon$ . 131 We make the denoising model aware of the level of noise through conditioning on a scalar  $\gamma$ . The proposed objective function for training  $f_{\theta}$  is 133

$$\mathbb{E}_{(\hat{\mathbf{y}},\mathbf{y}_0)(\epsilon,\gamma)} \left[ f_{\theta} \left( x, \sqrt{\gamma} \mathbf{y}_0 + \sqrt{1 - \gamma} \epsilon \, \middle| \, \mathbf{y}_t, \gamma \right) - \epsilon \right], \tag{14}$$

where  $(\hat{\mathbf{y}}, \mathbf{y}_0)$  is sampled from the training dataset and  $\gamma \sim p(\gamma)$ . The distribution of  $\gamma$  has a big 134 impact on the quality of the model and the generated outputs. For our training noise schedule, we 135 use a piecewise distribution for  $\gamma$ ,  $p(\gamma) = \frac{1}{T} \sum_{t=1}^{T} U(\gamma_{t-1}, \gamma_t)$  (Nanxin 2021). Specifically, during training, we first uniformly sample a time step  $t \sim \{0, ..., T\}$  followed by sampling  $\gamma \sim U(\gamma_{t-1}, \gamma_t)$ . 136 137 We set T = 100 in all our experiments. 138

# 4.3 Optimization of the DeepSTORM architecture

A first pass at localization treats localization as a binary classification problem, such that 0 denotes 140 a vacant pixel and 1 denotes an occupied pixel containing an emitter. Direct learning of pixel-wise 141 classification with cross-entropy loss leads to an imbalance of occupied and unoccupied pixels in 142 dense localization problems (Nehme 2020). CE loss is usually either weighted [51], replaced with a 143 Focal loss [52], or applied to a "blobbed" version of the desired boolean volume e.g. by placing a disk 144 around each GT position [53-55]. Alternative methods take a soft version of the binary classification 145 problem. That is, by placing a small Gaussian around each GT position (e.g. with std of 1 pixel), 146 and matching continuous heatmaps, backpropagation yields more meaningful gradients and eases the 147 learning process convergence. 148 Localization heatmaps thus form a natural encoding for SMLM images, which can be input to our 149 conditional diffusion model. Therefore, to encode raw data x into a more tractable representation, we 150 train the DeepSTORM architecture (Nehme 2020). Raw coordinates  $\theta$  are binned into an upsampled

$$\mathcal{L}(\mathbf{y}, \hat{\mathbf{y}}) = ||\mathbf{y} - \hat{\mathbf{y}}||^2$$

#### **Experiments** 153

image z.

139

151

152

Training data was simulated under the image degradation model, drawing coordinates uniformly over 154 a disc. We set T=100 for all experiments and treat forward process variances  $\beta_t$  as hyperparameters, with a linear schedule from  $\beta_0=10^{-4}$  to  $\beta_T=10^{-2}$ . These constants were chosen to be small relative to data scaled to [-1,1], ensuring that reverse and forward processes have approximately the same functional form while keeping the signal-to-noise ratio at  $x_T$  as small as possible ( $L_T$  =  $D_{KL}(q(x_T|x_0)||\mathcal{N}(0,I)) \approx 10^{-5}$  bits per dimension in our experiments).

- To represent the reverse process, we used the DDPM architecture based on a U-Net backbone (Ho
- 161 2020). Parameters are shared across time, which is specified to the network using the Transformer
- sinusoidal position embedding ?. We use self-attention at the  $16 \times 16$  feature map resolution ??.
- Details are in Appendix A.
- and the channel multipliers at different resolutions (see Appendix A for details). To condition the
- model on the input x, we up-sample the low-resolution image to the target resolution using bicubic
- interpolation. The result is concatenated with  $y_t$  along the channel dimension. We experimented with
- more sophisticated methods of conditioning, such as using, but we found that the simple concatenation
- yielded similar generation quality.

### 169 6 Related Work

# 170 6.1 Diffusion Models

- 171 Prior work of diffusion models ?? require 1-2k diffusion steps during inference, making generation
- slow for large target resolution tasks. We adapt techniques from ? to enable more efficient inference.
- Our model conditions on  $\gamma$  directly (vs t as in ?), which allows us flexibility in choosing the number
- of diffusion steps, and the noise schedule during inference. This has been demonstrated to work
- well for speech synthesis?, but has not been explored for images. For efficient inference, we set the
- maximum inference budget to 100 diffusion steps, and hyper-parameter search over the inference
- noise schedule. This search is inexpensive as we only need to train the model once?. We use FID on
- 178 held-out data to choose the best noise schedule, as we found PSNR did not correlate well with image
- 179 quality.

### 180 7 Conclusion

### 181 References

- 182 [1] Nehme, E., et al. DeepSTORM3D: dense 3D localization microscopy and PSF design by deep learning.
- 183 Nature Methods 17, 734–740 (2020).
- 184 [2] Ouyang, W., et al. Deep learning massively accelerates super-resolution localization microscopy. Nature
- 185 Biotechnology 36, 460–468 (2018).
- 186 [3] Speiser, A., et al. Deep learning enables fast and dense single-molecule localization with high accuracy.
- 187 Nature Methods 18, 1082–1090 (2021).
- 188 [4] Sohl-Dickstein J., et al. Deep unsupervised learning using nonequilibrium thermodynamics. ICLR (2015).
- [5] Ho J., et al. Denoising Diffusion Probabilistic Models. Advances in Neural Information Processing Systems
- 190 (2015)
- 191 [6] Nanxin C., et al. WaveGrad: Estimating Gradients for Waveform Generation . ICLR (2021).
- 192 [4] Chao, J., et al. Fisher information theory for parameter estimation in single molecule microscopy: tutorial.
- Journal of the Optical Society of America A 33, B36 (2016).
- 194 [5] Schermelleh, L. et al. Super-resolution microscopy demystified. Nature Cell Biology vol. 21 72–84 (2019).
- 195 [6] Zhang, B., et al. Gaussian approximations of fluorescence microscope point-spread function models. (2007).
- 196 [7] Smith, C.S., Fast, single-molecule localization that achieves theoretically minimum uncertainty. Nature
- 197 Methods 7, 373–375 (2010).
- 198 [8] Nieuwenhuizen, R., et al. *Measuring image resolution in optical nanoscopy*. Nature Methods 10. 557-562
- 199 (2013).
- 200 [9] Huang, F., et al. Video-rate nanoscopy using sCMOS camera-specific single-molecule localization algorithms.
- 201 Nat Methods 10, 653–658 (2013).
- 202 [10] Rust, M., et al. Sub-diffraction-limit imaging by stochastic optical reconstruction microscopy (STORM).
- 203 Nat Methods 3, 793–796 (2006).
- 204 [11] Betzig, E., et al. Imaging intracellular fluorescent proteins at nanometer resolution. Science 313, 1642–1645
- 205 (2006).

- 206 [12] Weigert, M., et al. Content-aware image restoration: pushing the limits of fluorescence microscopy. Nat.
- 207 Methods 15, 1090 (2018).
- 208 [13] Falk, T., et al. U-net: deep learning for cell counting, detection, and morphometry. Nat. Methods 16, 67–70
- 209 (2019).
- 210 [14] Boyd, N., et al. DeepLoco: fast 3D localization microscopy using neural networks. Preprint at bioRxiv
- 211 https://doi.org/10.1101/267096 (2018)
- 212 [15] Zelger, P., et al. Three-dimensional localization microscopy using deep learning. Opt. Express 26,
- 213 33166–33179 (2018)
- 214 [16] Zhang, P., et al. Analyzing complex single-molecule emission patterns with deep learning. Nat. methods 15,
- 215 913 (2018)
- 216 [17] Saharia, C., et al. Image Super-Resolution via Iterative Refinement. Preprint at arXiv
- 217 https://doi.org/10.48550/arXiv.2104.07636 (2021)