
Uncertainty-Aware Localization Microscopy by Variational Diffusion

Clayton W. Seitz
Department of Physics
Indiana University
Indianapolis, IN 46202
cwseitz@iu.edu

Abstract

Fast extraction of physically relevant information from images using deep neural networks has led to significant advances in fluorescence microscopy and its application to the study of biological systems. For example, the application of deep networks for kernel density (KD) estimation in single molecule localization microscopy (SMLM) has accelerated super-resolution imaging of densely-labeled structures in the cell. However, simple and interpretable uncertainty quantification is lacking in these applications, and remains a necessary modeling component in high-risk research. We propose a generative modeling framework for KD estimation in SMLM based on variational diffusion. This approach allows us to probe the structure of the posterior on KD estimates, creating an additional avenue toward quality control. We demonstrate that data augmentation with traditional SMLM architectures followed by a diffusion process permits simultaneous high-fidelity super-resolution with uncertainty estimation of regressed KDEs.

1 Introduction

Deep models have attracted tremendous attention from researchers in the natural sciences, with several foundational applications arising in microscopy [Weigert et al., 2018, Falk et al., 2019]. Recently, the application of deep image translation in single-molecule localization microscopy (SMLM) has received considerable interest. SMLM techniques are a mainstay of fluorescence microscopy, which localize “blinking” fluorescent molecules to produce a pointillist representation of biomolecules in the cell at diffraction-unlimited precision [Rust et al., 2006, Betzig et al., 2006]. Recently, the use of deep models to perform localization has been proposed as an alternative to traditional localization algorithms, in order to increase imaging speed and labeling density. In previous applications of deep models to localization microscopy, super-resolution images have been recovered from a sparse set of localizations with conditional generative adversarial networks [Ouyang et al., 2018] or localization itself can be performed using traditional convolutional networks [Nehme et al., 2020, Speiser et al., 2021]. In this paper, we perform localization indirectly by predicting kernel density (KD) estimates of a population of fluorescent molecules using a deep model.

Kernel density estimation in SMLM is necessarily performed using a single low-resolution image, and thus common measures of model performance are based on localization errors computed over ensembles of simulated images. Unfortunately, this choice precludes computation of uncertainty at test time under a fixed model. Bayesian probability theory is therefore an attractive alternative, which offers us mathematically grounded tools to reason about uncertainty.

We model a posterior on high-resolution KD estimates conditioned on a low-resolution image. Our approach is based on a type of score based generative model [Song et al., 2021], referred to as a denoising diffusion probabilistic model (DDPM) in the literature [Ho et al., 2020, Song et al., 2021].



Figure 1: Generative model of single molecule localization microscopy images. Low resolution images \mathbf{x} are generated from coordinates θ by integration of the optical transfer function O and sampling from the likelihood (1): $\mathbf{x} \sim p(\mathbf{x}|\theta) = \prod_k p(\mathbf{x}_k|\theta)$. A kernel density estimate \mathbf{y} is inferred from \mathbf{x}

We find that this technique is complementary to relevant existing approaches to uncertainty estimation, which would primarily address epistemic sources of uncertainty, using techniques such as ensembling [Lakshminarayanan et al., 2017] or Monte Carlo dropout [Gal et al., 2022]. The approach is inspired by recent variational perspectives on diffusion [Dirmeier et al., 2023, Ribeiro et al., 2024, Kingma et al., 2021, 2023]. Such techniques provide a mechanism for scalable variational inference, which can be trained using a variational bound written in terms of the signal-to-noise ratio of the diffused data, and a simple noise estimation loss. Indeed, recent efforts have shown that the variational bound can be reparameterized to give several more conventional diffusion losses [Kingma et al., 2021, 2023, Ribeiro et al., 2024].

In the remainder of this paper, we first introduce the likelihood of low-resolution images in localization microscopy, and show uncertainty quantification in a rudimentary example scenario. Then, we introduce KD estimation as an alternative to direct localization using low-resolution images, followed by demonstration of our variational diffusion model for measuring uncertainty KD estimation at scale.

2 Background

2.1 The Image Likelihood

The central objective of SMLM is to infer a set of molecular coordinates $\theta = (\theta_u, \theta_v)$ from measured low resolution images \mathbf{x} . The likelihood on a particular pixel $p(\mathbf{x}_k|\theta)$ is taken to be a convolution of Poisson and Gaussian distributions, due to shot noise $p(s_k) = \text{Poisson}(\omega_k)$ and sensor readout noise $p(\zeta_k) = \mathcal{N}(o_k, w_k^2)$

$$p(\mathbf{x}_k|\theta) = A \sum_{q=0}^{\infty} \frac{1}{q!} e^{-\omega_k} \omega_k^q \frac{1}{\sqrt{2\pi}w_k} e^{-\frac{(\mathbf{x}_k - g_k q - o_k)^2}{2w_k^2}} \approx \text{Poisson}(\omega'_k) \quad (1)$$

where A is some normalization constant. For the sake of generality, we include a per-pixel gain factor g_k , which is often unity. Sampling from $p(\mathbf{x}_k|\theta)$ is trivial; however, for computation of a lower bound on uncertainty in θ , the summation in (1) can be difficult to work with. Therefore, we choose to use a Poisson approximation for simplification, valid under a range of experimental conditions [Huang et al., 2013]. After subtraction of a known offset o_k of the pixel array, which can be easily measured, we have $\omega'_k = \omega_k + w_k^2$. The expectation of the Poisson process ω_k at each pixel of the image must then be computed from the optical impulse response $O(u, v)$, which is taken to be an isotropic Gaussian in two-dimensions. Consider an idealized scenario where an isolated fluorescent molecule exists in the image plane. For a particular pixel k of width δ centered at $\xi_k = (u_k, v_k)$, we define, along the first image dimension

$$\Delta E_u := \int_{u_k - \delta/2}^{u_k + \delta/2} O(u; \theta_u) du = \frac{1}{2} \left(\text{erf} \left(\frac{u_k + \frac{\delta}{2} - \theta_u}{\sqrt{2}\sigma_{\mathbf{x}}} \right) - \text{erf} \left(\frac{u_k - \frac{\delta}{2} - \theta_u}{\sqrt{2}\sigma_{\mathbf{x}}} \right) \right) \quad (2)$$

The expected value at each pixel is then $\omega_k \propto \Delta E_u(u_k, \theta_u, \sigma_{\mathbf{x}}) \Delta E_v(v_k, \theta_v, \sigma_{\mathbf{x}})$. Using this, sampling from the convolution distribution in (1) can be carried out by $\mathbf{x}_k = s_k + \zeta_k$ for

$s_k \sim \text{Poisson}(\omega_k)$, $\eta_k \sim \mathcal{N}(o_k, w_k^2)$. The complete generative process is depicted in (Figure 1). In this simple case, an estimate of the posterior $p(\theta|\mathbf{x})$ can be found by Markov Chain Monte Carlo (MCMC) sampling as shown in (Figure 2). Details of the MCMC algorithm used here are outlined in the Appendix.

2.2 Gaussian kernel density estimation

Direct optimization of the likelihood in (1) from observations \mathbf{x} alone is challenging when fluorescent emitters are dense within the field of view and fluorescent signals significantly overlap. However, convolutional neural networks (CNNs) have recently proven to be powerful tools fluorescence microscopy to extract parameters describing fluorescent emitters such as color, emitter orientation, z -coordinate, and background signal Zhang et al. [2018], Kim et al. [2019], Zelger et al. [2018]. For localization tasks, CNNs typically employ upsampling layers to reconstruct Bernoulli probabilities of emitter occupancy [Speiser et al., 2021] or KD estimates with higher resolution than experimental measurements [Nehme et al., 2020]. We choose to use KD estimates in our model, denoted by \mathbf{y} , which are latent in the low-resolution data \mathbf{x} . KDEs are the most common data structure used in SMLM, and can be easily generated from molecular coordinates, alongside observations \mathbf{x} .

Similar to the generative process on low resolution images \mathbf{x} , we generate KDEs \mathbf{y} by repurposing the generative model (1) on an unsampled image without noise. In other words, we cast Gaussian KD estimation as a noiseless image generation process on the domain of \mathbf{y} . Under a fixed configuration of N particles θ , the value of a non-normalized KDE pixel \mathbf{y}_k is given by

$$\mathbf{y}_k(\theta) = \sum_{n=1}^N \Delta E_u(u_k, \theta_u, \sigma_{\mathbf{y}}) \Delta E_v(v_k, \theta_v, \sigma_{\mathbf{y}}) \quad (3)$$

where the hyperparameter $\sigma_{\mathbf{y}}$ is a Gaussian kernel width.

3 Uncertainty-Aware Localization Microscopy by Variational Diffusion

We now consider more realistic datasets $(\mathbf{x}_i, \mathbf{y}_{0,i}, \hat{\mathbf{y}}_i)_{i=1}^N$ of observed images \mathbf{x}_i true KD images $\mathbf{y}_{0,i}$, and augmented low-resolution inputs $\hat{\mathbf{y}}_i = \phi(\mathbf{x}_i)$, where ϕ is a CNN. Observations \mathbf{x}_i are simulated under the convolution distribution (1) and KDEs are generated by (4).

3.1 Problem Statement

Kernel density estimates produced by the traditional deep architectures for localization microscopy produce strong results, but lack uncertainty quantification. Unfortunately, the posterior $p(\theta|\mathbf{x})$ has no known analytical form and can be difficult to compute at test time, since (i) molecules cannot be easily resolved and therefore θ is of unknown dimension and (ii) θ can be high dimensional and efficient exploration of the parameter space is challenging. The central goal of this paper is to instead model a conditional distribution on the latent \mathbf{y} : $p(\mathbf{y}|\mathbf{x})$, where \mathbf{y} is of known dimensionality. We choose to model $p(\mathbf{y}|\mathbf{x})$ with a diffusion model, given that the distribution $p(\mathbf{y}|\mathbf{x})$ is expensive to compute, even if $p(\theta|\mathbf{x})$ were known.

Recent advances in generative modeling, particularly diffusion models [Sohl-Dickstein et al., 2015, Ho et al., 2020, Song et al., 2021] present a unique opportunity to integrate uncertainty awareness into the localization microscopy toolkit. However, sampling from diffusion models can be computationally expensive, given that generation amounts to solving a complex stochastic differential equation, effectively mapping a simple base distribution to the complex data distribution. The solution of such equations requires numerical integration with very small step sizes, resulting in thousands of neural network evaluations [Saharia et al., 2021, Vahdat et al., 2021]. For conditional generation tasks in high-risk applications, generation complexity is further exacerbated by the need for the highest level of detail in generated samples. Therefore, we propose that sampling is preceded by an augmentation network ϕ , which in essence generates an initial estimate to guide the diffusion process. Reasoning for this choice in our application is two-fold:

Synthesis Speed. By training the augmentation network ϕ to obtain an approximate estimate of \mathbf{y}_0 , we can reduce the number of iterations, since the diffusion model only needs to model the remaining

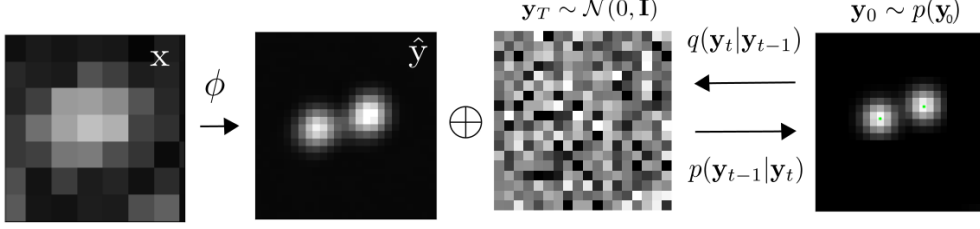


Figure 2: Conditional diffusion model for sampling kernel density estimates. An initial KDE estimate $\hat{\mathbf{y}}$ is found by the CNN ϕ , which is used as conditional input in the DDPM

mismatch, resulting in a less complex model from which sampling becomes easier. Speed is critical in SMLM applications, which can produce thousands of images in a single experiment.

Sample Fidelity. Since Langevin dynamics will often be initialized in low-density regions of the data distribution, inaccurate score estimation in these regions will negatively affect the sampling process. Moreover, mixing can be difficult because of the need of traversing low density regions to transition between modes of the distribution [Song et al., 2019].

3.2 Variational Diffusion

Diffusion models [Sohl-Dickstein et al., 2015, Ho et al., 2020, Song et al., 2021] are a class of generative models originally inspired by nonequilibrium statistical physics, which slowly destroy structure in a data distribution via a fixed Markov chain referred to as the *forward process*. In the present context, we leverage the variational interpretation of this model class [Kingma et al., 2021, 2023] to approximate the posterior $p(\mathbf{y}|\mathbf{x})$.

Diffusion Model. We use a forward process which gradually adds Gaussian noise to the latent \mathbf{y}_0 in discrete time, according to a variance schedule β_t :

$$q(\mathbf{y}_T|\mathbf{y}_0) = \prod_{t=1}^T q(\mathbf{y}_t|\mathbf{y}_{t-1}) \quad q(\mathbf{y}_t|\mathbf{y}_{t-1}) = \mathcal{N}\left(\sqrt{1-\beta_t}\mathbf{y}_{t-1}, \beta_t \mathbf{I}\right) \quad (4)$$

An important property of the forward process is that it admits sampling \mathbf{y}_t at an arbitrary timestep t in closed form [Ho et al., 2020]. Using the notation $\alpha_t := 1 - \beta_t$ and $\gamma_t := \prod_{s=1}^t \alpha_s$, we have $q(\mathbf{y}_t|\mathbf{y}_0) = \mathcal{N}(\sqrt{\gamma_t}\mathbf{y}_0, (1-\gamma_t)\mathbf{I})$ or $\mathbf{y}_t = \sqrt{\gamma_t}\mathbf{y}_0 + \sqrt{1-\gamma_t}\epsilon$ for $\epsilon \sim \mathcal{N}(0, \mathbf{I})$. The signal to noise ratio (SNR) as defined in [Kingma et al., 2023], at a time step t reads $\text{SNR}_t = \gamma_t/(1-\gamma_t)$.

The usual procedure is then to learn a parametric representation of the *reverse process*, and therefore generate samples of the latent \mathbf{y}_0 from $p(\mathbf{y}_0|\mathbf{x})$. Formally, $p_\psi(\mathbf{y}_0|\mathbf{x}) = \int p_\psi(\mathbf{y}_{0:T}|\mathbf{x}) d\mathbf{y}_{1:T}$ where \mathbf{y}_t is a latent representation with the same dimensionality of the data and $p_\psi(\mathbf{y}_{0:T}|\mathbf{x})$ is a Markov process, starting from a noise sample $p_\psi(\mathbf{y}_T) = \mathcal{N}(0, \mathbf{I})$. Writing this Markov process gives

$$p_\psi(\mathbf{y}_{0:T}|\mathbf{x}) = p_\psi(\mathbf{y}_T) \prod_{t=1}^T p_\psi(\mathbf{y}_{t-1}|\mathbf{y}_t, \mathbf{x}) \quad p_\psi(\mathbf{y}_{t-1}|\mathbf{y}_t, \mathbf{x}) = \mathcal{N}(\mu_\psi(\mathbf{y}_t, \gamma_t), \beta_t \mathbf{I}) \quad (5)$$

where we reuse the variance schedule of the forward process [Ho et al., 2020]. From (5) it can be seen that the learnable parameter in the reverse process is the expectation of the transition μ_ψ where ψ is a neural network.

Learning the reverse process can be approached by either regressing noise ϵ from the forward process, or the true latent \mathbf{y}_0 , as there is a deterministic relationship between them. We adopt the former for consistency with other work, and define ψ as a neural denoising function which regresses the noise ϵ from a noisy \mathbf{y}_t . A relation between the noise estimate ϵ_ψ and μ_ψ is given in the Appendix, which gives an intuition for sampling. The proposed sampling scheme is depicted in (Figure 3).

Variational Objective. Following [Kingma et al., 2021], we interpret the reverse process as a hierarchical generative model that samples a sequence of latents \mathbf{y}_t , with time running backward. Training of the model is achieved through the variational bound

$$-\log p(\mathbf{y}_0) \leq -\mathbb{E}_{q(\mathbf{y}_{1:T}|\mathbf{y}_0)} \log \left(\frac{p_\psi(\mathbf{y}_{1:T}, \mathbf{y}_0)}{q(\mathbf{y}_{1:T}|\mathbf{y}_0)} \right) \quad (6)$$

$$= D_{KL}(q(\mathbf{y}_T|\mathbf{y}_0)||p(\mathbf{y}_T)) + \mathbb{E}_{q(\mathbf{y}_1|\mathbf{y}_0)} \log p(\mathbf{y}_0|\mathbf{y}_1) + \mathcal{L}_\psi \quad (7)$$

where we have omitted conditioning on the low-resolution \mathbf{x} to simplify the notation. Note that, this is similar to a hierarchical VAE, but in a diffusion model $q(\mathbf{y}_{1:T}|\mathbf{y}_0)$ is fixed by the forward process rather than learned. The so-called diffusion loss \mathcal{L}_ψ is shown in Appendix A, and is the term of interest as the first two terms do not contribute meaningfully to the loss [Ho et al., 2020]. Furthermore, it has become standard to use simplified forms of \mathcal{L}_ψ , such as a noise estimation loss, as this has shown superior performance. Importantly, \mathcal{L}_ψ is simply a reweighted variant of a family of diffusion objectives [Kingma et al., 2021, 2023]. We use the following Monte Carlo estimate of \mathcal{L}_ψ , which demonstrates that the variational bound can be written in terms of the common noise-estimation loss

$$\mathcal{L}_\psi = \mathbb{E}_{\epsilon \sim \mathcal{N}(0, I), t \sim U(1, T)} \left[\left(\frac{\text{SNR}_{t-1}}{\text{SNR}_t} - 1 \right) \|\epsilon - \epsilon_\psi\|_2^2 \right] \quad (8)$$

A full derivation of this objective is outlined in the Appendix. Note that SNR_t is monotonically decreasing with t , and thus $\frac{\text{SNR}_{t-1}}{\text{SNR}_t} = \frac{\gamma_{t-1}}{\gamma_t} \frac{1-\gamma_t}{1-\gamma_{t-1}} \geq 1$, ensuring $\mathcal{L}_\psi \geq 0$. In this paper, we choose to use a uniformly weighted loss and leave the exploration of the weighted loss to future work.

4 Experiments

All training data consists of low-resolution 20×20 images, setting $\sigma_x = 0.92$ in units of low-resolution pixels, for consistency with common experimental conditions with a 60x magnification objective lens and numerical aperture (NA) of 1.4. We multiply ω_k by a constant $i_0 = 200$ for experiments for consistency with typical fluorophore emission rates. All KDEs have dimension 80×80 , are scaled between $[0, 1]$, and are generated using $\sigma_y = 3.0$ pixels in the upsampled image. For a typical CMOS camera, this results in KDE pixels with lateral dimension of $\approx 27\text{nm}$. Initial coordinates θ were drawn uniformly over a two-dimensional disc with a radius of 7 low-resolution pixels.

Localization RMSE. In order to verify the initial predictions made by the augmentation model ϕ , we simulated a dataset $(\mathbf{x}_i, \mathbf{y}_{0,i}, \hat{\mathbf{y}}_i)_{i=1}^N$ with $N = 1000$. Objects in the KDE $\hat{\mathbf{y}}_i$ are detected using the Laplacian of Gaussian (LoG) detection algorithm [Kong et al., 2013], which permits more direct comparison of model predictions to the Cramer-Rao lower bound on localization error, compared to other image similarity measures. Localization is carried out from scale-space maxima directly in LoG, as opposed to fitting a model function to KDEs. A particular LoG localization in the KDE is paired to the nearest ground truth localization and is unpaired if a localization is not within 5 KDE pixels of any ground truth localization. In addition to localization error, we measure a precision $P = \text{TP}/(\text{TP} + \text{FP}) = 1.0$ and recall $R = \text{TP}/(\text{TP} + \text{FN}) = 0.85$, where TP denotes true positive localizations, FP denotes false positive localizations, and FN denotes false negative localizations.

Variational Diffusion. We set $T = 100$ for all experiments and treat forward process variances β_t as hyperparameters, with a linear schedule from $\beta_0 = 10^{-4}$ to $\beta_T = 10^{-2}$. These constants were chosen to be small relative to ground truth KDEs, which are scaled to $[-1, 1]$, ensuring that forward process distribution $\mathbf{y}_T \sim q(\mathbf{y}_T|\mathbf{y}_0)$ approximately matches the reverse process $\mathbf{y}_T \sim \mathcal{N}(0, I)$ at $t = T$. Example KD estimates from low-resolution images and the marginal variances obtained from sampling $N = 100$ samples from $p_\psi(\mathbf{y}_0|\mathbf{x})$ are shown in (Figure 4).

Super-resolved immunofluorescence. To test our model in an experimental context, we performed immunofluorescence staining of endogenous BRD4 protein in HeLa cells. Cells were grown in a 35mm dish and fixed with Formaldehyde in 1X PBS at room temperature incubator for 10 minutes, and then permeabilized in 70 percent ethanol. Cells were blocked for 1h with 0.3 percent (v/v) Triton-X100 (Sigma-Aldrich) and 5 percent (w/v) nonfat dry milk at 4C. Cells were incubated

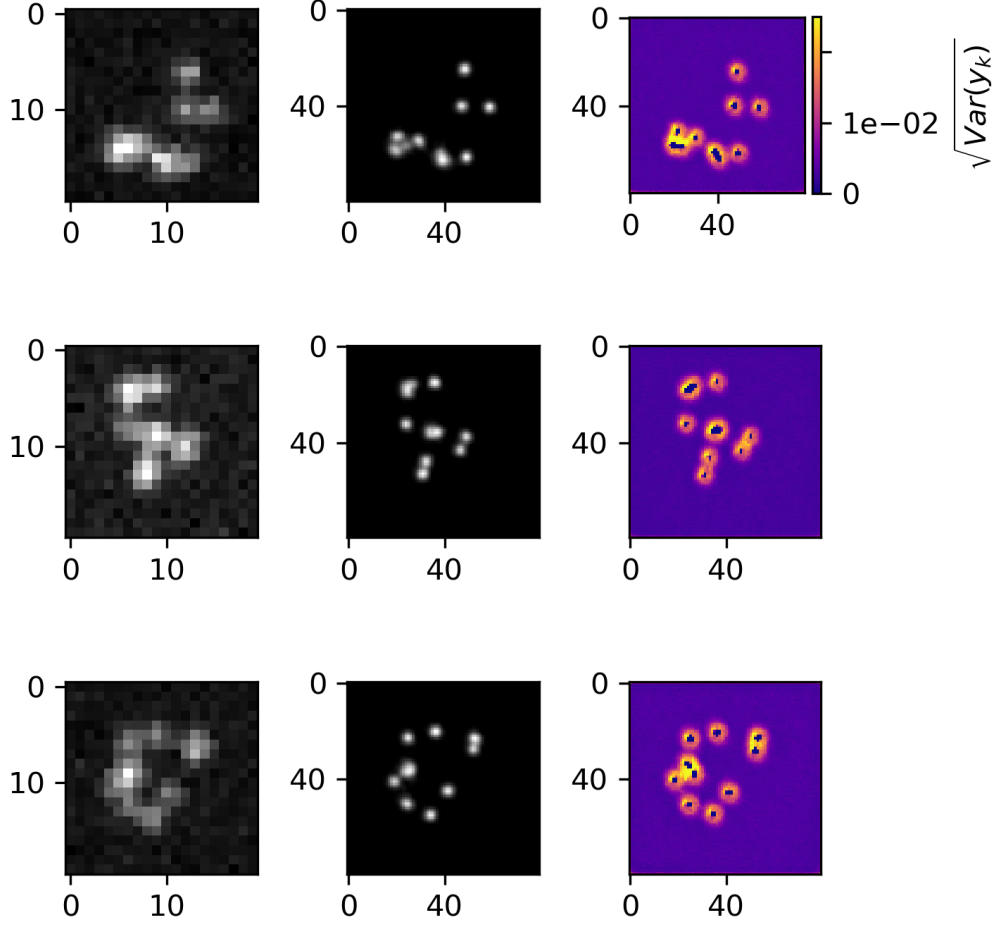


Figure 3: Non cherry-picked estimation of marginal variances. A low-resolution image \mathbf{x} (left column) is transformed by ϕ to produce a KDE estimate $\hat{\mathbf{y}}$ (middle column) and a DDPM ψ computes a map of marginal variances (right column)

overnight at 4C using anti-BRD4 primary antibody (Cell Signaling, clone E2A7X; 1:1000) in blocker. Cells were then washed and stained with secondary antibodies for BRD4 (Cell Signaling Anti-Mouse IgG-Alexa488, 1:1000). For imaging, cells were imaged using a 60X 1.4NA oil immersion objective. Images were projected onto a CMOS Fusion camera (Hamamatsu) with a 40ms exposure time, using a 488nm laser aligned for oblique illumination. After background subtraction, the encoding network and diffusion model were used to estimate the super-resolved immunofluorescence image and its corresponding uncertainty (Figure 4).

5 Conclusion

We proposed a variational diffusion model for uncertainty-aware localization microscopy. Our approach builds on recent advancements in conditional diffusion models, to model the posterior distribution on high-resolution KD estimates from low-resolution inputs. This tractable posterior distribution is constructed by first augmenting low resolution inputs to a KD estimate using the DeepSTORM architecture with minor modifications [Nehme et al., 2020]. Conditioning a diffusion model on this initial estimate permits sampling with relatively fewer samples than most existing

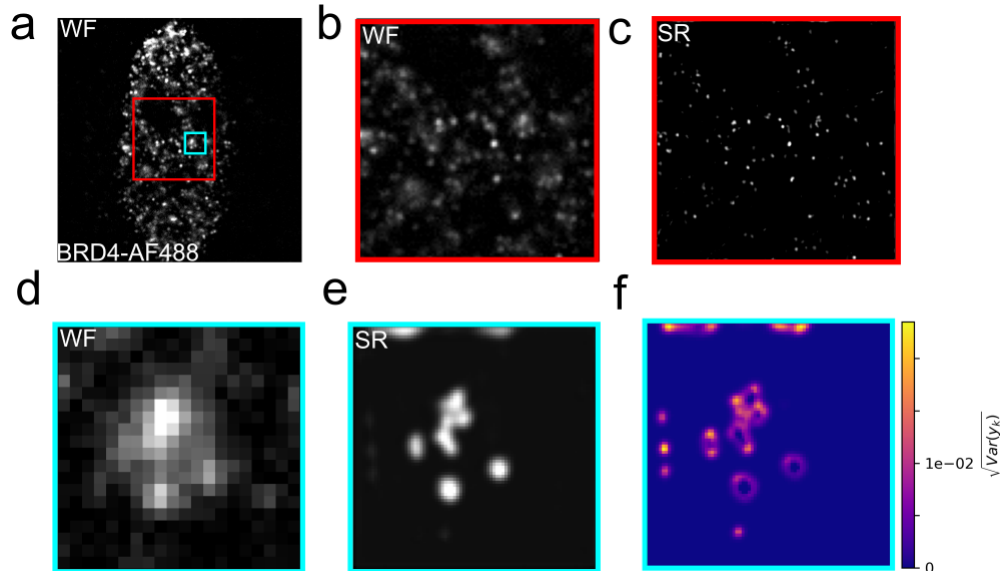


Figure 4: Super-resolution of BRD4 puncta in a HeLa cell nucleus. (a,b) Widefield immunofluorescence image of a BRD4 nucleus stained with Alexa488 secondary antibody (c) Super-resolved immunofluorescence image obtained by application of the deep model ϕ only (d-f) 20×20 inset region of the immunofluorescence image, pixel-wise mean of 100 samples obtained by the diffusion model, and the pixel-wise variance over 100 samples.

diffusion models in similar applications, thereby making computation of marginal variances possible. Our approach made three core contributions: (i) we derived a relationship between the posterior on kernel density estimates with the posterior on molecular locations, and (ii) we demonstrated that a diffusion model can model a distribution on KDEs with qualitatively similar marginal variances expected from predictions made using MCMC. By using a recently discovered relationship of the variational lower bound to a traditional noise-estimation objective, we can confidently approximate the true posterior.

6 Broader Impact

The development of a method for uncertainty estimation in super-resolution imaging, as proposed here, holds implications beyond its immediate application in SMLM. By leveraging diffusion models for uncertainty estimation, this approach not only enhances the reliability of super-resolution image reconstructions but also extends its utility to a diverse array of domains. The incorporation of a guided diffusion process facilitates efficient reconstruction while maintaining interpretation of the underlying uncertainty. Importantly, the principles underlying this method resonate across various fields, suggesting its potential applicability in domains beyond microscopy. For instance, the extension of similar techniques to general image processing tasks highlights the potential to address uncertainty in a wide range of applications in bioimaging or medical imaging. Moreover, the utilization of diffusion models for uncertainty estimation aligns with a broader trend in leveraging probabilistic frameworks for enhancing deep learning applications, with implications extending to fields such as natural language processing, computer vision, and autonomous systems. By bridging these interdisciplinary boundaries, this method not only addresses a critical need in localization microscopy but also contributes to the advancement of uncertainty-aware deep learning methodologies.

References

Eric Betzig et al. Imaging intracellular fluorescent proteins at nanometer resolution. *Science*, 313: 1642–1645, 2006.

- Jin Chao et al. Fisher information theory for parameter estimation in single molecule microscopy: tutorial. *Journal of the Optical Society of America A*, 2016.
- Simon Dirmeyer et al. Diffusion models for probabilistic programming. In *Advances in Neural Information Processing Systems*, 2023.
- Thorsten Falk et al. U-net: deep learning for cell counting, detection, and morphometry. *Nature Methods*, 16:67–70, 2019.
- Yarin Gal et al. Bayesian uncertainty quantification for machine-learned models in physics. *Nature Physics*, 2022.
- Sarah Frisken Gibson and Frederick Lanni. Diffraction by a circular aperture as a model for three-dimensional optical microscopy. *J. Opt. Soc. Am. A*, 6:1357–1367, 1989.
- Jonathan Ho et al. Denoising diffusion probabilistic models. In *Advances in Neural Information Processing Systems*, 2020.
- Fang Huang et al. Video-rate nanoscopy using sCMOS camera-specific single-molecule localization algorithms. *Nature Methods*, 10:653–658, 2013.
- Tae Hyun Kim et al. Information-rich localization microscopy through machine learning. *Nature Communications*, 10:1996, 2019.
- Diederik Kingma et al. Variational diffusion models. In *Advances in Neural Information Processing Systems*, 2021.
- Diederik Kingma et al. Understanding diffusion objectives as the elbo with simple data augmentation. In *Advances in Neural Information Processing Systems*, 2023.
- Hui Kong et al. A generalized laplacian of gaussian filter for blob detection and its applications. *IEEE Transactions on Cybernetics*, 43:1719–1733, 2013.
- Balaji Lakshminarayanan et al. Simple and scalable predictive uncertainty estimation using deep ensembles, 2017.
- Elias Nehme et al. Deepstorm3d: Dense 3d localization microscopy and psf design by deep learning. *Nature Methods*, 17:734–740, 2020.
- Wei Ouyang et al. Deep learning massively accelerates super-resolution localization microscopy. *Nature Biotechnology*, 36:460–468, 2018.
- Filipe Ribeiro et al. Demystifying variational diffusion models. 2024.
- Richards and Wolf. Electromagnetic diffraction in optical systems. *Proceedings of the Royal Society A*, 253:1358–379, 1959.
- Michael Rust et al. Sub-diffraction-limit imaging by stochastic optical reconstruction microscopy (storm). *Nature Methods*, 3:793–796, 2006.
- Chinmay Saharia et al. Image super-resolution via iterative refinement. 2021.
- Carlos Smith et al. Fast, single-molecule localization that achieves theoretically minimum uncertainty. *Nature Methods*, 7:373–375, 2010.
- Jascha Sohl-Dickstein et al. Deep unsupervised learning using nonequilibrium thermodynamics. In *International Conference on Learning Representations*, 2015.
- Yang Song et al. Generative modeling by estimating gradients of the data distribution. In *Advances in Neural Information Processing Systems*, 2019.
- Yang Song et al. Score-based generative model through stochastic differential equations. In *International Conference on Learning Representations*, 2021.
- Adam Speiser et al. Deep learning enables fast and dense single-molecule localization with high accuracy. *Nature Methods*, 18:1082–1090, 2021.

- Arash Vahdat et al. Score-based generative modeling in latent space. In *Advances in Neural Information Processing Systems*, 2021.
- Martin Weigert et al. Content-aware image restoration: pushing the limits of fluorescence microscopy. *Nature Methods*, 15:1090, 2018.
- Philipp Zelger et al. Three-dimensional localization microscopy using deep learning. *Opt. Express*, 26:33166–33179, 2018.
- Bo Zhang et al. Gaussian approximations of fluorescence microscope point-spread function models. *Applied Optics*, 46:1819–1819–1829, 2007.
- Peng Zhang et al. Analyzing complex single-molecule emission patterns with deep learning. *Nature Methods*, 15:913, 2018.

A Appendix

A.1 Sampling

Sampling from the reverse process $p_\psi(\mathbf{y}_{t-1}|\mathbf{y}_t, \mathbf{x})$ is achieved by estimation of the noise ϵ_ψ from \mathbf{y}_t by the denoising model ψ , and therefore estimation of \mathbf{y}_0

$$\hat{\mathbf{y}}_0 = \frac{1}{\sqrt{\gamma_t}}(\mathbf{y}_t - \sqrt{1 - \gamma_t}\epsilon_\psi) \quad (9)$$

followed by sampling from the forward process $\mathbf{y}_{t-1} \sim q(\mathbf{y}_{t-1}|\hat{\mathbf{y}}_0) = \mathcal{N}(\sqrt{\gamma_{t-1}}, (1 - \gamma_{t-1})I)$.

A.2 Derivation of the variational bound

We now derive the so-called diffusion loss \mathcal{L}_ψ , written in (8) in the main text. Similar derivations can be found in [Kingma et al., 2021, Ribeiro et al., 2024], and we include it here only for completeness

$$\begin{aligned} -\log p(\mathbf{y}_0) &\leq -\mathbb{E}_{q(\mathbf{y}_{1:T}|\mathbf{y}_0)} \log \frac{p(\mathbf{y}_{0:T})}{q(\mathbf{y}_{1:T}|\mathbf{y}_0)} \\ &= -\mathbb{E}_{q(\mathbf{y}_{1:T}|\mathbf{y}_0)} \log \frac{p(\mathbf{y}_T)p(\mathbf{y}_0|\mathbf{y}_1) \prod_{t=2}^T p(\mathbf{y}_{t-1}|\mathbf{y}_t)}{q(\mathbf{y}_T|\mathbf{y}_0) \prod_{t=2}^T q(\mathbf{y}_{t-1}|\mathbf{y}_t, \mathbf{y}_0)} \\ &= -\mathbb{E}_{q(\mathbf{y}_{1:T}|\mathbf{y}_0)} \left[p(\mathbf{y}_0|\mathbf{y}_1) + \log \frac{p(\mathbf{y}_T)}{q(\mathbf{y}_T|\mathbf{y}_0)} + \sum_{t=2}^T \log \frac{p(\mathbf{y}_{t-1}|\mathbf{y}_t)}{q(\mathbf{y}_{t-1}|\mathbf{y}_t, \mathbf{y}_0)} \right] \\ &= -\mathbb{E}_{q(\mathbf{y}_{1:T}|\mathbf{y}_0)} [p(\mathbf{y}_0|\mathbf{y}_1)] + D_{KL}(q(\mathbf{y}_T|\mathbf{y}_0)||p(\mathbf{y}_T)) \\ &\quad + \sum_{t=2}^T \mathbb{E}_{q(\mathbf{y}_t|\mathbf{y}_0)} D_{KL}(q(\mathbf{y}_{t-1}|\mathbf{y}_t, \mathbf{y}_0)||p(\mathbf{y}_{t-1}|\mathbf{y}_t)) \end{aligned}$$

As before, we omit conditioning on \mathbf{x} to simplify the notation. The first term is typically ignored, as it does not contribute meaningfully to the loss [Ribeiro et al., 2024]. Furthermore, the second term is approximately zero by construction. Therefore we are left with the last term, called the diffusion loss \mathcal{L}_ψ . The KL-divergence of q and p is between two Gaussians with identical variances $\sigma^2 = \frac{(1-\gamma_{t-1})(1-\alpha_t)}{1-\gamma_t}$, and expectations

$$\mu = \frac{\sqrt{\gamma_{t-1}}(1-\alpha_t)}{1-\gamma_t}\mathbf{y}_0 + \frac{\sqrt{\alpha_t}(1-\gamma_{t-1})}{1-\gamma_t}\mathbf{y}_t \quad \mu_\psi = \frac{\sqrt{\gamma_{t-1}}(1-\alpha_t)}{1-\gamma_t}\hat{\mathbf{y}}_0 + \frac{\sqrt{\alpha_t}(1-\gamma_{t-1})}{1-\gamma_t}\mathbf{y}_t$$

for a fixed noise schedule [Saharia et al., 2021]. Therefore, we have

$$\begin{aligned}
D_{KL}(q(\mathbf{y}_{t-1}|\mathbf{y}_t, \mathbf{y}_0)||p(\mathbf{y}_{t-1}|\mathbf{y}_t)) &= \frac{1}{2\sigma^2} \|\mu - \mu_\psi\|_2^2 \\
&= \frac{1}{2} \frac{\gamma_{t-1}(1 - \alpha_t)}{(1 - \gamma_{t-1})(1 - \gamma_t)} \|\mathbf{y}_0 - \hat{\mathbf{y}}_0\|_2^2 \\
&= \frac{1}{2} \frac{\gamma_{t-1}((1 - \gamma_t) - \alpha_t(1 - \gamma_{t-1}))}{(1 - \gamma_{t-1})(1 - \gamma_t)} \|\mathbf{y}_0 - \hat{\mathbf{y}}_0\|_2^2 \\
&= \frac{1}{2} \frac{\gamma_{t-1}((1 - \gamma_t) - \frac{\gamma_t}{\gamma_{t-1}}(1 - \gamma_{t-1}))}{(1 - \gamma_{t-1})(1 - \gamma_t)} \|\mathbf{y}_0 - \hat{\mathbf{y}}_0\|_2^2 \\
&= \frac{1}{2} \left(\frac{\gamma_{t-1}}{1 - \gamma_{t-1}} - \frac{\gamma_t}{1 - \gamma_t} \right) \|\mathbf{y}_0 - \hat{\mathbf{y}}_0\|_2^2 \\
&= \frac{1}{2} (\text{SNR}_{t-1} - \text{SNR}_t) \|\mathbf{y}_0 - \hat{\mathbf{y}}_0\|_2^2
\end{aligned}$$

Reparameterizing the loss in terms of the noise, using $\|\mathbf{y}_0 - \hat{\mathbf{y}}_0\|_2^2 = \frac{1-\gamma_t}{\gamma_t} \|\epsilon_0 - \epsilon_\psi\|_2^2$ [Ribeiro et al., 2024], we arrive at

$$\mathcal{L}_\psi = \frac{1}{2} \sum_{t=2}^T \mathbb{E}_{q(\mathbf{y}_t|\mathbf{y}_0)} \left(\frac{\text{SNR}_{t-1}}{\text{SNR}_t} - 1 \right) \|\epsilon - \epsilon_\psi\|_2^2$$

Using a Monte Carlo estimate of \mathcal{L}_ψ [Kingma et al., 2023] which optimizes random terms of the summation to avoid calculating all terms simultaneously, we arrive at the objective written in the main text (8)

$$\mathcal{L}_\psi = \mathbb{E}_{\epsilon \sim \mathcal{N}(0, I), t \sim U(1, T)} \left[\left(\frac{\text{SNR}_{t-1}}{\text{SNR}_t} - 1 \right) \|\epsilon - \epsilon_\psi\|_2^2 \right]$$

A.3 Optical impulse response

It is common to describe the optical impulse response of a microscope as a two-dimensional isotropic Gaussian [Zhang et al., 2007]. This is an approximation to the more rigorous diffraction models given by [Richards and Wolf, 1959, Gibson and Lanni, 1989]. Over a continuous domain, the impulse response reads

$$O(u, v) = \frac{1}{2\pi\sigma_x^2} e^{-\frac{(u-\theta_u)^2 + (v-\theta_v)^2}{2\sigma_x^2}}$$

The above expression can be interpreted as a probability distribution over locations where a photon can be detected. Therefore, for discrete detectors, we discretize this expression by integrating over pixels. The number of photon arrivals will follow Poisson statistics, with expected value

$$\omega_k = i_0 \left(\int_{u_k - \delta/2}^{u_k + \delta/2} O(u; \theta_u) du \right) \left(\int_{v_k - \delta/2}^{v_k + \delta/2} O(v; \theta_v) dv \right)$$

The scalar quantity i_0 represents the amplitude of the signal, which is proportional the quantum efficiency of a pixel η , the duration of exposure, Δ , and the number of photons emitter by a fluorescent molecule N_0 . With no loss of generality, $\Delta = \eta = 1$ and there is a single free parameter N_0 . A simple change of variables $u' = u - \theta_u$ and $v' = v - \theta_v$ gives

$$\omega_k = i_0 \left(\int_{u_k - \delta/2 - \theta_u}^{u_k + \delta/2 - \theta_u} O(u) du \right) \left(\int_{v_k - \delta/2 - \theta_v}^{v_k + \delta/2 - \theta_v} O(v) dv \right)$$

One of these terms can be written as

$$\begin{aligned} \int_{u_k - \delta/2 - \theta_u}^{u_k + \delta/2 - \theta_u} O(u) du &= \int_0^{u_k + \delta/2 - \theta_u} O(u) du - \int_0^{u_k - \delta/2 - \theta_u} O(u) du \\ &= \frac{1}{2} \left(\operatorname{erf} \left(\frac{u_k + \frac{\delta}{2} - \theta_i}{\sqrt{2}\sigma_x} \right) - \operatorname{erf} \left(\frac{u_k - \frac{\delta}{2} - \theta_i}{\sqrt{2}\sigma_x} \right) \right) \end{aligned}$$

where we have used the common definition $\operatorname{erf}(z) = \frac{2}{\sqrt{\pi}} \int_0^z e^{-t^2} dt$.

A.4 Metropolis-Hastings MCMC

To obtain numerical estimates of $p(\theta|\mathbf{x}) \propto p(\mathbf{x}|\theta)p(\theta)$ and therefore $p(\mathbf{y}|\mathbf{x})$, for the isolated fluorescent molecule as shown in (Figure 2), we used Metropolis-Hastings Markov Chain Monte Carlo (MCMC) to estimate the posterior on coordinates. Under the Poisson approximation in (1), the model negative log-likelihood is

$$\ell(\mathbf{x}|\theta) = -\log \prod_k \frac{e^{-(\omega'_k)} (\omega'_k)^{n_k}}{n_k!} = \sum_k \log n_k! + \omega'_k - n_k \log (\omega'_k) \quad (10)$$

where n_k is the observed number events at a pixel. MCMC is asymptotically exact, which is not guaranteed by variational methods which may rely on a Laplace approximation around the MLE. We choose a uniform prior $p(\theta)$, and Metropolis-Hastings is run for 10^4 iterations, the first 10^3 iterations are discarded as burn-in. A proposal $\theta' = \theta + \Delta\theta$ was generated with $\Delta\theta \sim \mathcal{N}(0, \sigma^2 I)$ where $\sigma^2 = 0.05$. The acceptance probability is

$$\alpha = e^{\beta(\ell(\theta) - \ell(\theta'))}$$

We choose $\beta = 0.2$ to achieve a target acceptance rate of 0.5.

A.5 Cramer-Rao Lower Bound

Reliable inference of θ from \mathbf{x} in general requires performance metrics for model selection. We use the Fisher information as an information theoretic criteria to assess the quality of the data augmentation model ϕ tested here, with respect to the root mean squared error (RMSE) of our predictions of θ [Chao et al., 2016]. The Poisson log-likelihood $\ell(\mathbf{x}|\theta)$ is also convenient for computing the Fisher information matrix [Smith et al., 2010] and thus the Cramer-Rao lower bound, which bounds the variance of a statistical estimator of θ , from below i.e., $\operatorname{var}(\hat{\theta}) \geq I^{-1}(\theta)$. The Fisher information is straightforward to compute under the Poisson log-likelihood in (1). In general, the Fisher information is given by the expression

$$I_{ij}(\theta) = \mathbb{E} \left(\frac{\partial \ell}{\partial \theta_i} \frac{\partial \ell}{\partial \theta_j} \right) \quad (11)$$

For an arbitrary parameter, we find that, for a Poisson log-likelihood ℓ

$$\begin{aligned} \frac{\partial \ell}{\partial \theta_i} &= \frac{\partial}{\partial \theta_i} \sum_k \log n_k! + \omega'_k - n_k \log (\omega'_k) \\ &= \sum_k \frac{\partial \omega'_k}{\partial \theta_i} \left(\frac{\omega'_k - n_k}{\omega'_k} \right) \end{aligned}$$

Using this result, we can compute the Fisher information matrix $I(\theta)$

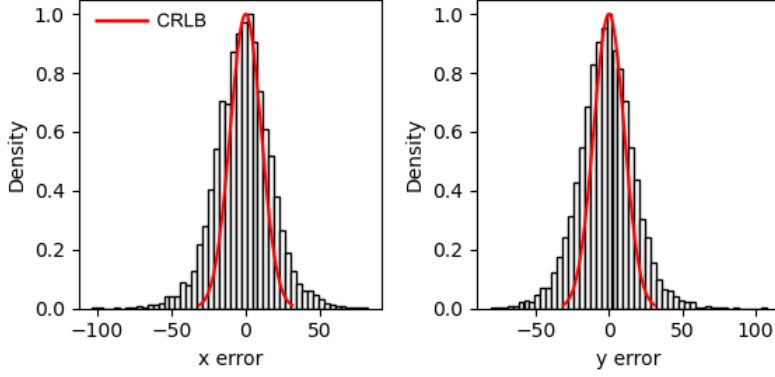


Figure 5: Localization errors of the trained model ϕ . The Cramer-Rao lower bound is shown in red, computing by taking the diagonal elements of $I^{-1}(\theta)$.

$$I_{ij}(\theta) = \mathbb{E}_{\theta} \left(\sum_k \frac{\partial \omega'_k}{\partial \theta_i} \frac{\partial \omega'_k}{\partial \theta_j} \left(\frac{\omega'_k - n_k}{\omega'_k} \right)^2 \right) = \sum_k \frac{1}{\omega'_k} \frac{\partial \omega'_k}{\partial \theta_i} \frac{\partial \omega'_k}{\partial \theta_j}$$

A fundamental lower bound on the variance in our estimates of θ then is found from its inverse: $\text{CRLB} = I^{-1}(\theta)$. This result is used to show in (Figure 5), that the data augmentation model ϕ efficiently estimates molecular coordinates under the experimental conditions tested here.

A.6 Neural Networks ψ, ϕ

DeepSTORM CNN ϕ . The DeepSTORM CNN, for 3D localization, can be viewed as a deep kernel density estimator, reconstructing kernel density estimates \mathbf{y} from low-resolution inputs \mathbf{x} . We utilize a simplified form of the original architecture [Nehme et al., 2020] for 2D localization, which we denote ϕ in this paper, which consists of three main modules: a multi-scale context aggregation module, an upsampling module, and a prediction module. For context aggregation, the architecture utilizes dilated convolutions to increase the receptive field of each layer. The upsampling module is then composed of two consecutive 2x resize-convolutions, computed by nearest-neighbor interpolation, to increase the lateral resolution by a factor of 4. Additional details regarding this architecture can be found in the original paper Nehme et al. [2020]. The terminal prediction module contains three additional convolutional blocks for refinement of the upsampled image, followed by an element-wise HardTanh. The architecture is trained using the objective $\mathcal{L}_{\phi} = \frac{1}{N} \sum_{n=1}^N (\mathbf{y}_{0,n} - \hat{\mathbf{y}}_n)^2$.

DDPM ψ . To represent the reverse process, we used a DDPM architecture originally proposed in [Saharia et al., 2021]. We chose the U-Net backbone to have channel multipliers $[1, 2, 4, 8, 8]$ in the downsampling and upsampling paths of the architecture. In this architecture, parameters are shared across time, which is specified to the network using the Transformer sinusoidal position embedding, and uses self-attention at the 16×16 feature map resolution. To condition the model on the input $\hat{\mathbf{y}}$, we concatenate the $\hat{\mathbf{y}}$ estimated by DeepSTORM along the channel dimension, which are scaled to $[0, 1]$, with $\mathbf{y}_T \sim \mathcal{N}(0, I)$. Others have experimented with more sophisticated methods of conditioning, but found that the simple concatenation yielded similar generation quality [Saharia et al., 2021].