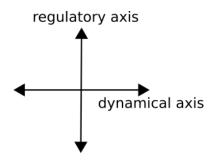
Establishing a quantitative framework for analyzing inducible gene expression in HeLa cells

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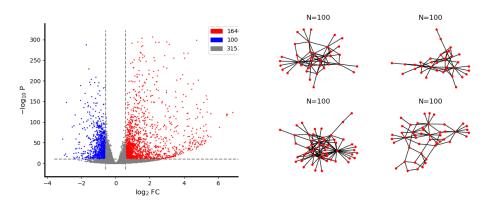
The three stages

- Select a set of genes that are differentially expressed after interferon treatment
- ▶ Mark genes which have special relevance in the literature e.g., PDL-1
- ▶ Iterative RNA FISH experiments, apply bursting models, spatial analysis



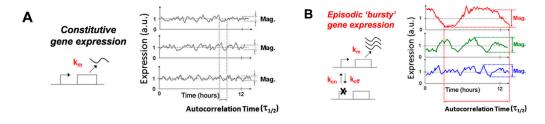
Interferon- γ induces differential expression of many genes

Single cell transcriptome measurements of polyA mRNA for naïve HeLa cells (N=90), induced with interferon gamma (50 ng/mL) for 24h



Randomly selected N=100 genes with edges drawn for $I(X;Y) \geq 0.1$ using Kraskov's method: $I(X,Y) \approx \psi(k) - \langle \psi(n_x+1) + \psi(n_y+1) \rangle + \psi(N)$

Promoter models are necessary for non-constitutive gene expression



Single-state models

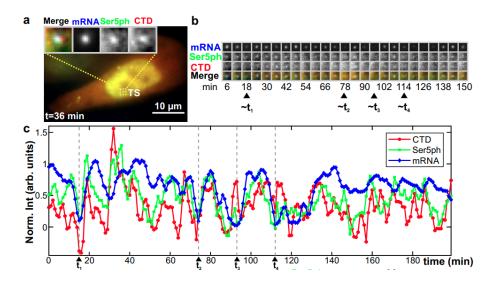
- RNAs are 'born' at a fixed rate
- RNA counts are Poisson

Multi-state models

- Promoter can be in multiple states (switching behavior)
- ► RNA counts are not Poissonian

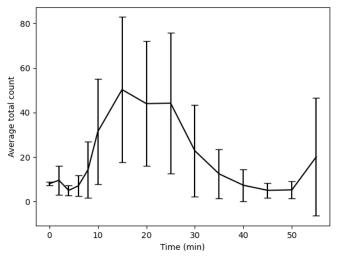
Single-state models tend to underestimate variance in RNA counts

Gene expression is stochastic (live-cell MS2-MCP)



Forero-Quintero, et al. Live-cell imaging reveals the spatiotemporal organization of endogenous RNAPII phosphorylation at a single gene. Nat Commun 2021

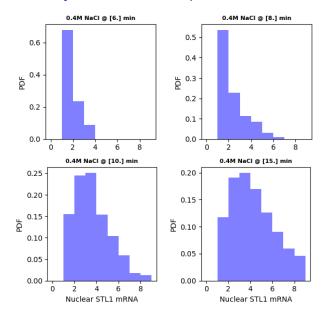
Example: variability in STL1 mRNA counts per cell at 0.4M NaCl



Error bars represent standard deviations from the mean Cells marked ON for > 3 STL1 mRNA in yeast

Assessing STL1 mRNA count variability at the transcription site

- Brightest spot in the nucleus defined as putative TS
- ► TS marked ACTIVE if 1 > 2 * med
- Nascent mRNA count is round(I/med)
- Count variability suggests asynchrony



A spatial model for induced gene expression

Let X represent an arbitrary RNA transcript of an induced gene G. Assume two promoter states (on and off)

Gene activation : $G_{off} \stackrel{k_{on}}{\rightarrow} G_{on}$

Gene inactivation : $G_{on} \stackrel{k_{off}}{\rightarrow} G_{off}$

Transcription : $G_{on} \stackrel{k_t}{\rightarrow} G_{on} + X_{\text{nuc}}$

RNA Export : $X_{\text{nuc}} \stackrel{k_{\text{exp}}}{\to} X_{\text{cyt}}$

RNA degradation : $X_{\text{cvt}} \stackrel{\gamma}{\to} \emptyset$

Raw data collected post induction can be used to infer parameters

$$\theta = (\textit{k}_{\textit{on}}, \textit{k}_{\textit{off}}, \textit{k}_{\textit{t}}, \textit{k}_{\textit{exp}}, \gamma)$$

Bayesian inference of model parameters

It is well-known that using just means and variances gives poor estimates of the model parameters (Munsky et al. PNAS 2018)

Let $\theta = (k_{on}, k_{off}, k_t, k_{exp}, \gamma)$. Using Bayes Rule:

$$P(\theta|X) = \frac{P(X|\theta)P(\theta)}{\int P(X|\theta)P(\theta)} \propto P(X|\theta)P(\theta)$$

Can infer θ if we know the likelihood $P(X|\theta)$ (the hard part) and specify a prior $P(\theta)$

Generally we have to resort to Monte Carlo methods to find $P(X|\theta)$

Kolmogorov's forward equation (chemical master equation)

To calculate the likelihood $P(X|\theta)$ one has to address the forward Kolmogorov equation

$$\frac{dP(x,t|x_0)}{dt} = \sum_k T_k(x-\nu_k)P(x-\nu_k,t) - T_k(x)P(x,t)$$

It is possible to find $P(x, t|x_0)$ in two main ways: (1) Finite state projection (2) Monte Carlo simulation

The former is exact, the latter is an approximation (see approximate Bayesian computation)