

# Galaxy Workflow ' gatk-variant-call-se-dataset-collection'

Step	Annotation
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Step 1: Input dataset collection	
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<b>Single end reads in dataset collection</b> <i>select at runtime</i>	
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Step 2: Input dataset	
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<b>input</b> <i>select at runtime</i>	
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Step 3: FASTQ Groomer	
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<b>File to groom</b> Output dataset 'output' from step 1  <b>Input FASTQ quality scores type</b> Sanger & Illumina 1.8+  <b>Advanced Options</b> Hide Advanced Options	
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Step 4: Bowtie2	
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<b>Is this single or paired library</b> Single-end  <b>FASTQ file</b> <i>select at runtime</i>  <b>Write unaligned reads (in fastq format) to separate file(s)</b> False  <b>Write aligned reads (in fastq format) to separate file(s)</b> False  <b>Will you select a reference genome from your history or use a built-in index?</b> Use a genome from the history and build index  <b>Select reference genome</b> Output dataset 'output' from step 2  <b>Set read groups information?</b> Set read groups (SAM/BAM specification)  <b>Auto-assign</b> False  <b>Read group identifier (ID)</b> gid  <b>Auto-assign</b> False  <b>Read group sample name (SM)</b> gsn  <b>Platform/technology used to produce the reads (PL)</b>	
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ILLUMINA

**Auto-assign**

False

**Library name (LB)**

Empty.

**Sequencing center that produced the read (CN)**

Empty.

**Description (DS)**

Empty.

**Date that run was produced (DT)**

Empty.

**Flow order (FO)**

Empty.

**The array of nucleotide bases that correspond to the key sequence of each read (KS)**

Empty.

**Programs used for processing the read group (PG)**

Empty.

**Predicted median insert size (PI)**

Not available.

**Platform unit (PU)**

Empty.

**Select analysis mode**

1: Default setting only

**Do you want to use presets?**

No, just use defaults

**Save the bowtie2 mapping statistics to the history**

False

#### Step 5: Realigner Target Creator

**Choose the source for the reference list**

History

**BAM file**

Output dataset 'output' from step 4

**Using reference file**

Output dataset 'output' from step 2

**Known Variants**

**Basic or Advanced GATK options**

Basic

**Basic or Advanced Analysis options**

Basic

#### Step 6: Indel Realigner

**Choose the source for the reference list**

History

**BAM file**

Output dataset 'output' from step 4

**Using reference file**

Output dataset 'output' from step 2

**Restrict realignment to provided intervals**

Output dataset 'output\_interval' from step 5

**Known Variants****LOD threshold above which the realigner will proceed to realign**

5.0

**Use only known indels provided as RODs**

False

**Basic or Advanced GATK options**

Basic

**Basic or Advanced Analysis options**

Basic

**Step 7: Unified Genotyper****Choose the source for the reference list**

History

**BAM files****BAM file 1****BAM file**

Output dataset 'output\_bam' from step 6

**Using reference file**

Output dataset 'output' from step 2

**Provide a dbSNP Reference-Ordered Data (ROD) file**

Don't set dbSNP

**Genotype likelihoods calculation model to employ**

BOTH

**The minimum phred-scaled confidence threshold at which variants not at 'trigger' track sites should be called**

30.0

**The minimum phred-scaled confidence threshold at which variants not at 'trigger' track sites should be emitted (and filtered if less than the calling threshold)**

30.0

**Basic or Advanced GATK options**

Basic

**Basic or Advanced Analysis options**

Basic