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**Software implementation of the quality
assurance tool for magnetic resonance imaging
distortion assessment**

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1 Introduction

1.1 Preface

Medicine has always been a scientific area where the crossing of different fields of research accounted for breakthroughs leading to a better understanding and, consequently, improved therapeutical methods. The success of modern approaches and the development future techniques owes to the increased interdisciplinary research and work done by both medical personnel such as doctors and nurses, but also scientists like biologists, chemists, physicists and engineers. As the knowledge about our own human body grows, the problems that we face are becoming more and more complex and need the interdisciplinary expertise. Even though some therapeutical questions appear easy to answer, because they are easily understood on a general level, the actual treatment of a real patient is something entirely different. Coming up with a treatment plan becomes exceedingly complex as we try to improve its precision and go towards targeted therapies which are tailor fit to the needs of an individual patient. Improving their quality of life and chance of survival has always come with increased costs and effort as trade-off. To make the best treatments available for everyone and, in the long run, also reduce the cost of used resources: This work shall be a small contribution to this development.

1.2 Photon - matter interactions

As light (visible and invisible wavelengths) passes through matter, its intensity decreases. This phenomenon is due to photons interacting with electrons, nuclei and their electric fields. All processes either change the direction they travel in, alter their energy, or result in the disappearance of single photons. The probability of these interactions differ for each material (dependent on its density; proton number Z) and photon energy ($h\nu$).

If a photon's energy exceeds the binding energy of an orbital electron, the photoelectric interaction can occur. Also known as 'photo effect', it describes a photon being completely absorbed by a tightly bound orbital electron which then is ejected from its atom. The now free electron is called 'photo-electron'. Its kinetic energy is the difference of the photon's energy and the electron's binding energy:

$$E_{kin} = h\nu - E_{binding} \quad (1.1)$$

Instead of being absorbed, photons might also just 'bounce off' electrons or entire atoms, transferring momentum and, in some cases, part of their energy to the particle they collide with. Rayleigh (coherent) scattering happens when a photon interacts with a tightly bound orbital electron (transferring momentum to the entire atom). This event can be seen as elastic, because only a negligible part of the photon's energy is transferred.

The Compton effect (incoherent scattering) involves a essentially free electron, such as an orbital electron with a relatively small binding energy compared to the photon's energy. Due to the weak binding, momentum is transferred only to the electron. This 'recoil electron' (or 'Compton electron') leaves its atom with a significant kinetic energy, which originated from the scattered photon. Since the photon loses part of its energy, the event is considered inelastic.

When a photon with an energy above 1.022 MeV passes through the electric field of a nucleus, it might disappear to create an electron-positron pair. This effect is called pair production. The threshold of 1.022 MeV equals exactly the rest mass $E_m = 2m_e c^2$ for the two equally heavy particles (electron and positron). The new particles travel in

opposite directions with the same kinetic energy:

$$E_{kin} = \frac{h\nu - 1.022 \text{ MeV}}{2} \quad (1.2)$$

A photon with energy of the order of 2 MeV or higher can also interact directly with the nucleus. Such a Photonuclear reaction is similar to the photo effect, in the sense that the photon is completely absorbed. Its energy is transferred to the nucleus resulting in the emission of either a proton or neutron.

Attenuation

The aforementioned interactions result in a gradual decrease of radiation intensity as it travels through matter. The combined effect is described by Beer's law:

$$I(x) = I_0 e^{-\mu(h\nu, Z)x} \quad (1.3)$$

where x is the thickness of a homogeneous material and μ its linear attenuation coefficient. The different probabilities for the interactions to occur is implicitly considered by the attenuation coefficient $\mu(h\nu, Z)$ (see Figure 1.1).

For a photon being transmitted through matter with varying properties, the attenuation coefficient changes, too. After travelling a distance d , the intensity can be expressed as:

$$(1.4)$$

Where $\mu(x)$ describes the attenuation at every distance x . (For whole chapter 1.2: [42, 28])

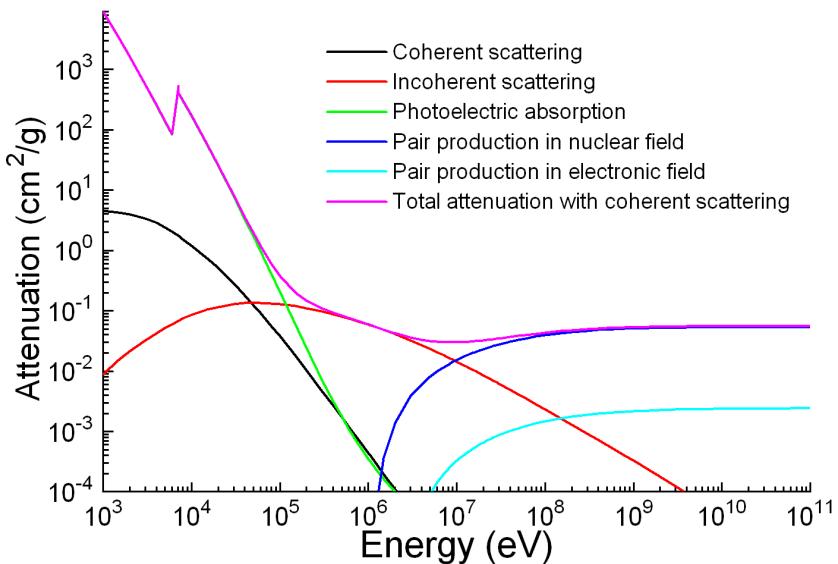


Figure 1.1: Photon attenuation for iron; [Materialscientist]

1.3 Basics of Radiobiology

1.3.1 The human cell

All higher organisms consist of cells working together to form what is called tissue. A collection of tissues which perform one or more functions is considered an organ.

Even though different types of cells exhibit distinctive traits which set them apart, all of them originated from the same totipotent zygote containing a original set of DNA. A zygote is a stem cell, it has the ability to replicate indefinitely, passing its DNA on to the resulting daughter cells. At the same time, it can change into any type of body cell. This feature is why it is called 'totipotent'. As soon as the zygote has divided into a sufficient number of identical cells, all of them differentiate into the various human tissues. In favour of becoming more specialised, cells lose their totipotency. During the early stages of an embryo they are still capable of developing into a number of different cell types, but already restricted to their own tissue type; either nerve, skin, or blood & muscle tissue. As those cells further specialise, they limit their potential even more. In a fully grown human body there are still stem cells present, such as bone marrow stem cells. Other than the zygote, bone marrow can only give rise to blood cells, but not to e.g. nerve or skin cells. A blood cell itself cannot replicate, it is considered a 'mature cell'.

The whole process follows guidelines dictated by the DNA. Every cell inherited its own personal copy of the original set. Inevitably, mistakes happen during its replication resulting in changes to the DNA called 'mutation'. Most of these alterations are repaired or do not lead to changes in the cells behaviour.

The time needed for a reparation process to be completed is not the same as the period between cell reproduction. Changes to the DNA (e.g. mutations) occurring less frequently than one reparation cycle are less probable to result in permanent alterations, than those taking place more frequently. This is the reason why cells in tissues and organs that divide more frequently (i.e. gonads) are more prone to cancer development than those reproducing more slowly (i.e. bones).

As the human body ages, the repair mechanism loses some of its efficiency and mutations accumulate. At one point, a cell may be reprogrammed to act in a unpredictable way, giving up its duties and duplicating without restraint, possibly forming tumours. Also, external factors are known to influence cell behaviour and induce such 'malign' cells (carcinogenesis). Cancer cells usually replicate more frequent than healthy cells, eventually leading to characteristic symptoms.

Different approaches have been developed to treat cancer, not all of which are suited to tackle every type of tumour. If the tumour's location is unknown or metastases have formed already in many places, chemotherapy might be considered. An easily accessible tumour could be removed in a surgery. Non invasive therapies also include radiotherapy, destroying cancer cells using radiation. (see chapter 1.3.2.)

Generally, early treatments have high chances of success, but tumours are often not noticed until they reached a certain stage. Reliable ways of diagnosing tumours are made possible by imaging techniques visualising the interior of the human body.
[**Basic Clinical Radiobiology (4th edition)**, 4]

1.3.2 Effects of radiation

Cell damage could either be caused by radiation interacting directly with the critical target in a cell or with other molecules and atoms within the cell. For X-rays, two thirds of the biological damage is attributed to indirect action. As described in 1.2, radiation transfers some of its energy to the medium it passes through. Most interactions, such as the photo effect, incoherent (Compton) scattering and pair production, result in free electrons. If the electron has a sufficiently high kinetic energy, it may free additional orbital electrons from atoms in its vicinity. The remaining ions are positively charged,

with a single unpaired valence electron. This type of chemical and free electrons are both referred to as 'radicals' and considered extremely reactive. As the human body consists mainly of water, the radicals created are often H_2O^+ (water ion) and $OH\bullet$ (hydroxyl radical). They are likely to take part in chemical events leading to the breakage of chemical bonds. Such processes can induce changes in DNA sequences and eventually produce biological damage.

A irradiated cell can be affected in various ways ranging from no effect to immediate cell death. The cell might also survive containing a minor mutation. A more fundamental mutation might lead to carcinogenesis. Irradiated cells might also send signals to their neighbours, inducing genetic damage known as 'bystander effects'. On the other hand, surviving cells can also react to irradiation and becoming more resistant.

One classification separates cell damage into lethal, sublethal or potentially lethal. Sublethal damage can be repaired, provided it occurs only once before the repair cycle is complete. Potentially lethal damage can be manipulated by repair, provided the cell is allowed to remain in a non-dividing state.

The relative biological effectiveness (RBE), which describes the damage done by a specific type of radiation (compared to a reference test radiation) to a certain type of tissue, is dependent on various factors, including the rate at which dose is delivered. In the case of radiation causing sublethal damage, for instance, the dose rate significantly affects the RBE. If the average time between two sublethal damages in a single cell is longer than the time necessary for a full repair cycle, the cell will have a fair chance of survival. Does it sustain damage more frequently, the cell will die with a much higher probability. Increasing the radiation rate above this threshold results in a jump of the RBE.

Raising the RBE does not automatically correspond in better treatment. Only if a differential effect reduces the RBE for healthy tissue compared to the tumour, there is a therapeutic advantage. Fortunately, tissues react differently to the same type of radiation. This behaviour can be used to increase the RBE for tumours and reduce it for healthy tissue.

On a larger scale, the sum of damages done to individual cells gives rise to characteristic symptoms. Generally, these harmful effects by radiation are defined as either

stochastic or deterministic. The probability of stochastic effects is directly proportional to the dose, but their severity in affected individuals is not. These effects arise in single cells (e.g. carcinogenesis), and it is assumed that the probability for such an occurrence is always greater than zero, even for small doses. If many cells show mutations, the probability of cancer development is higher, but the symptoms of growing tumours will not be worse. For deterministic effects, on the other hand, the severity scales with the dose. They are connected to tissue reaction caused by damage to a population of cells. The higher the dose, the more cells die, the graver are the biological consequences.

Changes to the DNA might not become apparent ever, others take years until they result in biological effects. The same goes for tissue effects, which could either be acute (soon after exposure) or delayed (chronic). A well known long term consequence of ionising radiation is leukaemia. Damage to germ cells (sperm/egg) might even result in genetic damages expressed in subsequent generations.

While imaging modalities utilising X-rays are designed to apply a dose as little as possible to keep effects of irradiation low, radiotherapy makes use of the lethal effects targeting cancer cells. [42, 28]

1.4 Imaging modalities

The imaging of human body's interior has diametrically changed medicine. It has its use in almost all medical disciplines. Currently, there are many ways to acquire section images (or also volumes) of our organism without causing serious and sustained side effects. They differ not only in size of the depicted volume and the image quality, but also in what additional information they provide besides purely morphologic data. These other features can be functional, for example describing effectiveness of a metabolic process; or even molecular, revealing pathways of a certain molecule's distribution. In the next chapters, X-ray planar imaging and the imaging modalities used for this thesis (computer tomography and magnetic resonance imaging) will be explained in more detail.

1.4.1 X-ray projection imaging

A widely used imaging technique based on photon interactions is X-ray projection. Its setup is made up by a radiation source, the object of interest and a detector. Since the technique is about projection, a patient needs to be placed between an X-ray tube and

the detector (usually a film-cassette or digital sensor). In the first stage of the imaging process, X-ray photons emitted by the tube enter the body. Next, while travelling through human tissue, they interact with its atoms in various ways as described above (see 1.2). These processes govern how much radiation is absorbed or scattered. Finally, photons which make it through the patient are recorded as they reach the detector on the opposite side. This results in a negative greyscale image, where brightness values correspond to the intensity reduction. Low intensity (= high absorption) leads to bright spots on the image and vice-versa. The whole process could also be described as 'the projection of attenuation shadows onto the detector', since the radiation absorption directly depends on the attenuation coefficient. The attenuation, on the other hand, depend on the tissue's properties (e.g. atomic number Z, density, etc). Consequently, the attenuation shadows depict a projection of the patient.

Soft tissue contrast

Soft tissue such as brain matter and muscles absorb only little radiation, casting a lighter shadow (dark areas on image) than bone which absorbs more photons (bright areas). Practically, in the human body, anything other than bone differs only slightly in attenuation, owing to the relatively small difference in atomic numbers and density. For this reason, X-ray projection imaging is considered reliable when it comes to diagnose bone fractures, while at the same time, it is not suited to clearly delineate soft tissue structures. The use of contrast agents, which effectively increase the density (atomic number) of certain structures or fluids, can help tackle this shortcoming. Such substances fill e.g. the bloodstream with heavier atoms, which can be clearly seen against the dark background of surrounding soft-tissue. In CT angiography, for instance, iodine is administered intravenously enhancing vessel to vessel-wall contrast. In studies of the abdomen a diluted iodine solution or barium compounds swallowed by the patient leads to improved visibility of the gastrointestinal tract. For some examinations, the patient inhales a contrast agent.

For patients allergic to those chemicals, a number of alternative agents have been developed. Unfortunately, most introduce minor, sometimes serious side-effects. There is ongoing research to find materials yielding enhanced contrast while at the same time minimising adverse reactions, a promising candidate being gold nano particles. [42, 28]

Other imaging modalities are potentially better suited for soft tissue imaging, like medical ultrasound and Magnetic Resonance Imaging (MRI), to name a few. They

are preferred for non invasive soft tissue examinations. The choice of suitable imaging modality depends a lot on the particular diagnostic needs and capabilities. It is for the clinician to decide how detailed the information needs to be and how fast it has to be provided. Often less accurate and/or cheaper methods are used first and, if necessary, followed by more sophisticated ones.

1.4.2 Computer Tomography - CT

Computer Tomography (CT) is a three-dimensional (3-D) imaging modality based on the measurement of X-ray planar projections. The technique has evolved from 2-D X-ray scanning. By mounting source and detector on a rotary ring with a patient at the centre, projections from any angle can be obtained. However, in contrast to 2-D projection methods, the detector resembles an arc made up by several hundreds of neighbouring detector elements. A single 'image' taken by the detector is therefore only in 1-D. Yet, by repeating this process from a sufficient number of different angles and along the entire patient (z-axis) a 3-D model can be computed (based on 'Radon transformation'). In contrast to 2-D methods, where the patients interior is projected/compressed onto a flat image, CT preserves the exact location information. This feature led to a radical improvement in diagnostics.

Since its clinical introduction in 1971 by Godfrey Hounsfield, CT has become a widely used 3-D imaging modality for a range of applications including radiation oncology. Especially in radiation therapy, knowledge of the exact geometry is crucial, which is why CT plays such a pivotal role in treatment planning (see 1.5). [42, 28]

3-D image reconstruction

As a photon passes through the patient, it encounters different materials associated with characteristic linear attenuation coefficients. It is practical to think of the scanned body as a collection of $N = N_X \cdot N_Y \cdot N_Z$ finite size cubes (Δx cube length) called 'voxels' (analogous to pixels in a 2-D digital photograph). The entire model can then be regarded as a 3-D matrix, with the attenuation coefficients μ_i of the voxels as its entries. Figure 1.2 represents a (4, 4, 1) matrix. It depicts the path an X-ray may follow passing through voxels with different values μ_i . This discretisation allows us to change equation 1.4 to:

$$I(x) = I_0 e^{-\sum_{i=1}^{N_X} \mu_i \Delta x} \quad (1.5)$$

The initial and final intensities can be read off the settings of the X-ray tube and the detected signal. Based on these values, image reconstruction algorithms derive the three-dimensional linear attenuation coefficient matrix. For convenience, the computed numbers are converted to Hounsfield Units which are displayed in the final image. [42, 28]

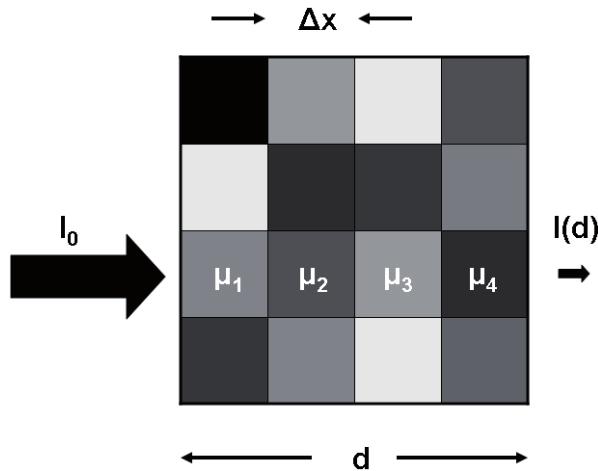


Figure 1.2: Simplified attenuation matrix (4,4,1); [28]

Hounsfield Units

In a final CT scan, voxel values are recorded in Hounsfield Units (HU), which relate to the attenuation of water at room temperature:

$$HU_{material} = \frac{\mu_{material} - \mu_{water}}{\mu_{water}} \cdot 1000 \quad (1.6)$$

Table 1.1 lists types of human tissue and their values on the HU scale. Generally, HU values range from -1024 to +3071 (12 bit), but the upper limit can be extended to 15,359 (14 bit) if materials with even higher attenuation need to be visualised (e.g. implants).

Table 1.1: Average HU values for various types of human tissue

Substance	HU
Air	-1000
Lung	-750 (-950 to -600)
Fat	-90 (-100 to -80)
Water	0
Muscle	+25 (+10 to +40)
Brain, white matter	+25 (+20 to +30)
Kidneys	+30 (+20 to +40)
Brain, grey matter	+35 (+30 to +40)
Blood	+55 (+50 to +60)
Liver	+60 (+50 to +70)
Compact bone	+1000 (+300 to +2500)

Typically, CT scans are displayed on Computer monitors, which imposes the need to map the HU values to a 8-bit greyscale (256 steps of luminosity). Since the number of possible values (dynamic range) on the HU scale is 16 times the shades of grey on a screen ($12-8 = 4$ bit difference; equivalent to a factor of 2^4) the screen cannot convey all details at the same time. A linear mapping would result in 16 neighbouring HU values being compressed to the same brightness on the screen. This way, the brightest (bone) and darkest parts (soft tissue) of the image would be clearly distinguishable. At the same time small differences (<16 HU) would appear to have exactly the same intensity. However, most of the time, the doctor's focus might lie either on soft tissue or bone material. Bearing in mind that soft tissue values range only from 10 HU to 70 HU at most (see table 1.1), such a compression would make distinguishing tissues using CT very unreliable. Instead of showing detail from the lowest to the highest value, a range of values - a so called window - can be chosen. Let's assume, for example, a range from -100 to 155 HU to be of interest. This selected range can be mapped directly and uncompressed to a 8-bit greyscale. Any values above 155 HU will be assigned the brightest value (white = 255), below -100 the darkest (black = 0). While showing very good soft tissue contrast, all bones would be depicted with exactly the same brightness (255), even though they might have a varying HU values. For bone structures, a range from 300 to 2500 HU might show sufficient contrast. Standard computer programs used to display CT images allow the user to change the window interactively to any value range. [42, 28]

Image acquisition

The time necessary to collect 1-D attenuation projections from sufficient angles is called 'acquisition time'. In 2-D X-ray scanning only one picture is taken, while a 3-D CT model is made up of a photo sequence. If the patient moves during the imaging process, the final model would show motion artefacts which might lead to wrong conclusions. Consequently, CT scanners are designed to minimise acquisition time while ensuring sufficient image quality. Very fast CT protocols result in smaller resolution, because less images are taken. It has to be said, though, that CT acquisition time is usually already significantly shorter than MRI. [42, 28]

Image quality

Additionally to the relatively short acquisition time, CT scans show little distortion compared MRI (see 1.4.3), which is why they are often used as 'gold standards' (reference scans used for MRI distortion assessment).

While bone structures are clearly visible in CT scans, 'soft tissue contrast' is relatively low compared to MRI. In other words, parts of the body which are considered as 'soft tissue' (intestines, brain, blood vessels, etc.) differ little in brightness and are therefore hard to distinguish. See 1.4.1 for more information.

Another aspect of image quality is the 'low contrast resolution' of the scan. It directly relates to how much structures and their surroundings have to differ in signal intensity to be clearly distinguishable by doctors. The quality of the 'low contrast resolution' is mainly limited by noise. Noise is a random pattern underlying the actual signal and is always present to some extent. Its prominence in the final image is described by the Signal to Noise Ratio (SNR). If the SNR is too low, fine structures blend with the noise and cannot be distinguished. Strategies to achieve a high SNR include raising the initial photon flux (intensity) or employing contrast agents. The intensity is governed by the tube current, which is limited by the heat capacity of the tube and health considerations regarding the patient dose.

Alternatively, the spatial resolution can be decreased, effectively combining neighbouring image slices. This way the SNR for the combined slices is increased, but fine structures along the z-axis might be lost due to the reduced resolution. [42, 28]

Health considerations

CT scans describe the attenuation throughout a patient, which is directly related to how much energy is transferred from photons to matter. Only because X-rays are absorbed by the human body, this imaging modality gives insight in the density distribution of a body's interior. However, this transferred energy is capable of causing biological damage. (see 1.3.2)

While the radiation dose administered during a single CT scan is relatively small (typically not more than 15 mSv) and almost negligible compared to the dose administered during a potential radiation therapy, patients receiving this dose regularly end up with a potentially harmful accumulated amount of radiation. Cancer patients, for instance, need to be imaged frequently during treatment planning. However, patients might die before those consequences come into effect. Therefore, it's typically children (who received a great number of CTs) to suffer from induced cancer occurring up to 40 years later. So while the benefit from using CT for diagnostics far outweighs the damage, there have been major efforts to reduce dose while maintaining reasonable image quality. [35, 6, 57, 56, 30, 17]

1.4.3 Magnetic Resonance Imaging - MRI

Magnetic Resonance Imaging (MRI) is a 3-D imaging modality based on Nuclear Magnetic Resonance (NMR), a phenomenon discovered by physicist Isidor I. Rabi in 1938. Atomic particles such as protons have an inherent quantum mechanic feature called 'spin', which is associated with a magnetic moment μ . Without an external field, a proton's spin is oriented in a random direction in space and so is its magnetic moment. The sum of magnetic moments belonging to a number of protons results in a net magnetisation. Due to their random orientation, the net magnetisation will be zero for a sufficiently high number of particles. This is because, on average, for every proton's spin there is always another particle's spin oriented exactly the opposite way cancelling its magnetic moment.

In the case of an applied external magnetic field (this static field is often called B_0), the spins will either align parallel (pointing in the same direction) or anti-parallel (opposite direction) to this field, where their energy reaches a local minimum. Parallel protons have an even lower energy than those pointing the other way. In a collection of many spins, the number of parallel spins will therefore slightly dominate, resulting in a net

magnetisation greater than zero (see figure 1.3). In other words, only the amount of protons that is not compensated by those looking in the opposite direction contributes to a detectable magnetic field.

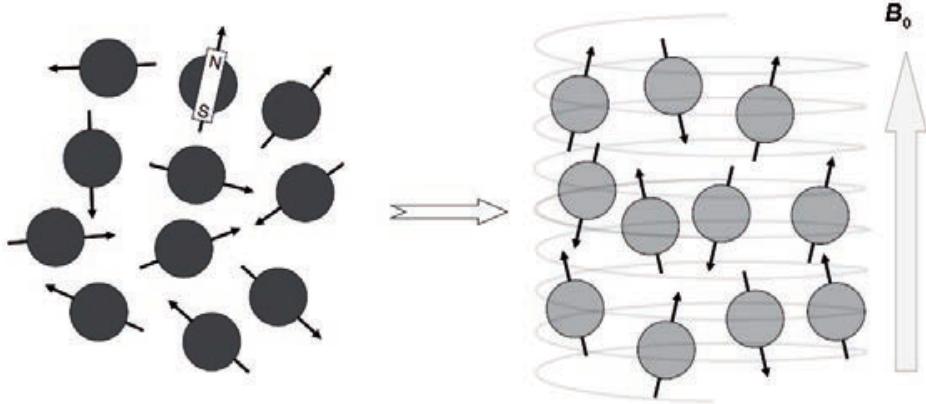


Figure 1.3: The spins, initially oriented randomly in space, become aligned either parallel or antiparallel to an externally applied magnetic field B_0 . [28]

By applying a short radio frequency pulse (often referred to as B1), the total external magnetic field changes and the magnetic moments start precessing around that new external field. The pulse duration is usually chosen as to flip the spins by a 90° angle. They are now oriented in the transverse plane to the original external field, and so is the resulting net magnetisation. Similar to how a spinning top rotating at an angle to the direction of gravity precesses, the magnetic moments will now precess about the direction of the external field with a frequency linearly proportional to the external field strength. This precession movement can be detected as induced current in a receiver coil, because the net magnetisation still follows the spins' orientation. Again, the particles would like to minimise their energy by aligning their spins to the external field, but in order to do so they need to give away the surplus energy, transferring it to the surrounding lattice. Those spin-lattice interactions happen with different efficiency depending on the tissue. The time it takes the spins to align is expressed in a material specific time constant T_1 . Shortly after applying the radio frequency pulse, regions of the body where magnetic moments align quickly (short T_1) have a stronger net magnetisation (in the direction of the external field) than those where energy is being transferred slowly (long T_1).

At the same time, the spins interact with each other, affecting the local magnetic field and spins in their vicinity. The magnetic moments, which started out precessing in phase directly after the radio frequency pulse flipped them, will precess at slightly different

frequencies, due to the small fluctuations of the local magnetic field. The differences cause the collection of magnetic moments to 'de-phase' and the net magnetisation in the transverse plane to vanish. This process caused by spin-spin interactions is described by the material specific time constant T_2 . Eventually, all spins will be again aligned either parallel or anti-parallel to the external field, just as they were before the RF-pulse.

Applying the RF pulse with a homogeneous field strength along the whole body would excite all spins simultaneously. In order to localise differences in tissue magnetisation, the RF pulse is instead combined with a linear magnetic gradient field 'selecting' a slice to be imaged at a time. The rest of the body is unaffected, the signal measured directly after such a pulse was applied originates only from the chosen slice. Eventually, by collecting data on the net magnetisation throughout the body in different locations ('scanning' the patient slice by slice), a 3-D image can be computed.

Depending on the information required from the examination, Doctors can choose to create images that reflect spin-lattice interactions (T_1 weighted) or spin-spin interactions (T_2 weighted).

Governed by the chosen settings, particular tissues will display varying contrasts. For example, areas with increased water level will be dark on T_1 weighted (T_{1w}) MRI whereas the same areas will be bright on T_{2w} MRI. Additionally, certain types of materials (tissues) can be intentionally not imaged (suppressed) to reveal others that are in their close proximity. This is often done with fatty tissue that can cover relevant parts of the field of view. There are various methods how this can be achieved. MR imaging gives practically endless possibilities in terms of selective imaging and is mostly limited to imaging time and technical characteristics of the scanner. Medical physicists and scanner vendors are incessantly working on new MR imaging methods and applications.

The set of parameters governing how the tissue is excited and data acquired is called 'image sequence'. Delineating tumours or lesions is often accomplished by looking at both T_{1w} and T_{2w} weighted images and drawing the right conclusions.

Soft tissue contains a lot of water, which is made up by oxygen and hydrogen. Hydrogen nuclei are single protons and their nuclear magnetic resonance is what MRI is tuned to visualise. This is why soft tissue appears as bright areas in MRI, whereas bone material has only little contrast. [11]

Most scanners are built to house a receiver coil in the gantry and they are able to measure the signal using only this one coil. However, a stronger signal could be obtained by placing the coil closer to its source, the patient. As the region of interest

(ROI) is usually limited to a specific organ, receiver coils are available in different sizes and shapes, often designed to fit the patient with a comfortable but narrow space in between. To get even closer, so called 'surface coils' can be placed on the patient. 'Spine coils' are sometimes hidden in the table on which the patient lies during the examination. Typically, for creating images of a patient's head, coils with a fixed arc-like geometry are used. This type of coil was also used for the data acquisition of this thesis.

Studies have shown that delineating certain types of tumours, for example prostate cancer, is more accurate using MR images than using CT. [47, 13, 48]

In diagnostics, MR images prove to be very useful. Also for radiation therapy treatment planning, the superior soft tissue contrast is exploited during the definition of organs at risk and targets. Unfortunately, due to the physical principles of MRI, it indirectly provides the information about the proton density, whereas CT can provide information about electron density. It is knowledge of the electron density which is necessary in the treatment planning process. During this, the dose distributions are calculated based on the applied beam geometry and the distribution of matter on its way.

Image Quality

Contrary to CT, MRI is prone to distortion due to field inhomogeneities. Organs might appear shifted, elongated or shrunk. The effect is most prominent along the outer edges of the scanner's field-of-view (FOV). In the isocentre (middle) of the scanner, the distortion is smaller, because here the field is least aberrant. For most applications, small position shifts and deformations are of minor importance. MRI scanners usually come equipped with an internal distortion correction algorithm. Figure 1.4 shows the unmodified and the corrected version of an image coming from such a scanner. Those methods are developed by the company designing the scanners. Knowing the technical details enables them to write tailor-fit scripts which drastically reduce the distortion.

However, for Radiotherapy treatment planning this might not be enough and it is necessary to additionally monitor the distortion and, if necessary, take additional corrective measures.

While its soft tissue contrast is superior to CT, a relatively long acquisition time is necessary to achieve a sufficient high SNR. This leads to the risk of motion artefacts (patients moving during the scanning procedure). To tackle this issue, resolution can be reduced, effectively combining signal from several voxels to create a single voxel,

reducing the overall noise. The trade-off is that fine structures might get lost.

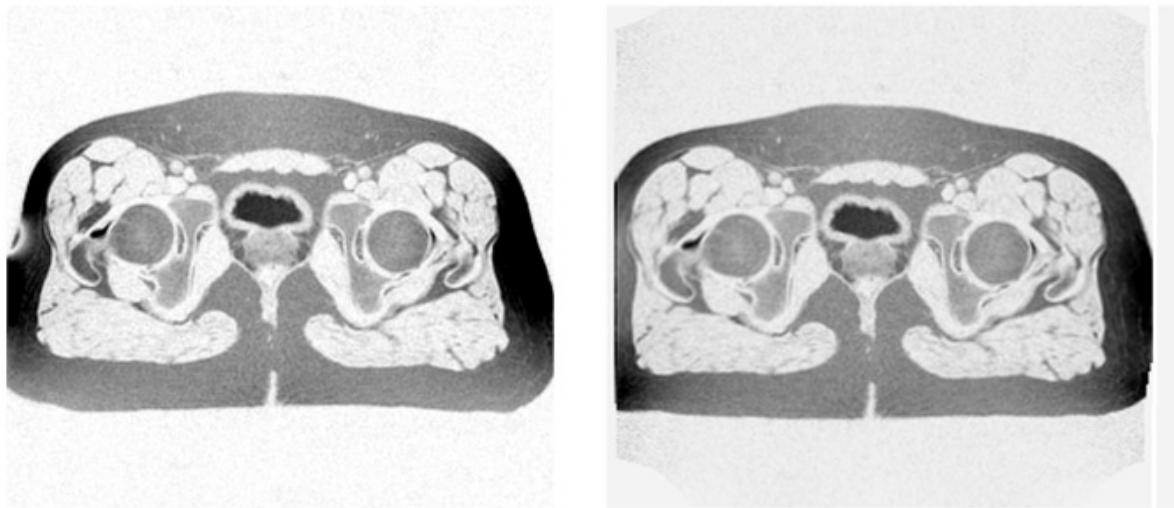


Figure 1.4: On the left is the original image, on the right the automatically corrected version of the same MR image (inverted colours)

Health considerations

Strong, static magnetic fields (typically up to $3T$) are present around MRI scanners at all times, and precautions have to be taken to ensure safety for patients and medical personnel. Ferromagnetic materials (such as steel and iron) can become dangerous projectiles in vicinity of the MRI scanner. They must be excluded from the room housing the magnet without exception.

The RF pulses repeatedly put spins in excited states, transferring energy to the human body. MRI scanners are designed to limit the rise of a patient's body temperature to $0.5^{\circ}C$ during standard imaging. Only combined with either medical or appropriate psychological monitoring the limit can be raised to $1^{\circ}C$. An ethics committee approval is necessary for even higher values. In general, patients should be exposed to RF fields only as strong as their thermoregulatory system is capable to cope with.

Finally, magnetic field gradients are applied together with the RF pulse. They are switched at high frequencies leading to induced currents in conducting body tissue. In principle, those currents stimulate nerves which might result in muscle twitching or pain.

However, gradient levels are set to avoid stimulation. During have reported some subtle biological effects, but there was no evidence pointing towards harm caused by short term exposures. At the same time, patients suffering from epilepsy might show increased sensibility to induced electric fields in the cortex and should be imaged with caution. [28] Some patients might be allergic to contrast agents used for specific MRI examinations. Here, safety procedures are similar to those performed during administration of regular drugs.

Open bore MRI scanners

The radiation oncology department of the Vienna General Hospital (AKH) is equipped with an $0.35T$ open-bore, c-arm MRI scanner (see Section 2.1.1 for more details). This open design has proven to drastically improve the well-being of patients experiencing anxiety in closed-bore scanners. For this reason, the number of incomplete MR examinations due to a claustrophobic events is relatively low. [16, 3] Besides, patients who would not fit in closed designed scanners can be imaged. Furthermore, brachytherapy patients can be placed in the scanner with applicators attached (see chapter ?? for more details).

This scanner's magnetic field is weaker than the fields of closed-bore scanners that are widely used (typically 1-3 Tesla). High field strengths result in greater resolution, better Signal to Noise (SNR) ratio and faster imaging time. Generally, diagnostics benefit from greater image quality. However, at some point, diagnostic accuracy stops increasing with field strength. At the same time significant improvements can be achieved at low fields. A “combination of field independent polarisation [...] with frequency optimized MRI detection coils [...] results in low-field MRI sensitivity approaching and even rivalling that of high-field MRI.” [9]

Low field MRI scanners are also typically characterised by less distorted images.
[\[https://link.springer.com/article/10.1007/PL00002385 \]](https://link.springer.com/article/10.1007/PL00002385)

Apart from the often satisfactory image quality, there are considerable cost advantages to the use of lower field MRI. The initial purchase price and the ongoing maintenance expenses are considerably lower than those of high field scanners, which often use superconducting magnets cooled with liquid helium. [50] Permanent magnets might be weaker, but do not require constant cooling. Also, low fields allow facilities to build smaller rooms and magnetic objects are less dangerous.

Diffusion Weighed Imaging - an example for non morphologic Imaging

Despite the fact that this type of MR imaging was not used for this thesis, it will be mentioned here as an example of the vast range of measurements possible with MRI techniques.

Diffusion weighted (DW) imaging quantifies molecular diffusion in the body. This imaging technique uses MRI technology differently: Additionally to the gradients needed to select a slice (strength $3-5\text{ mT/m}$, duration $2-4\text{ ms}$), the sequence for DW imaging applies two long, strong, consecutive and opposite gradients (strength $30-50\text{ mT/m}$, duration 20 ms) during which molecules may move due to diffusion. After those two gradients (which, usually, will be applied in all three Cartesian directions), the remaining signal (net magnetisation) is measured by the receiver coil.

Molecules which are restricted in their movement will experience two equally strong but opposite magnetic fields. The first will cause them to precess with a certain speed (linear with field strength), effectively changing their phase. The second will cause them to precess exactly the other way around (same strength, but other direction), returning them to their initial state. Now they will again be in phase and result in a net magnetisation which is not zero, but visible as bright areas on the scan.

Those molecules which are free to move, however, will not experience a constant field strength, because the field has a gradient. As they move through the body they will precess at varying speeds during the first and then at different speeds during the second gradient. As a result, they will be out of phase when the net magnetisation is measured by the receiver coil, and will not cause bright areas in the scan.

DW imaging can be used to diagnose acute strokes (brain infarct), because areas with restricted diffusion (blocked blood flow) show a strong signal compared to healthy tissue with normal diffusion. Another interpretation of low diffusion (high measured signal) can be the increased cellular density (so dense that free diffusion of water molecules is suppressed) which is characteristic for cancerous tissue.

Since the time necessary to allow the molecules to move during the two gradients is relatively long, the image will naturally be T_2 weighted. This is taken into account by creating a second image which is also T_2 weighted, but does not apply two consecutive opposite gradients. The difference between the DW and the not DW weighted sequences reflects the actual contribution of diffusion (apparent diffusion coefficient - ADC).

1.5 Radiation therapy

Radiation therapy utilizes ionizing radiation to damage and kill cancer cells in order to stop them from multiplying. This prevents the growth of tumours, makes them shrink in size and hopefully cures the patient.

During radiotherapy treatment planning (RTP), 3-D models of the patient are used to define targets (regions where the dose should go) and organs at risk (where the dose should not go). This ensures that vulnerable organs are spared from radiation while making sure the tumours receive sufficient dose. Moreover, methods to quantify the amount of radiation that different body parts absorbed are needed, because the actual treatment might differ from the plan.

While travelling through matter most types of radiation release energy mainly due to coulomb interactions with the outer shell electrons of atoms. Knowing the electron density of the targeted tissue area is therefore essential. In order to reach a specific penetration depth, the particles' initial energy has to be chosen accordingly. The necessity to treat the tumour with a required amount of radiation leads to a radiation therapy treatment plan.

There are two well established methods for applying the radiation. External beam radiotherapy (EBRT) is performed from afar: Gantryes are able to position the radiation source around the patient in a way covering virtually any possible angle. During a treatment session, fractions of the total dose are administered from many different locations. The sum of those individual treatments results in the required dose distribution. Figure 1.6 shows an example of two differently calculated treatment plans.

In conventional EBRT, photons (X-rays) in the range of 4MeV to 20MeV are used to deposit the necessary dose at the location of the tumour. Unfortunately, radiation interacts with all cells it passes until it is fully absorbed. It releases its energy along its entire path while travelling through the patient. This behaviour may result in energy being transferred to cells all the way from the point of entry to the point where the (weakened) ray leaves the patient. Other types of ionising radiation are also used, but less common. Electrons and low energy X-rays are favoured for some superficial tumour; rare methods using neutrons and even muons also exist. Charged particle therapy (using e.g. protons or carbon ions) is on the rise, but far from reaching the popularity of X-rays. This type of radiation minimises the damage done to healthy tissue due to its distinctive behaviour in energy loss called "Bragg Peak". They release most of their en-

ergy only shortly before being stopped completely. [36] This effect can be used to spare tissue lying behind the tumour from radiation entirely and also reduce the amount of energy transferred to organs located before. [39] A comparison between the behaviour of X-rays and protons is shown in figure 1.5.

Brachytherapy, on the other hand, is when the radiation source is placed close to or inside of the patient. The source is either moved close to the target area using applicators (temporary treatment) or implanted permanently. The latter method is done by inserting so called "seeds" (sealed metal containers with radioactive material) directly into the target area where they release high amounts of radiation. Over time they become less active and eventually the treatment stops automatically. These rice grain sized implants can remain in the body without causing any harm. Treatment of prostate and cervix cancer is often done with this technique. In comparison to EBRT, Brachytherapy allows higher doses while at the same time minimising the radiation reaching organs at risk; precise dose distributions can be achieved. At the same time, not all cancer types can be treated this way. For some, a non-invasive method (e.g. EBRT) is a better alternative.

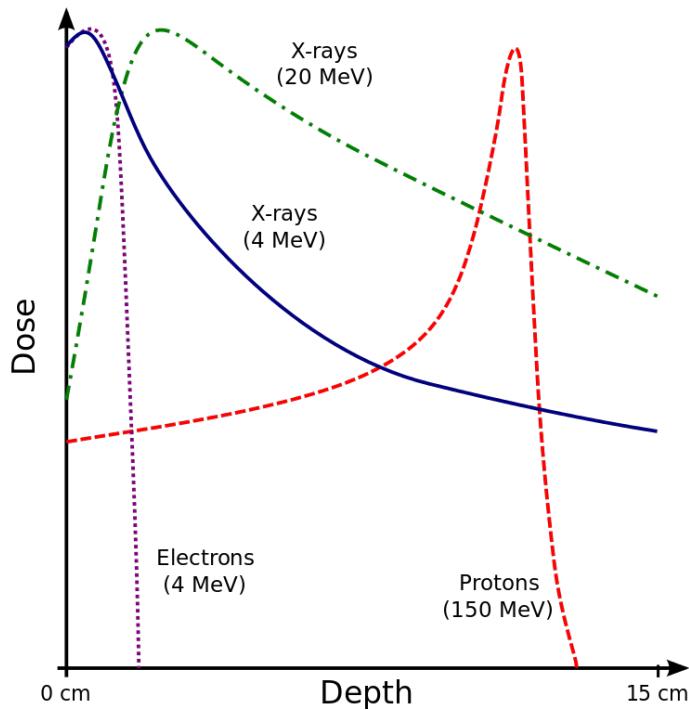


Figure 1.5: energy release of ionising radiation; [Cepheiden]

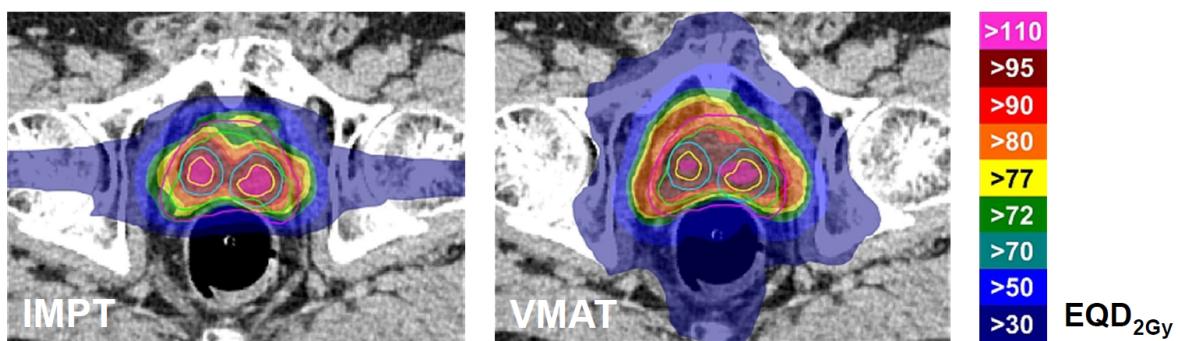


Figure 1.6: Example of Radiotherapy treatment plan: coloured areas represent dose values deposited during treatment. The plans were calculated using different planning techniques (IMPT and VMAT). (image source: [piotr])

1.5.1 Role of CT

Until recently, radiotherapy treatment planning (RTP) relied almost entirely on CT. There are two main reasons for this:

Firstly, calculating the electron density using data obtained with CT is an straightforward task. Secondly, CT images generate 3D images with little distortion. Exact geometries are needed for correct RTP. It is the most reliable approach to create precise radiotherapy treatment plans. [10, 52]

1.5.2 Role of MRI

MR images also record luminosity values, but they do not correspond to radiodensity. Due to the better visibility of tumours on MR images, RTP often uses combined data from both imaging modalities. Additional information derived from DW MRI can also support response prediction and assessment. However, there are some difficulties arising from combining CT and MRI for EBRT: In order to profit from separately acquired data, the resulting images must be aligned (registered) either manually or automatically. This is a hard task since non-rigid objects (organs) change their shape and location between measurements which may lead to inaccuracies. Algorithms supporting non-rigid registrations are already under development, but there is still room for improvement. For now, only local rigid registration is capable of reliable target and organ at risk definition.

Alternatively, MRI-only radiation therapy protocols are being developed: One way of doing this is to use MRI data to create a Pseudo-CT, which contains information about electron density. Comparisons to using CTs and MRI-based pseudo CTs have shown acceptable deviations for X-ray therapy. In charged particle therapy the resulting dose gain in healthy tissue and dose loss in cancer regions owed to inaccurately assigned electron density values is bigger. However, further improvement of accuracy promises to reduce time and money needed for RTP when CT is no longer needed. Furthermore, patients would be spared the additional dose of CT examinations. [46, 58, 37, 18, 8]

1.6 Aim of this work

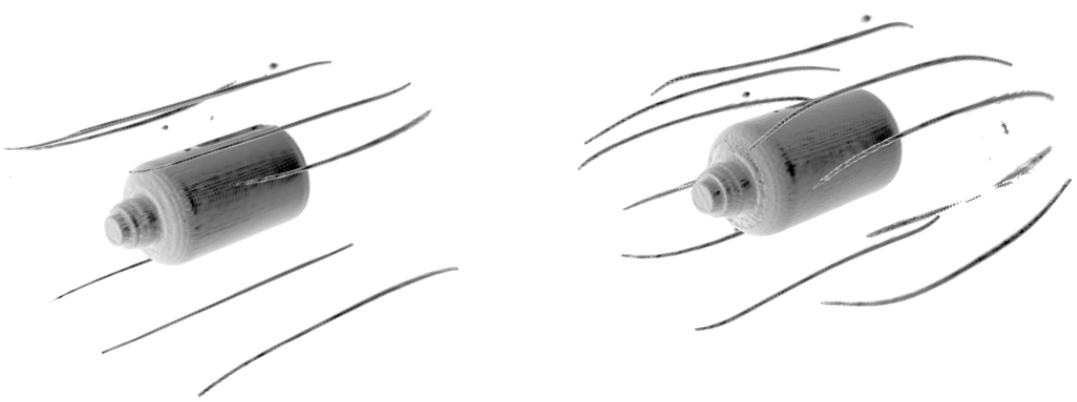
The idea of only using MRI for treatment planning is approaching the clinics, but there are still some issues that need to be addressed.

Due to the possible image distortion, great care needs to be taken and the MR images must be verified before they are used for RT target definition and dose calculation. The available MRI scanner at the AKH is equipped with an on board correction algorithm which is supposed to reduce distortion. See figure 1.7 for an example of how this correction affects an image. Distorted images might lead to wrong calculations of how much energy is needed for the radiation to accumulate exactly at the target region. If, for example, bone structure is depicted as thicker than it really is, RTP would suggest a treatment which would deposit more energy behind the tumour than intended. The opposite holds for cases where tissue appears to be thinner, which would result in areas lying before the tumour being irradiated.

The goal of this work is to: find an optimal (providing satisfactory image quality and convenience of use) liquid filling for the rod cavities in an already existing custom designed distortion phantom provided by the AKH Vienna to be able to acquire the reference CT image and test MR acquisitions.

develop and implement a method to assess and illustrate the distortion of the MR image based on tracking of the distortion of one of the phantom rods, with an option to further extend the tool for mutli-rod tracking.

Therefore, this work focuses mainly on the assessment of the acquired imaging data (using the implemented software tools), choosing which liquids to fill the phantom with, but not its entire design. However, possible fillings have to be produced and tested. Similar approaches are being used for distortion correction by other facilities. [43, 40, 59, 63, 64, 34]



(a) MR scan with no distortion correction (b) MR scan after internal distortion correction

Figure 1.7: Rendered MR scan showing the difference before (a) and after (b) applying the build-in distortion correction.

2 Material and methods

2.1 Scanners

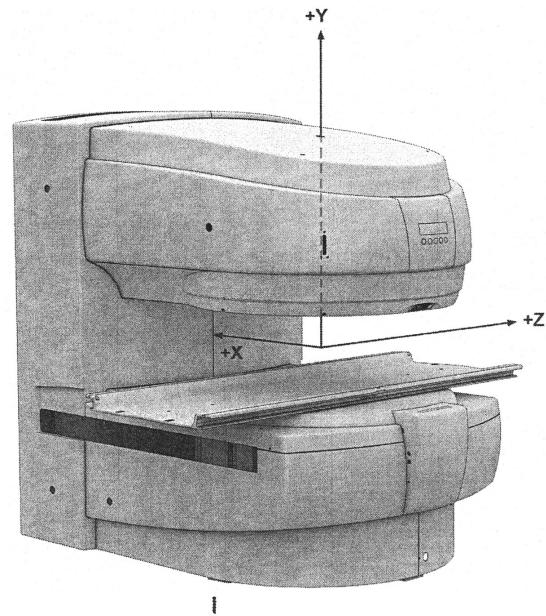
The radiation oncology department of the Vienna General Hospital (AKH) owns a open-bore, c-arm MRI scanner and a CT scanner (used as reference). They are listed in Table 2.1.

System	product name	company	coil [internal W x H]
MRI	Magnetom C!	Siemens	Body/Spine Array Coil XL [50 x 30.5 cm (19.7 x 12 in)]
CT			—

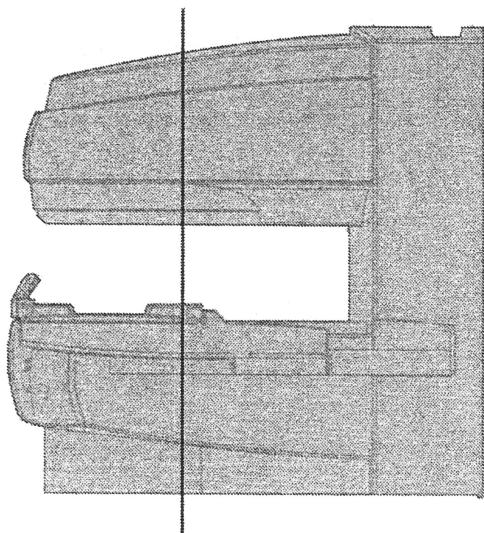
Table 2.1: used scanners

2.1.1 MRI scanner - field distribution

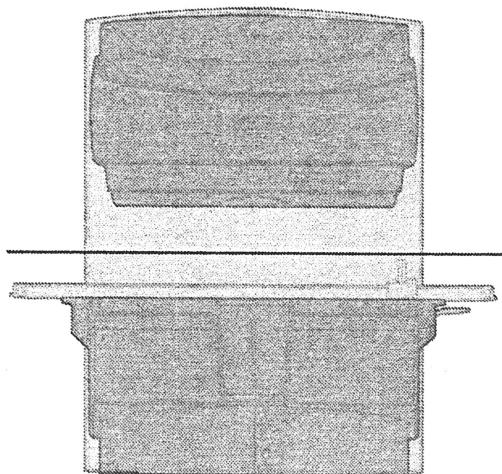
The Magnetom C! MRI scanner comes with a handbook which contains diagrams of the magnetic field strength (fig. 2.2) and its gradient (fig. 2.3, 2.5 and 2.4). Figure 2.1 shows an image and different views of the scanner and the planes along which the field strength and gradient are displayed. Patient are positioned as to place the ROI at the isocentre, the area where the B0 field is most homogeneous (where there is a low gradient).



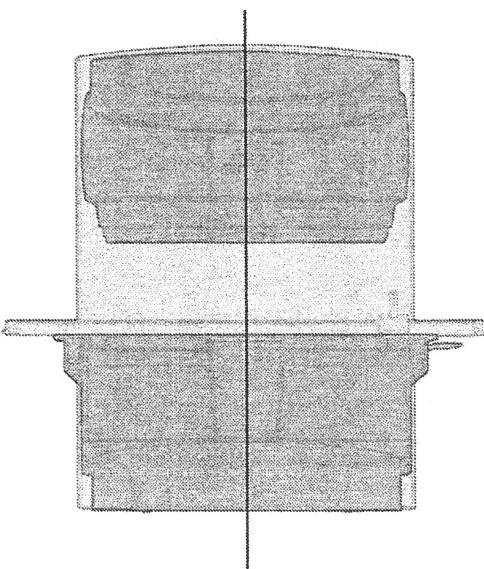
(a) Image of scanner
x,y, & z -axis



(b) side view along z-axis,
black line represents plane for
front view (see fig. 2.4)



(c) front view along x-axis,
black line represents plane for top
view (see fig. 2.5)



(d) front view along x-axis,
black line represents plane for side
view (see fig. 2.3 and 2.2)

Figure 2.1: Magnetom C! [magnetom_handbook]

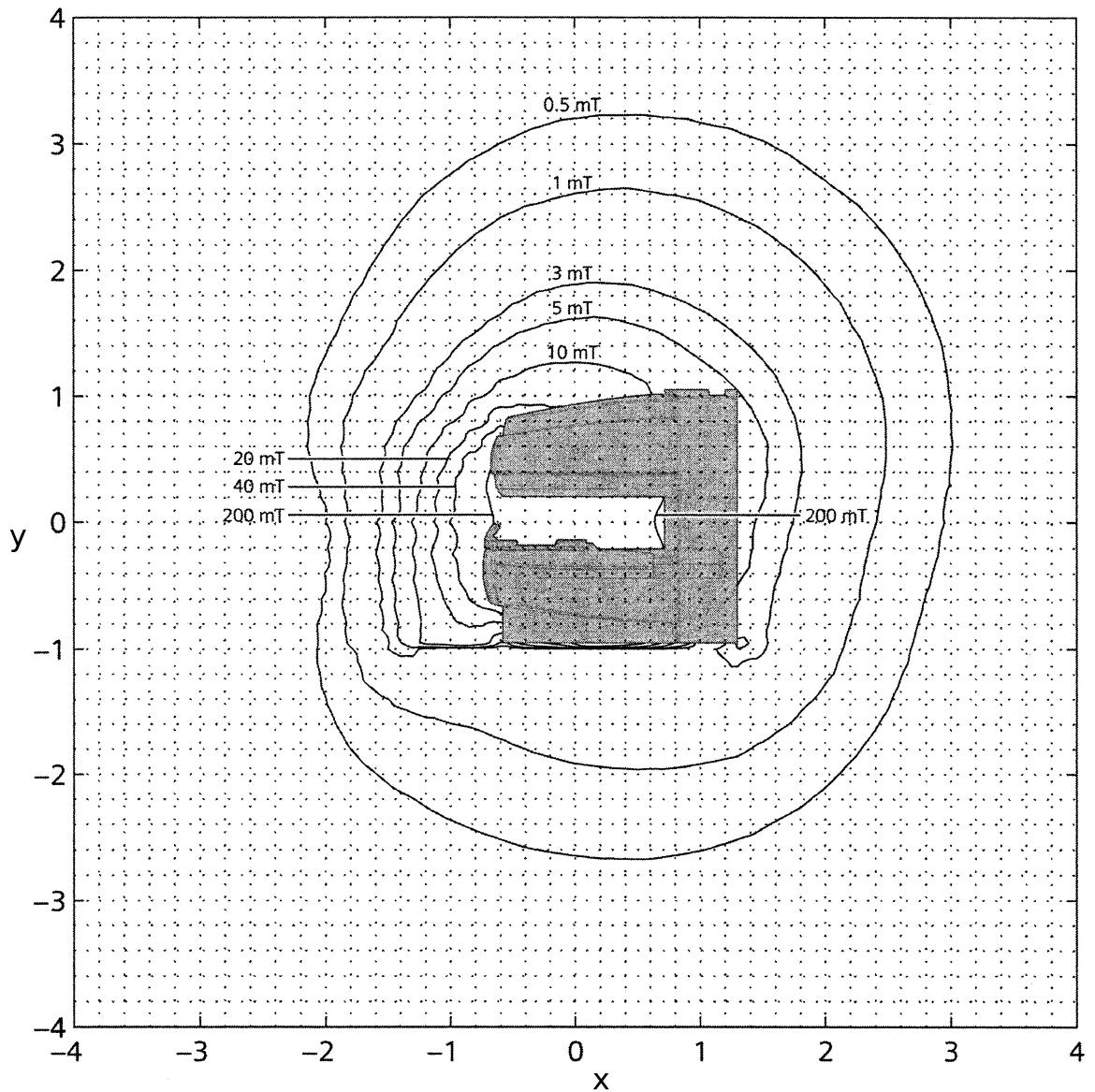


Figure 2.2: Magnetom C! field strength
side view along z-axis (see fig. 2.1d) [[magnetom_handbook](#)]

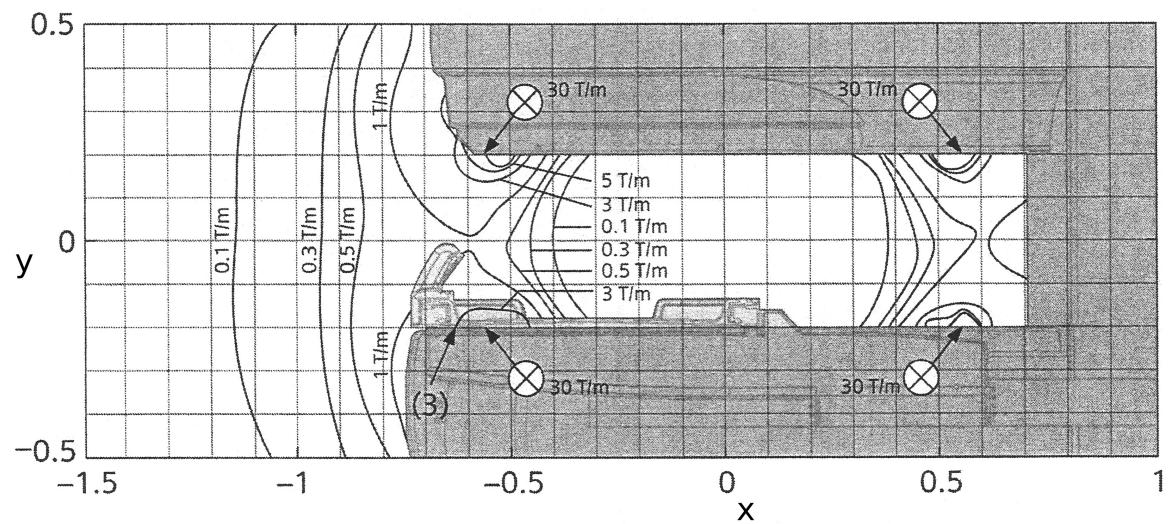


Figure 2.3: Magnetom C! field gradient
side view along z-axis (see fig. 2.1d) [[magnetom_handbook](#)]

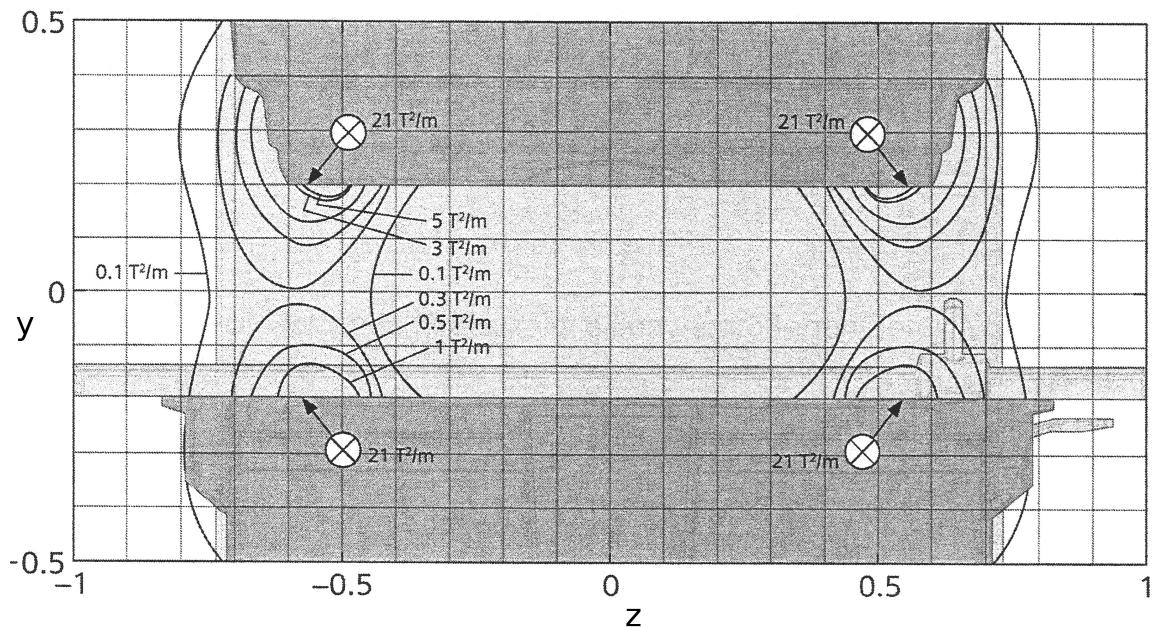


Figure 2.4: Magnetom C! field gradient
front view along x-axis (see fig. 2.1b) [[magnetom_handbook](#)]

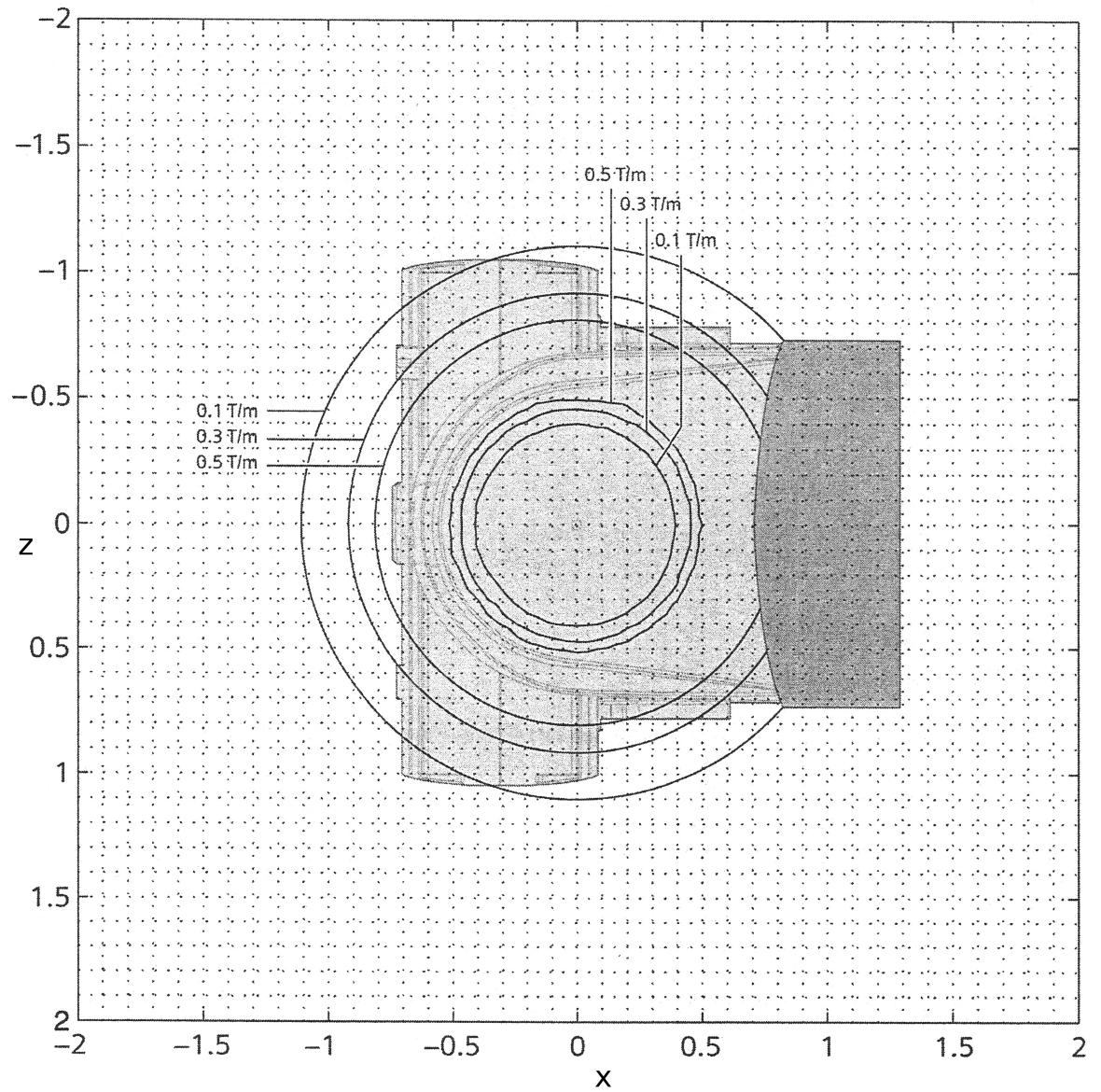


Figure 2.5: Magnetom C! field gradient
top view along y-axis (see fig. 2.1c) [[magnetom_handbook](#)]

2.2 Custom build phantom

To compare images from different scanners and asses occurring distortion, a rigid object with known dimensions is necessary. Such a 'phantom' is often made from plastics filled with liquids which are easy to handle and typically well seen on MR and CT images. The AKH's design is made up from an array of replaceable, fillable plastic rods.



Figure 2.6: Comparison (inverted colours): MRI only shows liquid filling, CT also the plastic rod and pane (horizontal black bar crossing middle and right rod);
(a:) *MRI* - filled rod, plastic not visible (field of view too small to show entire rod); (b:) *CT* - empty rod, plastic visible; (c:) *CT* - filled rod, plastic and filling visible

2.2.1 Frame and rods

The phantom was build to fit the largest available rigid coil for the MRI scanner. Three parallel acrylic glass panes in the shape of the coil serve as a frame for the plastic rods. In the middle an empty area was reserved for an optional additional smaller phantom (not used for this work). Figure 2.7 shows a rendered CT picture of the phantom. See also figure 2.8 showing a CT image of one pane (with no rods inserted).

More than 300 plastic rods (length: 50cm, outer diameter: 8mm, inner diameter: 4mm, volume: approx. 6ml) could be placed in the phantom. See figure 2.9 for a schematic sketch of one rod. The bottom part of each rod was sealed with a glued plastic plug,

the top could be closed with a plastic screw. Frame and rods were already build and assembled before the author started working on this project.

Additionally to the rods which would be used to assess the distortion, a number of vitamin-e pills were attached to the frame as reference markers (see figure 2.10). These pills are visible in both CT and MRI images and were used to align them (see section 2.4). This way there is another way of checking the alignment during the prototyping process in addition to the rods.

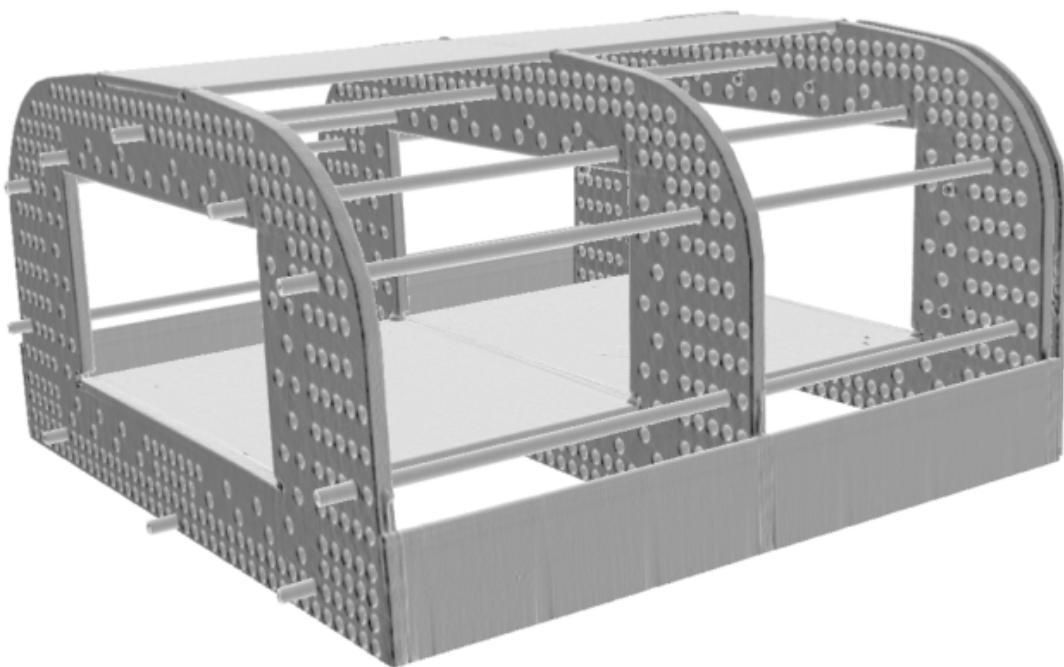


Figure 2.7: Rendered CT image of phantom



Figure 2.8: An axial view of one of three plastic panes that make up the frame of the phantom. This scan shows how the pane looks like with no rods inserted.

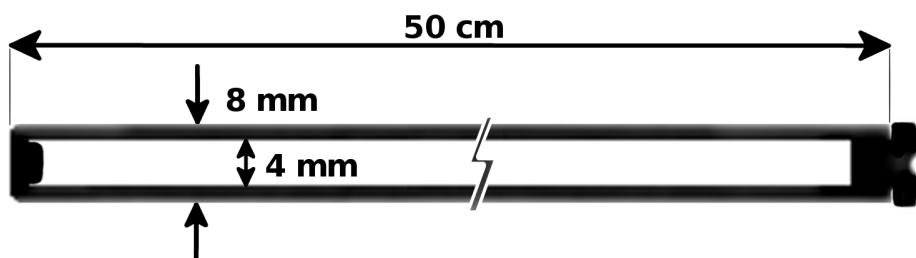


Figure 2.9: A schematic of an empty plastic rod as they are used for the phantom (inverted colours). On the left the rod ends with a glued plastic stopper, on the right hand side a plastic screw seals it. The figure does not show true proportions.

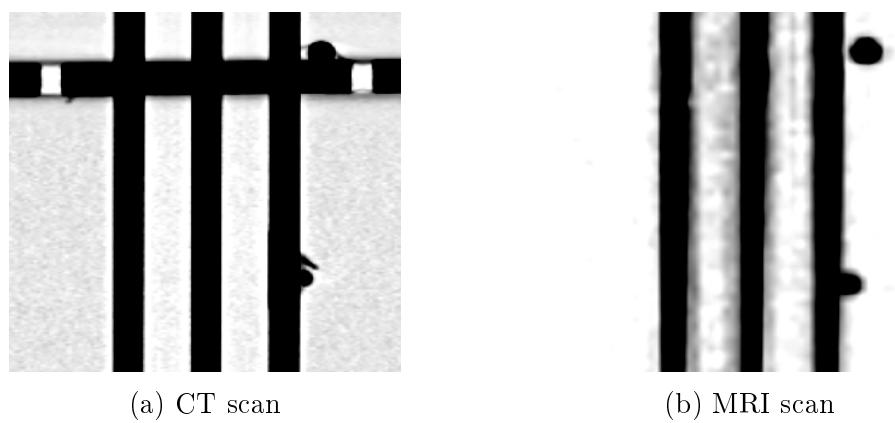


Figure 2.10: CT (a) MR (b) coronal images (inverted colours) showing rods and 2 attached vitamin-e pills visible on the right hand side of the rods. In the CT scan the plastic pane and adhesive tape used to hold the pills in place are also visible.

2.2.2 Rod fillings

For this study 17 different liquids were produced to be tested as possible fillings. They are listed in Table 2.2.

No.	$NaCl$	$CuSO_4 \cdot 5H_2O$	Soap	Ascorbic Acid	Agar	Primovist [volume-%]
#1						
#2	3.6		1.96			
#3	3.6		3.92			
#4	3.6		19.6			
#5	3.6		1.96	1		
#6	3.6		1.96	5		
#7	3.6		1.96	20		
#8	3.6		1.96		0.36	
#9	3.6		1.96		3.6	
#10	3.6		1.96		36	
#11	3.6					0.1%
#12	3.6					1%
#13	3.6					10%
#14	3.6		1.96		0.5	
#15	3.6		1.96		20	
#16			Motor Oil:	<i>Castrol Power1</i>		
#17			Silicon Oil:	<i>Charge: 15HLVY023</i>		

Table 2.2: The composition of all tested solutions.
(components in g/L ; exception: Primovist in volume-%)

- #1 distilled water
- #2 $NaCl + CuSO_4 \cdot 5H_2O$
- #3 increased concentration of $CuSO_4 \cdot 5H_2O$
- #4 further increased concentration of $CuSO_4 \cdot 5H_2O$
- #5 generic washing-up soap added to #2
- #6 increased soap concentration
- #7 further increased soap concentration
- #8 ascorbic acid added to #2
- #9 increased ascorbic acid concentration
- #10 further increased ascorbic acid concentration
- #11 Primovist
- #12 increased amount of Primovist
- #13 further increased amount of Primovist
- #14 agar
- #15 increased agar concentration
- #16 synthetic motor oil
- #17 silicon oil

Being closed at one end and having a capillary shape (small diameter) makes it impossible to fill the rods by simply pouring the liquid through the opening. Instead of adding the fluid at the top, it has to be injected starting at the bottom. This way the contained air is pushed out by the injected liquid through the opening at the top. A long, thin plastic tube was inserted and used for injection, leaving enough room for the gas to escape. Between injections of different liquids, the tube was flushed with #1 (distilled water) or #2 (main component of most solutions).

In order to minimise the amount of gas dissolved, the liquids were brought to boil shortly before injecting. Gas solubility generally decreases with rising temperature [20, 51]. After injecting the solution in the rods, they were left to cool down. Before closing, the rods were topped up completely (no trapped air bubbles). The oil based liquids, #16 and #17, were not brought to boil.

2.3 Sequences

(TODO: elaborate!) Following the suggestions given in the Report of AAPM MR Subcommittee TG1 "MR Acceptance Testing and Quality Control" [22], T1 weighted se-

quences were chosen to evaluate the possible solutions. (Table 2.3)

System	—	—	—	—
MRI	-	-	-	-
CT	-	-	-	-

Table 2.3: used sequences

2.4 Pre-processing MRI and CT scans

Prior to analysing the data, the scans had to be prepared. Figure 2.11 explains the image processing workflow.

- Step 1** After loading the CT and MRI scan into *MIRADA*, they were aligned using the vitamin-e pills, yielding maximum overlap in the centre of the image.
- Step 2** Next, as the MRI image had a lower resolution than the CT scan, the MRI scan was resampled. Its voxel's size were changed to match the CT voxels and both scans exported.
- Step 3** Both layers (MRI and CT) were loaded into *3D Slicer* (Versions: Slicer-4.5.0-1-linux-amd64, Slicer-4.6.2-win-amd64)
- Step 4** Its module 'annotations' was used to set a new region of interest (ROI) to include only a single rod.
- Step 5** With the module called 'crop volume' (setting: voxel based cropping) the scans were reduced to show only the selected ROI.
- Step 6** Using the module 'resample scalar volume' a number of interpolated (setting: 'bspline') higher resolution pairs (CT/MRI) were created.
- Step 7** All new pairs and the cropped original CT/MRI pair were exported with 'create a dicom series' and saved in a separate folder each.

After this procedure a number of pairs based on the original CT and MRI were available, all of which had the same number of slices along the z-axis parallel to the phantom's rods. They only differ in the number of pixels making up each slice, their resolution varying from the original up to a hundred times finer. Each pair (CT/MRI) has the same pixel spacing (and resolution) in x and y direction. See table 2.4 for more details. Figure 2.12 depicts 3 CT/MRI scans of a single rod (axial) with different resolutions. "x1" stands for the original CT scan resolution (MRI resampled to match). "x4" is a resolution caused by 1 pixel being split in 4 smaller pixels, "x9" in 9, and so

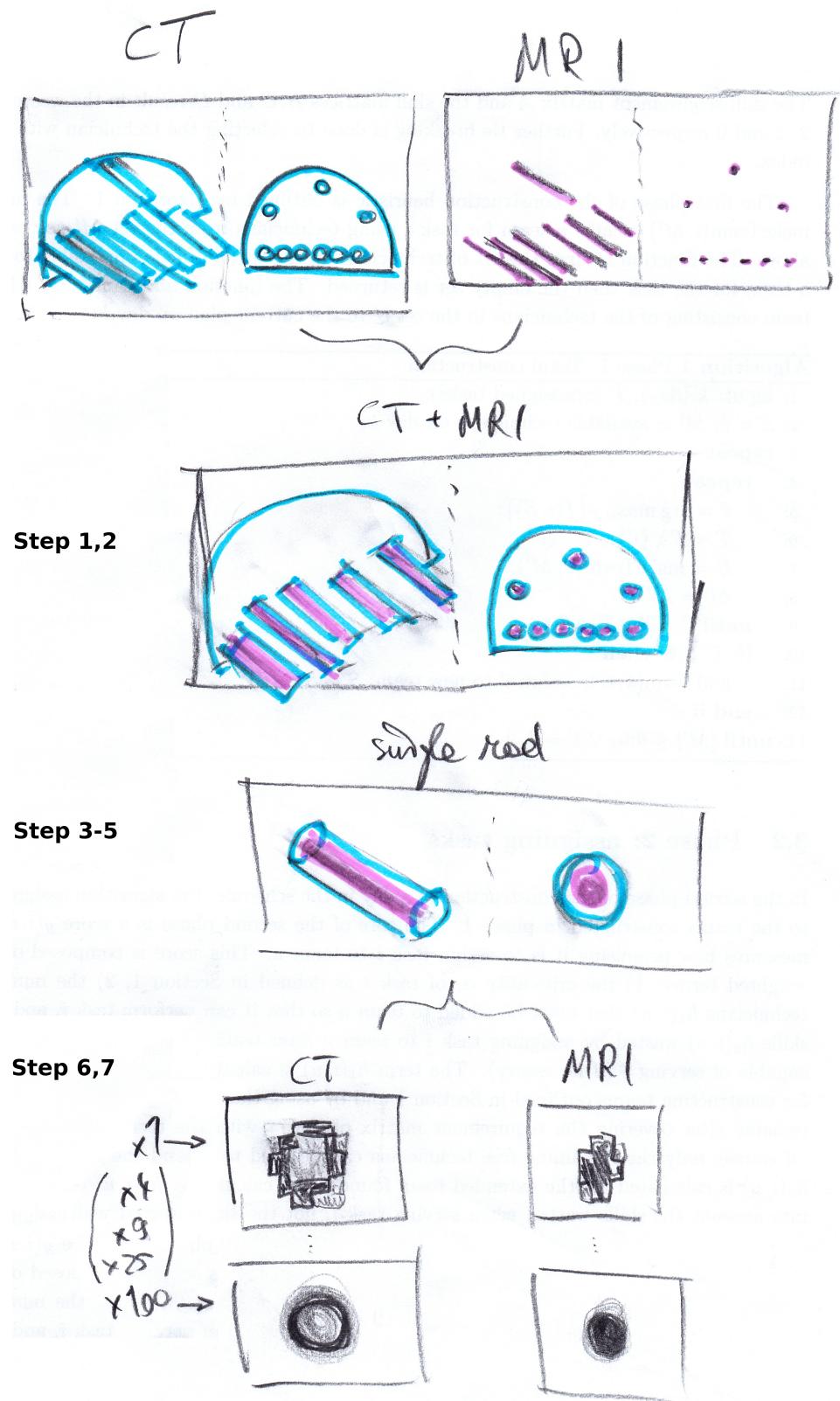
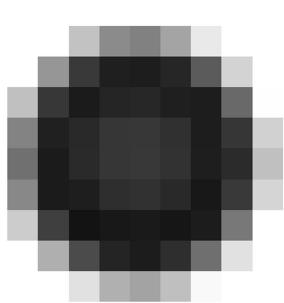


Figure 2.11: Steps performed before analysing data with script (TODO: remake)

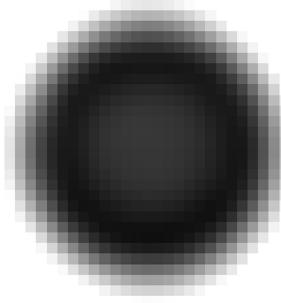
on and so forth. For better visibility, images shown as figures in this work are printed with inverted colours. Dark pixels have a high density/intensity value, white pixels are equivalent to air (low density/intensity).

resample factor	z (not affected)	y (same as x)	x
x1	0.60	0.98	0.98
x4	0.60	0.49	0.49
x9	0.60	0.33	0.33
x25	0.60	0.2	0.2
x100	0.60	0.2	0.1

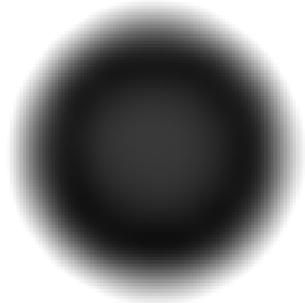
Table 2.4: pixel Spacing (rounded values) [mm]



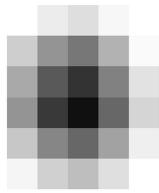
(a) CT x1



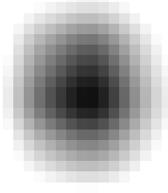
(b) CT x9



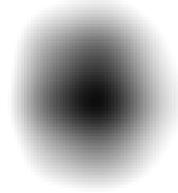
(c) CT x100



(d) MRI x1



(e) MRI x9



(f) MRI x100

Figure 2.12: CT/MRI axial images of the same rod (filling #5, inverted colours). The images on the very left show the original (CT) resolution, the resolution of those in the middle and the right were increased by resampling (9 times and 100 times finer)

2.5 Developed software tool

In order to asses the distortion of the MRI scanner, a software tool was programmed. It is written in Python 2.7 and uses the *SimpleITK* package to read and process *DICOM* ("Digital Imaging and Communications in Medicine") files. [45, 14] *SimpleITK* is a object-oriented "C++ library with wrappers for Python, Java, CSharp, R, Tcl and Ruby". [53, 55] It's versatility is one of the reasons why this approach was favoured. It is a simplified layer built on top of the National Library of Medicine Insight Segmentation and Registration Toolkit (ITK). SimpleITK is also used by Applications like *3D Slicer*, a "free and open source software package for visualisation and medical image computing". [1, 24] For this work *3D Slicer* was used to crop and resample images, quickly read values and visualise the results. Documentation and code examples of SimpleITK can be found at [54, 26] An alternative way to handle DICOM data in Python would be Pydicom. [44, 25]

This is an extensive list of python packages used to process data after using *3D Slicer*:

- SimpleITK
- numpy
- scipy
- matplotlib.pyplot [21]
- skimage.draw
- datetime
- os

Capabilities - Overview

The developed software tool is not able to automatically detect individual rods in a CT or MRI scan depicting the whole phantom. Instead the acquired 3D images have to be cropped so they include only a single rod (see section 2.4).

The python script can be used to:

- separate bright areas which are not connected ('masking' might be used as future method to automatically detect individual rods)
- find and mark slices which show irregularities
- calculate the centroid coordinates along the rod
- measure the local distortion described by the location shift (referred to as "warp")

dice coefficient (the "DC" refers to an object's roundness)

- visualise individual rod slices
- plot the average/peak brightness, warp, or DC along the rod
- write warp and DC values for each slice in a combined ".txt" file
- export a rod shaped scan where the pixel values reflect the distortion occurring in each slice instead of their brightness as a ".mha" file (useful for visualisation, see figure ?? ([TODO: ref to colour map](#))).

2.5.1 Detection of irregularities

The method to find irregularities is described in figure 2.13. After loading the image data, the script calculates the mean brightness of a reference slice (which was chosen by the user) (1.). This reference slice will be used to decide if and which other slices might show irregularities, for example air bubbles, markers, or plastic panes. It is the user's responsibility to make sure the reference is free from any such objects. Ideally, it is located near the iso centre and has a brightness that is representative for the whole image.

The script will compare each slice of the volume to the reference slice individually, starting at one end (2.). To decide whether a particular slice is "irregular", its average brightness will be compared to the reference slice's (4.). If the difference exceeds a certain value (5b.), the current slice will be marked as irregular and consequently won't be used to calculate its DC or centre of mass (COM). A value of 40 was found to be yield good results for CT scans. In the future a more sophisticated approach should be implemented where the script chooses a suitable value based on the image modality and the image size.

Irregular slices and their COM and DC values

As irregular slices cannot be used to asses distortion caused by the scanner, numbers describing their distortion are not of interest. Instead of calculating DC, COM and warp numbers, they will all be set to "-1". The position of the COM cannot lie outside the image, yet coordinates "(-1, -1)" would indicate this. Similarly, the value referred to as *warpMagnitude* and the DC are defined to be positive numbers. All three are therefore easily understood to be invalid, indicating that the particular slice was marked as "irregular". The values representing x- and y-shift, on the other hand, are

allowed to be negative or positive. To be consistent, they are still set to "-1". It is essential to bear this in mind when interpreting the script's output.

Measuring distortion

Since the rods have a cylindrical shape, distortion can only be assessed in radial direction. The z-axis is parallel to the rods, x and y are radial. Ideally, each slice ($z = \text{const.}$) should depict the bright circular profile of the liquid (and of the plastic rod in CT) surrounded by black pixels (air).

Two phenomena were chosen to reflect the amount of distortion occurring in each slice of the MRI scans. The distance which the rod appears to be shifted in the MRI slice compared to the CT slice is referred to as "**warp**". The rod's deformation (deviation from circular profile) is described using the dice-coefficient "**DC**" (also known as Sorensen-Index).

2.5.2 Calculation: dice coefficient (DC)

The DC was chosen as indicator for the deviation from a circular profile. The implementation as python function is based on the open source python package "Medpy". [31] A part of it's module called "metric" was adapted. [32]

The calculation of the DC is performed for each slice individually. Additionally, to asses the overall distortion occurring along the rod, the average of all those values is also saved. As this aspect of distortion does not need a reference scan, the DC is measured for CT and MRI images separately. The dice coefficient or Sorensen index [33] is defined as:

$$DC = \frac{2|A \cap B|}{|A| + |B|} \quad (2.1)$$

It compares a binary image (input A) to a circle (reference B). In a binary image there are only 2 possible pixel values: "0" and "1". However, in the original image, values lie in a range between 0 (or -1024 in CT) and 1000 or higher. In order to reduce the true image to a binary image A, the script needs to split the pixels in 2 groups. A copy of the original picture is created where all pixels with a value above a certain threshold are set to the value of "1". These are regarded as part of the rod. Those which are darker

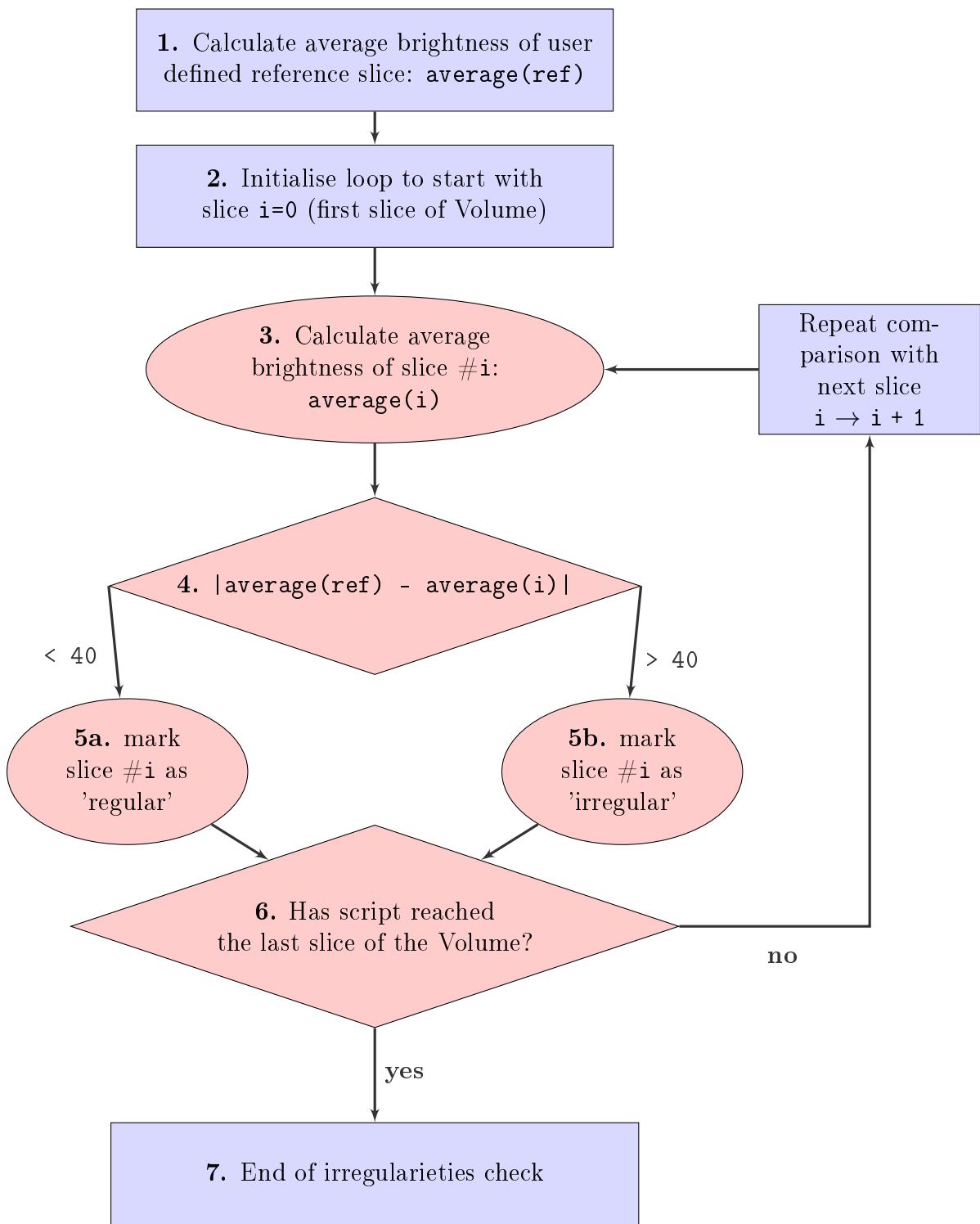


Figure 2.13: Check for irregularities

are set to "0". (See section 2.5.3 for details on how the threshold is calculated). Ideally, this procedure separates the surrounding dark area from the bright rod. This way the new binary image A still shows where the rod is, but it lost all information on the actual brightness.

Reference B is a circle with its midpoint placed at the centre of mass (COM) of the rod. The COM is calculated using those pixels which exceed the threshold, but weighted with their actual brightness (not using the binary image)

This way the script is supposed to place the reference circle B in the centre of the input image A. The position of the circle's centre and its radius highly influence the outcome.

The DC ranges from 0 to 1. A value of 1 indicates a perfect circular shape. A low DC, on the other hand, means the shape differs greatly from a circle and could be caused by many things such as: little overlap (e.g. a ring or crescent shape); a very dark image hindering delineation of the rod from background; a small circle with a radius close to a only a few pixels.

The obvious choice for the radius of the reference circle B is to use the true size of the physical rod. For CT images this would be 4mm, for MRI images it would be 2mm. The script calculates the DC using various radii close to those values and returns the result yielding in the highest average DC for the whole rod. For CT images it chooses values between 3.5 and 4.5, for MRI it varies from 1.8 to 3.8. These boundaries could be changed in the future.

2.5.3 Calculation: warp & centre of mass (COM)

To calculate the location shift between rods shown in CT and MRI, the coordinates of the centre of mass (COM) were subtracted. The x- and y-shift (*warpXY*) measured in each slice was saved in an array. Furthermore, the absolute value of the coordinate shift (*warpMagnitude*) was calculated.

The calculation of the COM is done with help of the "scipy" python package. Its module "ndimage" contains the function "*center_of_mass()*", which returns the COM's coordinates of a given input array. The values assigned to voxels in CT images lie in the range from -1024 HU (air) to around 200 HU (plastic rod). Before a meaningful result can be obtained, the values need to be shifted to be > 0 . Additionally, only pixels representing the rod or the liquid should be used for the calculation. Otherwise the

almost black voxels surrounding the rod would influence the result. To be regarded as part of the rod, the pixels' value has to reach a certain threshold. In order to find the relevant pixels two methods were developed:

- 1** calculating the number of pixels based on rod size
- 2** finding a COM resulting in good DC

Both methods rely on a single reference slice to calculate the threshold. This reference should be representative for the whole scan, because the threshold deduced from it will be used to find pixels belonging to the rod at any other slice.

add 1: The inner ($2mm$) and outer ($4mm$) radius of the plastic rods are known. So is the pixel *spacing*, the size of a voxel in real space (mm). Calculating the number of pixels which make up the more or less circular profile of the rod in a slice is calculated as follows:

$$pixelNumber = (radius^2 \cdot \pi) / (spacing^2) \quad (2.2)$$

For CT images $radius = 4mm$, in MRI scans $radius = 2mm$. *spacing* is the pixel spacing in x and y direction. Next the pixels are sorted by brightness. The top *pixelNumber* pixels are then used to calculate the COM. The value of the darkest pixel that is still counted as part of the rod is saved as threshold for future calculations (e.g. finding the DC associated with the COM, see section 2.5.2). **add 2:** This algorithm is an iteration method. Figure 2.14 shows in what order the scripts executes individual steps during the iteration.

To begin with, it looks at the whole range of possible *pixelNumbers*, from 0% to 100% **(1.)**. As a reasonable first guess it assumes that 50% of all pixels belong to the rod **(2.)**. Now, in the first iteration **(3.)**, to find out whether more or less pixels would result in a better DC, it considers two new guesses: One halfway from the lower limit (0%) to its current guess (50%) which is:

$$\frac{0 + 50}{2} = 25\% \quad (2.3)$$

and one halfway from the upper limit (100%) to its current guess (50%) which is:

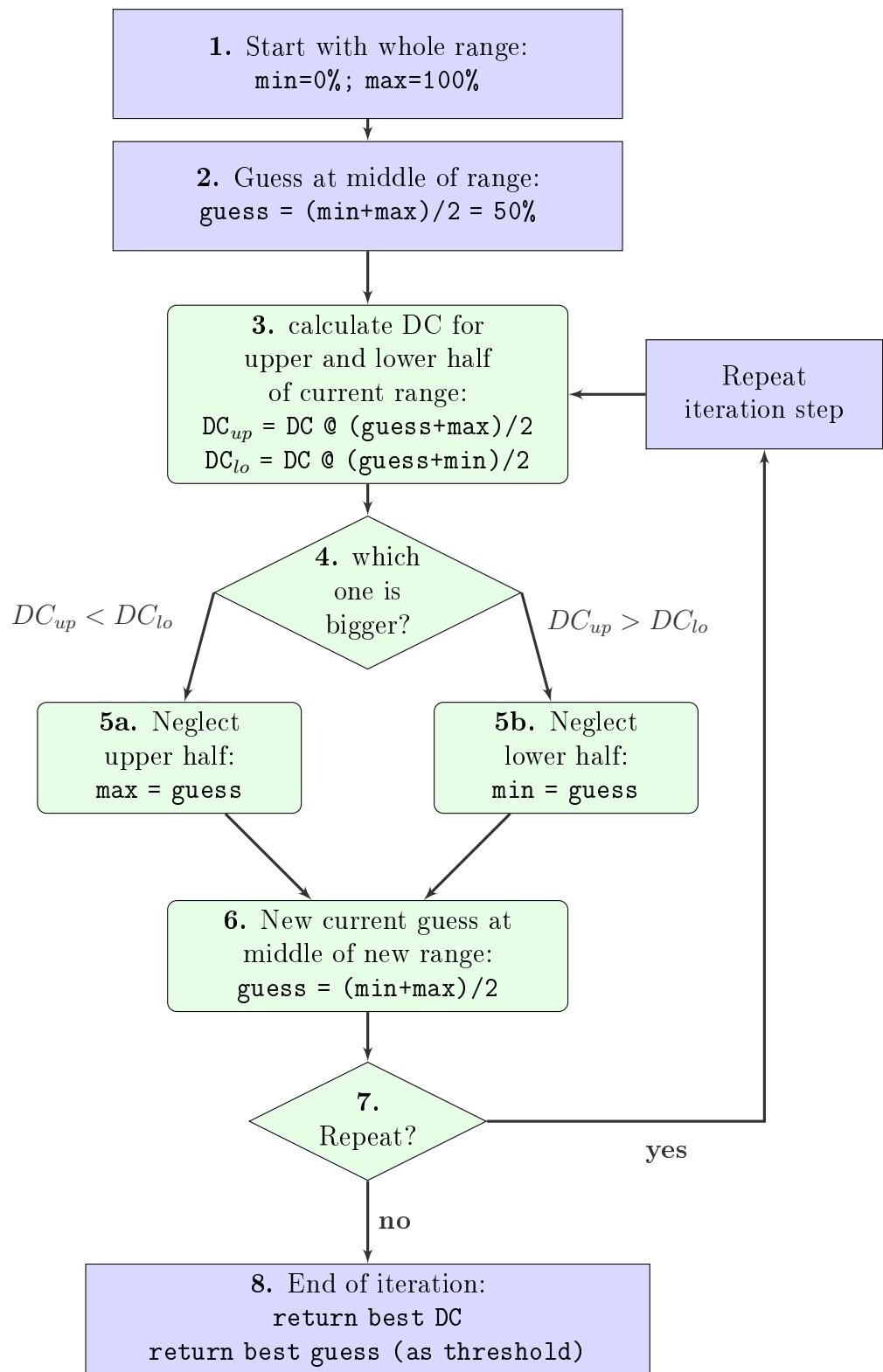


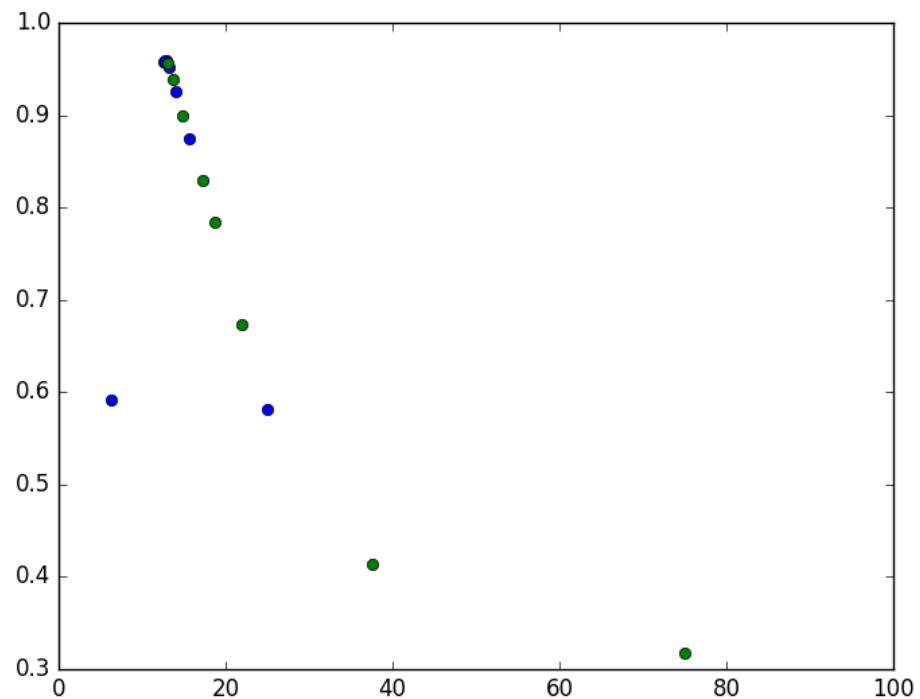
Figure 2.14: Iteration method, finding COM

$$\frac{100 + 50}{2} = 75\% \quad (2.4)$$

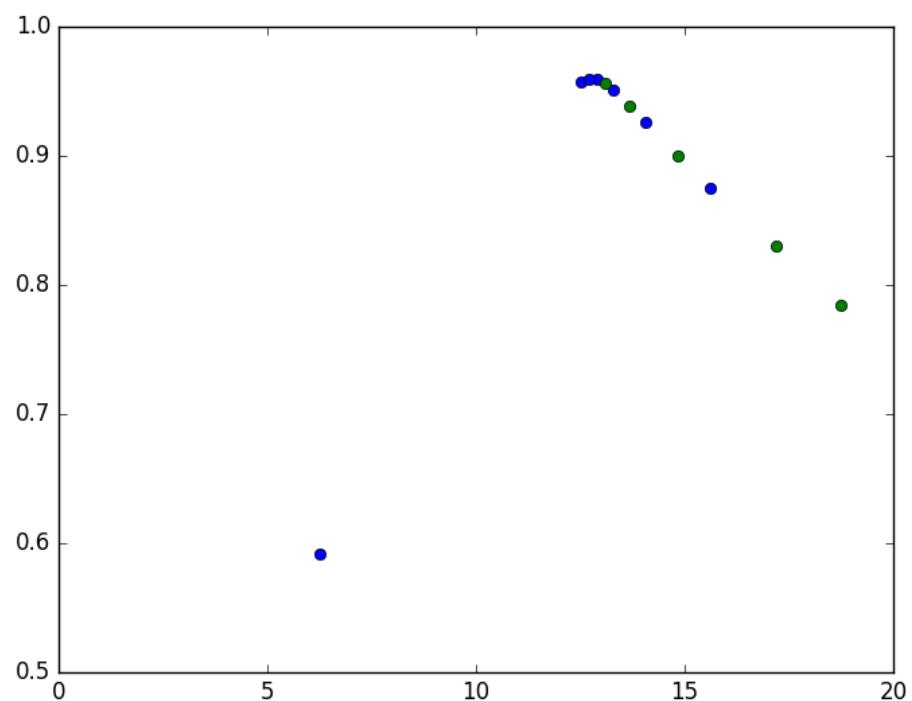
This numbers correspond to a threshold each separating the chosen number of pixels from all darker pixels in the slice. Using these thresholds, the script calculates the COM and DC for both possible guesses. Comparing the two DCs will decide which guess is closer to representing the rod better (4.). If the lower number of pixels yields a better DC (5a.), the upper half of the range will be neglected in the next iteration or in other words, the new upper limit takes the value of the former guess (50%). If the higher percentage yields a better DC (5b.), the lower half of the range will be neglected in the next iteration, the new lower limit takes the value of the former guess (50%). At the end of the iteration (6.), the percentage resulting in the higher DC is saved as the new current guess. After this first iteration (7.) the script can either repeat steps 3-6 or end it by returning the best guess and its DC (8.).

To get a better understanding, let's suppose the iteration is repeated. At the start of the second iteration, the range is smaller (half of the entire range) and the current guess is set exactly in its middle. If, for example, the DC for 25% was higher than for 75%, the next guess will be 25%, because the new range goes from 0% to 50%. In that case, DC values for the lower half of that range (12,5%) and the upper half (18,5%) will be calculated and compared to decide which half to eliminate in the third iteration. If, on the other hand, the DC for 75% was higher, the next guess will then be 75%, because the new range goes from 50% to 75%. In that case, DC values for 62,5% and 82,5% will be compared.

The iteration continues until further steps yield no better DC values or a set number of steps has been performed. After the iteration process, the algorithm will return the COM which resulted in the best DC. The percentage of pixels that led to this DC is equivalent to a threshold value which is saved for future calculations. Figure 2.15 shows the DC values found in the course of trying different percentages during the iteration method.



(a) full iteration process



(b) close up

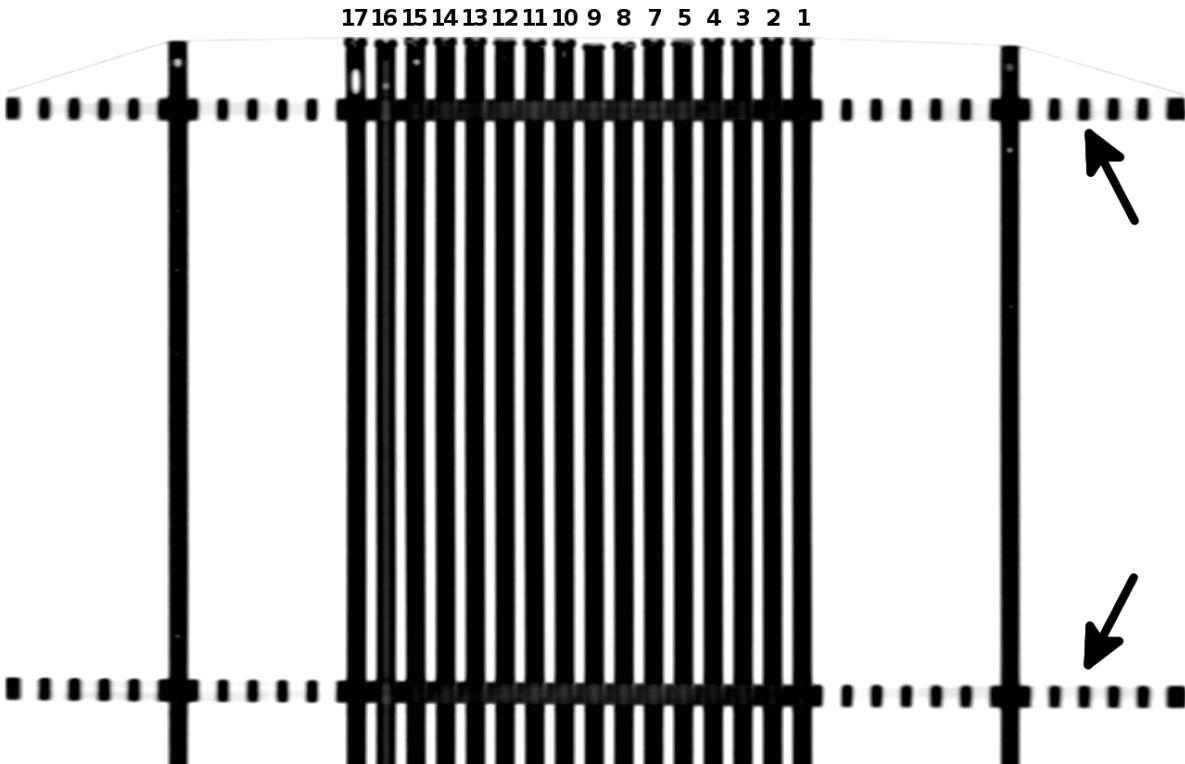
Figure 2.15: COM iteration method, 8 repetitions

3 Results

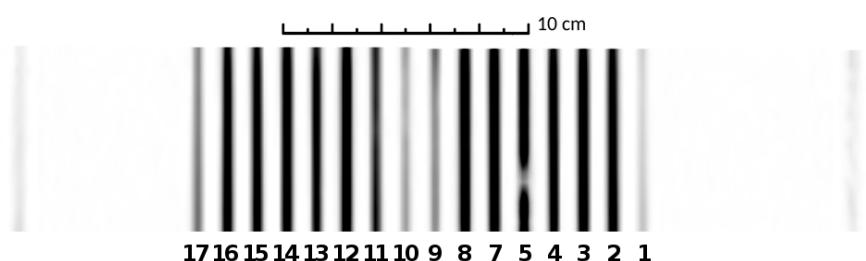
On the second day of working with the filled rods the one containing liquid #6 broke (leakage). It happened when delicately knocking it against on the table while standing upright. This was intended to mobilise bubbles that stucked to the wall and make them travel vertically to on end of the rod. (see table 3.2) The plastic stopper on the lower end came loose. The rod containing filling #6 was not replaced. Consequently, all CT/MRI images used to asses signal intensities of the tested solutions show only 16 rods.

3.1 Obtained MRI and CT scans

Figure 3.1 shows a coronal view of the 16 rods filled with the tested liquids and a reference rod on either side. In Figure 3.1b a trapped air bubble is clearly visible at the lower half of rod #5. Figure 3.2 shows an axial view of the tested rods and some surrounding rods. In figure 3.2b a water filled plastic bottle placed in the middle of the phantom is also visible. This was necessary, because the MRI scanner needs sufficient signal to start imaging. Without the bottle, the limited number of rods used for this scan would not have created enough signal. During a future distortion assessment where all available rods (over 300) would be used, they are supposed to result in the required signal strength on their own.

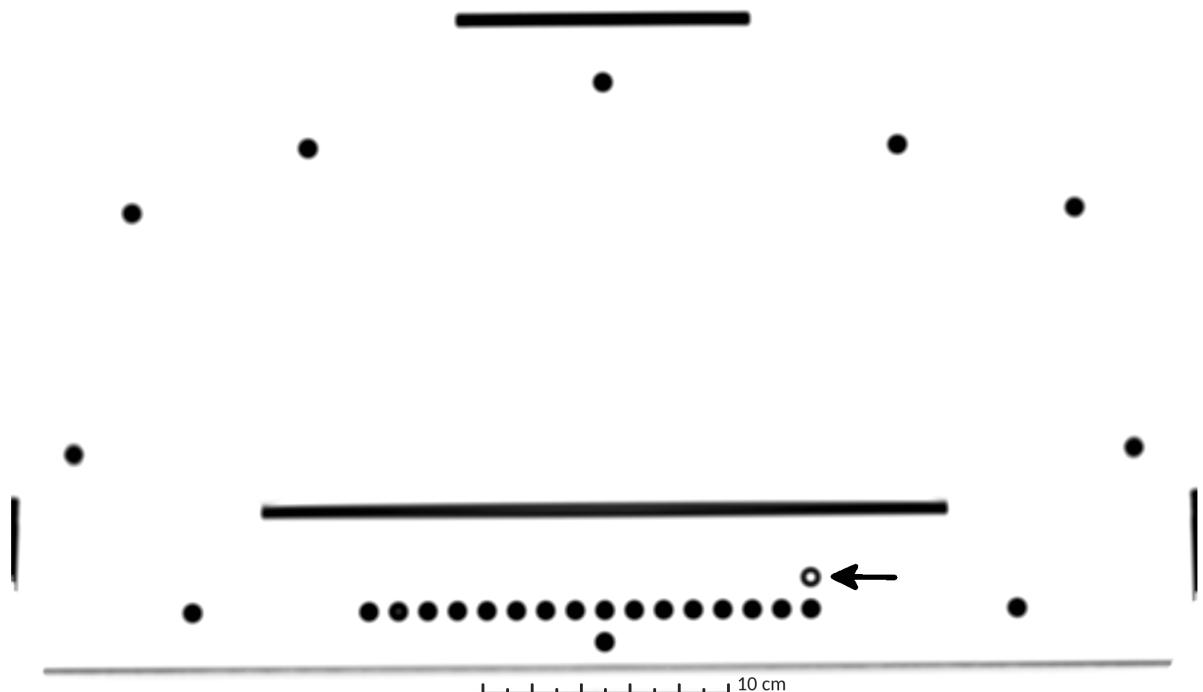


(a) CT: The periodic black lines in upper and lower part of image (indicated with arrows) depict the plastic panes and their holes from above; The faint line across upper end of rods shows adhesive tape used to hold the rods in place; In rod #17 an air bubble is clearly visible close to the upper end.

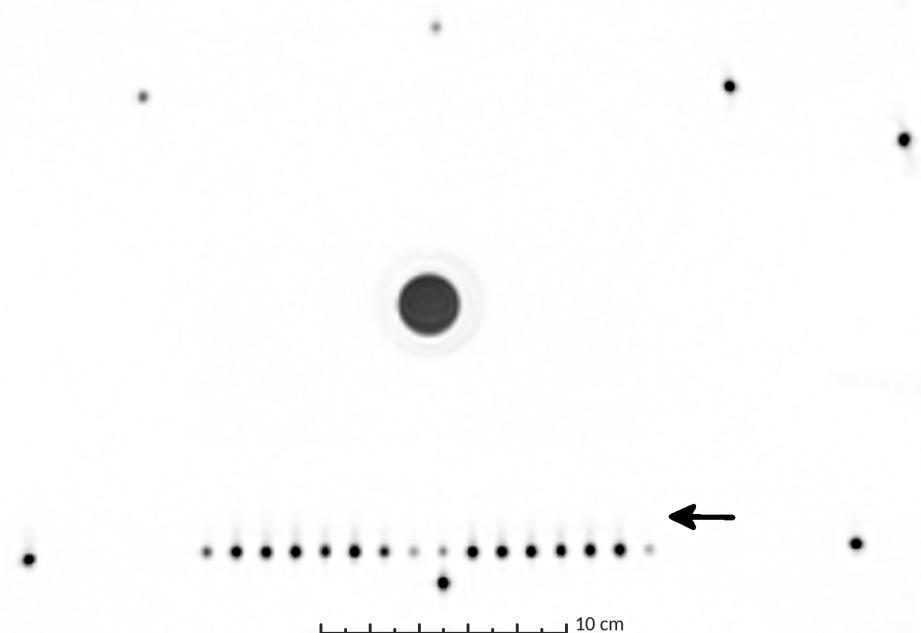


(b) MR: The rods appear to be thinner than in the CT scan, because only the liquid filling is visible. The tested liquids result in different signal intensity (brightness); all plastic parts (rods and panes) are not visible; there is a trapped air bubble visible in rod #5.

Figure 3.1: Coronal CT/MRI (inverted colours; same scale; cropped images) images of 16 rods (tested liquids, numbering starting from the right, #6 excluded) + 2 reference rods (filled with water) on the sides.



(a) CT: The black bars visible just above the 16 tested rods, at the very top, and to the sides show plastic parts of the phantom holding it together. The faint grey line running below the tested rods represents the table on which the phantom was positioned during imaging.



(b) MRI: The black circle in the centre depicts a water bottle which was placed in the middle of the phantom (necessary for MRI scanner to start imaging).

Figure 3.2: axial CT/MRI (inverted colours, same scale) images of the 16 rods filled with tested liquids (numbering starting from the right, #6 excluded); surrounded by reference rods (filled with water) and one empty rod (marked with arrow) which is not visible on the MRI scan

3.2 Tested solutions

3.2.1 Visibility on CT/MRI scans

It should be noticed that in the CT image there is little difference in brightness between the tested liquids. The plastic rods themselves result in brighter pixels than any of the tested solutions.

On MRI scans most liquids had a mean and a max brightness value above 1000 (see table 3.1).

Only #1, #9, #10, & #17 resulted in significantly less signal (below < 1000).

No.	Min	Max	Mean	Median	σ
1	182	371	288	269	69,8
2	1044	1921	1443,8	1405	312,9
3	941	2075	1451,2	1394,5	413,2
4	1176	1709	1440	1437,5	232,5
5	1125	2111	1583,8	1549,5	355
7	971	2241	1466,8	1316	471,2
8	1459	1947	1704	1705	180,5
9	385	584	486,8	489	93
10	247	502	343,6	266	111
11	830	1268	1036,2	1023,5	163,2
12	1158	2211	1648,8	1613	394,2
13	836	1657	1146,8	1047	321,2
14	800	2062	1383	1335	473,1
15	1156	1829	1476,2	1460	272,7
16	1102	1967	1509	1483,5	325,8
17	356	938	629,6	602	223,6

Table 3.1: liquid visibility on MRI scan

3.2.2 Mechanical properties of solutions

The liquids were filled in a rod each and observed for several months. Number #14 could be injected without problems, the solution remained fluid even after reaching room temperature. Number #15 on the other hand changed to a gel like consistence and clogged the injection tube at the quickly after the rod was filled. The tube could not be used again.

Each rod was free of bubbles directly after sealing. All rods containing water based solutions contained some air after 2 months and the amount of liquid continued to decrease further (see table 3.2). In some of the tested rods the produced air bubbles would stick to the wall. Only after gently hitting the rod they would start moving. Knowing the inner diameter d of the rods and measuring the length l of trapped air bubbles, we can approximate the volume of contained gas:

$$V = \frac{d^2}{4} \cdot \pi \cdot l \quad (3.1)$$

While adding generic washing soap (#5, #6 and #7) did not hinder air bubbles from forming, it significantly improved their mobility. Not only did they move quickly when the rod was tilted, large quantities of air also did not block the entire diameter of the rod. Instead they formed large but cohesive bubbles that could be moved to one end of the rod easily and at no point sticked to the plastic wall.

The ascorbic acid present in #8 (concentration of 0.36 g/L corresponds to approx. 0.00204 mol/L), #9 (3.6g/L), and #10 (36g/L) seemed to have held back the formation of air bubbles for up to one week. After two months of observation, however, the rods also contained some air. It should be noted that all three liquids turned brown, the colour being more saturated for higher concentrations of ascorbic acid.

The rods filled with Primovist (#11 to #13) were filled with some air bubbles after at least two days. Moreover, the bubbles sticked to the walls of the rod and only shaking it violently made them move to one side of the rod.

It took more than a week until the rod containing the low concentration of agar (#14) contained an air bubble. The viscous consistency made it impossible to coerce it to either end of the rod. Liquid #15 on the other hand did not form bubbles at the middle of the rod, but seemed to have dried starting at the end with the plastic stopper.

No.	after 1 day		after 2 days		after 1 week	
	bubbles	hit req.	bubbles	hit req.	bubbles	hit req.
#1	yes	no	no		no	
#2	yes	yes	no		no	
#3	yes	yes	no		no	
#4	yes	yes	no		no	
#5	yes	no	yes	no	no	
#6	yes	no	<i>rod was leaking</i>			
#7	yes	no	yes	no	yes	no
#8	no		no		no	
#9	no		no		no	
#10	no ¹		yes	yes	yes	yes
#11	no		yes,	<i>sticked to wall</i>	yes	yes
#12	yes	yes	yes,	<i>sticked to wall</i>	yes	yes
#13	yes	yes	yes,	<i>sticked to wall</i>	yes	yes
#14	no		yes	no	yes	yes
#15	no		no		no	
#16	no		no		no	
#17	no		no		no	

No.	after 2 months	
	length of trapped bubble l [mm]	approx. volume V [mm ³]
#1	2	25.13
#2	1.8	22.62
#3	1+1 (air blockage, at lower end)	25.13
#4	4	50.27
#5	1.5 (many small bubbles)	18.85
#6	<i>rod was leaking</i>	
#7	2 (many small bubbles)	25.13
#8	2.3	28.90
#9	3	37.70
#10	2.4	30.16
#11	2	25.13
#12	2	25.13
#13	2.3	28.90
#14	1.5+0.5 (big immobile bubble, at center)	25.13
#15	3.4 (agar gel dried)	42.73
#16	0	0.00
#17	0.5	6.28

Table 3.2: Observations regarding the mechanical properties of the tested solutions.

3.3 Distortion assessment

All results were obtained by manually cropping the 3D image to depict only a single rod.

3.3.1 rod #5

The first scans were not only used to asses the brightness of the different solutions, but also to calculate occurring distortion. As an example, the rod containing liquid #5 was analysed. Table 3.3 shows the most interesting areas of the rod.

The obtained dice coefficient varies not only because of the circle's centre and the radius, it also depends on the images resolution. Figure ?? and ?? show the DC (optimised) obtained using CT and MRI scans over resample rate.

slice	dist	$warp_x$		$warp_y$		$warp_M$		DC_{CT}		DC_{MR}		$DC_{MR(CT-COM)}$		$warp_x^*$		$warp_y^*$		$warpM^*$		DC^{*}_{CT}		DC^{*}_{MR}		$DC^{*}_{MR(CT-COM)}$	
		$warp_x$	$warp_y$	$warp_x$	$warp_y$	DC_{CT}	DC_{MR}	$DC_{MR(CT-COM)}$	$warp_x^*$	$warp_y^*$	$warpM^*$	DC^{*}_{CT}	DC^{*}_{MR}	$DC^{*}_{MR(CT-COM)}$	$warp_x^*$	$warp_y^*$	$warpM^*$	DC^{*}_{CT}	DC^{*}_{MR}	$DC^{*}_{MR(CT-COM)}$	$warp_x^*$	$warp_y^*$	$warpM^*$	DC^{*}_{CT}	DC^{*}_{MR}
0	-183	-0.1046	-1.115	1.1199	0.981	0.846	0.5982		-0.1104	-1.0595	1.0653	0.9906	0.9477	0.5982				0.9906	0.9487	0.5982	0.9863	0.9613	0.8938		
1	-182	-0.1205	-1.1132	1.1197	0.9832	0.8551	0.6037		-0.1245	-1.0514	1.0588	0.9906	0.9487	0.6037				0.9906	0.9487	0.6037	0.9852	0.9617	0.8932		
2	-181	-0.1278	-1.1107	1.1181	0.9831	0.8678	0.6109		-0.1313	-1.0485	1.0567	0.9846	0.948	0.5813				0.9846	0.948	0.5813	0.9839	0.9621	0.8958		
:	:																								
171	-12	0.0025	-0.6891	0.6891	0.9847	0.9412	0.7571		-0.0313	-0.6969	0.6976	0.9869	0.9451	0.7571				0.9869	0.9451	0.7571	0.9822	0.9458	0.7571		
172	-11	0.0089	-0.6825	0.6825	0.9865	0.9412	0.7571		-0.0272	-0.7012	0.7017	0.9822	0.9458	0.7571				0.9822	0.9458	0.7571	0.9504	-1			
173	-10	-1	-1	-1	-1	0.9409	-1		-1	-1	-1	-1	-1	-1				-1	0.946	-1					
174	-9	-1	-1	-1	-1	0.9413	-1		-1	-1	-1	-1	-1	-1				-1	0.9458	-1					
:	:																								
192	9	-1	-1	-1	-1	-1	-1		-1	0.9464	-1	-1	-1	-1				-1	0.95	-1					
193	10	-1	-1	-1	-1	-1	-1		-1	0.9479	-1	-1	-1	-1				-1	0.9504	-1					
194	11	0.0379	-0.709	0.71	0.989	0.9484	0.7649		0.0028	-0.7153	0.7153	0.9805	0.9496	0.7649				0.9805	0.9496	0.7649	0.9821	0.9498	0.7661		
195	12	0.0364	-0.7048	0.7058	0.989	0.948	0.7661		0.0033	-0.7156	0.7156	0.9821	0.9498	0.7661				0.9821	0.9498	0.7661					
:																									
301	118	0.2997	-0.8702	0.9204	0.9785	0.4295	0.3962		0.1128	-0.8025	0.8104	0.9841	0.8861	0.3962				0.9841	0.8861	0.3962	0.9841	0.8666	0.1423		
302	119	0.3943	-1.1317	1.1984	0.9818	0.1453	0.1423		0.1073	-0.8906	0.8971	0.9841	0.8666	0.1423				0.9841	0.8666	0.1423	0.9841	0.8317	0		
303	120	-1	-1	-1	-1	0.9822	-1		0.1066	-1.0077	1.0133	0.9841	0.8317	0			0.9841	0.8317	0	0.9841	0.8317	0			
304	121	-1	-1	-1	-1	0.9832	-1		0.0896	-1.1827	1.1861	0.9857	0.7845	0			0.9857	0.7845	0	0.9857	0.7845	0			
305	122	-1	-1	-1	-1	0.9828	-1		0.0852	-1.4164	1.419	0.9854	0.7153	0			0.9854	0.7153	0	0.9854	0.7153	0			
306	123	-1	-1	-1	-1	0.9824	-1		0.0769	-1.5832	1.5851	0.9849	0.6758	0			0.9849	0.6758	0	0.9849	0.6758	0			
307	124	-1	-1	-1	-1	0.9833	-1		0.0788	-1.5436	1.5456	0.9851	0.7156	0			0.9851	0.7156	0	0.9851	0.7156	0			
308	125	-1	-1	-1	-1	0.982	-1		0.0826	-1.5257	1.5279	0.9862	0.7501	0			0.9862	0.7501	0	0.9862	0.7501	0			
309	126	-1	-1	-1	-1	0.9837	-1		0.0868	-1.4826	1.4826	0.9859	0.7831	0			0.9859	0.7831	0	0.9859	0.7831	0			
310	127	0.3786	-2.0028	2.0383	0.9805	0.0754	0.011		0.0718	-1.4467	1.4485	0.9865	0.8135	0.011			0.9865	0.8135	0.011	0.9862	0.8422	0.0434			
311	128	0.3434	-1.9836	2.0131	0.9837	0.1648	0.0629		0.0775	-1.4081	1.4102	0.9862	0.8422	0.0434			0.9862	0.8422	0.0434	0.9862	0.8422	0.0434			
:																									
393	210	0.1083	-0.2563	0.2783	0.9819	0.9334	0.8938		0.0968	-0.2638	0.281	0.9863	0.9613	0.8938				0.9863	0.9613	0.8938	0.9852	0.9617	0.8932		
394	211	0.1147	-0.263	0.2869	0.9828	0.932	0.8932		0.0993	-0.2653	0.2832	0.9852	0.9617	0.8932				0.9852	0.9617	0.8932	0.9839	0.9621	0.8958		
395	212	0.1214	-0.2454	0.2738	0.9836	0.9338	0.8984		0.1041	-0.2532	0.2738	0.9839	0.9621	0.8958				0.9839	0.9621	0.8958	0.9839	0.9621	0.8958		

Table 3.3: rod #5: script generated data

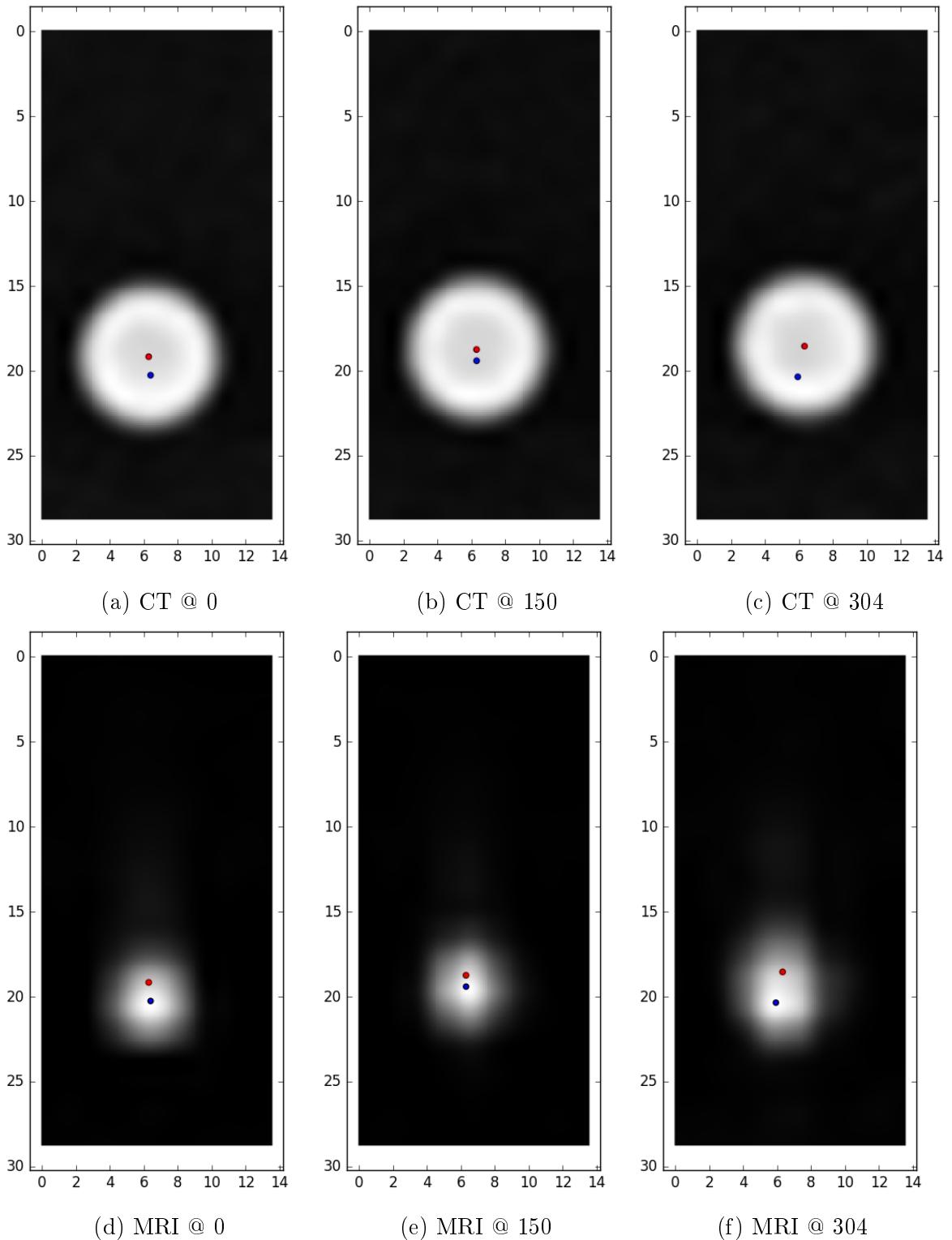


Figure 3.3: MRI x100 scan of rod filled with #5 (true colours). The dark blue dot represents the MRI centroid, the bright red dot represents the CT centroid. Slice 150 is located approximately at the isocentre, 0 on the very end of the image, 304 on the other but also close to an air bubble.

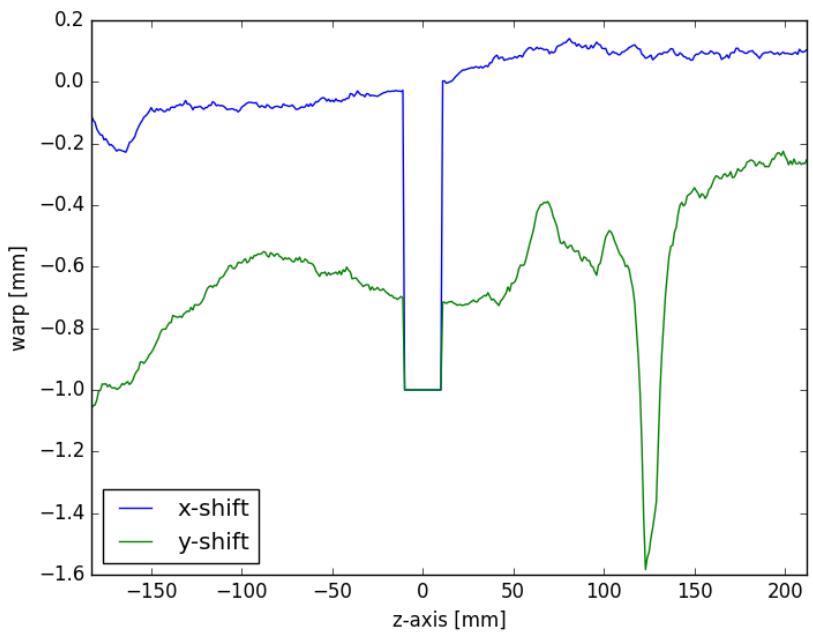


Figure 3.4: Rod #5: warp XY [mm] (iteration method), CT-MRI x100

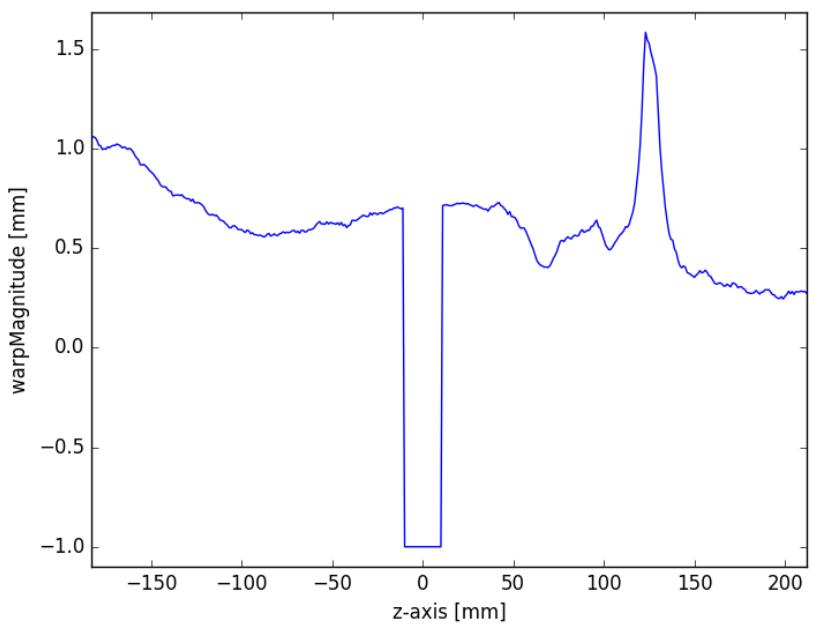


Figure 3.5: Rod #5: warp Magnitude [mm] (iteration method), CT-MRI x100

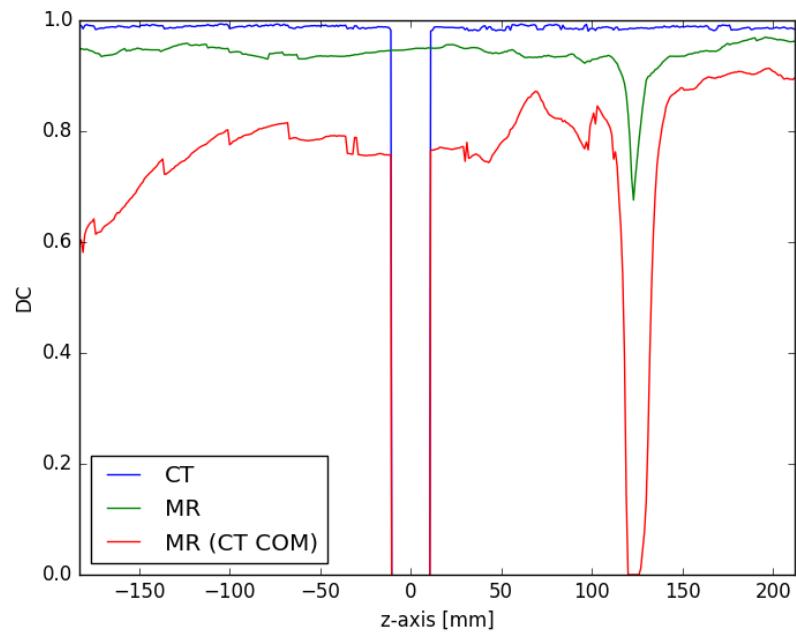


Figure 3.6: Rod #5: DC (iteration method) for CT & MRI & MRI (using CT COM)

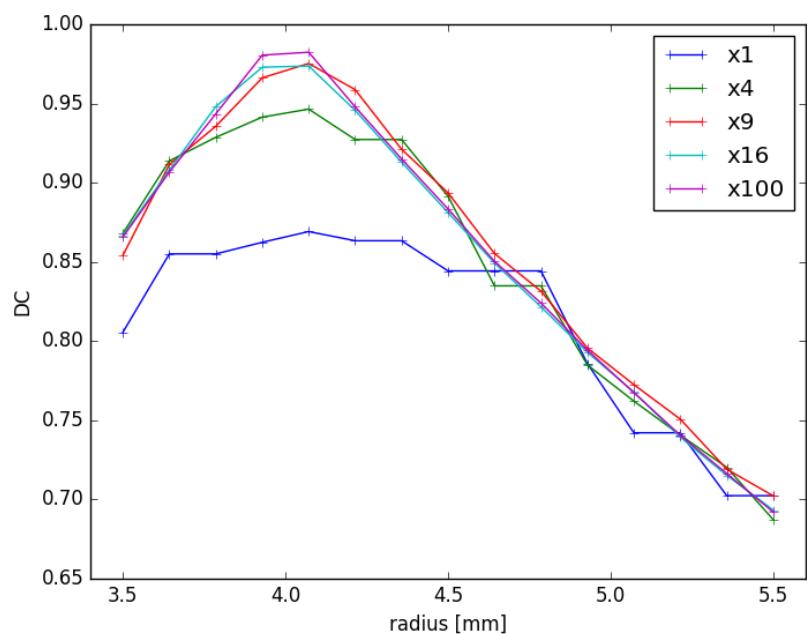


Figure 3.7: Rod #5: CT DC of varied radii & resolutions

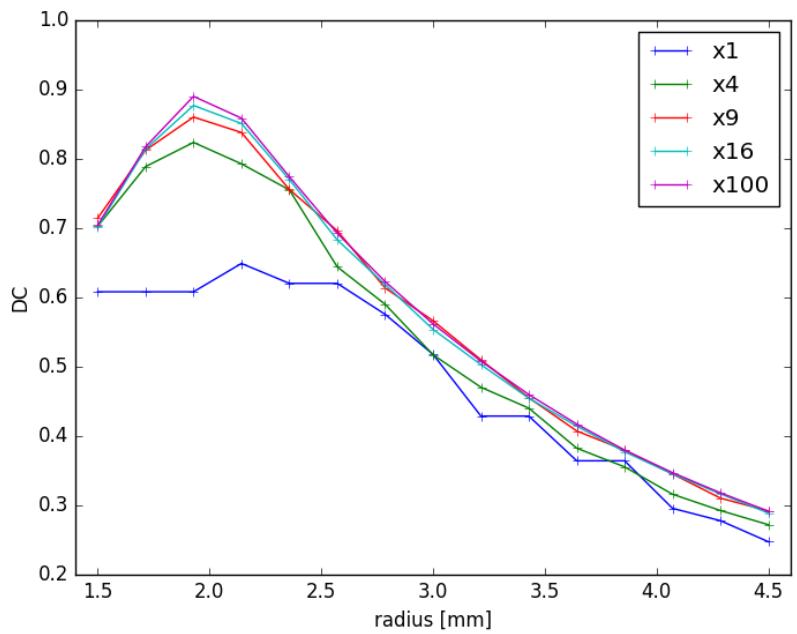


Figure 3.8: Rod #5: MRI DC of varied radii & resolutions

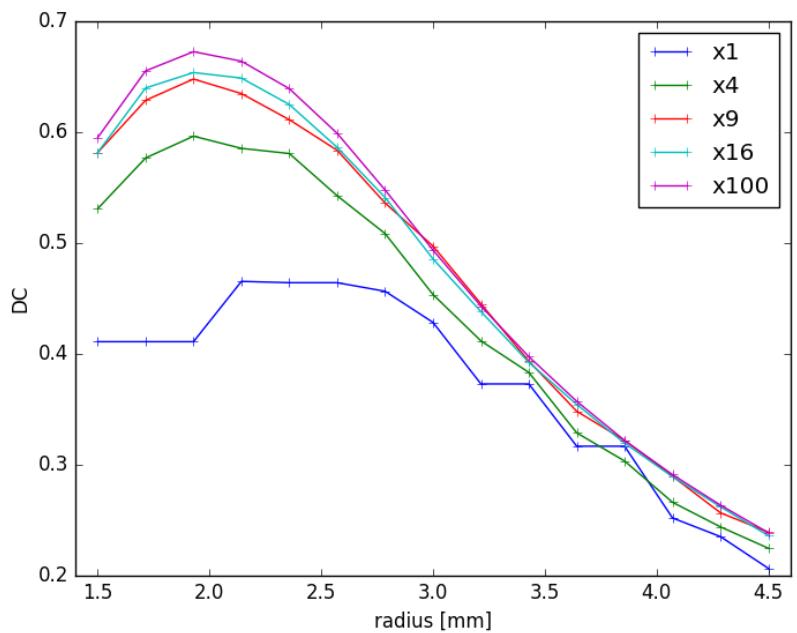


Figure 3.9: Rod #5: MRI DC of varied radii & resolutions (using CT-COM)

3.3.2 rod #17

After deciding which of the possible liquids might be a suitable choice for future use of the phantom, another set of MRI/CT scan was taken. This second data set shows the rod containing liquid #17. Again it was analysed using the developed script. Table 3.4 shows the most interesting areas of the rod.

(TODO: PIOTR: create colour coded images)

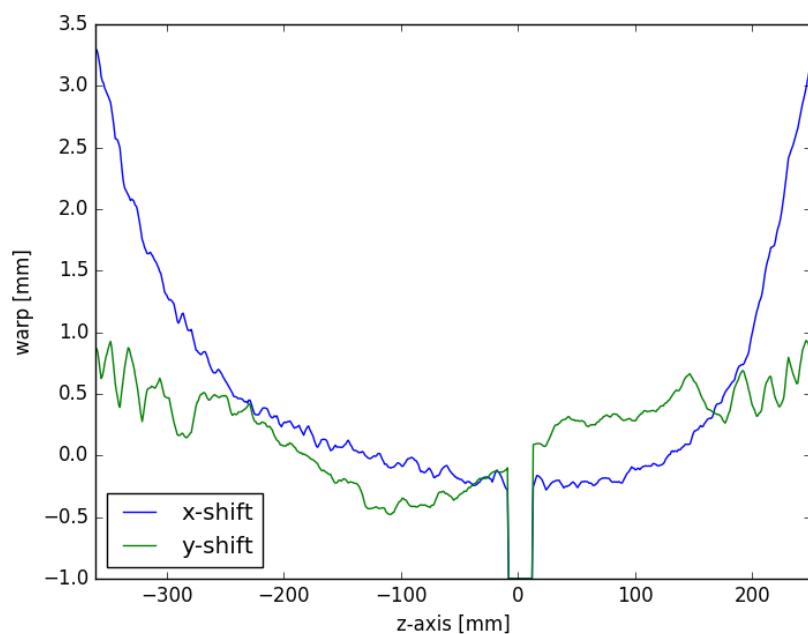


Figure 3.10: Rod #17: warp XY [mm] (iteration method), CT-MRI x100

slice	dist	$warp_x$	$warp_y$	$warpM$	DC_{CT}	DC_{MR}	$DC_{MR(CT-COM)}$	$warp_x^*$	$warp_y^*$	$warpM^*$	DC_{CT}^*	DC_{MR}^*	$DC_{MR(CT-COM)}^*$
0	-361	2.9415	0.4286	2.9725	0.967	0.0286	0.0077	3.2985	0.8734	3.4122	0.9406	0.603	0.0131
1	-360	2.94	0.4294	2.9712	0.9698	0.0286	0.0077	3.2943	0.8677	3.4067	0.9396	0.604	0.0131
2	-359	2.9441	0.4224	2.9742	0.9717	0.0338	0.0092	3.2666	0.8416	3.3732	0.9416	0.6014	0.0145
:	:												
351	-10	-0.2049	-0.0356	0.208	0.9854	0.9307	0.9045	-0.2709	-0.1155	0.2945	0.9907	0.932	0.9045
352	-9	-0.2217	-0.0174	0.2223	0.9853	0.9307	0.9056	-0.2813	-0.0996	0.2984	0.9832	0.9313	0.9056
353	-8	-1	-1	-1	-1	0.9302	-1	-1	-1	-1	-1	0.9304	-1
354	-7	-1	-1	-1	-1	0.9296	-1	-1	-1	-1	-1	0.9295	-1
355	-6	-1	-1	-1	-1	0.9276	-1	-1	-1	-1	-1	0.9262	-1
356	-5	-1	-1	-1	-1	0.9253	-1	-1	-1	-1	-1	0.9187	-1
357	-4	-1	-1	-1	-1	0.9198	-1	-1	-1	-1	-1	0.9185	-1
358	-3	-1	-1	-1	-1	0.9174	-1	-1	-1	-1	-1	0.9178	-1
359	-2	-1	-1	-1	-1	0.9142	-1	-1	-1	-1	-1	0.9179	-1
360	-1	-1	-1	-1	-1	0.912	-1	-1	-1	-1	-1	0.9184	-1
361	0	-1	-1	-1	-1	0.909	-1	-1	-1	-1	-1	0.9079	-1
362	1	-1	-1	-1	-1	0.8999	-1	-1	-1	-1	-1	0.9055	-1
363	2	-1	-1	-1	-1	0.9007	-1	-1	-1	-1	-1	0.9031	-1
364	3	-1	-1	-1	-1	0.9011	-1	-1	-1	-1	-1	0.9008	-1
365	4	-1	-1	-1	-1	0.9012	-1	-1	-1	-1	-1	0.8981	-1
366	5	-1	-1	-1	-1	0.9028	-1	-1	-1	-1	-1	0.8961	-1
367	6	-1	-1	-1	-1	0.9052	-1	-1	-1	-1	-1	0.9006	-1
368	7	-1	-1	-1	-1	0.908	-1	-1	-1	-1	-1	0.9045	-1
369	8	-1	-1	-1	-1	0.9109	-1	-1	-1	-1	-1	0.9024	-1
370	9	-1	-1	-1	-1	0.9138	-1	-1	-1	-1	-1	0.9057	-1
371	10	-1	-1	-1	-1	0.9159	-1	-1	-1	-1	-1	0.9116	-1
372	11	-1	-1	-1	-1	0.9216	-1	-1	-1	-1	-1	0.9177	-1
373	12	-1	-1	-1	-1	0.9233	-1	-1	-1	-1	-1	0.9219	-1
374	13	-0.2506	0.1242	0.2797	0.9859	0.9284	0.9142	-0.2553	0.0875	0.2699	0.9899	0.9284	0.9142
375	14	-0.2181	0.1166	0.2473	0.9838	0.9354	0.9207	-0.2258	0.091	0.2434	0.9926	0.9287	0.9207
:	:												
609	248	3.0734	0.7814	3.1712	0.9803	0.9316	0.363	3.0791	0.9091	3.2105	0.9825	0.9225	0.363
610	249	3.1422	0.7546	3.2315	0.9796	0.9345	0.3446	3.1281	0.8819	3.2501	0.9822	0.9329	0.3446
611	250	3.2214	0.7399	3.3053	0.9832	0.9363	0.3085	3.1826	0.8566	3.2958	0.9828	0.9358	0.325

Table 3.4: rod #17: script generated data

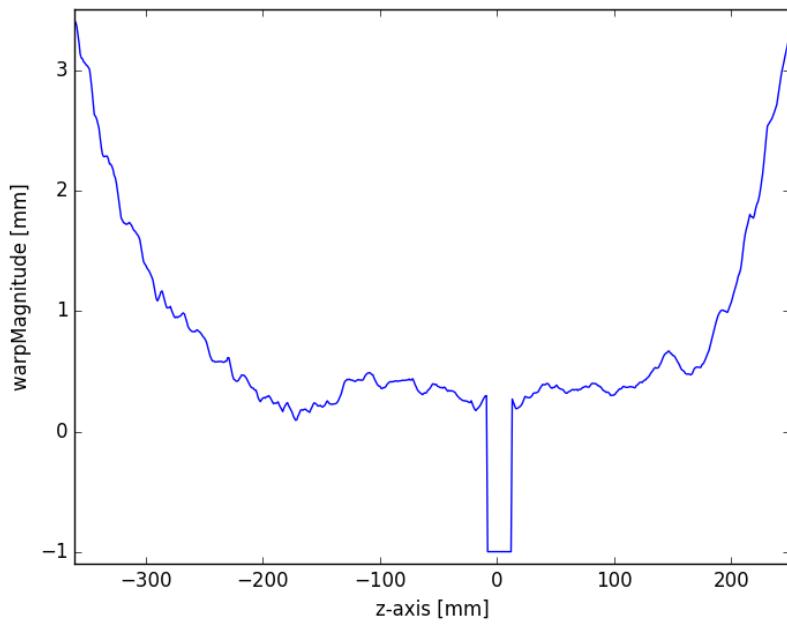


Figure 3.11: Rod #17: warp Magnitude [mm] (iteration method), CT-MRI x100

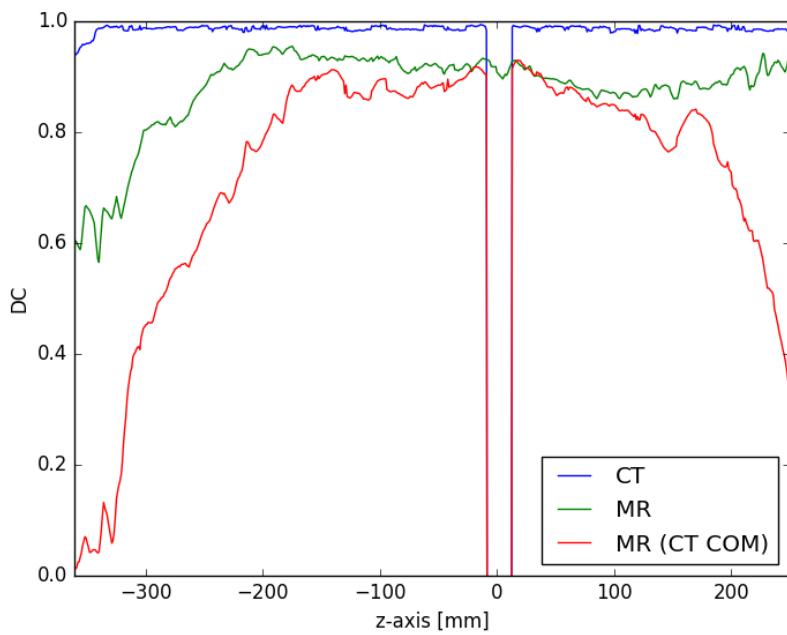


Figure 3.12: Rod #17: DC (iteration method) for CT & MRI & MRI (using CT COM)

4 Discussion

4.1 Phantom design

To measure distortion the scanners must image a rigid object with known dimensions. Such phantoms are commercially available, but often expensive and designed for a specific calibration protocol. Some institutions build their own to fulfil exactly the requirements of a given application. The scanner used at the AKH is a relatively rare model, which is why there are only few available off-the-shelf phantoms that would fit its coil.

For the AKH it was important to create a lightweight phantom which can be imaged by CT and MRI scanners. Due to the underlying physics, plastics are not visible in MRI scans. CT however, also shows plastic. See figure 2.6 for a comparison (MRI/CT visibility). Therefore, they decided on plastic rods with a suitable fluid filling. Such a liquid should be easily produced, non-toxic, and yielding sufficient signal in MRI scans.

Commercially available phantoms often resemble water filled tanks containing plastic grids as a reference. This design results in stronger signal, but exceeds practical weight. There are few brands offering solutions utilising liquid fiducial markers in the shape of pellets. They are arranged in a regular pattern surrounded by air or plastic. The AKH's design however relies on replaceable rods, which makes it a novelty.

4.1.1 observed issues

Interestingly, all water based solutions seemed to have evaporated partly. As the rod filled with liquid #15 seemed to have dried starting at the end with the plastic stopper, it is likely that the plastic stopper is not closing the rod in a way that makes them airtight. It might also be that the rod itself does not hinder volatile liquids from escaping slowly. An airtight container might have lead to other conclusions regarding the formation of air bubbles. It is hard to tell if they were caused by evaporation only or if dissolved gases played a role, too.

4.2 Tested solutions

For measuring the position of the rods in the CT scans, the plastic rods without filling would be enough already. Hollow plastic rods would not be visible on the MRI scans, though. That is why the visibility of the liquids on CT is not important at all. From now on 'signal (strength)' or 'visibility' will refer to MRI scans only (see table 3.1).

4.2.1 Thoughts about choosing possible candidates

The selection of solutions tested were chosen for a number of reasons:

1. Generally, imaging techniques aims for a high signal-to-noise ratio. Therefore, Liquids resulting in brighter pixels are favoured.
2. The amount of gas in the rods should be minimised.
3. If air bubbles form, tilting the entire phantom slightly should be enough to move them to one side. The FOW of the MRI scanner is too small to show the entire phantom anyway.
4. Most tested fluids are based on water, because this makes them easy to empty and clean. They could then be filled again with a different liquid if needed.
5. Preferably, the components which are chosen to be used for the entire phantom should be either non-toxic or harmless if not swallowed.

Rod #1 was filled with plain distilled water as reference. It was clear from the beginning that it would not result in high signal and was never considered a possible filling.

Aiming for high SNR

To achieve a better SNR, liquids #2 to #10 and #14 to #15 are based on a solution of sodium chloride ($NaCl$ concentration of 0.36 g/L) and copper(II) sulfate pentahydrate ($CuSO_4 \cdot 5H_2O$ concentration of 1.96 g/L as suggested by AAPM MR Subcommittee [22]; #3 and #4 contain double and ten-fold the concentration) in distilled water. Most of these liquids resulted in an about 5 times brighter signal than plain distilled water. Regarding the toxicity of $CuSO_4 \cdot 5H_2O$, the minimum dose to have caused acute toxic effects in humans is reported to be $11mg/Kg$. As the concentration of $CuSO_4 \cdot 5H_2O$ is only 1.96 g/L , even a young patient of 4 years and $15kg$ would need to drink more than

80mL to reach that threshold. Also, ingestion usually leads to vomiting triggered by its irritating effect on the gastrointestinal tract, which would stop further (more severe) toxic effects from happening. Keeping the scanner bed clean and checking the phantom for leakage before and after use should be sufficient to prevent an intoxication.

Primovist (#11 to #13) is a common contrast agent used for MRI scans [60, 49, 41] intended to yield a strong signal.

Handling dissolved gas

Unfortunately, dissolved gases may eventually leave the liquid and form air bubbles.

To improve the mobility of trapped air bubbles, generic washing-up soap was added (#5, #6, and #7; suggestion by Data Spectrum Corporation [12]). The higher concentrations of soap were tested as reference. If the liquid should happen to leak from the rod, the relatively low concentration of soap would not add to its toxicity. For those reasons, and because the liquid is cheap and easy to produce, it appears to be a promising candidate.

Liquids #8 (0.36g/L), #9 (3.6g/L), and #10 (36g/L) contain ascorbic acid. Adding this is supposed to reduce forming of air bubbles by binding dissolved oxygen and eventually degrade to dehydro-ascorbic acid and water. The amount suggested by [2, 5] is 0.00204 mol/L which corresponds to approx. (0.36g/L).

In an attempt to limit the forming of gas, *agar* was used in solutions #14 and #15. Agar and agarose are commonly used as basic reference material for MRI phantoms [7, 29]

Non water-based liquids

As an alternative 2 oils were proposed. Since air is not soluble in oil, once a rod is completely with oil, no air bubbles should form. Yet, oil is not as easily removed from a rod as a water based liquid. At the same time, it might not be necessary to ever replace the oil. Once filled, the rods could be used until the surrounding plastic breaks or starts leaking. Using vegetable oil would be a non-toxic solution, but has been ruled out as

a filling from the beginning, because it would eventually rot. Mineral oil on the other hand does not rot, however, it might be dangerous if not handled properly as it might leave toxic traces on the MRI scanner bed.

4.2.2 Choosing a promising candidate

As all rods containing water-based liquids continued to lose liquids due to evaporation, only the early forming of air bubbles might indicate whether solutions effectively hinder dissolved gases to result in trapped air bubbles. Apart from the solutions containing ascorbic acid (#8 (0.36g/L), #9 (3.6g/L)) all water based liquids produced some air bubbles after at least two days (see table 3.2.2). Considering the low visibility of #9, the only suitable water based liquid capable of staying free from air bubbles might be #8. At the same time, long-term observations might have shown that even #8 only delays the formation.

The reason why the rod containing the highest concentration of ascorbic acid showed an unexpected colouring is not known. Whether this is due to a contamination or a unforeseen chemical reaction involving the high concentration of ascorbic acid is unclear. As the high concentration in #9 and #10 led to a radical reduction in signal with the tested MRI sequence, this solution is not considered a suitable filling anyway.

Primovist (#11 to #13) lead to a good signal, but its limited mobility of air bubbles rules it out as a candidate.

If the forming of a small amount of gas is not considered a problem, adding soap appears to be a reasonable solution. The smallest tested amount of soap (#5, 1 g/L) was already enough to result in sufficient mobility of air bubbles, and an even lower concentration might also be acceptable. Interestingly, the rod filled with this solution contained the least amount of gas after 2 months, but this might be because the particular rod closed better than the others. The visibility recorded was among the higher candidates, too. For those reasons, and because the liquid is cheap and easy to produce, it is a promising candidate.

Solutions containing agar (#14 and #15) are even harder to remove from rods if not impossible. As they might lead to air bubbles, too, which cannot be moved to either side of the rod, agar is not suited for this phantom.

Finally, the generic motor (#16) oil resulted in the highest signal intensity of all candidates. Besides the question of its toxicity, it seems to be a good alternative to water based liquids. The silicon oil (#17) on, the other side, had a low signal compared

to most candidates and is therefore not suited.

4.3 Sequence

(TODO: why T1 weighted sequence? why this particular sequence? Influence on signal of water/oil, components.)

4.4 Distortion

4.4.1 Irregularities

In the case of a slice located at a plastic pane, the CT image will be mostly very bright, as the plastic results in a high intensity. Consequently, this slice will be marked as irregular as the average brightness differs greatly from the reference slice's. However, the MRI scan will not show such a increase in brightness at that location, as plastic is not visible for the MRI scanner. Here the MRI data will not be marked as irregular, provided there is no other cause for a change in brightness, like an air bubble. Such an absence of liquid would result in a drastic decrease of brightness and result in an irregular slice. In CT, on the other hand, an air bubble would hardly make a difference, as the surrounding plastic is far brighter than the contained liquid.

4.4.2 COM calculation

DC values calculated with the COM iteration method do not exceed those calculated with method 1. However, both methods choose very similar thresholds for their calculations. This is good, as the iteration method does not rely on additional information (true dimensions, radii of rods), but yields similar results. The iteration method might yield better results if tweaked slightly. At the moment it does not take into account the steepness of the DC curve. This leads to the left hand side close to very low percentages to be neglected sometimes. This happens when the perfect guess lies roughly in the middle of the current range, but a little bit to the left. Because the slope is steeper on the left (close to 0%), a value representing the left hand side (also close to 0%) yields a much lower result than the value on the right hand side (flat slope). This should be taken into account during further improvement of the method.

To get a idea of the occurring distortion one should look at both the absolute value of coordinate shift and the dice-coefficient (DC). An air bubble might lead to a incorrect COM. Without looking at the dice-coefficient it's hard to tell why this distortion only appears to be present in a few slices. If both indicators show unexpected local irregularities, a conclusion might be easier to draw.

(TODO: discuss what is visible on the graphs in results section) (TODO: interpret data table spit out by script)

5 Conclusion and Outlook

5.1 Liquid for phantom

As topping up over 300 rods regularly is too time consuming, and all water-based liquids continued to evaporate, they are not long-term solution for this phantom. For short time experiments with this phantom (not airtight rods) #5 is recommended, because the soap allows the air to be moved out of the FOW. If a new set of (airtight) rods was obtained, adding ascorbic acid to the solution (a combination of #5 and #8) might be an even better filling:

- distilled water
- 0.36 g/L of NaCl
- 1.96 g/L of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$
- 1 g/L of soap
- 0.36 g/L of ascorbic acid

In that case, the forming of air bubbles might be either avoided entirely (due to the ascorbic acid) or delayed and then easily taken care of by tilting the whole phantom slightly to move the bubbles out of the FOW. The long-term behaviour of the mix might lead to adverse properties and should be tested, though.

In the current set-up (not airtight rods), it seems best to fill the rods of the phantom type of oil that does not rot, but yields high signal. The tested generic motor oil (#16) fits those requirements. However, a non toxic alternative would be preferred.

5.2 Future improvement of software tool

The developed tool took a simplified approach to the problem by looking only at a single rod. Future improvements should enable the software to take into account all the rods automatically. This could be done by a auto-trace function which detects individual rods and applies the already implemented algorithms to each of them separately.

Distortion assessment can be performed using the generated tool, however, it needs to be implemented on more general scale to be able to measure the distortion for all the rods automatically

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