

Immunopathogenesis of Rheumatoid Arthritis

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<http://dx.doi.org/10.1016/j.immuni.2017.02.006>

Rheumatoid arthritis (RA) is the most common inflammatory arthropathy. The majority of evidence, derived from genetics, tissue analyses, models, and clinical studies, points to an immune-mediated etiology associated with stromal tissue dysregulation that together propagate chronic inflammation and articular destruction. A pre-RA phase lasting months to years may be characterized by the presence of circulating autoantibodies, increasing concentration and range of inflammatory cytokines and chemokines, and altered metabolism. Clinical disease onset comprises synovitis and systemic comorbidities affecting the vasculature, metabolism, and bone. Targeted immune therapeutics and aggressive treatment strategies have substantially improved clinical outcomes and informed pathogenetic understanding, but no cure as yet exists. Herein we review recent data that support intriguing models of disease pathogenesis. They allude to the possibility of restoration of immunologic homeostasis and thus a state of tolerance associated with drug-free remission. This target represents a bold vision for the future of RA therapeutics.

Immune dysregulation was first implicated in the pathogenesis of rheumatoid arthritis (RA) by the discovery of anti-immunoglobulin G (IgG) antibodies known as rheumatoid factors, initially by Erik Waaler and then more fully described by H.M. Rose in the 1940s. However, concepts of how immune responses contribute to disease have evolved dramatically over the last 50 years. Autoreactivity as a pivotal step dominates the conceptual landscape, although other mechanisms, both immunologic and tissue-derived, clearly contribute to disease pathogenesis. Thus, RA is characterized by evidence of disordered innate immunity, including immune complex-mediated complement activation, adaptive immune responses against “self”-antigens comprising predominantly post-translationally modified proteins, dysregulated cytokine networks, osteoclast and chondrocyte activation, and imprinting of resident stromal cells that in turn develop semi-autonomous features that support disease progression (Arend and Firestein, 2012; Firestein, 2003).

Based on substantial new data identifying the immunologic and metabolic events that precede onset of clinical disease, even by years (i.e., “pre-RA”), and the increasing impact of the application of the advanced molecular “omics” revolution to disease investigation, an integrated hypothesis for disease pathology is emerging (Catrina et al., 2016; Tan and Smolen, 2016). In this review, we will posit that RA starts with a high-risk genetic background that, in combination with epigenomic marks that contribute to heritability and disease chronicity and stochastic environmental exposures that create neo-epitopes, launches a cascade of events inducing synovitis and ultimately chronic destructive arthritis (Figure 1). Based on the diversity of clinical responses to highly targeted therapeutic agents, we propose that RA probably does not comprise a single entity. Rather, it is perhaps more appropriately considered a syndrome with a common clinical phenotype arising from diverse pathways operating variably, albeit often with overlap, in individual patients (Firestein, 2014).

Genetics of Rheumatoid Arthritis

Genetic factors clearly play a role in RA risk, severity, and progression. Monozygotic twins share RA on about 12%–15% of occasions compared to 1% for the general population and around 2%–5% for fraternal twins or other first-degree relatives. This relatively low concordance implicates many other factors, including those in the environment and the microbiome in pathogenesis. Note also that gene sequences are not the sole determinant of heritability, and epigenetic marks probably also contribute, especially for monozygotic twins (Kaminsky et al., 2009).

The most important genetic risk allele for RA resides in the class II major histocompatibility (MHC) locus, accounting for about 40% of the genetic influence. The odds ratio of developing RA in individuals with MHC class II HLA-DR4 alleles is about 5:1. This link between HLA-DR and RA was initially described in the 1970s with the observation that HLA-DR4 is present in 70% of RA patients, compared with about 30% of controls. A so-called shared “susceptibility epitope” (SE) was identified in amino acids 70 through 74 in the third hypervariable region of the DR β chain. The sequence associated with disease is generally glutamine-leucine-arginine-alanine-alanine (QKRAA), which is present in some DR4 and DR14, in addition to DR1 β chains. The SE with the closest links to RA include DRB*0401, DRB*0404, DRB*0101, and DRB*1402. More than 90% of patients with RA express at least one of these variants (Weyand et al., 1992). The SE is also associated with increased disease severity, such as extra-articular manifestations and progression of erosions.

The SE region predominantly faces away from the antigen-binding groove that binds processed peptides for presentation to T cells, which has raised some questions about their precise contributory role (Firestein and Zvaifler, 2002). RA-specific peptides that bind to QKRAA-containing molecules have been difficult to identify (Kirschmann et al., 1995). This observation led to the notion that SE might also partially contribute by shaping the

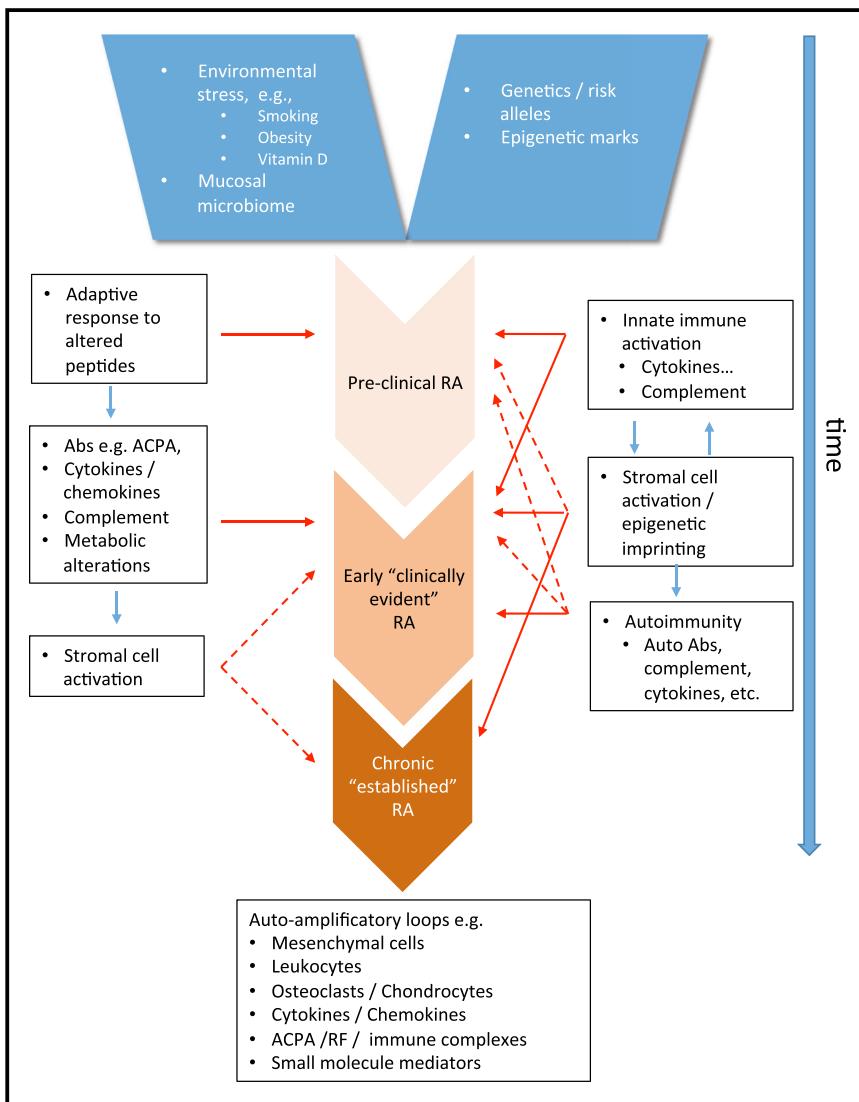


Figure 1. Sequence of Events Leading to the Development of Clinically Detectable Rheumatoid Arthritis: Two Potential Models

In model A (left), a pre-RA phase comprises early generation of autoantibodies (ACPAs) that can bind post-translationally modified self-proteins, particularly via citrullination. This is followed by amplification of the range of specificities of ACPA and by the elaboration of cytokines and chemokines, complement, and metabolic disturbance in the months prior to clinical development of disease. A transition event that requires a “second hit” (as yet poorly understood) permits the development of synovitis. The latter is characterized by frank inflammation, stromal compartment changes, and tissue modification leading to articular damage. In model B (right), which is not mutually exclusive, there is an early interaction between innate immune activation and stromal factors that lead to stromal cell alteration, including epigenetic modifications that initiate a cycle of inflammatory stromal-mediated damage. Autoimmunity can arise as a result of these interactions that in turn can contribute directly or in an amplification loop to disease perpetuation.

citrullinated peptides are also characterized by the presence of “anti-citrullinated peptide antibodies” (ACPAs), observed in 80%–90% of RA patients.

Together these data support the notion that HLA-DR risk for RA is based at least in part on the increased efficiency of antigen presentation for *altered* peptides rather than native proteins (van der Helm-van Mil et al., 2006). Citrullination of peptides in the presence of environmental stress is ubiquitous in mammalian cells and is not a unique feature of RA. Instead, production of antibodies recognizing citrullinated peptides differentiates individuals at risk. The emergence of numerous other post-transla-

T cell repertoire in the thymus, altering intracellular HLA-DR trafficking and antigen loading, or serving as an autoantigen.

More recently, linkage disequilibrium and informatics-based studies to understand the role of the SE identified additional amino acids located in the base of the antigen binding groove as a likely explanation for antigen selection, especially leucine or valine variants at amino acid 11 (Raychaudhuri et al., 2012). Serine at amino acid 11 confers decreased risk for RA. The risk alleles, especially deep within the cleft, could directly increase risk via presentation of arthritogenic antigens, such as citrullinated proteins. For example, citrullinated vimentin peptides bind much more avidly to the RA HLA-DR than does native protein; in contrast, other MHCs do not exhibit this differential binding (Hill et al., 2003; Law et al., 2012). Subsequent crystal studies have characterized in detail the nature of the citrullinated peptide binding to HLABR1 (Scally et al., 2013). RA-associated alleles present citrullinated peptides efficiently to T cells, which, in turn, produce higher amounts of cytokines IL-17 and IFN- γ than to native peptide. Adaptive immune responses to

tionally modified protein targets, e.g., via carbamylation or acetylation, recognized by autoantibodies in RA is consistent with the notion of altered presentation of post-translationally modified peptides; other families of altered peptides could be implicated in discrete subsets of patients.

Overall, genome-wide association studies (GWASs) and meta-genomic analyses have identified more than 100 single-nucleotide polymorphisms (SNPs) and genes associated with ACPA⁺ RA beyond the HLA. RA-associated SNPs tend to cluster around immune genes (Okada et al., 2014). The genetic architecture of RA has recently been extensively reviewed (Kim et al., 2017). In brief, individual SNPs usually provide modest contribution to risk with odds ratios typically in the 1.05- to 1.2-fold range, though this need not infer low functional impact. Combinations of these genes can potentially interact to increase risk. For example, a relatively rare combination of *HLA-DR*, *PTPN22*, and *TRAF1-C5* SNPs increases risk more than 40-fold. While there are undoubtedly additional combinations or rare variants with high penetrance, the contribution of newly

identified genes to overall risk is growing increasingly small. Notably, meta-analyses of patients with ACPA⁻ RA show some differences from ACPA⁺ disease, consistent with the idea that “seronegative RA” should be considered a distinct clinical pathologic entity.

Intriguing individual non-MHC linkages are associated with peptidyl arginase deiminase (*PADI*) and *PTPN22*. *PADI* gene products are enzymes that convert arginine to citrulline, creating new potential antigens that bind to RA-associated MHC proteins. An extended *PADI4* haplotype prolongs its mRNA stability and could, therefore, increase peptide citrullination (Suzuki et al., 2003). Up to a 2-fold increase in risk of RA was observed with *PADI4* SNPs in Asian and North American cohorts. The *PTPN22* allele with an amino acid substitution (R620W) doubles the risk of developing ACPA⁺ but not ACPA⁻ RA. This SNP is also associated with many other immune-mediated diseases, including systemic lupus erythematosus (SLE) and type 1 diabetes (Rawlings et al., 2015). The R620W allele is a gain-of-function change mutation in the *PTPN22* protein, a phosphatase that regulates (among other signal proteins) Lck and ZAP70. Thus, modified T cell receptor signaling could alter either the T cell repertoire in the thymus or immune responses, or tolerance in the periphery. The R620W allele, however, also positively regulates TLR-induced type 1 IFN expression in myeloid cells via reduced efficiency of *PTPN22W*-mediated TRAF3 ubiquitination and thus could amplify innate cytokine expression to promote disease (Wang et al., 2013).

A number of genes involved with adaptive immunity have RA-associated SNPs in a various populations (Yamamoto et al., 2015). In addition to *PTPN22*, the co-stimulation receptors CTLA and CD28 have emerged from GWASs. Other genes associated with B cell function and/or antigen presentation such as *BTLA* (B and T lymphocyte attenuator), Fc receptors, and *CD40* have been identified. Signal transduction genes and pathways that regulate immune function such as *TRAF1-C5* and *STAT4*, cell migration (*ELMO1*), and fetal development (*LBH*) are also identified. Finally, multiple gene polymorphisms have been identified in cytokine or cytokine receptor coding loci, including TNF. This is especially relevant in light of the pivotal role of cytokines in synovitis described below and the consequent efficacy of targeted anti-cytokine, and cytokine receptor signal pathway therapeutics. In some cases, the causative mutations can alter expression or function. For instance, RA-associated TNF promoter polymorphisms at positions –238 and –308 can potentially modulate TNF gene expression (Fonseca et al., 2007). An IL-6 receptor polymorphism that is functionally implicated in the intensity of IL-6 signaling is also associated with RA (Ferreira et al., 2013). Thus, the genetic clues to RA pathogenesis implicate inflammation and adaptive and innate immunity at the core of pathogenesis.

Interactions between Genes and Environment: RA as a “Mucosal Disease”?

RA is properly considered an immune-mediated disease with a strong genetic influence. However, its origins may involve the interface between external influences and the immune system, manifest especially at mucosal surfaces. Three sites have been associated particularly with RA, namely (1) the lungs, (2) the oral mucosa, and (3) the gastrointestinal tract. Whereas precise

mechanisms that enhance risk are not fully understood for each, it is likely that local tissue stress leads to post-translational modification of peptides with subsequent antibody formation serving as a common mechanism.

Epidemiologic/genetic studies, particularly those arising from impressive Swedish registries, have highlighted the role of cigarette smoking as a prominent environmental risk factor for RA, pointing to a role for pulmonary mucosal biology in disease etiology. The amount and duration of smoking are important, with the highest risk observed with greater than 20 pack-years of exposure. Risk declines slowly back to normal within 10 years of smoking cessation (Källberg et al., 2011), suggesting that effects are not solely toxin dependent but may also arise by altering immunologic function. Notably, other pulmonary exposures can replace smoking in increasing risk, e.g., silica or textile dust. Nevertheless, mechanisms underlying the relationship with smoking are uncertain (Catrina et al., 2014). Exposure to inhaled toxic chemicals in cigarette smoke can potentially increase *PADI* expression in the airway and increase protein citrullination (Vassallo et al., 2014). Smoking-related risk interacts in a synergistic manner with MHC risk alleles. The QKRAA-containing HLA chains alone modestly increase the likelihood of developing RA 4- to 6-fold, but this risk increases up to 20- to 40-fold when combined with smoking (Lundström et al., 2009).

Thus in individuals carrying risk HLA alleles, such peptides could be more efficiently presented and ACPAs generated via subsequent T-B cell help. Alternatively, “innate” B cell subsets could initiate responses to modified self-peptides, and thereafter present peptide to T cells. In this model, production of “autoantibodies” could result from “auto-reactive” T cell clones that are available to respond to modified peptides because they were not deleted during development since the altered proteins/peptides were not present in the thymus (Linn-Rasker et al., 2006). A corollary to this would be that a major immunologic regulatory deficit in RA patients is a failure to regulate T cell recognition of post-translationally modified self-proteins across a range of specificities—as observed above, this is compatible with the expression of a relatively broad autoantibody spectrum in clinical disease.

Broncho-alveolar immunohistology shows high levels of citrullinated proteins in pulmonary macrophages and smoking increases such citrullinated proteins in broncho-alveolar lavage fluid (Makrygiannakis et al., 2008). However, citrullinated proteins can also be present in the context of other diseases or even in clinically normal lungs (Catrina et al., 2014). Thus, risk from smoking might be more complex than via *PADI* induction and may include promotion of epigenetic alterations, notably DNA methylation with inflammatory potential (Tsaprouni et al., 2014). Perhaps the location of citrullination and the stochastic combination of altered peptides, innate leukocyte subsets, and mucosal inflammation conspire a permissive microenvironment for the creation of ACPAs. High-resolution CT studies demonstrate associations between ACPA formation and bronchial thickening in the pre-RA stages and support the idea that inflammatory responses are required (Reynisdottir et al., 2014). More recent studies have shown signs of local inflammation in bronchial tissues of untreated RA patients (Reynisdottir et al., 2016). Using B cell barcoding techniques, the frequency of circulating IgA plasmablasts was found to be elevated in at-risk subjects without RA who are ACPA positive, providing further

evidence for a mucosal origin for at least a proportion of the autoimmunity detected in RA (Kinslow et al., 2016).

The relationship between the microbiome and RA has been suspected for many years and supported by pre-clinical studies. For example, arthritis susceptibility and severity in a variety of rodent strains is decreased when maintained germ-free or in environments with a restricted bacterial flora (Liu et al., 2016). These data suggest that bacteria provide complex adjuvant functions that enhance autoimmunity, either directly (e.g., bacterial cell walls or lipopolysaccharides) or by critically altering the immunoregulatory mucosal environment. The latter could support particularly the generation of type 17 responses implicated in several animal models of RA (Miossec and Kolls, 2012).

In humans, inflammation of the oral mucosa, particularly periodontitis, is associated with increased susceptibility to RA. The most common bacteria implicated in periodontitis is *P. gingivalis*, since it can express PADI. *P. gingivalis* could potentially citrullinate peptides in the oral mucosa that, in the context of inflammation, could promote ACPA generation (Wegner et al., 2010). While this concept is attractive, a clear association between *P. gingivalis* compared other bacteria is not consistently observed in unbiased microbiome studies. More recent data suggest that *P. gingivalis* or other bacteria could act, instead, by ligating Toll-like receptor 2 (TLR2) (de Aquino et al., 2014). This process could increase IL-1 production and stimulate local Th17 cell differentiation. An intriguing recent study identified that *A. actinomycetemcomitans* initiates hypercitrullination through the effect of a toxin, leukotoxin A (LtxA), mediated on neutrophils and was detected in RA patients' oral microbiome wherein it could act as a bacterial trigger to disease (Konig et al., 2016a).

In the colon, *Prevotella copri* species were enriched in an unbiased analysis of the gastrointestinal microbiome in early RA patients. *Bacteroides* species were decreased in the same patients (Scher et al., 2013). While not fully understood, the overrepresentation of *Prevotella* was not observed in chronic RA or in other forms of arthritis. Subsequent studies in the Chinese population have extended these observations to include novel microbial species and demonstrated common expression in oral and GI mucosal sites in individuals that, remarkably, was correlated with ACPA levels and CRP. Moreover, further alterations in the microbiome occurred upon therapeutic immune modification (Zhang et al., 2015).

Epigenetics: Integrating Environmental Stress and the Genome

Epigenetic modifications decorate genomic DNA and contribute to regulation of gene expression. The role of these marks had been widely viewed as key determinants of orderly gene expression during development as well as lineage-dependent RNA transcription throughout life. Their contribution to pathogenesis is now more clearly understood in many diseases, especially cancer. A variety of epigenetic mechanisms have been implicated in RA, including DNA methylation, histone modification, and microRNA expression. Because the science of epigenetics in autoimmunity is still relatively young, it is uncertain whether some or all of these disease-specific alterations are inherited, develop during life due to stochastic exposures, cause disease, or are actually caused by disease. Regardless, stable epigenetic

marks have been identified in RA that alter cell function and permanently imprint some lineages, most notably synovial fibroblast-like synoviocytes (FLS).

Most information on epigenetics in RA comes from studies of RA joint samples, including synovial tissue and FLS. Candidate gene approaches showed that the *IL6*, *CXCL12*, and *EFNB1* promoters are hypomethylated in rheumatoid FLS while others, like *DR3*, *EBF2*, and *IRX1*, are hypermethylated (Karouzakis et al., 2011). Genome-wide approaches have identified a large number of differentially methylated loci in RA FLS compared with normal or osteoarthritis (OA) FLS (Nakano et al., 2013a). The methylation pattern in RA FLS is stable over many cell passages in tissue culture and involves genes and pathways related to cell adhesion, matrix regulation, immune function, and cytokine signaling. While synovial cytokines can transiently alter the methylome by regulating DNA methyl transferase (DNMT) expression, the changes are reversible in culture and suggest that cytokines alone are not solely responsible for the ex vivo profile observed (Nakano et al., 2013b). The cells, therefore, appear to be "imprinted," which could contribute particularly to the persistent aggressive phenotype of RA FLS.

The functional relevance of differential methylation was confirmed for several candidate genes identified via integrative analysis of various "omics" platforms. For example, the development gene *LBH* was identified because it has several RA-associated SNPs and differentially methylated loci (DMLs) (Hamaker et al., 2016). *LBH* regulates the cell cycle—a functional SNP in an *LBH* enhancer was identified adjacent to a DML in FLS, and modification of the enhancer methylation status altered *LBH* transcription (Ekwall et al., 2015). In addition, a DML in *PTPN11*, the gene encoding the oncogene SHP2, led to discovery of a novel intron enhancer that regulates *PTPN11* expression and contributes to high levels of SHP2 in RA FLS (Zhang et al., 2015). In future, incorporating epigenetic marks into the systematic analysis of cell function should help identify key pathogenic genes and provides a rich source of potential therapeutic targets.

We still do not know when in the evolution of RA the epigenetic imprinting occurs. Recent data in FLS from patients with early RA (<1 year of symptoms) show that they also have an abnormal methylation pattern. However, the early RA methylome can be distinguished from late RA, especially for pathways related to cell growth and differentiation (Ai et al., 2015). DNA methylation signatures in RA FLS probably occur very early or even antedate clinical disease. However, it is not static and can evolve over time due to synovial inflammatory environment.

Perhaps even more intriguing, DNA methylation patterns, as well as transcriptomes, can vary between joints (Ai et al., 2016). Early studies using histology and immunohistochemistry suggested that the various joints involved with RA are similar. However, recent studies show that the DNA methylation patterns and transcriptomes differ between RA hip and knee FLS (Ai et al., 2016). Of interest, the pathways involved are related to therapeutic targets, including IL-6-JAK-STAT signaling. It is not clear whether cells are imprinted elsewhere and then migrate to the joint or whether they attain a local phenotype after arriving. These data provide a possible explanation of why some joints improve in RA while others do not when patients are treated with a given targeted agent.

Sequencing studies suggest that the pattern of microRNA (mir) in RA compared with OA leukocytes and FLS exhibit global differences in expression. A growing number of mirs have been evaluated using a candidate gene approach. One example is mir124a, which is dysregulated in RA and can suppress cell cycle and chemokine gene expression. Increasing mir124a levels in RA FLS suppresses MCP-1 production (Nakamachi et al., 2009). On the other hand, mir203 overexpression increases MMP and IL-6 expression (Stanczyk et al., 2011), while mir155 knockdown decreases the same gene expression in RA FLS. mir155 is also implicated in monocyte activation and cytokine production in part via regulation of TLR signaling. mir16, mir124a, mir202, mir134a, mir23, mir346, and mir15a (and perhaps several others) are differentially expressed in RA-derived cells, although their pathogenic role still needs to be defined (Bottini and Firestein, 2013a).

There is considerably less genome-wide information on histone marks in RA. There are a variety of ways that histones can be modified, including methylation, acetylation, and citrullination. Histone deacetylases, which remove acetyl groups and remodel chromatin, are expressed in RA FLS and can regulate their function (Huber et al., 2007). For example, induced HDAC1 or 2 deficiency increases p53 tumor suppressor gene and matrix metalloproteinase 1 (MMP1) expression in RA synovial cells (Horiuchi et al., 2009). Pro-inflammatory cytokines can increase HDAC1 expression in FLS and contribute to chromatin remodeling. HDAC inhibitors suppress adjuvant arthritis in rats and collagen-induced arthritis in mice, supporting the notion that histone modifications can participate in inflammatory joint disease (Joosten et al., 2011).

Pre-RA: An Evolving Immunologic Response?

As noted before, protein modification through citrullination is actually common, especially in the context of inflammation or stress. What is strikingly different in RA is the generation of antibodies against these epitopes, likely arising due to the unique properties of the binding pocket of RA-associated HLA-DR alleles and presumably other regulatory deficits (e.g., in T cells themselves) that permit emergence of autoreactivity over time. The teleologic explanation for induction of PADIs in normal immune responses is not known. PADIs have other potential functions that regulate cell migration and gene expression through citrullination of chemokines and histones (Christophorou et al., 2014). PADI2, for example, migrates to the nucleus and probably modifies many proteins involved in gene expression. Thus, protein citrullination could have a variety of functions that regulate homeostasis and cell responses to stress.

The presence of citrullinated peptides in the lungs or other mucosal sites could thus represent the “original sin” in a pathway that ultimately leads to inflammatory joint disease. ACPAs, along with other antibody systems, are detected years or perhaps even decades before clinical disease is apparent. Over time, the serum levels of ACPAs gradually increase and reach a peak at the time of disease onset through the first year of symptoms. A concomitant rise in blood chemokines and cytokines and other altered metabolic parameters suggest an ongoing and gradually accelerating subclinical systemic inflammatory pathology (Deane et al., 2010). ACPA specificities at all stages are quite broad and increase during the “pre-RA phase.”

The specificity of antibodies varies widely but include citrullinated epitopes on fibrinogen, vimentin, fibronectin, collagen, enolase, histones, and many others (Anzilotti et al., 2006). There does not appear to be a single dominant specificity of ACPA, nor a particular pattern that predicts disease onset, phenotype, or severity (van Beers et al., 2013). Notably, in patients with early undifferentiated inflammatory arthritis, ACPAs can predict individuals who will progress to RA.

In established disease, ACPAs are present in the serum of 80% to 90% of RA patients and are independently associated with greater disease activity, extra-articular manifestations, and joint damage. The antibodies alone are not especially pathogenic and do not cause arthritis when injected into normal mice. However, they can exacerbate disease in mice with arthritis, perhaps because of the presence of citrullinated substrates that can ultimately lead to local complement fixation. They likely act together with rheumatoid factors (RFs), which are autoantibodies that bind to the Fc portion of IgG. ACPAs are more sensitive and specific for RA than RFs, with specificity approaching 90%. ACPAs are also present in nearly 20% of unaffected first-degree relatives and more than 10% of more distant relatives in Native American RA patients (El-Gabalawy et al., 2009). ACPAs and RFs are also produced by synovial tissue B cells and can be detected in synovial fluid, suggesting that they could play a role in local innate immunity activation and complement fixation. Evaluation of the phylogenetic tree of RF-producing cells from the synovium suggests that affinity maturation occurs in the joint (Randen et al., 1992).

A subset of patients with RA develops antibodies against other classes of modified proteins. For example, lysine and cyanate are converted to homocitrulline and can form neo-epitopes through non-enzymatic processes analogous to conversion of arginine to citrulline, giving rise to anti-carbamylated protein antibodies (ACarPs) present in a minority of patients with RA, including about 30% of ACPA⁻ patients (Shi et al., 2011). A variety of substrates are carbamylated and can lead to antibody formation with multiple specificities, such as fibronectin. As with ACPAs, they are observed about 40% of RA patients prior to the onset of clinical synovitis (Shi et al., 2014). Like ACPAs, the ACarPs do not cause arthritis, but immunizing mice with carbamylated peptides is arthritogenic, and transferring lymphocytes from immunized animals can transfer disease to naive mice (Mydel et al., 2010). Autoantibodies that identify acetylated proteins have also been described (Juarez et al., 2016). Finally, there are data that challenge the notion of loss of tolerance to citrullination as a definitive early event—e.g., anti-native RA33 and citrullinated RA33 were detected as a continuum in a recent analysis with native reactivity dominant in earlier disease (Konig et al., 2016b). Further sequential studies of detailed antibody profiles against the spectrum of the “citrullome” in RA would be helpful in resolving these issues.

Evolution from a High-Risk State to Clinical Synovitis

The foregoing demonstrates that in high-risk individuals with a characteristic genetic (and possibly epigenetic) background, stochastic events especially at mucosal surfaces can lead to protein modification and ultimately antibody formation recognizing altered peptides. The site of antibody production could be either at the site of stress or in lymphoid organs defined by

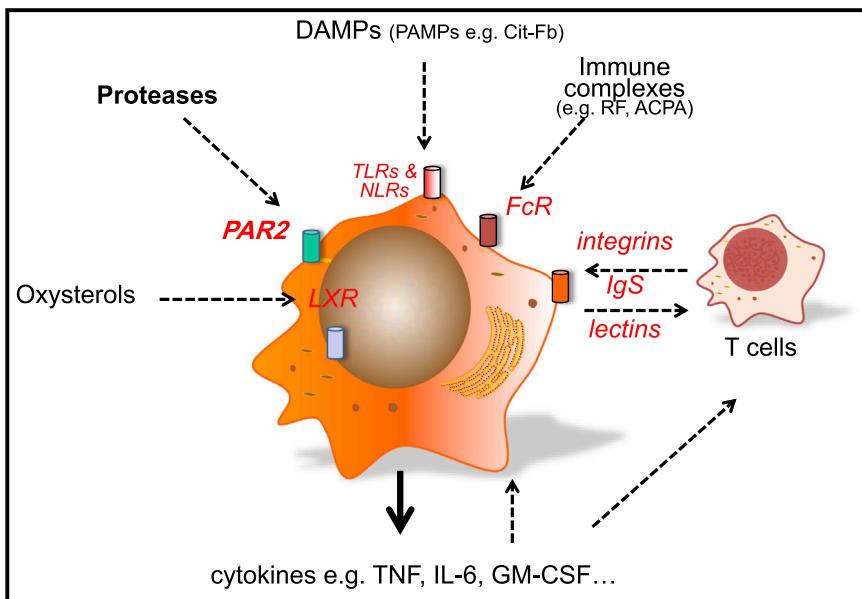


Figure 2. Myeloid Cells, Particularly Macrophages, Are Central to the Perpetuation of Disease Pathogenesis

Pathways that can activate macrophages in RA synovitis are shown that promote cytokine release—IL-6 and TNF in particular, and probably GM-CSF, are of central importance as defined by the clear therapeutic benefits achieved upon their inhibition.

dendritic cells that process arthritogenic peptide at mucosal surfaces and migrate centrally to relevant draining lymph nodes. Lymph nodes in pre- and early RA are not normal (Rodríguez-Carrio et al., 2016). By definition, however, disease emerges in the joints. That ACPAs, rheumatoid RFs, ACarPs, and perhaps other antibody systems do not typically lead to inflammatory joint disease per se suggests that a “second hit” is required, at least in model systems for tissue localization.

The nature of this second signal is not well defined but could comprise some combination of vascular, neuro-regulatory, microtrauma, or transient infection-dependent pathways. In addition, there have long been suggestions that articular infections, e.g., Epstein-Barr virus, could further contribute (Tan and Smolen, 2016). In animal models, the formation of immune complexes can trigger synovitis. The inciting complexes do not have to be specific for altered peptides but can actually be irrelevant; even immunocomplexes with antibody and horseradish peroxidase is sufficient in some mouse models. Thus, a variety of random tissue insults (e.g., microtrauma, viral infection) could lead to synovial vascular activation in the presence of the requisite size and charge of complexes that permits Fc receptor engagement in the synovium. Triggered synovial innate cells like mast cells could release vasoactive mediators and increase antibody access to the joint. Complement activation across this range of effector pathways could act to accelerate this response. Other mechanisms like Toll-like receptor engagement or triggering via various danger-sensing molecules could theoretically cause a similar cascade. Recent data suggest a potential role for ACPAs beyond immune complexes depending upon their chemical properties and cellular specificity. Intriguing studies suggest that ACPA recognition of osteoclast (OC) membrane epitopes, e.g., citrullinated vimentin, can promote osteoclastogenesis—thus an early route to articular involvement via systemic ACPA expression may be via local OC activation (Harre et al., 2012). Moreover, one consequence of ACPA OC binding is the induction of pain and release of IL-8,

providing an enticing route to subsequent leukocyte recruitment and the initiation of a vicious cycle of articular inflammation (Krishnamurthy et al., 2016). In addition, ACPAs most likely to progress disease exhibit an altered glycosylation state (Rombouts et al., 2015), though the functional consequences of these observations remains less well understood.

Once in the joint in established disease, autoantibodies may bind to antigen, fix complement, release

chemotactic fragments, and initiate a cascade of events that activates other resident cells, recruits new innate and adaptive immune cells, and promotes stromal cell activation (Tan and Smolen, 2016). These, in turn, can produce additional cytokines and chemokines to create a positive feedback loop and ultimately a self-perpetuating process with inadequate negative regulators required for termination (Figures 2 and 3). The joint is uniquely susceptible to promote such a series of events. For example, the vasculature is already porous and facilitates ingress of proteins and cells. The synovial intimal lining, which forms the surface of the synovium and contains macrophage-like and fibroblast-like synoviocytes (FLS), is not an effective barrier because there are no tight junctions or an organized basement membrane. Once in the joint, antibodies can be displayed on cartilage to promote complement fixation. Murine models involving antibodies to type II collagen or glucose-6-phosphate isomerase are particularly effective at inducing synovitis because of their propensity to bind to these articular surfaces. The former is pathogenic because it is a critical component of hyaline cartilage and the latter most likely adheres to negatively charged cartilage surfaces (Matsumoto et al., 2002).

One of the important lessons of this prolonged and stepwise pre-RA process is that the sequence of events does not require autoimmunity against native proteins in the earliest stages. The immune response could be more properly considered a normal appropriate adaptive immune response to altered antigens that are viewed as “foreign” because they were not generated during thymic development and pathogenic T cells could not be deleted. The high-risk individuals with “unfortunate” HLA-DR alleles happen to bind to these modified proteins more avidly than the native molecule. Later, perhaps through epitope spreading, more traditional autoimmunity develops and leads to production of antibodies commonly detected in chronic RA that bind to collagens and many other native proteins.

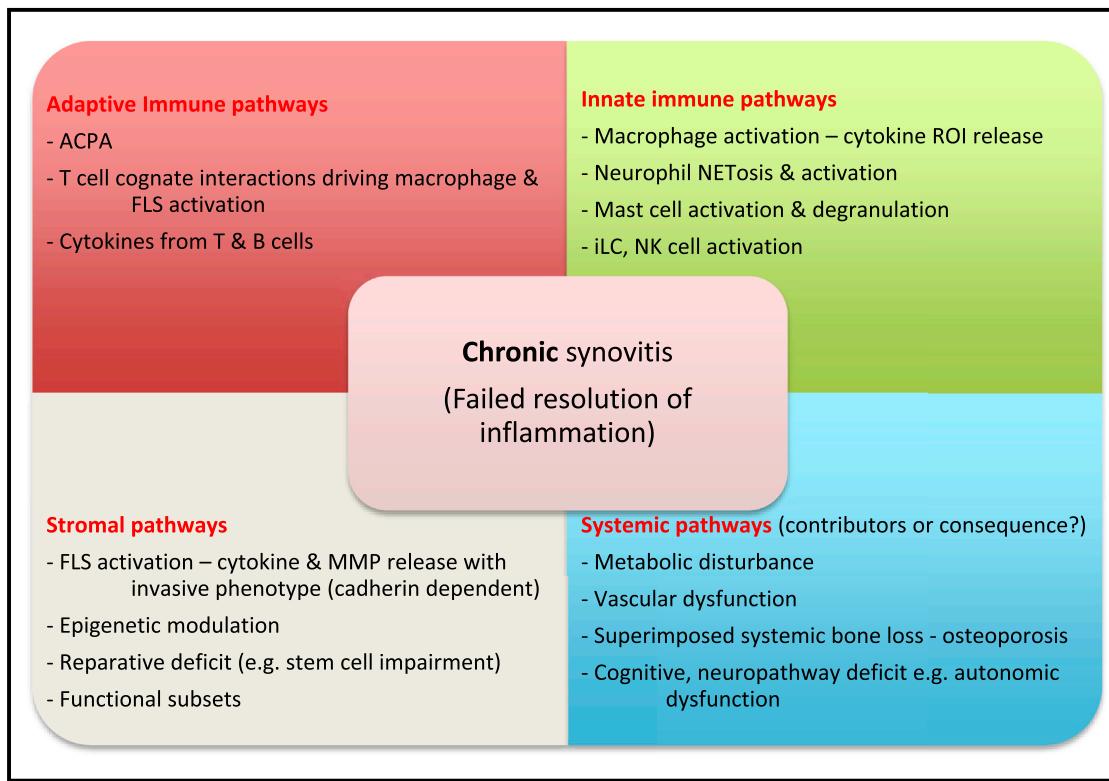


Figure 3. Mechanisms that Drive the Chronicity of RA

Distinct pathways combine to mediate failed resolution of disease—these include adaptive, innate, and stromal components with additional impact of the systemic features of disease. The latter may be both consequence of the chronic inflammation and could promote the chronicity, e.g., via creation of pro-inflammatory lipid profiles, promotion of bone remodeling, and secondary osteoarthritis.

Early Role for Stromal Cells as Important Pathogenic Factors

Compelling data implicate adaptive immune responses in the pathogenesis of RA, but alternative hypotheses propose that stromal elements can also play a critical role. Early pioneering studies by Gay and colleagues showed that FLS assume an autonomous aggressive phenotype that is maintained for months after removal from the patient (Müller-Ladner et al., 1996). These data and the destructive potential of fibroblasts led to the notion that FLS are imprinted or permanently altered in RA and could either initiate or exacerbate disease (Firestein, 1996; Bottini and Firestein, 2013b). Numerous mechanisms for “partial transformation” have been implicated, including abnormal DNA methylation (see below), somatic mutations in various genes (e.g., p53, mitochondria, vimentin), defective PTEN expression, and high levels of senescent (Pap et al., 2000; Franz et al., 2000; Firestein et al., 1997; and reviewed in Bottini and Firestein, 2013b). In addition, activated FLS can play an active role in antigen presentation and contribute to local adaptive immune responses (Tran et al., 2007). Finally, the synovial fibroblasts are not a uniform population but segregate into different phenotypes based, in part, on their cytokine profiles (Croft et al., 2016). The role of mesenchymal cells in disease initiation and progression represents an interesting therapeutic opportunity because no approved agents currently target these cells (see Figure 1 for

alternate models of contribution of stromal cells and autoimmunity in a notional hierarchy—note that these are not mutually exclusive).

Pathologic Characteristics of Clinical Synovitis

The histologic appearances of RA synovitis have been extensively characterized. Normal synovial membrane is relatively acellular, comprising scattered macrophages within a stromal tissue curated by FLS and sparse blood vessels. Its primary function is to lubricate and nourish the articular surface, which lacks a blood supply. During the pre-RA phase, the synovial appearance is not markedly altered; biopsy studies or ACPA⁺ arthralgia are remarkably benign, with no obvious leukocyte infiltrates nor transcriptional changes (de Hair et al., 2014). However, with the onset of clinically evident articular swelling (including that detected by MRI or power-Doppler ultrasound imaging), the synovium assumes a rather different appearance (reviewed by McInnes and Schett, 2011). The lining layer, normally 2–3 cells thickness, expands up to 10–12 cells depth, comprising macrophages and FLS. Underlying this is a dense interstitial cellular infiltrate containing cells characteristic of innate (e.g., macrophages, dendritic cells, mast cells, natural killer [NK] cells, innate lymphoid cells) and adaptive (e.g., B and T lymphocytes, plasma cells) responses. This is associated with significant neovascularization, with some lymphangiogenesis.

The immunologic properties of RA synovium are now well characterized. Synovitis studies relied originally on arthroplasty-derived tissues, which were usually by definition end-stage disease, but have been expanded by the use of synovial biopsies obtained via arthroscopy or ultrasound methodologies facilitating investigation of all stages of disease (Kelly et al., 2015). Macrophages, mainly of M1 phenotype, are potent contributors to effector pathology by virtue of release of cytokines (e.g., TNF, IL-1, IL-6), chemokines, MMPs, vasoactive peptides, and oxygen and nitrogen intermediates. Numerous activation pathways of macrophages have been described including TLR ligands, soluble proteases (via PAR2), cholesterol derivatives, and immune complexes (Figure 2). The latter, containing RF and ACPA, act via FcgR1 and FcgR3 that are highly expressed on synovial macrophages. Coincident binding of ACPAs to soluble citrullinated TLR ligands, e.g., cit-fibronectin, may enhance these effects—by this model ACPAs can bind both TLR and FcR on the same cell to optimize activation (Sokolove et al., 2011), leading to high levels of cytokine release. The glycosylation status, particularly sialylation, of ACPAs also alters their capacity to promote inflammation (Rombouts et al., 2015). Cognate interaction with memory T cells is a further intriguing activation pathway—acting via a non-antigen-dependent manner, the cytokine-rich synovial milieu, particularly IL-15, IL-7, and IL-6, can induce integrin and Ig-superfamily-dependent interactions between CD45RO⁺ memory T cells and macrophages to drive subsequent macrophage cytokine and chemokine release (McInnes and Schett, 2007).

Most interest in the contribution of T cells has however focused on their antigen-driven role in initiation and perpetuation of disease. CD45RO T cells are abundant in synovial membrane and exhibit an activated phenotype. Fascinating studies have established that RA peripheral blood CD45RO naive T cells exhibit unique anabolic metabolic features characterized by low glucose metabolism and lower icATP levels associated with PFKFB3 deficiency that may play a role in their relative senescence and likely therefore their chronic activation state (Yang et al., 2016). Synovial biopsies from very early RA contain expanded T cell clones that probably represent enrichment for self-reactive (or least altered-peptide reactive) clones—this feature disappears with disease progression, presumably reflecting dilution of these clones by secondarily recruited polyclonal memory T cells (Klarenbeek et al., 2012). The differentiation state of the synovial T cell is also uncertain. Murine arthritis models strongly suggest a pivotal role for T helper 17 (Th17) cells (Miossec and Kolls, 2012; Lubberts, 2015). Human data have rather challenged this proposition though no firm conclusions can yet be drawn as the critical pre-RA phase could well be Th17 cell driven but not manifest in the effector stages of disease amenable to direct examination. Nevertheless, one report suggests that cytokine profiles in lavage fluid from early RA are paradoxically rich in IL-4, IL-13, and IL-15, whereas those T cells detected in established disease have features of both Th1 and Th17 (Raza et al., 2005). A recent study in which citrullinated tetramer responses were explored in RA patients found most T cells that were cit-specific were of Th1 cell phenotype (Snir et al., 2011). IL-17 targeting therapeutics including IL-17A and IL-17RA inhibitors have as yet not been particularly effective in established RA trials despite remarkable responses being

achieved in psoriasis (Langley et al., 2014) and to some extent spondyloarthropathies (McInnes et al., 2015).

Future studies using single-cell-based sequencing methodologies and the availability of detailed transcriptional definitions of T cells are underway to facilitate characterization of T cell subset differentiation in the phases of RA development. Understanding those factors that define T cell differentiation in RA or perhaps other cell lineages that could contribute to the evolution of this unique disease might be critically important because the detection of Treg cells in synovium has also been extensively documented—clearly their functional impact is insufficient, or they may have a functional deficit. Some evidence suggests that inadequate Treg cell function is characteristic of RA and is corrected by TNF blocker therapy (Byng-Maddick and Ehrenstein, 2015). However, it is not clear whether this defect is primary or secondary. Re-establishing local T cell homeostatic balance via expansion or recovery of Treg cell function is an attractive approach to future therapeutics, particularly as we seek restoration of tolerance as the clinical target (Rossetti et al., 2017; Benham et al., 2015).

Clearly autoantibody production is a hallmark of RA. B cell lineages at each stage of development are detected in synovium, including the presence of mature plasma cells. The synovium provides a cytokine-rich milieu for B cell survival, e.g., via BAFF, APRIL, IL-6 expression. A particular role for FLS (which are the primary source of IL-6 in the RA synovium) in maintaining this milieu has recently emerged, exemplifying the tissue dependency of the articular immune response (Bombardieri et al., 2011). In a proportion of tissues there are ectopic lymphoid structures (ELSs) that have been associated with more rapid erosive progression. Prior studies demonstrated that ELSs in RA tissue contain the enzymatic machinery necessary for antigen-driven selection, differentiation, somatic hypermutation, and class switching (Pitzalis et al., 2014). Recently, single-cell sequencing of synovial B cells suggests extensive self-reactivity particularly to citrullinated histones H2A and H2B and other citrullinated self-proteins—neutrophil could form a focal event permitting expansion of such clones (Corsiero et al., 2016). Thus, in addition to antigen presentation and autoantibody formation and release, B cells should be considered important sources of local antigen presentation and of cytokine release into the milieu.

Finally, it is vital to consider the role of the local stromal tissues in synovium. FLS have been long recognized to contribute to the matrix deposition and tissue remodeling that characterizes RA (Zvaifler et al., 1994). They exhibit anchorage independence, migratory activity (potentially even between joints), and local proliferation and release high levels of MMP and TIMPs (tissue inhibitors of metalloproteinases), cytokines and chemokines together with collagens and other ECM components (Filer 2013). They appear to be particularly dependent upon cadherin 11 for their effector function (Kiener et al., 2009). As such they should be considered a critical component of the RA inflammatory response rather than as responding bystanders. Other studies suggest the existence subsets of FLS, although the functional significance of these remains uncertain. Further complexity is emerging since FLS exhibit distinct features even across different diarthrodial joints, e.g., hips and knees, described above. FLS may also migrate across anatomically

Table 1. Major Drug Classes that Have Been Informative of the Pathogenesis of RA

Immune Target	Drugs	Key Modes of Action
TNF	Adalimumab Etanercept Golimumab Certolizumab Infliximab Biosimilars	reduced endothelial, stromal cell, and chondrocyte activation reduced osteoclast differentiation/activation modified cellular migration reduced cytokine expression, e.g., IL-6 reduced metabolic/CVD risk
IL-6	Tocilizumab Multiple Abs in development	reduced endothelial, stromal cell activation reduced osteoclast activation reduced cytokine/chemokine expression altered lipid metabolism
Co-stimulation (CD28-CD80/86)	Abatacept	reduced T cell activation diminished dendritic cell activation, cytokine production, and antigen presentation reduced osteoclast activation
B cell depletion (anti-CD20)	Rituximab	depletion of CD20 ⁺ B cell lineages, sparing plasma cells reduced cytokine production reduced antigen presentation
JAK inhibitors	Tofacitinib Baricitinib multiple compounds in development	inhibition of pivotal cytokine receptors with predictable downstream effects; JAK1, JAK2, and JAK3 thus far targeted
IL-1	Anakinra	decreased endothelial cell, stromal cell activation decreased cellular migration altered T cell differentiation innate immunity activation
Other cytokines (GM-CSF, IL-17, and others)	multiple	multiple

discrete articular sites (Lefèvre et al., 2009). Presumably reflecting the epigenetic changes described above, FLS thus play a critical role in recruiting, retaining, activating, and sustaining leukocytes, e.g., via type 1 interferons, CXCL12 (SDF-1) in the synovium (e.g., Perlman et al., 2003). Finally, in combination with endothelial cells, they may become permissive for ELS formation (Pitzalis et al., 2014) further enhancing an “auto-reactive-prone” articular microenvironment.

Articular destruction, often reflected in radiographic evidence of bone erosion, is a further hallmark of RA. Mechanisms driving damage are reviewed elsewhere (McInnes and Schett, 2007). In brief, FLS drive damage primarily via MMP (particularly MMP1 and MMP3), which mediates direct cartilage damage or by promoting chondrocyte activation and tissue catabolism through cytokine release in concert with macrophages. IL-1, TNF, and IL-17 are particularly relevant in this respect. Bone damage arises from more complex interactions that sustain osteoclast precursor maturation mediated particularly via RANKL, M-CSF, and TNF pathways. Recent insights into the participation of sclerostin, DKK, ACPAs, and other osteoclast and osteoblast co-regulators suggest the intriguing possibility of promoting bone healing—this has not yet been possible on the basis of existing therapeutics (Diarra et al., 2007; McInnes and Schett, 2011). Other factors in the synovial milieu including ACPAs are also implicated via their ability to bind directly to osteoclasts and promote their differentiation and activation.

One further development of note is the recent observation by some groups of synovial tissue micro-heterogeneity in several

cohort studies. Thus, subsets of patients can potentially be defined by myeloid, lymphoid, and stromal-rich synovial pathotypes (Dennis et al., 2014). The longer-term functional implications of these subsets are currently unclear but carry the potential to offer subtype-specific therapeutics in due course—clinical trials are underway to test this hypothesis (Astori et al., 2015; Kelly et al., 2015). If confirmed this would suggest that distinct pathologic processes can induce pathotypes that are pathologically discrete and of functional importance in identifying dominant effector pathways and tissue responses.

Lessons from Therapeutic Targeting as to the Hierarchy of Pathways and Heterogeneity of Responses

The utility of immune targeting in RA represents a major advance in disease management (Smolen et al., 2016; Smolen and Aletaha, 2015). Thus, inhibitors of TNF and IL-6 are particularly potent in delivering clinical responses. Co-stimulatory blockade with abatacept (CTLA4:Ig) is similarly effective, as is depletion of CD20 lineages by rituximab (Nam et al., 2014; Schiff et al., 2014). Notably, those blocking IL-17, IL-1, BAFF, IL-21, and IL-20 (for example) appear less effective. More recently, small molecule inhibitors of the JAK pathways, e.g., tofacitinib (targeting JAK1 and JAK3) and baricitinib (JAK1 and JAK2), have demonstrated considerable efficacy, especially if used in early disease (Genovese et al., 2016; Lee et al., 2014).

Immune therapeutics have taught us much (Table 1): (1) critical immune checkpoints or functional nodes must exist within the complex synovial lesion that are amenable to blockade and promote collapse of the majority of the lesion; (2) upstream immune

dysregulation apparently manifests primarily in cytokine (especially TNF and IL-6)-mediated effector mechanisms; and (3) murine models, despite offering exquisite mechanistic insight and proof of possibility, poorly reflect the complexity of the human RA state and therefore the therapeutic potential of any given agent. Moreover, the discrete effects of cytokine inhibitors across the wider range of chronic inflammatory diseases is increasingly pointing to a new molecular disease taxonomy that can be therapeutically defined—RA might represent a TNF and IL-6-dependent axis whereas psoriasis might reflect more TNF, IL-23, and IL-17-dependent pathways.

The diversity of responses to highly targeted agents like cytokine inhibition, B cell depletion, or co-stimulation blockade suggest that RA is not a single disease entity. Instead, it is a syndrome with a common clinical presentation but multiple pathogenic pathways, any one of which (or combination) might be relevant for a particular patient (Firestein, 2014). One of the key challenges going forward will be how we can stratify patients so that we can make better treatment selections. Thus far there are no studies that have separated responder RA populations to distinct interventions. A variety of molecular and cellular profiling approaches in peripheral blood have not been successful as yet. As noted above, preliminary studies suggest that synovial histologic patterns might correlate with responses to therapy, although other careful assessments of synovial structure and biomarkers have not demonstrated sufficient power for clinical utility (Dennis et al., 2014). These in turn could presage the advent of biomarkers that enrich for response. We also need to begin considering more robust clinical trial instruments to take into account joint-specific differences in clinical responses.

Kinetically distinct phases of immune pathway utilization as RA progresses also should be considered—in essence we use all immune therapeutics across the various phases of RA development without consideration to the dominant pathogenesis at that time point (McInnes et al., 2016). Immunologic adaptation is a characteristic of chronic infectious responses and it seems plausible that similar alterations in effector pathways will evolve as damage accrues in patients. Should we now consider “pathogenesis-appropriate” timing of therapeutic interventions? For example, could there be a role for immune tolerance-inducing therapies in very early RA, or even pre-RA? This supposes that pre-RA can be identified in a meaningful population and at a stage when immune homeostasis or re-programming can be achieved and that molecular outcome measures can be derived to define therapeutic success. Various therapeutics are being explored in this respect, including B cell depletion or co-stimulatory blockade in high-risk populations. The PRAIRI study administered a single dose of rituximab to people at high risk of developing RA—formal reporting of outcome is awaited (NTR2442 Netherlands Trial Register). A similar study is ongoing in which abatacept is being given to high risk pre-RA (APIPRA Study; ISRCTN46017566). Finally, a broader approach to identification of pre-RA comprises the StopRA study in which hydroxychloroquine is being given to healthy subjects with an elevated ACPA (NCT02603146). Finally, exciting cell transfer clinical trials using tolerogenic dendritic cells, with thus far encouraging results in terms of immune function modulation, are also ongoing (Benham et al., 2015).

Processes Driving Co-morbidities

RA is associated with increased cardiovascular risk, metabolic syndrome, psychosocial deficit including psychiatric disease, osteoporosis, and increased cancer rates. The mechanisms underlying these are increasingly understood to depend on cytokines and other circulating immune moieties, primarily using immune therapeutics as molecular scalpels. For example, IL-6 and to some extent TNF directly regulate cholesterol metabolism, such that active RA is associated with paradoxical low cholesterol levels that are replenished with effective therapy—the latter appears to operate via direct IL-6-mediated, STAT3-dependent modulation of cholesterol ester catabolism (Charles-Schoeman et al., 2015; Robertson et al., 2013). Similarly, the atherosclerotic lesion and RA synovium exhibit remarkable similarities leading to the suggestion that TNF and IL-6-driven pathways may enhance the progression of vascular disease. TNF inhibition in patients leads to reduced vascular dysfunction and to improved vascular mortality commensurate with this notion (McKellar et al., 2009). Moreover, RA atherosclerotic plaques exhibit exaggerated cytokine expression compared to non-RA controls (Ahmed et al., 2016). Similarly, circulating TNF, RANKL, and IL-1 are implicated in systemic osteoporosis (McInnes and Schett, 2011), and TNF has been implicated in the hippocampal dysfunction that likely underlies depression in RA (Krishnadas et al., 2016).

Concluding Remarks

RA represents a remarkable example of successful immunologic interrogation of tissue culture cells, pre-clinical models, and human translational studies that has delivered enormous therapeutic advances. The penumbra of these advances has also benefited many other immune-mediated disease, such as psoriasis, spondyloarthropathies, inflammatory bowel disease (IBD), and inflammatory eye disease. It has taught us much about immune pathogenetic mechanisms in the context of human autoimmunity and chronic inflammation, not least in unraveling the complex multi-hit sequence of events that ultimately overcome immune homeostasis to manifest as tissue-destructive disease. Despite such successes there remains much unmet need, not least in the current absence of the capacity to predict, prevent, and cure the disease or even predict which patients will respond to a particular therapy. It is clear that further pathogenetic discovery must drive such progress and lead ultimately to the delivery of agents with homeostatic activity, used within a precision medicine-based therapeutic culture.

REFERENCES

- Ahmed, A., Hollan, I., Curran, S.A., Kitson, S.M., Riggio, M.P., Mikkelsen, K., Almdahl, S.M., Aukrust, P., McInnes, I.B., and Goodyear, C.S. (2016). Brief report: Proatherogenic cytokine microenvironment in the aortic adventitia of patients with rheumatoid arthritis. *Arthritis Rheumatol.* 68, 1361–1366.
- Ai, R., Whitaker, J.W., Boyle, D.L., Tak, P.P., Gerlag, D.M., Wang, W., and Firestein, G.S. (2015). DNA methylome signature in synoviocytes from patients with early rheumatoid arthritis compared to synoviocytes from patients with longstanding rheumatoid arthritis. *Arthritis Rheumatol.* 67, 1978–1980.
- Ai, R., Hammaker, D., Boyle, D.L., Morgan, R., Walsh, A.M., Fan, S., Firestein, G.S., and Wang, W. (2016). Joint-specific DNA methylation and transcriptome signatures in rheumatoid arthritis identify distinct pathogenic processes. *Nat. Commun.* 7, 11849.

- Anzilotti, C., Merlini, G., Pratesi, F., Tommasi, C., Chimenti, D., and Migliorini, P. (2006). Antibodies to viral citrullinated peptide in rheumatoid arthritis. *J. Rheumatol.* 33, 647–651.
- Arend, W.P., and Firestein, G.S. (2012). Pre-rheumatoid arthritis: predisposition and transition to clinical synovitis. *Nat. Rev. Rheumatol.* 8, 573–586.
- Astori, E., Nerviani, A., Bombardieri, M., and Pitzalis, C. (2015). Towards a stratified targeted approach with biologic treatments in rheumatoid arthritis: role of synovial pathobiology. *Curr. Pharm. Des.* 21, 2216–2224.
- Benham, H., Nel, H.J., Law, S.C., Mehdi, A.M., Street, S., Ramnoruth, N., Phau, H., Lee, B.T., Ng, J., Brunck, M.E., et al. (2015). Citrullinated peptide dendritic cell immunotherapy in HLA risk genotype-positive rheumatoid arthritis patients. *Sci. Transl. Med.* 7, 290ra87.
- Bombardieri, M., Kam, N.W., Brentano, F., Choi, K., Filer, A., Kyburz, D., McInnes, I.B., Gay, S., Buckley, C., and Pitzalis, C. (2011). A BAFF/APRIL-dependent TLR3-stimulated pathway enhances the capacity of rheumatoid synovial fibroblasts to induce AID expression and Ig class-switching in B cells. *Ann. Rheum. Dis.* 70, 1857–1865.
- Bottini, N., and Firestein, G.S. (2013a). Epigenetics in rheumatoid arthritis: a primer for rheumatologists. *Curr. Rheumatol. Rep.* 15, 372.
- Bottini, N., and Firestein, G.S. (2013b). Duality of fibroblast-like synoviocytes in RA: passive responders and imprinted aggressors. *Nat. Rev. Rheumatol.* 9, 24–33.
- Byng-Maddick, R., and Ehrenstein, M.R. (2015). The impact of biological therapy on regulatory T cells in rheumatoid arthritis. *Rheumatology (Oxford)* 54, 768–775.
- Catrina, A.I., Ytterberg, A.J., Reynisdottir, G., Malmström, V., and Klareskog, L. (2014). Lungs, joints and immunity against citrullinated proteins in rheumatoid arthritis. *Nat. Rev. Rheumatol.* 10, 645–653.
- Catrina, A.I., Joshua, V., Klareskog, L., and Malmström, V. (2016). Mechanisms involved in triggering rheumatoid arthritis. *Immunol. Rev.* 269, 162–174.
- Charles-Schoeman, C., Fleischmann, R., Davignon, J., Schwartz, H., Turner, S.M., Beysen, C., Milad, M., Hellerstein, M.K., Luo, Z., Kaplan, I.V., et al. (2015). Potential mechanisms leading to the abnormal lipid profile in patients with rheumatoid arthritis versus healthy volunteers and reversal by tofacitinib. *Arthritis Rheumatol.* 67, 616–625.
- Christophorou, M.A., Castelo-Branco, G., Halley-Stott, R.P., Oliveira, C.S., Loos, R., Radziszewska, A., Mowen, K.A., Bertone, P., Silva, J.C., Zernicka-Goetz, M., et al. (2014). Citrullination regulates pluripotency and histone H1 binding to chromatin. *Nature* 507, 104–108.
- Corsiero, E., Bombardieri, M., Carlotti, E., Pratesi, F., Robinson, W., Migliorini, P., and Pitzalis, C. (2016). Single cell cloning and recombinant monoclonal antibodies generation from RA synovial B cells reveal frequent targeting of citrullinated histones of NETs. *Ann. Rheum. Dis.* 75, 1866–1875.
- Croft, A.P., Naylor, A.J., Marshall, J.L., Hardie, D.L., Zimmermann, B., Turner, J., Desanti, G., Adams, H., Yemm, A.I., Müller-Ladner, U., et al. (2016). Rheumatoid synovial fibroblasts differentiate into distinct subsets in the presence of cytokines and cartilage. *Arthritis Res. Ther.* 18, 270.
- de Aquino, S.G., Abdollahi-Roodsaz, S., Koenders, M.I., van de Loo, F.A., Pruijn, G.J., Marijnissen, R.J., Walgreen, B., Helsen, M.M., van den Bersselaar, L.A., de Molon, R.S., et al. (2014). Periodontal pathogens directly promote autoimmune experimental arthritis by inducing a TLR2- and IL-1-driven Th17 response. *J. Immunol.* 192, 4103–4111.
- de Hair, M.J., van de Sande, M.G., Ramwadhoebe, T.H., Hansson, M., Landewé, R., van der Leij, C., Maas, M., Serre, G., van Schaardenburg, D., Klareskog, L., et al. (2014). Features of the synovium of individuals at risk of developing rheumatoid arthritis: implications for understanding preclinical rheumatoid arthritis. *Arthritis Rheumatol.* 66, 513–522.
- Deane, K.D., O'Donnell, C.I., Hueber, W., Majka, D.S., Lazar, A.A., Derber, L.A., Gilliland, W.R., Edison, J.D., Norris, J.M., Robinson, W.H., and Holers, V.M. (2010). The number of elevated cytokines and chemokines in preclinical seropositive rheumatoid arthritis predicts time to diagnosis in an age-dependent manner. *Arthritis Rheum.* 62, 3161–3172.
- Dennis, G., Jr., Holweg, C.T., Kummerfeld, S.K., Choy, D.F., Setiadi, A.F., Hackney, J.A., Haverty, P.M., Gilbert, H., Lin, W.Y., Diehl, L., et al. (2014). Synovial phenotypes in rheumatoid arthritis correlate with response to biologic therapeutics. *Arthritis Res. Ther.* 16, R90.
- Diarra, D., Stolina, M., Polzer, K., Zwerina, J., Ominsky, M.S., Dwyer, D., Korb, A., Smolen, J., Hoffmann, M., Scheinecker, C., et al. (2007). Dickkopf-1 is a master regulator of joint remodeling. *Nat. Med.* 13, 156–163.
- Ekwall, A.K., Whitaker, J.W., Hammaker, D., Bugbee, W.D., Wang, W., and Firestein, G.S. (2015). The rheumatoid arthritis risk gene LBH regulates growth in fibroblast-like synoviocytes. *Arthritis Rheumatol.* 67, 1193–1202.
- El-Gabalawy, H.S., Robinson, D.B., Hart, D., Elias, B., Markland, J., Peschken, C.A., Smolik, I., Montes-Aldana, G., Schroeder, M., Fritzler, M.J., et al. (2009). Immunogenetic risks of anti-cyclical citrullinated peptide antibodies in a North American Native population with rheumatoid arthritis and their first-degree relatives. *J. Rheumatol.* 36, 1130–1135.
- Ferreira, R.C., Freitag, D.F., Cutler, A.J., Howson, J.M., Rainbow, D.B., Smyth, D.J., Kaptoge, S., Clarke, P., Boreham, C., Coulson, R.M., et al. (2013). Functional IL6R 358Ala allele impairs classical IL-6 receptor signaling and influences risk of diverse inflammatory diseases. *PLoS Genet.* 9, e1003444.
- Filer, A. (2013). The fibroblast as a therapeutic target in rheumatoid arthritis. *Curr. Opin. Pharmacol.* 13, 413–419.
- Firestein, G.S. (1996). Invasive fibroblast-like synoviocytes in rheumatoid arthritis. Passive responders or transformed aggressors? *Arthritis Rheum.* 39, 1781–1790.
- Firestein, G.S. (2003). Evolving concepts of rheumatoid arthritis. *Nature* 423, 356–361.
- Firestein, G.S. (2014). The disease formerly known as rheumatoid arthritis. *Arthritis Res. Ther.* 16, 114.
- Firestein, G.S., and Zvaifler, N.J. (2002). How important are T cells in chronic rheumatoid synovitis?: II. T cell-independent mechanisms from beginning to end. *Arthritis Rheum.* 46, 298–308.
- Firestein, G.S., Echeverri, F., Yeo, M., Zvaifler, N.J., and Green, D.R. (1997). Somatic mutations in the p53 tumor suppressor gene in rheumatoid arthritis synovium. *Proc. Natl. Acad. Sci. USA* 94, 10895–10900.
- Fonseca, J.E., Cavaleiro, J., Teles, J., Sousa, E., Andreozzi, V.L., Antunes, M., Amaral-Turkman, M.A., Canhão, H., Mourão, A.F., Lopes, J., et al. (2007). Contribution for new genetic markers of rheumatoid arthritis activity and severity: sequencing of the tumor necrosis factor-alpha gene promoter. *Arthritis Res. Ther.* 9, R37.
- Franz, J.K., Pap, T., Hummel, K.M., Nawrath, M., Aicher, W.K., Shigeyama, Y., Müller-Ladner, U., Gay, R.E., and Gay, S. (2000). Expression of sentrin, a novel antiapoptotic molecule, at sites of synovial invasion in rheumatoid arthritis. *Arthritis Rheum.* 43, 599–607.
- Genovese, M.C., Kremer, J., Zamani, O., Ludivico, C., Krogulec, M., Xie, L., Beattie, S.D., Koch, A.E., Cardillo, T.E., Rooney, T.P., et al. (2016). Baricitinib in patients with refractory rheumatoid arthritis. *N. Engl. J. Med.* 374, 1243–1252.
- Hammaker, D., Whitaker, J.W., Maeshima, K., Boyle, D.L., Ekwall, A.H., Wang, W., and Firestein, G.S. (2016). Limb bud and heart development gene transcription is regulated by the interplay of an enhancer risk allele and DNA methylation in rheumatoid arthritis. *Arthritis Rheumatol.* 68, 2637–2645.
- Harre, U., Georgess, D., Bang, H., Bozec, A., Axmann, R., Ossipova, E., Jakobsson, P.J., Baum, W., Nimmerjahn, F., Szarka, E., et al. (2012). Induction of osteoclastogenesis and bone loss by human autoantibodies against citrullinated vimentin. *J. Clin. Invest.* 122, 1791–1802.
- Hill, J.A., Southwood, S., Sette, A., Jevnikar, A.M., Bell, D.A., and Cairns, E. (2003). Cutting edge: the conversion of arginine to citrulline allows for a high-affinity peptide interaction with the rheumatoid arthritis-associated HLA-DRB1*0401 MHC class II molecule. *J. Immunol.* 171, 538–541.
- Horiuchi, M., Morinobu, A., Chin, T., Sakai, Y., Kurosaka, M., and Kumagai, S. (2009). Expression and function of histone deacetylases in rheumatoid arthritis synovial fibroblasts. *J. Rheumatol.* 36, 1580–1589.
- Huber, L.C., Brock, M., Hemmatzad, H., Giger, O.T., Moritz, F., Trenkmann, M., Distler, J.H., Gay, R.E., Kolling, C., Moch, H., et al. (2007). Histone deacetylase/acetylase activity in total synovial tissue derived from rheumatoid arthritis and osteoarthritis patients. *Arthritis Rheum.* 56, 1087–1093.
- Joosten, L.A., Leoni, F., Meghji, S., and Mascagni, P. (2011). Inhibition of HDAC activity by ITF2357 ameliorates joint inflammation and prevents cartilage and bone destruction in experimental arthritis. *Mol. Med.* 17, 391–396.

- Juarez, M., Bang, H., Hammar, F., Reimer, U., Dyke, B., Sahbuddin, I., Buckley, C.D., Fisher, B., Filer, A., and Raza, K. (2016). Identification of novel antiacetylated vimentin antibodies in patients with early inflammatory arthritis. *Ann. Rheum. Dis.* 75, 1099–1107.
- Källberg, H., Ding, B., Padyukov, L., Bengtsson, C., Rönnelid, J., Klareskog, L., and Alfredsson, L.; EIRA Study Group (2011). Smoking is a major preventable risk factor for rheumatoid arthritis: estimations of risks after various exposures to cigarette smoke. *Ann. Rheum. Dis.* 70, 508–511.
- Kaminsky, Z.A., Tang, T., Wang, S.C., Ptak, C., Oh, G.H., Wong, A.H., Feldcamp, L.A., Virtanen, C., Halfvarson, J., Tysk, C., et al. (2009). DNA methylation profiles in monozygotic and dizygotic twins. *Nat. Genet.* 41, 240–245.
- Karouzakis, E., Rengel, Y., Jüngel, A., Kolling, C., Gay, R.E., Michel, B.A., Tak, P.P., Gay, S., Neidhart, M., and Ospelt, C. (2011). DNA methylation regulates the expression of CXCL12 in rheumatoid arthritis synovial fibroblasts. *Genes Immun.* 12, 643–652.
- Kelly, S., Humby, F., Filer, A., Ng, N., Di Cicco, M., Hands, R.E., Rocher, V., Bombardier, M., D'Agostino, M.A., McInnes, I.B., et al. (2015). Ultrasound-guided synovial biopsy: a safe, well-tolerated and reliable technique for obtaining high-quality synovial tissue from both large and small joints in early arthritis patients. *Ann. Rheum. Dis.* 74, 611–617.
- Kiener, H.P., Niederreiter, B., Lee, D.M., Jimenez-Boj, E., Smolen, J.S., and Brenner, M.B. (2009). Cadherin 11 promotes invasive behavior of fibroblast-like synoviocytes. *Arthritis Rheum.* 60, 1305–1310.
- Kim, K., Bang, S.Y., Lee, H.S., and Bae, S.C. (2017). Update on the genetic architecture of rheumatoid arthritis. *Nat. Rev. Rheumatol.* 13, 13–24.
- Kinslow, J.D., Blum, L.K., Deane, K.D., Demoruelle, M.K., Okamoto, Y., Parish, M.C., Kongpachith, S., Lahey, L.J., Norris, J.M., Robinson, W.H., and Holers, V.M. (2016). Elevated IgA plasmablast levels in subjects at risk of developing rheumatoid arthritis. *Arthritis Rheumatol.* 68, 2372–2383.
- Kirschmann, D.A., Duffin, K.L., Smith, C.E., Welpy, J.K., Howard, S.C., Schwartz, B.D., and Woulfe, S.L. (1995). Naturally processed peptides from rheumatoid arthritis associated and non-associated HLA-DR alleles. *J. Immunol.* 155, 5655–5662.
- Klarenbeek, P.L., de Hair, M.J., Doorenspleet, M.E., van Schaik, B.D., Esveldt, R.E., van de Sande, M.G., Cantaert, T., Gerlag, D.M., Baeten, D., van Kampen, A.H., et al. (2012). Inflamed target tissue provides a specific niche for highly expanded T-cell clones in early human autoimmune disease. *Ann. Rheum. Dis.* 71, 1088–1093.
- Konig, M.F., Abusleme, L., Reinholdt, J., Palmer, R.J., Teles, R.P., Sampson, K., Rosen, A., Nigrovic, P.A., Sokolove, J., Giles, J.T., et al. (2016a). *Aggregatibacter actinomycetemcomitans*-induced hypercitrullination links periodontal infection to autoimmunity in rheumatoid arthritis. *Sci. Transl. Med.* 8, 369ra176.
- Konig, M.F., Giles, J.T., Nigrovic, P.A., and Andrade, F. (2016b). Antibodies to native and citrullinated RA33 (hnRNP A2/B1) challenge citrullination as the inciting principle underlying loss of tolerance in rheumatoid arthritis. *Ann. Rheum. Dis.* 75, 2022–2028.
- Krishnadas, R., Nicol, A., Sassarini, J., Puri, N., Burden, A.D., Leman, J., Combet, E., Pimlott, S., Hadley, D., McInnes, I.B., and Cavanagh, J. (2016). Circulating tumour necrosis factor is highly correlated with brainstem serotonin transporter availability in humans. *Brain Behav. Immun.* 51, 29–38.
- Krishnamurthy, A., Joshua, V., Haj Hensvold, A., Jin, T., Sun, M., Vivar, N., Ytterberg, A.J., Engström, M., Fernandes-Cerdeira, C., Amara, K., et al. (2016). Identification of a novel chemokine-dependent molecular mechanism underlying rheumatoid arthritis-associated autoantibody-mediated bone loss. *Ann. Rheum. Dis.* 75, 721–729.
- Langley, R.G., Elewski, B.E., Lebwohl, M., Reich, K., Griffiths, C.E., Papp, K., Puig, L., Nakagawa, H., Spelman, L., Sigurgeirsson, B., et al.; ERASURE Study Group; FIXTURE Study Group (2014). Secukinumab in plaque psoriasis—results of two phase 3 trials. *N. Engl. J. Med.* 371, 326–338.
- Law, S.C., Street, S., Yu, C.H., Capini, C., Ramnoruth, S., Nel, H.J., van Gorp, E., Hyde, C., Lau, K., Pahau, H., et al. (2012). T-cell autoreactivity to citrullinated autoantigenic peptides in rheumatoid arthritis patients carrying HLA-DRB1 shared epitope alleles. *Arthritis Res. Ther.* 14, R118.
- Lee, E.B., Fleischmann, R., Hall, S., Wilkinson, B., Bradley, J.D., Gruben, D., Koncz, T., Krishnaswami, S., Wallenstein, G.V., Zang, C., et al.; ORAL Start Investigators (2014). Tofacitinib versus methotrexate in rheumatoid arthritis. *N. Engl. J. Med.* 370, 2377–2386.
- Lefèvre, S., Knedla, A., Tennie, C., Kampmann, A., Wunrau, C., Dinsen, R., Korb, A., Schnäker, E.M., Tarner, I.H., Robbins, P.D., et al. (2009). Synovial fibroblasts spread rheumatoid arthritis to unaffected joints. *Nat. Med.* 15, 1414–1420.
- Linn-Rasker, S.P., van der Helm-van Mil, A.H., van Gaalen, F.A., Kloppenburg, M., de Vries, R.R., le Cessie, S., Breedveld, F.C., Toes, R.E., and Huizinga, T.W. (2006). Smoking is a risk factor for anti-CCP antibodies only in rheumatoid arthritis patients who carry HLA-DRB1 shared epitope alleles. *Ann. Rheum. Dis.* 65, 366–371.
- Liu, X., Zeng, B., Zhang, J., Li, W., Mou, F., Wang, H., Zou, Q., Zhong, B., Wu, L., Wei, H., and Fang, Y. (2016). Role of the gut microbiome in modulating arthritis progression in mice. *Sci. Rep.* 6, 30594.
- Lubberts, E. (2015). The IL-23-IL-17 axis in inflammatory arthritis. *Nat. Rev. Rheumatol.* 11, 415–429.
- Lundström, E., Källberg, H., Alfredsson, L., Klareskog, L., and Padyukov, L. (2009). Gene-environment interaction between the DRB1 shared epitope and smoking in the risk of anti-citrullinated protein antibody-positive rheumatoid arthritis: all alleles are important. *Arthritis Rheum.* 60, 1597–1603.
- Makrygiannakis, D., Hermansson, M., Ulfgren, A.K., Nicholas, A.P., Zendman, A.J., Eklund, A., Grunewald, J., Skold, C.M., Klareskog, L., and Catrina, A.I. (2008). Smoking increases peptidylarginine deiminase 2 enzyme expression in human lungs and increases citrullination in BAL cells. *Ann. Rheum. Dis.* 67, 1488–1492.
- Matsumoto, I., Maccioni, M., Lee, D.M., Maurice, M., Simmons, B., Brenner, M., Mathis, D., and Benoist, C. (2002). How antibodies to a ubiquitous cytoplasmic enzyme may provoke joint-specific autoimmune disease. *Nat. Immunol.* 3, 360–365.
- McInnes, I.B., and Schett, G. (2007). Cytokines in the pathogenesis of rheumatoid arthritis. *Nat. Rev. Immunol.* 7, 429–442.
- McInnes, I.B., and Schett, G. (2011). The pathogenesis of rheumatoid arthritis. *N. Engl. J. Med.* 365, 2205–2219.
- McInnes, I.B., Mease, P.J., Kirkham, B., Kavanaugh, A., Ritchlin, C.T., Rahman, P., van der Heijde, D., Landewé, R., Conaghan, P.G., Gottlieb, A.B., et al.; FUTURE 2 Study Group (2015). Secukinumab, a human anti-interleukin-17A monoclonal antibody, in patients with psoriatic arthritis (FUTURE 2): a randomized, double-blind, placebo-controlled, phase 3 trial. *Lancet* 386, 1137–1146.
- McInnes, I.B., Buckley, C.D., and Isaacs, J.D. (2016). Cytokines in rheumatoid arthritis – shaping the immunological landscape. *Nat. Rev. Rheumatol.* 12, 63–68.
- McKellar, G.E., McCarey, D.W., Sattar, N., and McInnes, I.B. (2009). Role for TNF in atherosclerosis? Lessons from autoimmune disease. *Nat. Rev. Cardiol.* 6, 410–417.
- Miossec, P., and Kolls, J.K. (2012). Targeting IL-17 and TH17 cells in chronic inflammation. *Nat. Rev. Drug Discov.* 11, 763–776.
- Müller-Ladner, U., Kriegsmann, J., Franklin, B.N., Matsumoto, S., Geiler, T., Gay, R.E., and Gay, S. (1996). Synovial fibroblasts of patients with rheumatoid arthritis attach to and invade normal human cartilage when engrafted into SCID mice. *Am. J. Pathol.* 149, 1607–1615.
- Mydel, P., Wang, Z., Brisslert, M., Hellvard, A., Dahlberg, L.E., Hazen, S.L., and Bokarewa, M. (2010). Carbamylated-dependent activation of T cells: a novel mechanism in the pathogenesis of autoimmune arthritis. *J. Immunol.* 184, 6882–6890.
- Nakamachi, Y., Kawano, S., Takenokuchi, M., Nishimura, K., Sakai, Y., Chin, T., Saura, R., Kurosaka, M., and Kumagai, S. (2009). MicroRNA-124a is a key regulator of proliferation and monocyte chemoattractant protein 1 secretion in fibroblast-like synoviocytes from patients with rheumatoid arthritis. *Arthritis Rheum.* 60, 1294–1304.
- Nakano, K., Whitaker, J.W., Boyle, D.L., Wang, W., and Firestein, G.S. (2013a). DNA methylation signature in rheumatoid arthritis. *Ann. Rheum. Dis.* 72, 110–117.
- Nakano, K., Boyle, D.L., and Firestein, G.S. (2013b). Regulation of DNA methylation in rheumatoid arthritis synoviocytes. *J. Immunol.* 190, 1297–1303.

- Nam, J.L., Ramiro, S., Gaujoux-Viala, C., Takase, K., Leon-Garcia, M., Emery, P., Gossec, L., Landewe, R., Smolen, J.S., and Buch, M.H. (2014). Efficacy of biological disease-modifying antirheumatic drugs: a systematic literature review informing the 2013 update of the EULAR recommendations for the management of rheumatoid arthritis. *Ann. Rheum. Dis.* 73, 516–528.
- Okada, Y., Wu, D., Trynka, G., Raj, T., Terao, C., Ikari, K., Kochi, Y., Ohmura, K., Suzuki, A., Yoshida, S., et al.; RACI consortium; GARNET consortium (2014). Genetics of rheumatoid arthritis contributes to biology and drug discovery. *Nature* 506, 376–381.
- Pap, T., Franz, J.K., Hummel, K.M., Jeisy, E., Gay, R., and Gay, S. (2000). Activation of synovial fibroblasts in rheumatoid arthritis: lack of expression of the tumour suppressor PTEN at sites of invasive growth and destruction. *Arthritis Res.* 2, 59–64.
- Perlman, H., Bradley, K., Liu, H., Cole, S., Shamiyah, E., Smith, R.C., Walsh, K., Fiore, S., Koch, A.E., Firestein, G.S., et al. (2003). IL-6 and matrix metalloproteinase-1 are regulated by the cyclin-dependent kinase inhibitor p21 in synovial fibroblasts. *J. Immunol.* 170, 838–845.
- Pitzalis, C., Jones, G.W., Bombardier, M., and Jones, S.A. (2014). Ectopic lymphoid-like structures in infection, cancer and autoimmunity. *Nat. Rev. Immunol.* 14, 447–462.
- Randen, I., Brown, D., Thompson, K.M., Hughes-Jones, N., Pascual, V., Victor, K., Capra, J.D., Forre, O., and Natvig, J.B. (1992). Clonally related IgM rheumatoid factors undergo affinity maturation in the rheumatoid synovial tissue. *J. Immunol.* 148, 3296–3301.
- Rawlings, D.J., Dai, X., and Buckner, J.H. (2015). The role of PTPN22 risk variant in the development of autoimmunity: finding common ground between mouse and human. *J. Immunol.* 194, 2977–2984.
- Raychaudhuri, S., Sandor, C., Stahl, E.A., Freudenberg, J., Lee, H.S., Jia, X., Alfredsson, L., Padyukov, L., Klareskog, L., Worthington, J., et al. (2012). Five amino acids in three HLA proteins explain most of the association between MHC and seropositive rheumatoid arthritis. *Nat. Genet.* 44, 291–296.
- Raza, K., Falciani, F., Curnow, S.J., Ross, E.J., Lee, C.Y., Akbar, A.N., Lord, J.M., Gordon, C., Buckley, C.D., and Salmon, M. (2005). Early rheumatoid arthritis is characterized by a distinct and transient synovial fluid cytokine profile of T cell and stromal cell origin. *Arthritis Res. Ther.* 7, R784–R795.
- Reynisdottir, G., Karimi, R., Joshua, V., Olsen, H., Henvold, A.H., Harju, A., Engström, M., Grunewald, J., Nyren, S., Eklund, A., et al. (2014). Structural changes and antibody enrichment in the lungs are early features of anti-citrullinated protein antibody-positive rheumatoid arthritis. *Arthritis Rheumatol.* 66, 31–39.
- Reynisdottir, G., Olsen, H., Joshua, V., Engström, M., Forsslund, H., Karimi, R., Sköld, C.M., Nyren, S., Eklund, A., Grunewald, J., and Catrina, A.I. (2016). Signs of immune activation and local inflammation are present in the bronchial tissue of patients with untreated early rheumatoid arthritis. *Ann. Rheum. Dis.* 75, 1722–1727.
- Robertson, J., Peters, M.J., McInnes, I.B., and Sattar, N. (2013). Changes in lipid levels with inflammation and therapy in RA: a maturing paradigm. *Nat. Rev. Rheumatol.* 9, 513–523.
- Rodríguez-Carrio, J., Hähnlein, J.S., Ramwadhoebe, T.H., Semmelink, J.F., Choi, I.Y., van Lienden, K.P., Maas, M., Gerlag, D.M., Tak, P.P., Geijtenbeek, T.B., and van Baarsen, L.G. (2016). Altered innate lymphoid cells subsets in human lymph node biopsies during the at risk and earliest phase of rheumatoid arthritis. *Arthritis Rheumatol.* 69, 70–76.
- Rombouts, Y., Ewing, E., van de Stadt, L.A., Selman, M.H., Trouw, L.A., Deelder, A.M., Huizinga, T.W., Wuhrer, M., van Schaardenburg, D., Toes, R.E., and Scherer, H.U. (2015). Anti-citrullinated protein antibodies acquire a pro-inflammatory Fc glycosylation phenotype prior to the onset of rheumatoid arthritis. *Ann. Rheum. Dis.* 74, 234–241.
- Rossetti, M., Spreafico, R., Consolaro, A., Leong, J.Y., Chua, C., Massa, M., Saidin, S., Magni-Manzoni, S., Arkachaisri, T., Wallace, C.A., et al. (2017). TCR repertoire sequencing identifies synovial Treg cell clonotypes in the bloodstream during active inflammation in human arthritis. *Ann. Rheum. Dis.* 76, 435–441.
- Scally, S.W., Petersen, J., Law, S.C., Dudek, N.L., Nel, H.J., Loh, K.L., Wijeyewickrema, L.C., Eckle, S.B., van Heemst, J., Pike, R.N., et al. (2013). A molecular basis for the association of the HLA-DRB1 locus, citrullination, and rheumatoid arthritis. *J. Exp. Med.* 210, 2569–2582.
- Scher, J.U., Sczesnak, A., Longman, R.S., Segata, N., Ubeda, C., Bielski, C., Rostron, T., Cerundolo, V., Pamer, E.G., Abramson, S.B., et al. (2013). Expansion of intestinal *Prevotella copri* correlates with enhanced susceptibility to arthritis. *eLife* 2, e01202.
- Schiff, M., Weinblatt, M.E., Valente, R., van der Heijde, D., Citera, G., Elegbe, A., Maldonado, M., and Fleischmann, R. (2014). Head-to-head comparison of subcutaneous abatacept versus adalimumab for rheumatoid arthritis: two-year efficacy and safety findings from AMPLE trial. *Ann. Rheum. Dis.* 73, 86–94.
- Shi, J., Knevel, R., Suwannalai, P., van der Linden, M.P., Janssen, G.M., van Veelen, P.A., Levarht, N.E., van der Helm-van Mil, A.H., Cerami, A., Huizinga, T.W., et al. (2011). Autoantibodies recognizing carbamylated proteins are present in sera of patients with rheumatoid arthritis and predict joint damage. *Proc. Natl. Acad. Sci. USA* 108, 17372–17377.
- Shi, J., van de Stadt, L.A., Levarht, E.W., Huizinga, T.W., Hamann, D., van Schaardenburg, D., Toes, R.E., and Trouw, L.A. (2014). Anti-carbamylated protein (anti-CarP) antibodies precede the onset of rheumatoid arthritis. *Ann. Rheum. Dis.* 73, 780–783.
- Smolen, J.S., and Aletaha, D. (2015). Rheumatoid arthritis therapy reappraisal: strategies, opportunities and challenges. *Nat. Rev. Rheumatol.* 11, 276–289.
- Smolen, J.S., Aletaha, D., and McInnes, I.B. (2016). Rheumatoid arthritis. *Lancet* 388, 2023–2038.
- Snir, O., Rieck, M., Gebe, J.A., Yue, B.B., Rawlings, C.A., Nepom, G., Malmstrom, V., and Buckner, J.H. (2011). Identification and functional characterization of T cells reactive to citrullinated vimentin in HLA-DRB1*0401-positive humanized mice and rheumatoid arthritis patients. *Arthritis Rheum.* 63, 2873–2883.
- Sokolove, J., Zhao, X., Chandra, P.E., and Robinson, W.H. (2011). Immune complexes containing citrullinated fibrinogen costimulate macrophages via Toll-like receptor 4 and Fc γ receptor. *Arthritis Rheum.* 63, 53–62.
- Stanczyk, J., Ospeit, C., Karouzakis, E., Flier, A., Raza, K., Kolling, C., Gay, R., Buckley, C.D., Tak, P.P., Gay, S., and Kyburz, D. (2011). Altered expression of microRNA-203 in rheumatoid arthritis synovial fibroblasts and its role in fibroblast activation. *Arthritis Rheum.* 63, 373–381.
- Suzuki, A., Yamada, R., Chang, X., Tokuhiro, S., Sawada, T., Suzuki, M., Nagasaki, M., Nakayama-Hamada, M., Kawaida, R., Ono, M., et al. (2003). Functional haplotypes of PADI4, encoding citrullinating enzyme peptidylarginine deiminase 4, are associated with rheumatoid arthritis. *Nat. Genet.* 34, 395–402.
- Tan, E.M., and Smolen, J.S. (2016). Historical observations contributing insights on etiopathogenesis of rheumatoid arthritis and role of rheumatoid factor. *J. Exp. Med.* 213, 1937–1950.
- Tran, C.N., Davis, M.J., Tesmer, L.A., Endres, J.L., Motyl, C.D., Smuda, C., Somers, E.C., Chung, K.C., Urquhart, A.G., Lundy, S.K., et al. (2007). Presentation of arthritogenic peptide to antigen-specific T cells by fibroblast-like synoviocytes. *Arthritis Rheum.* 56, 1497–1506.
- Tsaprouni, L.G., Yang, T.P., Bell, J., Dick, K.J., Kanoni, S., Nisbet, J., Viñuela, A., Grundberg, E., Nelson, C.P., Meduri, E., et al. (2014). Cigarette smoking reduces DNA methylation levels at multiple genomic loci but the effect is partially reversible upon cessation. *Epigenetics* 9, 1382–1396.
- van Beers, J.J., Willemze, A., Jansen, J.J., Engbers, G.H., Salden, M., Raats, J., Drijfhout, J.W., van der Helm-van Mil, A.H., Toes, R.E., and Pruijn, G.J. (2013). ACPA fine-specificity profiles in early rheumatoid arthritis patients do not correlate with clinical features at baseline or with disease progression. *Arthritis Res. Ther.* 15, R140.
- van der Helm-van Mil, A.H., Verpoort, K.N., Breedveld, F.C., Huizinga, T.W., Toes, R.E., and de Vries, R.R. (2006). The HLA-DRB1 shared epitope alleles are primarily a risk factor for anti-cyclic citrullinated peptide antibodies and are not an independent risk factor for development of rheumatoid arthritis. *Arthritis Rheum.* 54, 1117–1121.
- Vassallo, R., Luckey, D., Behrens, M., Madden, B., Luthra, H., David, C., and Taneja, V. (2014). Cellular and humoral immunity in arthritis are profoundly influenced by the interaction between cigarette smoke effects and host HLA-DR and DQ genes. *Clin. Immunol.* 152, 25–35.
- Wang, Y., Shaked, I., Stanford, S.M., Zhou, W., Curtsinger, J.M., Mikulski, Z., Shaheen, Z.R., Cheng, G., Sawatzke, K., Campbell, A.M., et al. (2013). The autoimmunity-associated gene PTPN22 potentiates toll-like receptor-driven, type 1 interferon-dependent immunity. *Immunity* 39, 111–122.

- Wegner, N., Wait, R., Sroka, A., Eick, S., Nguyen, K.A., Lundberg, K., Kinloch, A., Culshaw, S., Potempa, J., and Venables, P.J. (2010). Peptidylarginine deiminase from *Porphyromonas gingivalis* citrullinates human fibrinogen and α -enolase: implications for autoimmunity in rheumatoid arthritis. *Arthritis Rheum.* 62, 2662–2672.
- Weyand, C.M., Hicok, K.C., Conn, D.L., and Goronzy, J.J. (1992). The influence of HLA-DRB1 genes on disease severity in rheumatoid arthritis. *Ann. Intern. Med.* 117, 801–806.
- Yamamoto, K., Okada, Y., Suzuki, A., and Kochi, Y. (2015). Genetics of rheumatoid arthritis in Asia—present and future. *Nat. Rev. Rheumatol.* 11, 375–379.
- Yang, Z., Shen, Y., Oishi, H., Matteson, E.L., Tian, L., Goronzy, J.J., and Weyand, C.M. (2016). Restoring oxidant signaling suppresses proarthritogenic T cell effector functions in rheumatoid arthritis. *Sci. Transl. Med.* 8, 331ra38.
- Zhang, X., Zhang, D., Jia, H., Feng, Q., Wang, D., Liang, D., Wu, X., Li, J., Tang, L., Li, Y., et al. (2015). The oral and gut microbiomes are perturbed in rheumatoid arthritis and partly normalized after treatment. *Nat. Med.* 21, 895–905.
- Zvaifler, N.J., Boyle, D., and Firestein, G.S. (1994). Early synovitis—synoviocytes and mononuclear cells. *Semin. Arthritis Rheum.* 23 (6, Suppl 2), 11–16.