

PyCon PL
2019-09-13

Biohack

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Epam Systems: data science for industry, Jagiellonian University: bioinformatics, lectures, classes



Bioinformatics



Image source: <https://blog.f1000.com/wp-content/uploads/2017/01/Header-Image-crop-1024x559.jpg>

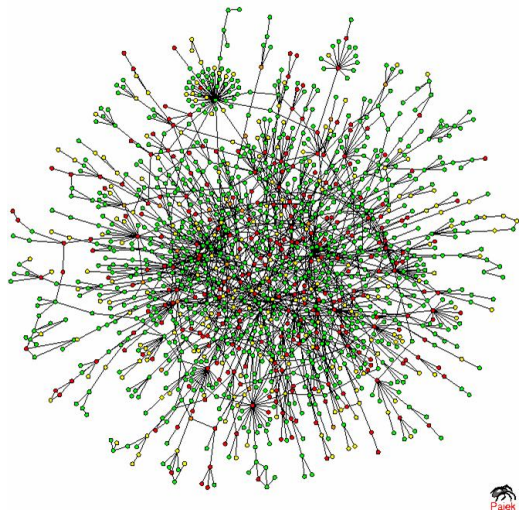
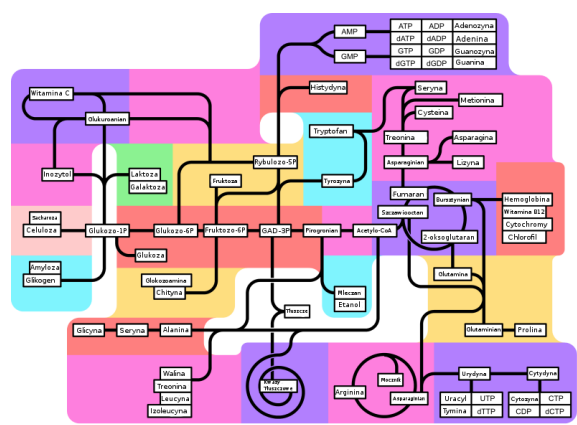
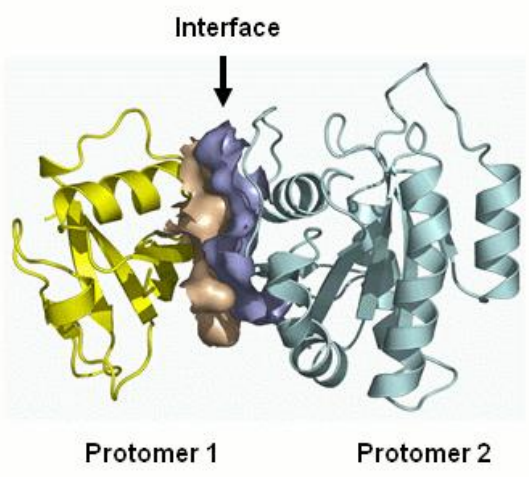
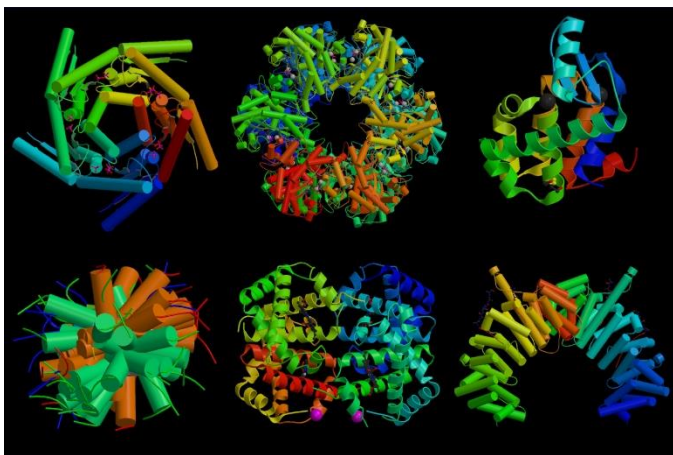
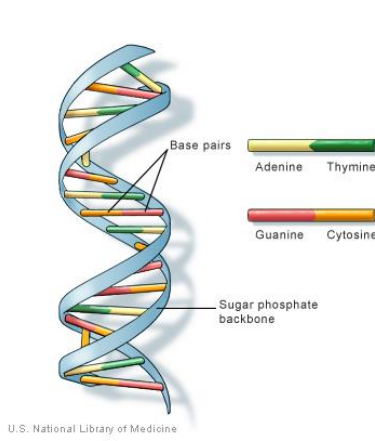
data
challenges
applications

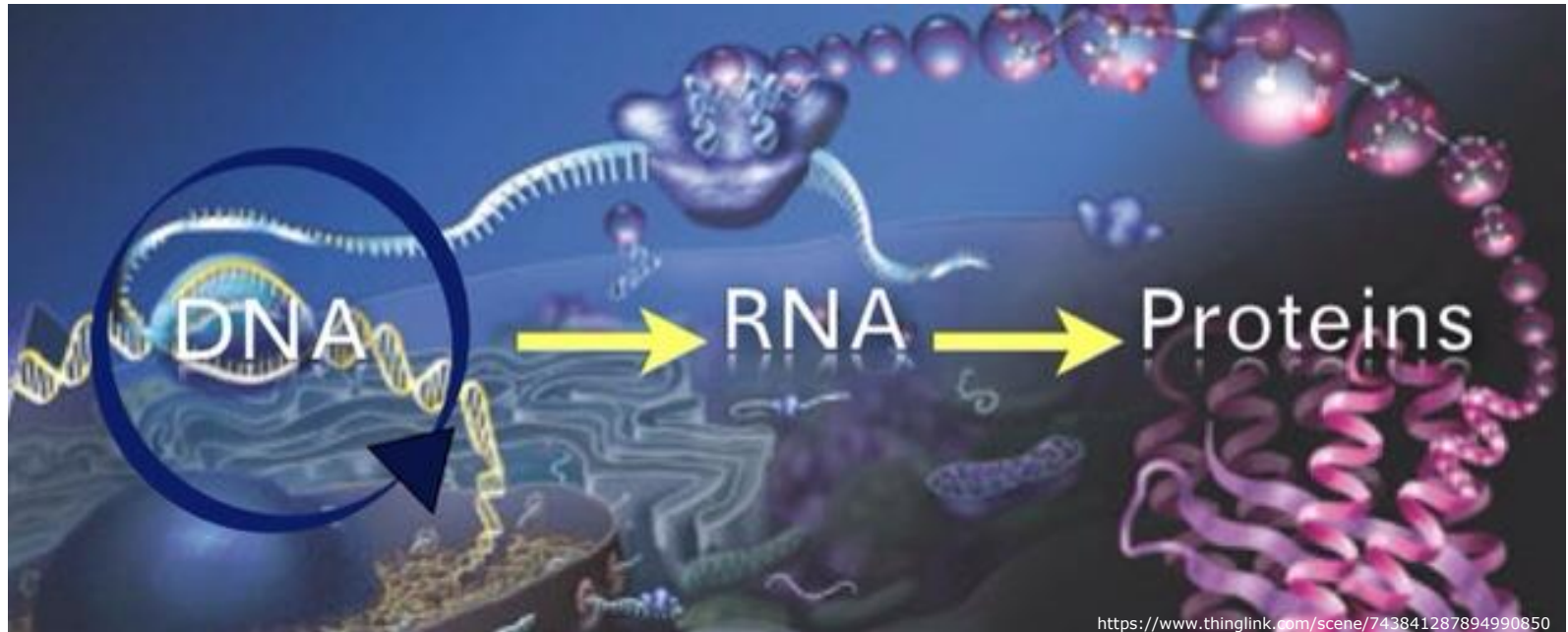
DNA sequence

> AY169899.1 *Morelia viridis* strain ABTC66386 cytochrome b gene, partial cds; mitochondrial gene for mitochondrial product

```
TTCGGCTCAATATTATTAACATGTTTAGCCCTACAAGTACTACCGGC  
TTCTTCTTAGCCGTCCACTACACAGCAAACATCAACCTAGCATTCT  
CATCCATTATCCATATCACTCGAGATGTCCCATACGGCTGAATAATA  
CAAAACCTACACGCCATCGGAGCATCCATATTCTTCATTTGCATTTA  
CATCCACATCGCACGAGGACTATACTACGGATCCTACCTCAACAAA  
GAGACTTGAATATCCGGGTATCACCCCTACTCATCACATTAATAGCAAC  
CGCCTTCTTTG
```

Data visualisation





Sequence → Structure → Function

Data availability

- Over 1500 bioinformatics databases listed by Nucleic Acid Research journal
- Main databases connected in network (meta-database)

NCBI Entrez:

<https://www.ncbi.nlm.nih.gov/search>

GenBank

Swiss Prot

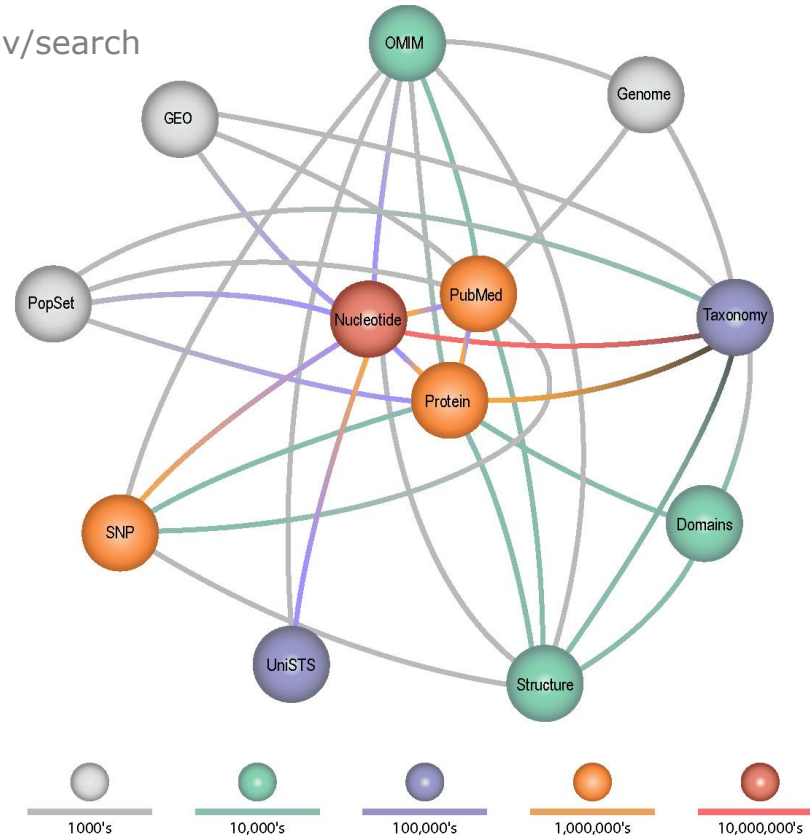
Protein Data Bank

PubMed

OMIM

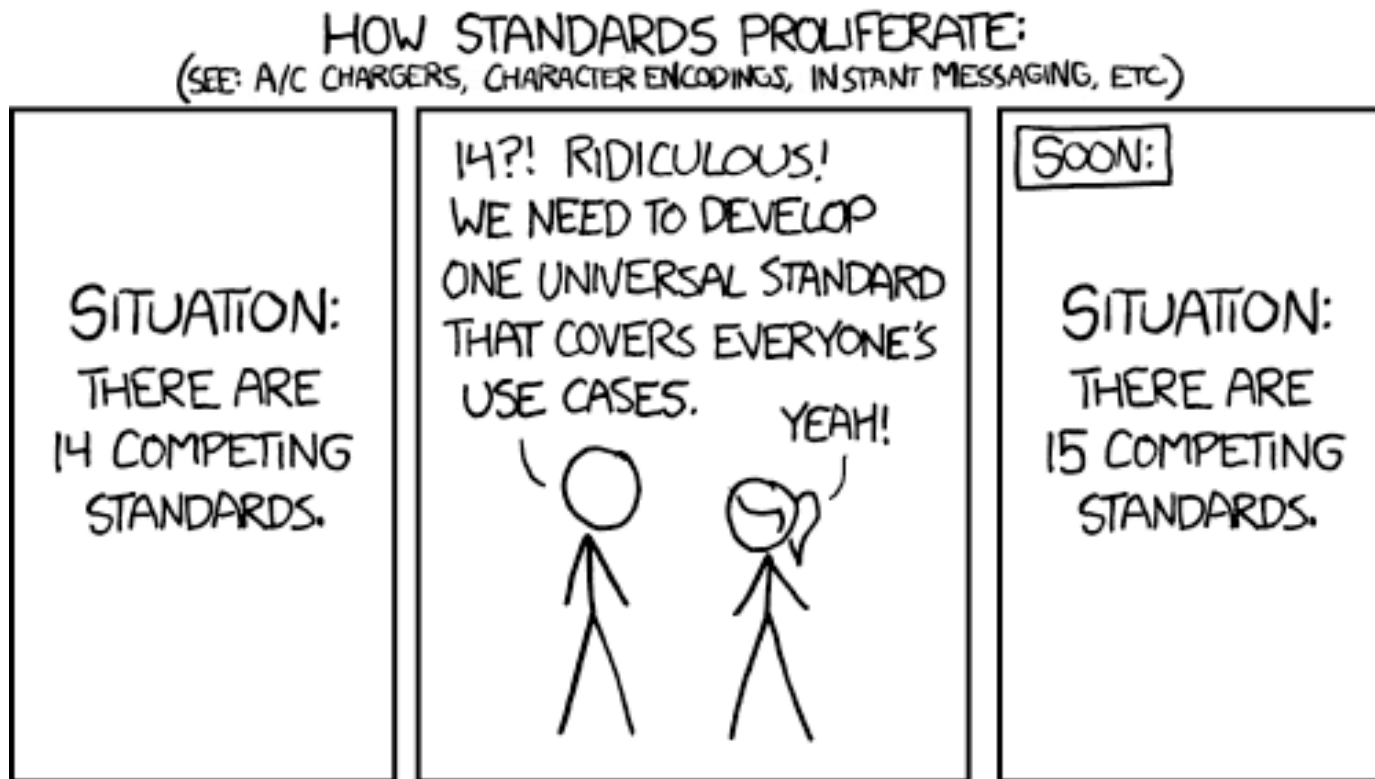
SNP

(...)



Data issues

- Corectness and reliability
- Inconsistency of data formats
- Outdated databases



Challenge: next generation sequencing (NGS)

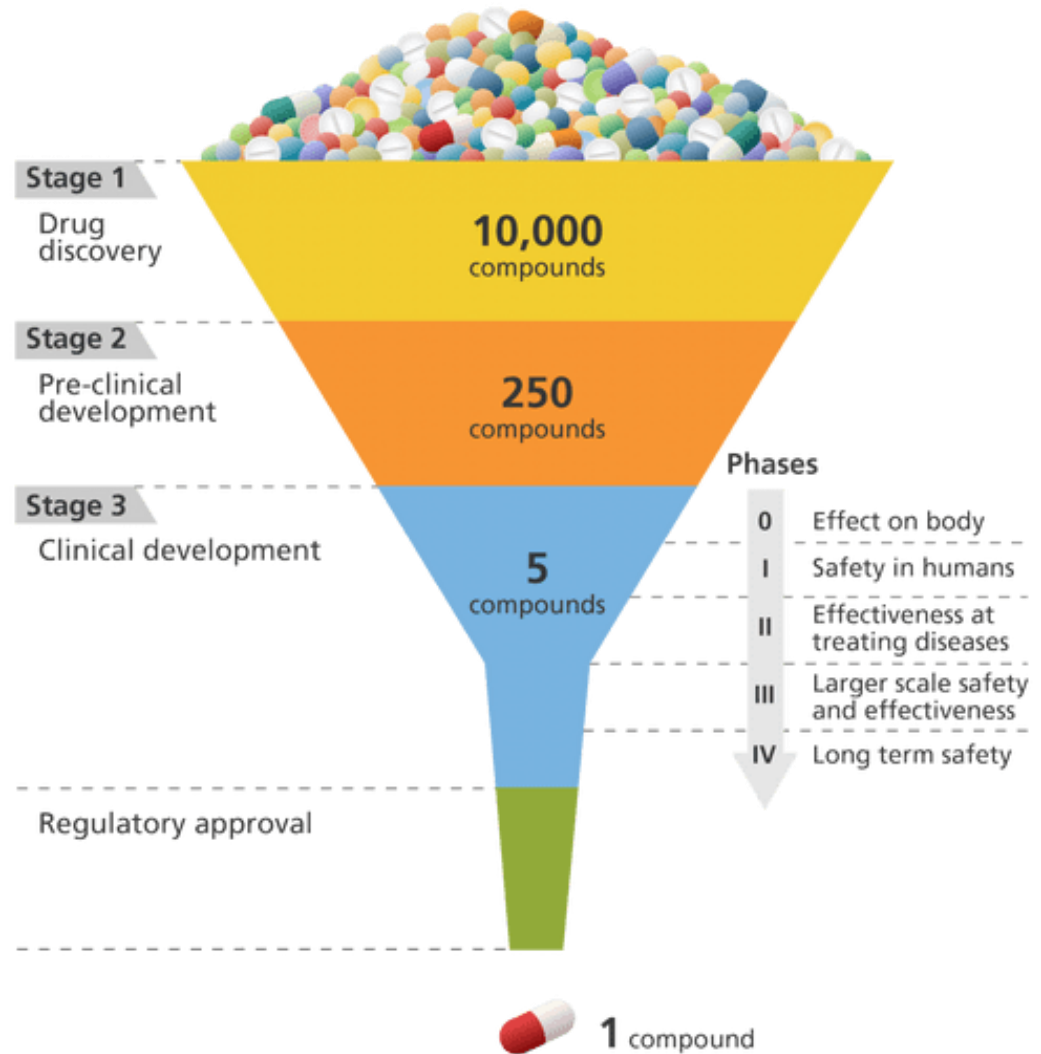


https://singularityhub.com/wp-content/uploads/2019/01/3d-illustration-dna-sequencing_shutterstock_430949605-1068x601.jpg

- sequence assembly
 - data storage
 - variations

Application example: drug design

- target identification
- structure determination
- lead search
- pharmacokinetics
- response prediction (ADME & toxicology)



Biohack



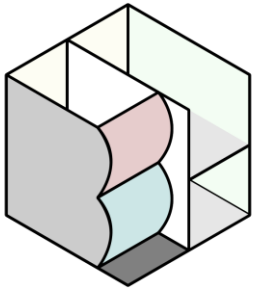
Bioinformatics hackathons

Hosted worldwide by several organisations.

Eg.: <http://biohackathon.org> (annual event since 2008)

<https://biohackathons.github.io>

<https://www.biohackathon-europe.org>



Biohack (past editions: 2018, 2019)

<http://biohack.com.pl/>

Bioninja (upcoming, 4-6.10.2019, registration still active 😊)

<http://www.bioninjachallenge.eu>

- onsite (Łódź)
- free participation
- 4-5 people interdisciplinary teams
(if you are alone, orga team creates team for you)
- tasks are selected randomly
- 24 h for coding
- integration part, board games, pizza, etc...

1. The analysis of bacteriophage genomes considering their lytic/lysogenic potential
2. Implementation of balance tree algorithm for abundance difference analysis in metagenomic research
3. Analysis of GWAS data using machine learning algorithms
4. Development of algorithm for translation of common names of disease entities into ICD-10 units (NLP approach preferred)
5. Development of local tool for annotation of bacteriophage genomes
6. Development of method for ordered storage of NGS-obtained data

1. Immunogenicity prediction of viral neoantigens in murine hosts.
2. Feature space engineering for optimal classification of virome dataset.
3. Predicting response to cancer immunotherapy

<http://biohack.com.pl/biohack-ii-exemplary-tasks/>

Predicting response to cancer immunotherapy

Treatment method:

the patient's immune system is stimulated to destroy tumor cells.

Only 20-40% of patients respond to immunotherapy.

Making prediction which patients will be responders is one of the challenges in cancer research.

Provided data

A dataset related to patients who underwent cancer immunotherapy:

- patients' clinical information
- RNA sequencing data
- response status

```
In [1]: import pandas as pd
```

```
In [2]: df_cv = pd.read_csv("../data//X_covariates.tsv", sep="\t")  
df_genes = pd.read_csv("../data//X_genes.tsv", sep="\t")  
df_y = pd.read_csv("../data//y.tsv", sep="\t", header=None)
```

17 parameters regarding patient interview, diagnosis results and treatment history.

```
In [3]: df_cv.shape
```

Out[3]: (200, 17)

```
In [4]: df_cv.head()
```

Out[4]:

| | FMOne mutation burden per MB | Neoantigen burden per MB | Enrollment IC | IC Level | TC Level | Immune phenotype | Sex | TCGA Subtype | Lund | Lund2 | Received platinum | Met Disease Status | Sample age | Sample collected pre- platinum | Intravesical BCG administered | Baselir ECO Scol |
|---|---------------------------------------|--------------------------------|------------------|-------------|-------------|---------------------|-----|-----------------|---------|--------------------|----------------------|--------------------------|--------------------------|---|-------------------------------------|------------------------|
| 0 | 8.0 | 0.803922 | IC2 | IC2+ | TC0 | inflamed | M | IV | MS2b1 | Infiltrated | N | LN Only | (less than) 1 year | NaN | N | |
| 1 | 4.0 | 0.764706 | IC0 | IC0 | TC0 | desert | F | III | MS2b2.2 | Basal/SCC- like | N | NaN | 1-2 years | NaN | N | |
| 2 | 5.0 | 0.117647 | IC1 | IC1 | TC0 | desert | M | III | MS2b2.2 | Basal/SCC- like | Y | Visceral | (less than) 1 year | N | N | |
| 3 | 4.0 | 0.294118 | IC2 | IC2+ | TC2+ | excluded | M | III | MS2b1 | Infiltrated | N | Visceral | more than 2 years | NaN | Y | |
| 4 | 10.0 | NaN | IC1 | IC1 | TC0 | desert | M | IV | MS2b1 | Infiltrated | Y | Visceral | (less than) 1 year | NaN | N | |

Expression level for 31085 genes.

```
In [6]: df_genes.shape
```

Out[6]: (200, 31085)

```
In [9]: df_genes.head(12)
```

Out[9]:

| | TPM_hugo_A1BG | TPM_hugo_A1BG-AS1 | TPM_hugo_A1CF | TPM_hugo_A2M | TPM_hugo_A2M-AS1 | TPM_hugo_A2ML1 | TPM_hugo_A2MP1 | TPM_hugo_A3GALT2 | TPM_ |
|----|---------------|-------------------|---------------|--------------|------------------|----------------|----------------|------------------|------|
| 0 | 1.564289 | 2.711834 | 0.000000 | 599.387994 | 2.354073 | 43.245808 | 0.0 | 0.000000 | |
| 1 | 3.487859 | 1.717013 | 0.000000 | 222.711937 | 2.288359 | 5.718716 | 0.0 | 0.564476 | |
| 2 | 0.613334 | 0.508520 | 0.000000 | 204.222937 | 0.627338 | 300.472716 | 0.0 | 0.000000 | |
| 3 | 2.385017 | 1.600782 | 0.000000 | 1851.589619 | 3.301540 | 1.346349 | 0.0 | 0.000000 | |
| 4 | 1.964353 | 0.791064 | 0.000000 | 982.752783 | 0.589165 | 85.088254 | 0.0 | 0.096887 | |
| 5 | 0.907173 | 0.349210 | 0.000000 | 684.072973 | 0.654054 | 113.067767 | 0.0 | 0.223721 | |
| 6 | 1.703079 | 1.925504 | 0.040575 | 1077.182707 | 0.625106 | 48.229871 | 0.0 | 0.000000 | |
| 7 | 0.476492 | 0.395063 | 0.000000 | 210.165565 | 0.128256 | 511.146137 | 0.0 | 0.091396 | |
| 8 | 0.633058 | 0.224946 | 0.000000 | 137.755978 | 0.219083 | 5.613576 | 0.0 | 0.000000 | |
| 9 | 1.047450 | 0.651337 | 0.009150 | 729.116811 | 0.380617 | 12.210105 | 0.0 | 0.000000 | |
| 10 | 0.636399 | 0.296799 | 0.013898 | 668.243637 | 1.252610 | 6.273756 | 0.0 | 0.137326 | |
| 11 | 2.505328 | 2.945830 | 0.039792 | 924.930657 | 1.177047 | 202.212006 | 0.0 | 0.000000 | |

12 rows × 31085 columns

Response status

Binary.

„1” – response for treatment
was observed

„0” – no response for treatment

```
In [31]: df_y.shape
```

```
Out[31]: (200, 1)
```

```
In [32]: df_y.head()
```

```
Out[32]:
```

| | 0 |
|---|---|
| 0 | 0 |
| 1 | 1 |
| 2 | 0 |
| 3 | 0 |
| 4 | 1 |

```
In [33]: df_y[df_y[0]==0].count()
```

```
Out[33]: 0    154
dtype: int64
```

```
In [34]: df_y[df_y[0]==1].count()
```

```
Out[34]: 0     46
dtype: int64
```

Our team



Dealing with incomplete data

```
In [11]: df_cv.apply(lambda x: 200 - x.count(), axis=0)
```

```
Out[11]: FMOne mutation burden per MB      45  
Neoantigen burden per MB      57  
Enrollment IC      0  
IC Level      1  
TC Level      1  
Immune phenotype      34  
Sex      0  
TCGA Subtype      0  
Lund      0  
Lund2      0  
Received platinum      0  
Met Disease Status      17  
Sample age      0  
Sample collected pre-platinum      49  
Intravesical BCG administered      0  
Baseline ECOG Score      0  
Tobacco Use History      0  
dtype: int64
```

```
In [13]: df_cv.dropna().shape
```

```
Out[13]: (68, 17)
```

Remove incomplete records?

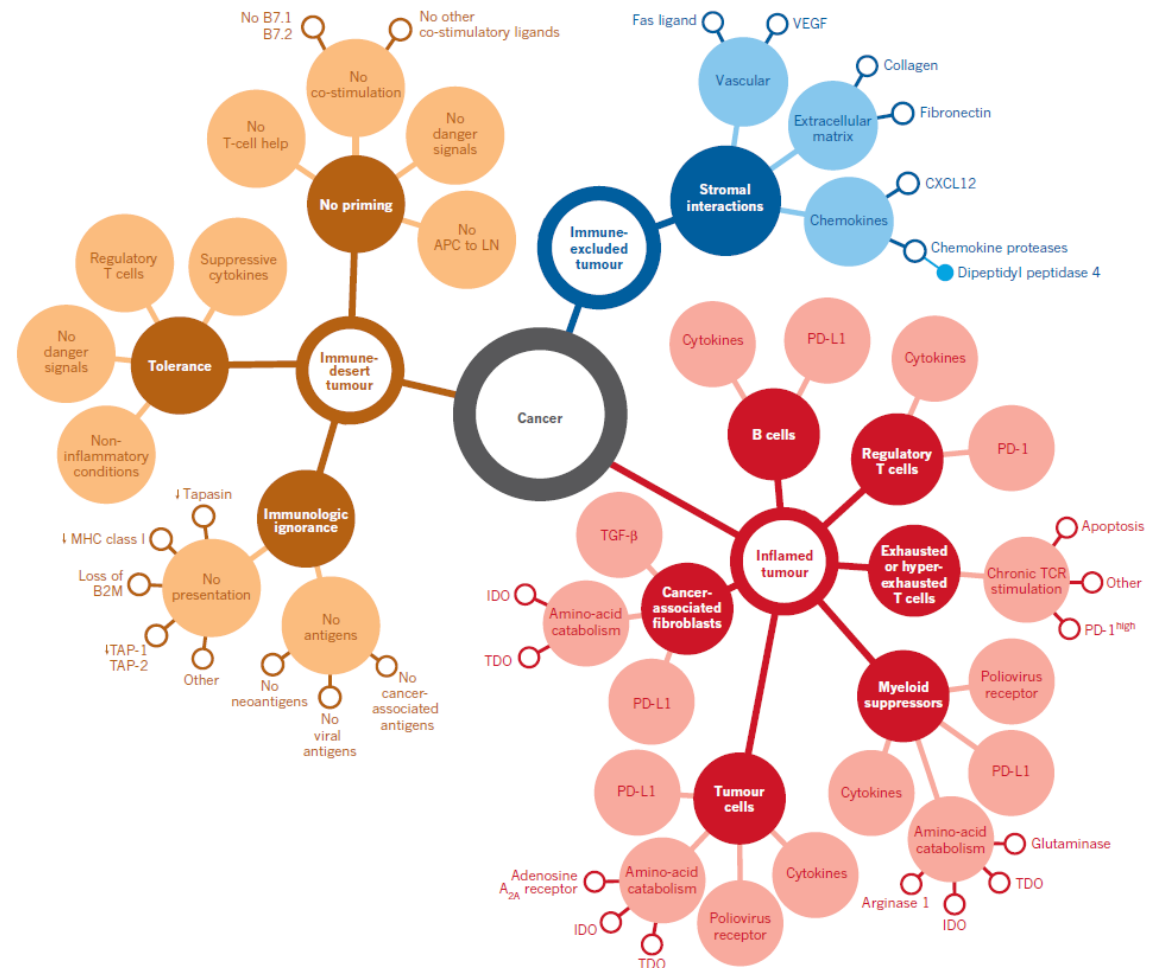
Fill with artificial value?

Try to estimate?

Features selection

Use clinical data? Genes? Both?

Are some columns more important?



Literature research

| <u>Immune checkpoints</u> | <u>DNA repair</u> | <u>Suppression cytokines</u> | <u>Inflammation cytokines</u> | <u>Interferon genes</u> | <u>Peptide binding complex</u> | <u>MHC class I</u> | <u>MHC class II</u> | <u>Macrophage-specific genes</u> | <u>Chemokine-related genes</u> | <u>Growth factor receptors</u> | <u>Chaperones</u> | <u>Autophagy-related genes</u> | <u>Extracellular matrix</u> | <u>Other</u> | <u>Clinical</u> |
|---------------------------|-------------------|------------------------------|-------------------------------|-------------------------|--------------------------------|--------------------|---------------------|----------------------------------|--------------------------------|--------------------------------|-------------------|--------------------------------|-----------------------------|--------------|--------------------|
| PDCD1 | MLH1 | IL4 | IL1A | IFNA1 | TAP1 | HLA-A | HLA-DPA1 | MSR1 | CCL4 | FGFR1 | HSP90AA1 | ATG10 | FN1 | FASLG | Mutation burden |
| CD274 | MSH2 | IL10 | IL1B | IFNA2 | TAP2 | HLA-B | HLA-DPB1 | CD58 | CCL8 | FGFR2 | HSP90AB1 | ATG101 | | DPP4 | Neoantigen burden |
| CD276 | MSH6 | IL13 | NOD1 | IFNA4 | B2M | HLA-C | HLA-DPB2 | CLEC7A | CXCL9 | EGFR | HSP90AB4P | ATG12 | | MARCH1 | Immune phenotype |
| CTLA-4 | PMS2 | IL17A | NOD2 | IFNA5 | TAPBP | HLA-E | HLA-DQA1 | FCGR1A | CXCL10 | ERBB2 | HSP90B1 | ATG13 | | PDCD1LG2 | |
| IDO1 | | TGFB | IL12A | IFNA6 | CALR | HLA-F | HLA-DQA2 | FCGR1B | CXCL11 | ERBB3 | HSP90B2P | ATG14 | | TIGIT | Met disease status |
| LAG3 | | | IL23A | IFNA7 | CANX | HLA-F-AS1 | HLA-DQB1 | FCGR1C | GZMA | ERBB4 | HSPA12A | ATG16L1 | | CD27 | |
| VTCN1 | | | TNF | IFNA8 | ERAP1 | HLA-G | HLA-DQB1-AS1 | FCGR3A | GZMB | VEGFA | HSPA12B | ATG16L2 | | CCL5 | |
| HAVCR2 | | | IL2 | IFNA10 | ERAP2 | HLA-L | HLA-DQB2 | | CXCR6 | VEGFB | HSPA13 | ATG2A | | PSMB10 | |
| TNFRSF4 | | | | IFNA13 | | HLA-H | HLA-DRA | | | | HSPA14 | ATG2B | | STAT1 | |
| IDO2 | | | | IFNA14 | | HLA-J | HLA-DRB1 | | | | HSPA1A | ATG3 | | NKG7 | |
| TDO2 | | | | IFNA16 | | | HLA-DRB3 | | | | HSPA1B | ATG4A | | CMKLR1 | |
| CD80 | | | | IFNA17 | | | HLA-DRB4 | | | | HSPA1L | ATG4B | | | |
| CD86 | | | | IFNA21 | | | HLA-DRB5 | | | | HSPA2 | ATG4C | | | |
| CD8A | | | | IFNA22P | | | HLA-DRB6 | | | | HSPA4 | ATG4D | | | |
| | | | | IFNB1 | | | HLA-DMA | | | | HSPA4L | ATG5 | | | |
| | | | | IFNE | | | HLA-DMB | | | | HSPA5 | ATG7 | | | |
| | | | | IFNG | | | HLA-DOA | | | | HSPA6 | ATG9A | | | |
| | | | | IFNG-AS1 | | | HLA-DOB | | | | HSPA7 | ATG9B | | | |
| | | | | IFNK | | | | | | | HSPA8 | | | | |
| | | | | IFNL1 | | | | | | | HSPA9 | | | | |
| | | | | IFNL2 | | | | | | | HSPB1 | | | | |
| | | | | IFNL3 | | | | | | | HSPB11 | | | | |
| | | | | IFNL4 | | | | | | | | | | | |
| | | | | IFNW1 | | | | | | | | | | | |

166 genes

117 directly involved in immune regulation

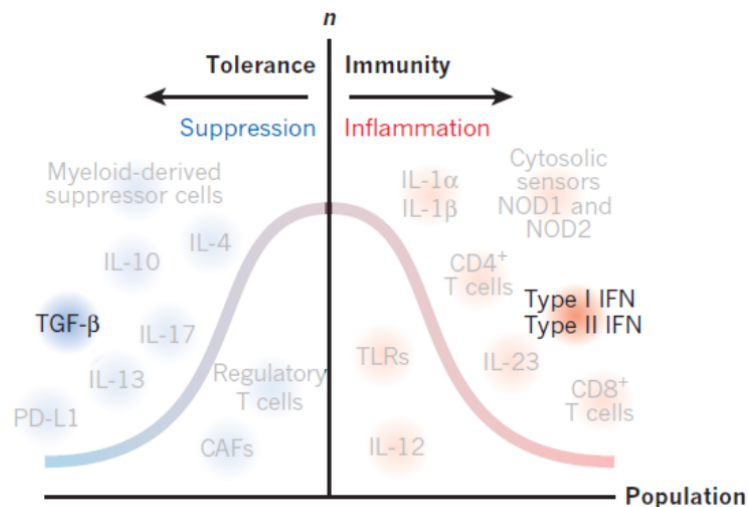
4 clinical features

Statistics revealed best markers

2 genes

IFNG

TGFB1



Chen et al., 2017

3 clinical features

MTB

Neoantigen burden

Met disease status

Basic neural network
10x cross-validated

Average accuracy: 0.76
Average ROC: 0.81

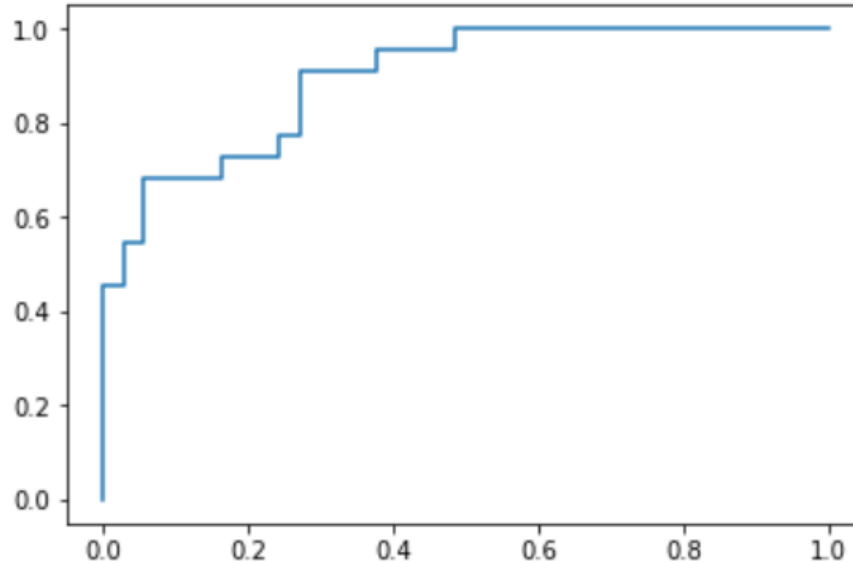
Area under ROC

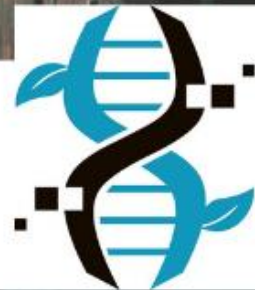
```
In [24]: # Scoring
score = roc_auc_score(y_test[0], predictions[1])
print("ROC AUC score: ", score)

fpr, tpr, thresholds = roc_curve(y_test[0], predictions[1], pos_label=1)

# ROC curve graph
plt.plot(fpr, tpr)
plt.show()
```

ROC AUC score: 0.8955773955773956



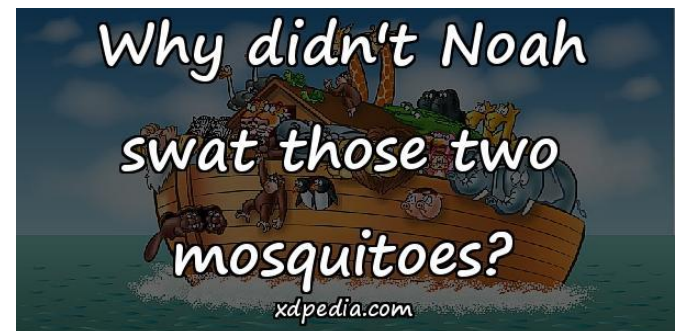


BioHack
bioinformatics hackathon



Thank you for your attention

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