

3. (a).

$$\dot{m}_i = r_{x,i} \bar{u}_i - (\mu + \theta_{m,i}) m_i$$

$$\dot{p}_i = r_{L,i} w_i - (\mu + \theta_{p,i}) p_i$$

At steady state. $\dot{m}_i = 0$. $\dot{p}_i = 0$.

$$\dot{m}_i = 0 \quad r_{x,i} \bar{u}_i = (\mu + \theta_{m,i}) m_i^*$$

$$m_i^* = \frac{r_{x,i}}{\mu + \theta_{m,i}} \bar{u}_i \simeq k_{x,i} \bar{u}_i$$

$$\dot{p}_i = 0 \quad r_{L,i} w_i^* = (\mu + \theta_{p,i}) p_i^*$$

$$p_i^* = \frac{r_{L,i}}{\mu + \theta_{p,i}} w_i^* \simeq k_{L,i} w_i^*$$

w_i^* describes the translation process and is related to m_i^*

$$w_i^* = w_i \cdot m_i^* \quad \text{Thus, } \underline{p_i^* \simeq k_{L,i} \cdot k_{x,i} \bar{u}_i w_i}$$

(b).
$$r_{L,i} = K_{E,i}^L R_{LT} \left(\frac{m_i}{\tau_{L,i} k_{L,i} + (\tau_{L,i} + 1) m_i} \right)$$

Since $\tau_{L,i} k_{L,i} \gg (\tau_{L,i} + 1) m_i$

$$\rightarrow r_{L,i} = K_{E,i}^L R_{LT} \frac{m_i}{\tau_{L,i} k_{L,i}}$$

$$r_{x,i} = K_{E,i}^X R_{XT} \left(\frac{g_i}{\tau_{x,i} k_{x,i} + (\tau_{x,i} + 1) g_i} \right)$$

To sum up:

$$P_i^* = \frac{k_{E,i}^L R_{LT}}{\underbrace{(\mu + \theta_{p,i}) (\tau_{L,i} k_{L,i})}_{k_{L,i}}} \cdot \underbrace{\frac{k_{E,i}^X R_{XT}}{\mu + \theta_{m,i}} \left(\frac{g_i}{\tau_{X,i} k_{X,i} + (\tau_{X,i} + 1) g_i} \right)}_{K_{X,i}} \bar{\mu}_i \cdot w_i$$

According to Prelim 1 Q1. $K_{X,i} = 0.575 \text{ nmol/gDW}$.

$$\bar{\mu}_i = \frac{k_1 + k_2 f_1}{1 + k_1 + k_2 f_1} \quad f_1 = \frac{1^n}{K^n + 1^n}$$

$$\text{cell dry weight} = 4.3 \times 10^{-13} \text{ g} \times 0.3 = 1.29 \times 10^{-13} \text{ g.} \quad V = 1 \mu\text{m}^3 = 10^{-3} \text{ L.}$$

$$K_{L,i} = 200 \mu\text{M} = 200 \mu\text{mol/L} \times 10^{-3} \text{ L} = 200 \text{ nmol} / (1.29 \times 10^{-13} \text{ g}) = 1.55 \times 10^{15} \text{ nmol/gDW}$$

$$\text{Translation Rate} = 14 \text{ AA/s} \quad \text{Length} = 333 \text{ AA} \quad \text{Time} = 333/14 \text{ s.}$$

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$$k_{E,i}^L = 14/333 = 0.04204 \text{ s}^{-1}$$

translation elongation rate

$$R_{LT} = 1000 \text{ ribosome/cell.}$$

ribosome conc.

$$\theta_{p,i} = \frac{\ln 2}{24 \text{ h}} = \frac{\ln 2}{(24 \times 3600) \text{ s}} = 8.0225 \times 10^{-6} \text{ s}^{-1}$$

degradation

$$\mu = \frac{\ln 2}{40 \text{ min}} = \frac{\ln 2}{(40 \times 60) \text{ s}} = 2.8881 \times 10^{-4} \text{ s}^{-1}$$

dilution

$$\tau_{L,i} = \frac{k_{E,i}^L}{k_1} = \frac{14/333 \text{ s}^{-1}}{1/1.5 \text{ s}^{-1}} = 7/111 = 0.06306$$

time constant

$$K_{L,i} = 1.45 \times 10^{-3} \text{ mmol/gDW}$$

$$P_i^* = 8.33 \times 10^{-4} \bar{\mu}_i$$

Plot P_i^* curve wrt $\bar{\mu}_i$ in Excel [Fig. 1].

(C).

If polysome amplification constant $k_p > 1$

$$k'_{L,i} = k_p k_{L,i} > k_{L,i}$$

p_i^* will be larger, the curve will move upward and become steeper as a function of \bar{u}_i .

Plot p_i^* curve with larger k' in Excel. [Fig.2]

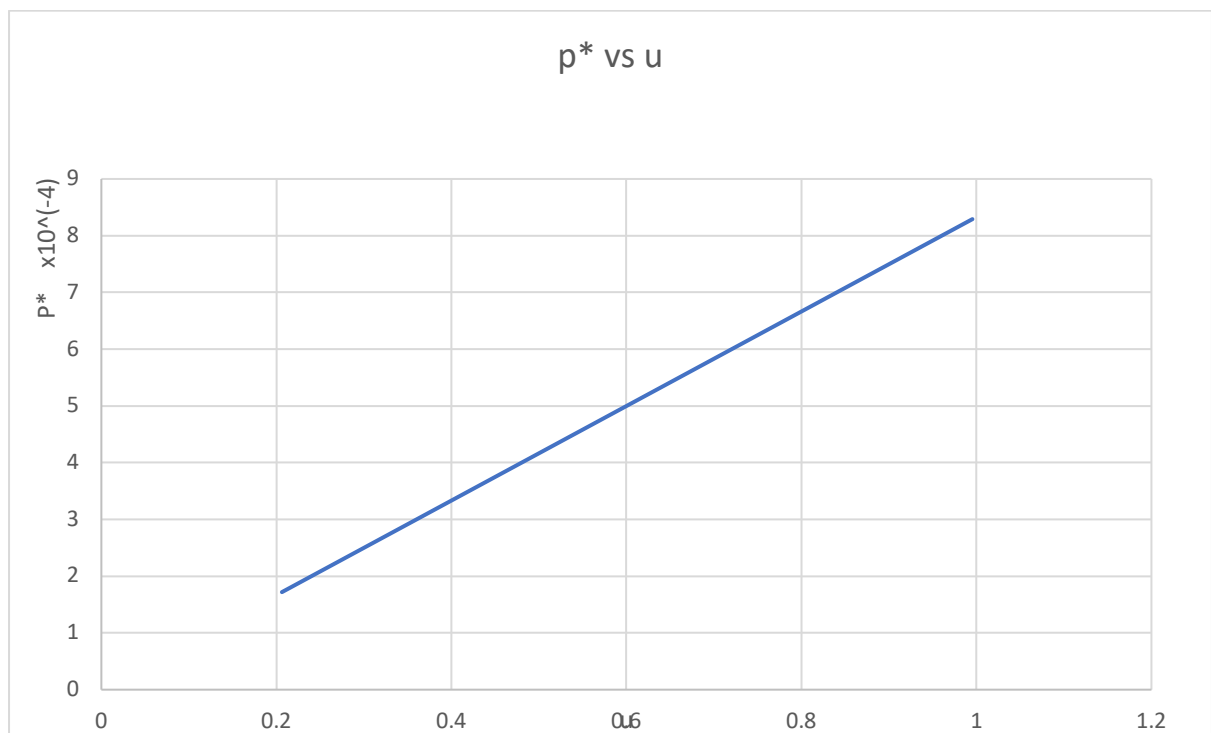


Fig.1

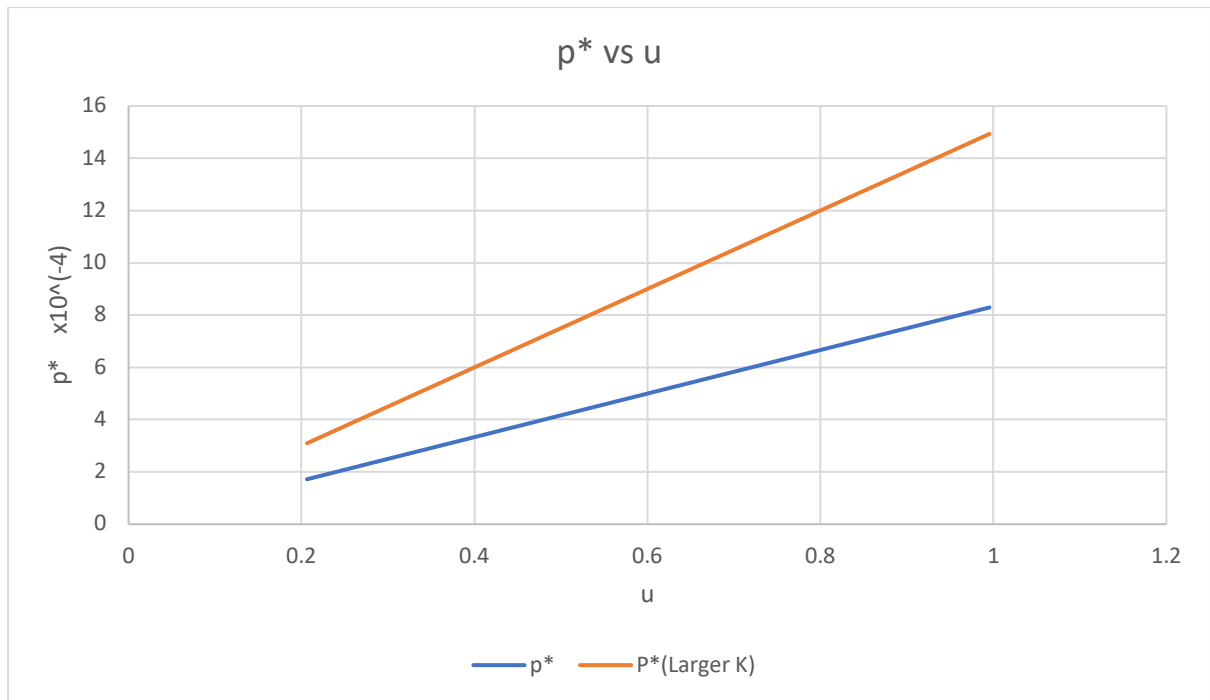


Fig.2

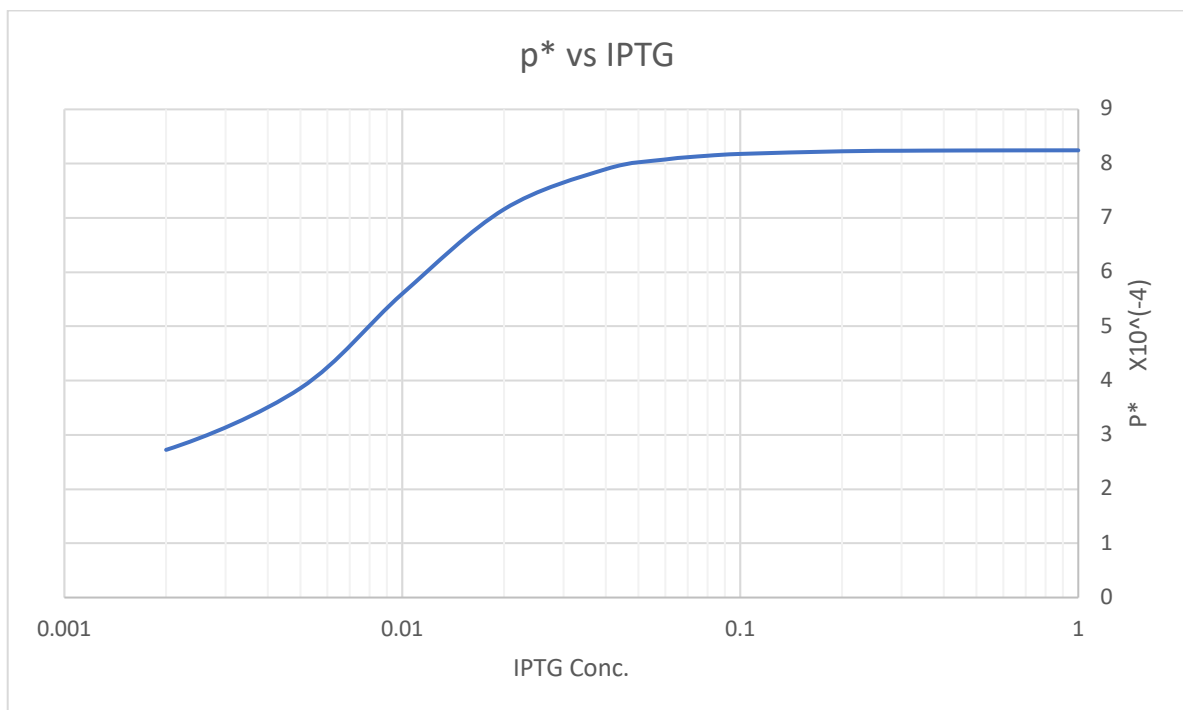


Fig.3

We can also plot the p^* with IPTG concentration. Referring to the data and equation obtained from Prelim Q1, the curve is shown in Fig.3.

Additional equation:

$$u = (W1+W2*f[I])/(1+W1+W2*f[I])$$

$$f[I] = ([I]^n)/(KD^n + [I]^n)$$

$$KD = 0.09$$

$$n = 1.85$$

$$W1 = 0.4$$

$$W2 = 98$$