Gibbs Sampling with Data Augmentation for Bayesian Analysis of Binary and Polychotomous Response Data

STAT 230 Final Report

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1 Abstract

Here's the abstract.

2 Introduction

We cite Albert and Chib (1993)

Here's the introduction.

3 Methodology

Here's the methodology.

4 Results

For three datasets, we apply the appropriate previously discussed model as well as standard maximum likelihood estimation and custom models written in Stan (Carpenter et al. 2017) and impelemented in the RStan package (Stan Development Team 2019).

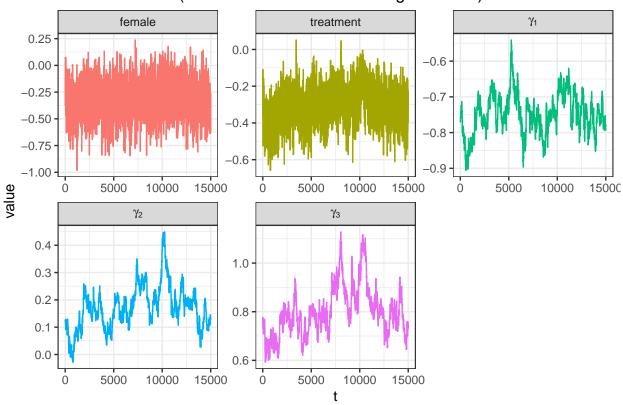
4.1 Small-Cell Carcinoma

The small-cell carcinoma data comes from our STAT 211 class. Small-cell carcinoma of the lung is an aggressive cancer that can be treated with chemotherapy. Patients with small-cell carcinoma were randomly assigned to one of two therapy options. The outcome of the therapy is recorded on an ordinal scale (1: Progressive, 2: No Change, 3: Partial Remission, 4: Complete Remission). We fit multinomial ordered probit regression models with with therapy option and sex as predictors of outcome. The model parameters are estimated with data augmented Gibbs sampling following the algorithm described in Albert and Chib (1993) and implemented in R, a custom model built in Stan, and traditional maximum likelihood estimation as implemented in the polr function in the MASS package (Venables and Ripley 2002). Both Bayesian methods are run to generate 4000 posterior samples, with the first 1000 discarded as burn-in samples, leaving 3000 samples for analysis. The Stan model took 1 minute and 13,9 seconds seconds to run, while the R model completed in 2.4 seconds.

Table 1: Posterior sample summaries for small-cell carcinoma data using R-Gibbs

Variable	mean	sd	2.5%	50%	97.5%
female	-0.3177006	0.1695306	-0.6483304	-0.3154434	0.0143534
treatment	-0.3081612	0.1039444	-0.5136893	-0.3092713	-0.1059862
gamma[1]	-0.7478856	0.0569057	-0.8576571	-0.7492453	-0.6420117
gamma[2]	0.1726966	0.0795173	0.0235177	0.1732758	0.3510630
gamma[3]	0.8011110	0.1027040	0.6390310	0.7890847	1.0615511

Posterior Trace (Small-Cell Carcinoma Using R-Gibbs)



Posterior Distribution (Small-Cell Carcinoma Using R-Gibbs)

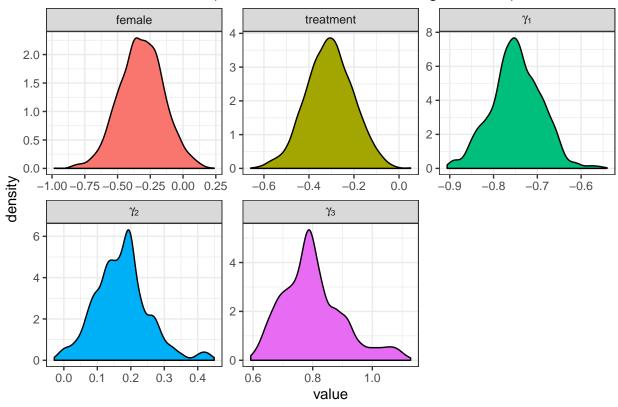
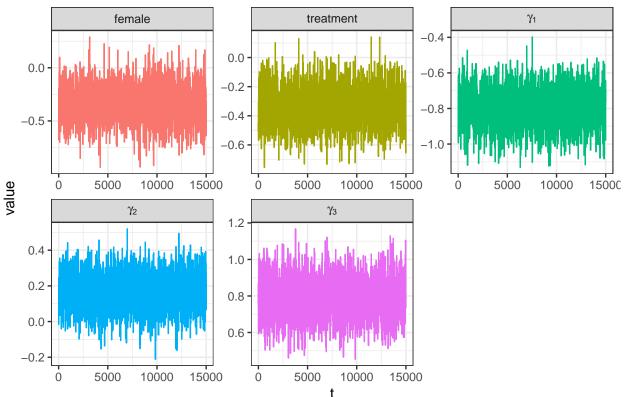


Table 2: Posterior sample summaries for small-cell carcinoma data using Stan $\,$

Variable	mean	sd	2.5%	50%	97.5%
female	-0.3383656	0.1702371	-0.6777031	-0.3369185	-0.0045383
treatment	-0.3311899	0.1282283	-0.5829356	-0.3309320	-0.0730097
gamma[1]	-0.7992210	0.1048007	-1.0079013	-0.7988986	-0.5917817
gamma[2]	0.1684474	0.1002620	-0.0302685	0.1692335	0.3625867
gamma[3]	0.7914654	0.1058342	0.5811806	0.7903738	1.0012122

Posterior Trace (Small-Cell Carcinoma Using Stan)



Posterior Distribution (Small-Cell Carcinoma Using Stan)

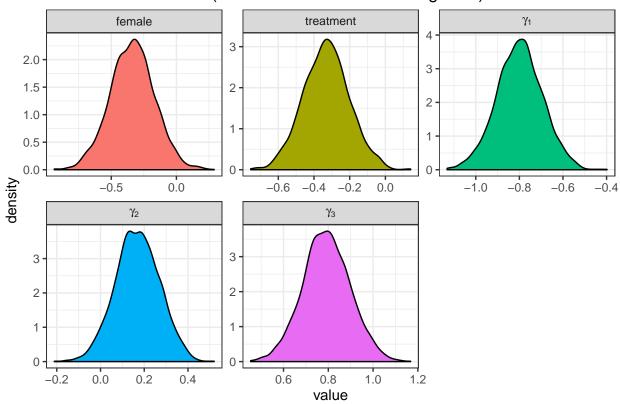


Table 3: MLE estimates for small-cell carcinoma data

Variable	Estimate	Std. Error
female	-0.3401606	0.1749021
treatment	-0.3344764	0.1254351
gamma[1]	-0.7994983	0.1053802
gamma[2]	0.1649279	0.0988553
gamma[3]	0.7818721	0.1068102

4.2 Breast Cancer Data

We evaluate the probit regression methods with the Wisconsin Breast Cancer dataset, obtained from the University of Wisconsin Hospitals, Madison from Dr. William H. Wolberg (Wolberg and Mangasarian 1990). The data reports 9 discrete measurements for 699 observations of clumps of breast cancer cells as well as the response variable, indicating whether or not the clump is malignant (1) or benign (0). The variables are Clump Thickness, Uniformity of Cell Size, Uniformity of Cell Shape, Marginal Adhesion, Single Epithelial Cell Size, Bare Nuclei, Bland Chromatin, Normal Nucleoli, and Mitoses. Only the Bare Nuclei variable included any missing data, so it was not included in this analysis. The model parameters are estimated with data-augmented Gibbs sampling following the algorithm described in Albert and Chib (1993) and implemented in R, a custom model built in Stan, and traditional maximum likelihood estimation as implemented in the glm function in R (R Core Team 2019). Both Bayesian methods are run to generate 4000 posterior samples, with the first 1000 discarded as burn-in samples, leaving 3000 samples for analysis. The Stan model took 6 minutes and 10.5 seconds seconds to run, while the R model completed in 7.8 seconds.

Table 4: Posterior sample summaries for breast cancer data using R-Gibbs

Variable	mean	sd	2.5%	50%	97.5%
(Intercept)	-5.2657995	0.4516886	-6.0869080	-5.2781142	-4.3524946
Clump Thickness	0.3030510	0.0570216	0.1891526	0.3017604	0.4131211
Uniformity of Cell Size	0.0229394	0.0891064	-0.1458150	0.0219254	0.2035911
Uniformity of Cell Shape	0.3118907	0.1020941	0.1119921	0.3143989	0.5132866
Marginal Adhesion	0.1532512	0.0538371	0.0546572	0.1515937	0.2611788
Single Epithelial Cell Size	0.0733985	0.0731480	-0.0708377	0.0732059	0.2141101
Bland Chromatin	0.2863431	0.0725573	0.1497924	0.2856310	0.4414197
Normal Nucleoli	0.0509178	0.0490190	-0.0488792	0.0528914	0.1421364
Mitoses	0.2845107	0.1376854	0.0269171	0.2745250	0.5957231



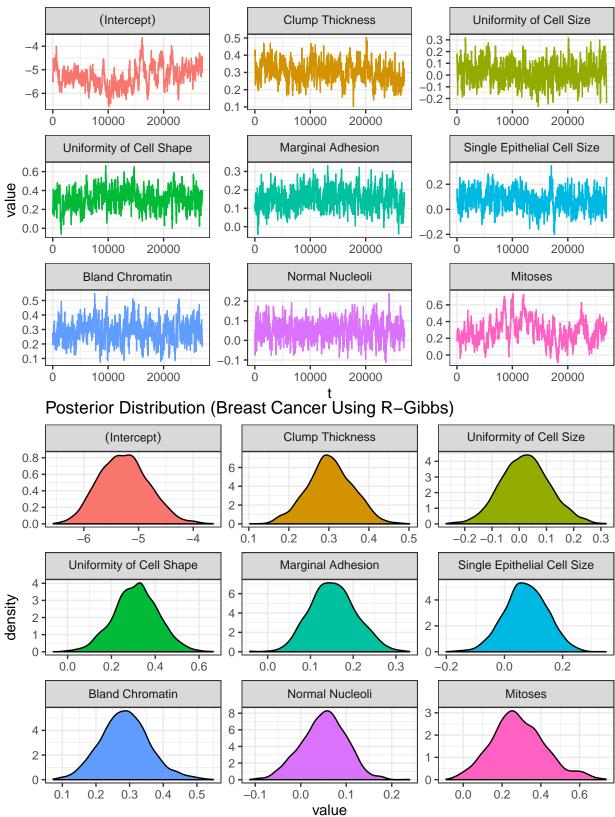
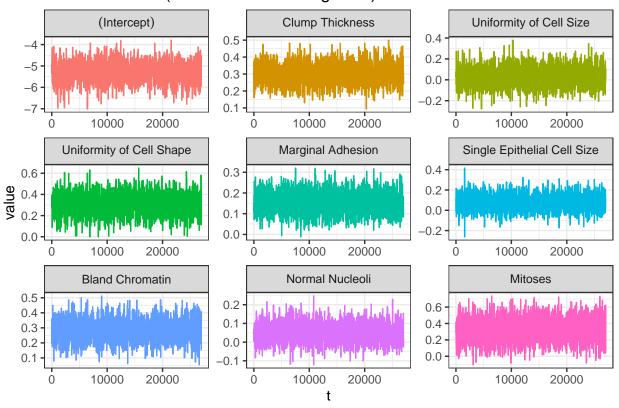


Table 5: Posterior sample summaries for breast cancer data using Stan $\,$

Variable	mean	sd	2.5%	50%	97.5%
(Intercept)	-5.2803740	0.4474952	-6.1788551	-5.2675580	-4.4570687
Clump Thickness	0.3020233	0.0581248	0.1891183	0.3010757	0.4166279
Uniformity of Cell Size	0.0283373	0.0899331	-0.1404464	0.0256264	0.2084892
Uniformity of Cell Shape	0.3066612	0.0950854	0.1178779	0.3088803	0.4907881
Marginal Adhesion	0.1525317	0.0502227	0.0576146	0.1520253	0.2512294
Single Epithelial Cell Size	0.0739874	0.0750008	-0.0656497	0.0713772	0.2254342
Bland Chromatin	0.2812440	0.0686278	0.1482095	0.2824382	0.4200620
Normal Nucleoli	0.0526038	0.0495117	-0.0427017	0.0537263	0.1511080
Mitoses	0.3191038	0.1369070	0.0561501	0.3168852	0.5875316

Posterior Trace (Breast Cancer Using Stan)



Posterior Distribution (Breast Cancer Using Stan)

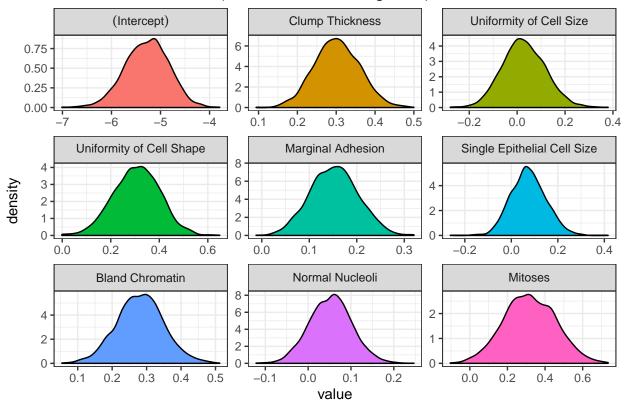


Table 6: MLE estimates for breast cancer data

Variable	Estimate	Std. Error
(Intercept)	-5.1320418	0.4516719
Clump Thickness	0.2897958	0.0572147
Uniformity of Cell Size	0.0156388	0.0871703
Uniformity of Cell Shape	0.3037402	0.0931808
Marginal Adhesion	0.1493079	0.0518700
Single Epithelial Cell Size	0.0715684	0.0743288
Bland Chromatin	0.2758152	0.0715357
Normal Nucleoli	0.0538840	0.0509148
Mitoses	0.3109816	0.1430757

4.3 Baseball

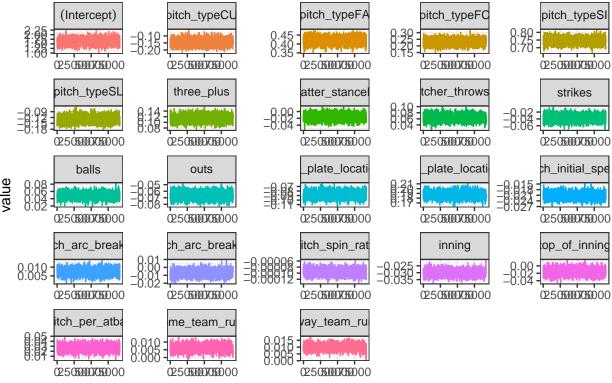
We evaluate the performance of data-augmented Gibbs sampling on a larger datasets (more predictors and observations than the breast cancer data), by fitting probit regression models on baseball data. The baseball data comes from Major League Baseball pitch tracking software and was prepared by the Los Angeles Dodgers for the purpose of predicting whether or not a pitch was put into play. There are 100,000 pitches (observations) thrown by Dodger's starting pitchers and 19 features of each pitch, including velocity, spin rate, and location. The binary response indiactes whether or not a pitch was put into play (0: not put into play, 1: put into play). Missing data was removed, leaving 99,254 observations with 18 covariates for analysis. The model parameters are estimated with data augmented Gibbs sampling following the algorithm described in Albert and Chib (1993) and implemented in R and traditional maximum likelihood estimation as implemented in the glm function in R (R Core Team 2019). Both Bayesian methods are run to generate 5000 posterior samples, with the first 1000 discarded as burn-in samples, leaving 4000 samples for analysis. We also attempted to fit

a custom model in Stan but the samples were generated very slowly (fewer than 500 samples per hour). In contrast, the R model generated all 5000 samples in 15 miutes and 41 seconds.

Table 7: Posterior sample summaries for baseball data using R-Gibbs

Variable	mean	sd	2.5%	50%	97.5%
(Intercept)	1.6419615	0.1641598	1.3275671	1.6432777	1.9577907
pitch_typeCU	-0.1427578	0.0223181	-0.1875506	-0.1427039	-0.0990722
$pitch_typeFA$	0.4187926	0.0181987	0.3820077	0.4188624	0.4544004
$pitch_typeFC$	0.2290787	0.0213407	0.1871727	0.2289236	0.2717159
$pitch_typeSI$	0.7452006	0.0191260	0.7073330	0.7452225	0.7822887
$pitch_typeSL$	-0.1282329	0.0162129	-0.1598217	-0.1282685	-0.0963311
three_plus	0.1138644	0.0114908	0.0918353	0.1136950	0.1355962
$batter_stanceR$	-0.0157872	0.0088088	-0.0331719	-0.0157398	0.0011848
$pitcher_throwsR$	0.0640037	0.0100326	0.0445198	0.0640910	0.0836733
strikes	-0.0369098	0.0091613	-0.0552714	-0.0368880	-0.0193004
balls	0.0487882	0.0079058	0.0331798	0.0489391	0.0639353
outs	-0.0660833	0.0050520	-0.0760450	-0.0661052	-0.0563236
$pitch_plate_location_x$	-0.0880404	0.0067681	-0.1014258	-0.0879550	-0.0749833
$pitch_plate_location_z$	0.1874257	0.0063152	0.1750398	0.1874246	0.1995573
$pitch_initial_speed$	-0.0211755	0.0016717	-0.0244022	-0.0212001	-0.0179145
$pitch_arc_break_x$	0.0073551	0.0017686	0.0038227	0.0073578	0.0107850
$pitch_arc_break_z$	-0.0068673	0.0038864	-0.0145743	-0.0068384	0.0005285
pitch_spin_rate	-0.0000956	0.0000100	-0.0001149	-0.0000956	-0.0000758
inning	-0.0301068	0.0020301	-0.0340363	-0.0301332	-0.0260424
top_of_inning	-0.0157989	0.0084155	-0.0318350	-0.0159557	0.0010927
pitch_per_atbat	0.0263585	0.0060101	0.0148513	0.0263168	0.0383177
$home_team_runs$	0.0062439	0.0020007	0.0022450	0.0062631	0.0100559
$away_team_runs$	0.0099700	0.0019942	0.0061492	0.0099993	0.0137278





Posterior Distribution (Baseball Using R-Gibbs)

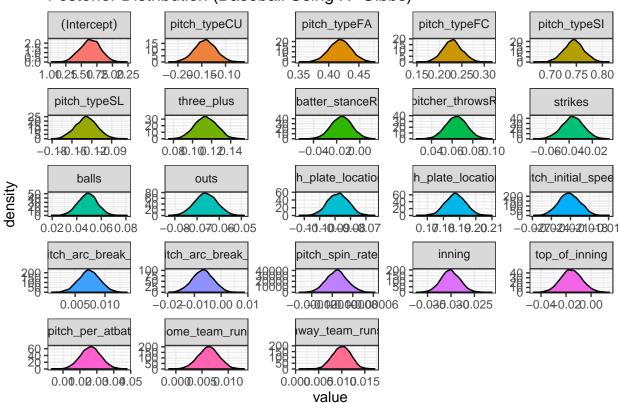


Table 8: MLE estimates for baseball data

Variable	Estimate	Std. Error
(Intercept)	1.6457301	0.1643384
pitch_typeCU	-0.1425477	0.0226587
pitch_typeFA	0.4184923	0.0185715
pitch_typeFC	0.2282183	0.0219511
pitch_typeSI	0.7453712	0.0188182
$pitch_typeSL$	-0.1279136	0.0167241
three_plus	0.1136658	0.0114023
$batter_stanceR$	-0.0156687	0.0088127
$pitcher_throwsR$	0.0637459	0.0099670
strikes	-0.0369765	0.0102701
balls	0.0489444	0.0088985
outs	-0.0661775	0.0050947
pitch_plate_location_x	-0.0881536	0.0068380
pitch_plate_location_z	0.1874354	0.0063953
pitch_initial_speed	-0.0212061	0.0016916
$pitch_arc_break_x$	0.0073317	0.0017433
pitch_arc_break_z	-0.0067832	0.0038488
pitch_spin_rate	-0.0000956	0.0000100
inning	-0.0300979	0.0020700
top_of_inning	-0.0160490	0.0083282
pitch_per_atbat	0.0262550	0.0070157
home_team_runs	0.0062091	0.0019981
away_team_runs	0.0100125	0.0020411
- There were 23 model</td <td>parameters –</td> <td>></td>	parameters –	>

5 Discussion

Here's the discussion.

6 References

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Wolberg, William H, and Olvi L Mangasarian. 1990. "Multisurface Method of Pattern Separation for Medical Diagnosis Applied to Breast Cytology." *Proceedings of the National Academy of Sciences* 87 (23). National Academy of Sciences: 9193–6. https://doi.org/10.1073/pnas.87.23.9193.

7 Appendix