Winter temperatures dominate spring phenological responses to warming

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1   **Abstract**

2   Decades of fundamental research on woody plant species highlight three major cues that shape spring

phenological events: chilling, forcing, and photoperiod *(e.g., 1; 2; 3)*. Increasing research on the phenological impacts of climate change has led to debate over how common these cues are across species, and—if prevalent—whether chilling and/or photoperiod cues may be slowing phenological responses to warming in recent years *(4; 5; 6)*. Here we use a global meta-analysis of all published controlled environment studies to test for the relative effects of these cues across 67 species. We find almost all species show strong responses to all three cues, with chilling being the strongest (1.9X greater than forcing), and photoperiod the weakest (0.7X forcing). Simple forecasts from our findings for a well-studied region (Central Europe) suggest that spring phenology will continue to advance, as stalling effects of chilling generally appear above 4°C warming for most locations. Our results unify both sides of the debate over phenological cues: while all species may respond to all cues strongly in experimental conditions, in current environmental conditions the dominant signal of climate change is from increased forcing. Further progress to improve budburst forecasts under future climate change will require fully separating chilling and forcing effects at the physiological-level.

Main text

For decades, plant phenology has been one of the most reported and consistent biological imprints of climate change *(7)*: many temperate plants are leafing and flowering days to weeks earlier with rising temperatures *(8; 9)*. Understanding such shifts is important as phenology shapes community assembly and a suite of ecosystem services, including pollination and carbon sequestration, and scales up to impact projections of climate change itself *(10)*.

As research interest in phenology has progressed, critical discrepancies and uncertainties in our understanding have emerged. Though responses to warming are consistent on average, they show substantial variation among species and sites *(11)*. Furthermore, long-term observational studies provide increasing evidence that sensitivities of phenology to temperature are weakening in recent decades *(12; 13; 14)*, especially in Europe, where researchers suggest that responses to multiple environmental cues underlie declining forcing temperature sensitivities *(5)*.

Fundamental research in phenology outlines how three major environmental cues, chilling (cool temperatures, generally occurring in the fall through late winter), forcing (warm temperatures, generally occurring in the late winter through early spring), and photoperiod (daylength), provide multiple routes to budburst each spring, depending on the environment *(15)*. For example, in some species a cool winter will lower the amount

of forcing required to trigger budburst, compared to a warmer winter *(16)*. Additionally, photoperiod may trigger budburst, given low chilling and/or forcing *(6; 17; 18)*. Research suggests that all three cues may affect spring phenology for many temperate woody species *(3; 17; 19)*, which could have critical forecasting implications—predicting delays in spring phenology as increased warming reduces chilling in some areas *(20)* or where earlier budburst shortens the experienced photoperiod. However, there is strong debate, with research suggesting some cues—such as photoperiod—may be effectively absent in some species, but dominate in others *(6; 21; 17; 22)*.

Resolving this debate requires overcoming major hurdles to estimate responses to each cue. Studies attempting to estimate cues using long-term observational data *(e.g., 6; 23)* generally fail to overcome the fundamental challenge that cues are strongly correlated in nature (*e.g.*, during the seasonal transition from winter to spring at temperate latitudes forcing and photoperiod usually increase in step for a given location; average chilling and spring (forcing) temperatures can be positively correlated in space, especially at high latitudes, Fig. **??**). In contrast to observational studies and experimental field warming studies designed to test higher temperatures in natural conditions *(11)*, controlled environment experiments can break down correlations between the cues. These experiments, which generally rely on dormant tree cuttings or dormant plants exposed to controlled temperature and light regimes in growth chambers (Fig. 1), have been shown to replicate whole-plant responses in nature *(24)*. Such experiments have been conducted for decades (though each experiment generally lasts under a year). They have produced contrasting results, however, potentially due to differences in focal species or study sites *(6; 18; 25; 26; 27)*. Resolving these discrepancies is critical to accurate predictions of spring phenology, especially as continued climate change will yield warmer temperatures than have been experienced in at least the last 150 years *(28; 29; 30; 31; 32)*.

Here, we leverage controlled environment studies in a meta-analysis to understand how chilling, forcing, and photoperiod determine budburst timing in woody species. We reviewed 201 papers from controlled environment studies, then extracted data from all experiments that reported budburst responses, yielding data from 72 studies and 203 species (Fig. **??**, Tables **??**, **??**). The resulting Observed Spring Phenology Responses in Experimental Environments (OSPREE) database includes studies of dormant plant tissue (grown in greenhouses or taken directly from the field) exposed to experimental conditions *(33)* for which we could identify forcing, photoperiod, and chilling treatments quantitatively. Most experiments reported forcing and photoperiod treatments, while chilling occurred mainly in the field, though some studies additionally applied chilling before moving plants into forcing conditions (Fig. 1). Because chilling was rarely reported, we calculated

an estimate of chilling (both in the field and in experimental conditions), using a common but approximate method *(34)*, based on a hypothesis of how chilling accumulates *(35)*, with no chilling accumulating below 1.4°C or above 12.4°C (throughout the main text we use the term 'chill unit,' see Supplemental Materials, especially Table **??**, for details).

We estimated the effects of chilling, forcing, and photoperiod using a Bayesian hierarchical model. Our model averages over interactive effects of predictors, including only main effects that we could more robustly estimate given current study designs (see *Methods* in Supplemental Materials). Species are modeled hierarchically, producing estimates of both species-level responses (generally yielding more accurate estimates for well-studied species, such as *Fagus sylvatica* and *Betula pendula*), and the distribution from which they are drawn, yielding estimates of the overall responses across species (see *Methods* in Supplemental Materials):

$$y_i = \alpha_{sp[i]} + \beta_{forcing_{sp[i]}} + \beta_{photoperiod_{sp[i]}} + \beta_{chilling_{sp[i]}} + \epsilon_i$$

,

$$\epsilon_i \sim N(0, \sigma_y^2)$$

The α and each of the three β coefficients were modeled at the species level, as follows:

$$\alpha_{sp} \sim N(\mu_\alpha, \sigma_\alpha)$$

$$\beta_{forcing_{sp}} \sim N(\mu_{forcing}, \sigma_{forcing})$$

$$\beta_{photoperiod_{sp}} \sim N(\mu_{photoperiod}, \sigma_{photoperiod})$$

$$\beta_{chilling_{sp}} \sim N(\mu_{chilling}, \sigma_{chilling})$$

where i represents each unique observation, sp is the species (or species complex grouping, explained below), α represents the intercept, β terms represent slope estimates, and y is the days to budburst since forcing conditions were applied. Some species were represented in only one dataset in the OSPREE database, making it impossible to statistically differentiate between species and dataset for these taxa. To address this, we focus

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76 on estimates (reported as mean with 95% uncertainty intervals, unless otherwise noted) from a model of 67

77 taxa, which were included in multiple datasets and treatments (generally this occurred at the species-level,

78 but in some cases we collapsed species found in only one study into "complexes" at the level of genera, see the

79 *The Observed Spring Phenology Responses in Experimental Environments (OSPREE) database* section in the

80 Supplemental Materials for details). Estimates from this model were generally similar to estimates from a

81 model of all 203 species (Tables **??**, **??**). To directly compare the effects of chilling, forcing and photoperiod

82 we fit models using standardized predictor variables *(*following 36, which we refer to as "standard units") and

83 predictors in their natural units (chill units, °C, hours). We further fit several additional models, including a

84 model testing provenance latitude effects, one testing effects of chilling study design, and one testing effects

85 of life-stage (see *Models* in the Supplemental Materials for model equations and other details).

86 Across studies, all cues—chilling, forcing, and photoperiod—advance budburst phenology (Fig. 2, Tables **??**,

87 **??**). Chilling was the strongest cue (-8.33 days/standard unit [-11.5 to -5.3] or -2.76 days per chill unit [-3.65

88 to -1.89]), followed by forcing (-4.35 days/standard unit [-6.56 to -2.08] or -0.8 days per °C of warming, [-1.18

89 to -0.43]), and photoperiod (-2.96 days/standard unit [-5.43 to -0.56] or -0.53 days per hour of daylight [-0.92

90 to -0.15]; Figs.3, **??**, **??**; Tables **??**, **??**; see Supplemental Materials for more details). While photoperiod

91 had the smallest effect among the three cues, our results contrast with the extensive literature suggesting

92 photoperiod is an unimportant cue for many species *(6; 37)*—instead we found it was surprisingly large, even

93 when accounting for its interaction with provenance latitude (*i.e.*, the latitude of origin for plant material;

94 see Supplemental Materials for details, especially Figs. **??**, **??**, Table **??**). It was also generally consistent

95 across species (variance = 5.18 days/standard unit), only deviating in *Fagus sylvatica*, a species well-known

96 for having a large response to photoperiod (which we also found, Figs. 2, **??**). Species responses to chilling

97 were slightly more variable (variance = 7.2 days/standard unit, Fig. 2) than responses to forcing (variance

98 = 5.72 days/standard unit Fig. 2, Table **??**).

99 As temperature is fundamentally altered by climate change, our finding that different ends of the temperature

100 spectrum—chilling and forcing—have the strongest effects on budburst suggests that understanding these two

101 cues will be critical for forecasting phenology with climate change. Many previous studies attribute advances

102 in budburst to increased forcing *(9; 16; 17; 38)*. Our results, however, suggest that, across 67 species and 72

103 controlled environment studies, chilling has a greater effect on budburst than forcing (Figs. 2, **??**, **??**; Tables

104 **??**-**??**). This has not been widely suggested previously, perhaps because little work has directly manipulated

105 chilling, and the few studies that have were designed to compare chilling versus photoperiod effects *(e.g.,*

6; 17; 19; 25), not chilling versus forcing effects. Process-based phenological models, however, that explicitly model chilling often find this cue to be most critical (e.g., 39? ; 40, but see **(author?)** (41)).

Despite its apparent importance, chilling and its related physiological stage of endodormancy, are not well understood (15). Physiologically, plants appear to accumulate forcing only after they have exited endodormancy (and entered ecodormancy, Fig. 1), which is generally thought to occur when chilling requirements have been met (15). Thus, while researchers generally define "chilling" and "forcing" treatments based on temperatures in controlled environments (including in the studies used here, see Fig. 1), fully separating out what plants experience as chilling versus forcing will likely require new methods to measure endo- and ecodormancy (42).

Until then, researchers must generally rely on modeled estimates of chilling, as we have used here. Models of how species accumulate chilling are poorly developed for forest trees, with few relevant tests evaluating the particular temperatures at which species do or do not accumulate chilling. Instead, researchers generally rely on models developed for perennial fruit trees (i.e., Utah, Table **??**, 34) and chill portions (43), both of which were developed for peach species. These models are themselves hypotheses for how chilling may accumulate and produce dormancy release, but are likely to be inaccurate for many species (35). We found that applying a different chilling model did not strongly affect our estimates, however (i.e., 95% uncertainty intervals of estimates for chilling, photoperiod, and forcing overlapped for models using Utah and chill portions, with standardized predictors, Table **??**).

Progress on developing chilling models for wild species may be especially slow as only a small portion of studies (13 of the total 72 studies) manipulate chilling directly. Instead many studies estimate chilling effects through sequential removal of tissue from the field followed by exposure to "forcing" conditions (Fig. 1A,B, 25 out of 72; the remaining 34 studies did not appear to manipulate chilling), with the assumption that tissues collected later experience more chilling (44). This method benefits from more natural chilling conditions but introduces other challenges: first, chilling duration may not always co-vary with the magnitude of total accumulated chilling (35), and, second, photoperiod and other factors also change over time. Indeed, we found that sequential-removal studies tended to result in later budburst, weaker effects of chilling, and stronger effects of forcing compared to estimates from studies that directly manipulated chilling (Fig. **??**, Table **??** (44; 45). This suggests that a study's design of chilling manipulation impacts both forcing and chilling estimates and further supports that an improved understanding of chilling could in turn alter our understanding of forcing.

Linking such short-term controlled experiments to natural conditions robustly will require more efforts to understand the complex interactions between chilling, forcing, and photoperiod that we were not able to quantify in this meta-analysis. Most experimental studies do not test for interactions between all three cues (Table **??**). Further, many additional factors can affect phenological responses, including ontogeny (Table **??**) *(46)*, provenance latitude (Fig. **??**), and air humidity *(47)*.

Despite these limitations, a simple interpretation of our results does support the widespread hypotheses that chilling and/or photoperiod cues may underlie declining sensitivities to warming in long-term Central European data *(5; 12; 13)*. Under these hypotheses, warming increases forcing and thus advances budburst, but such advances become muted if warming also causes important declines in chilling and shorter photoperiods experienced near the timing of budburst *(39)*. This basic agreement between our results and long-term observational trends integrates across experimental conditions that encompass more extreme scenarios than may be seen in nature (Fig. **??**, **??**). A more robust comparison requires examining predictions under conditions closer to those found in nature.

Reinterpreting our model using climate and phenology data that have led to observations of declining temperature sensitivities in Central Europe suggests that chilling and photoperiod are unlikely to cause the observed declines. Our model predicts such declines only at extreme warming for most sites (see Supplemental Materials). In contrast to the common hypothesis that plants experience less chilling with global warming, we found that—for many sites—total estimated chilling increased with warming (Fig. 4A, C), though this varied with local climate prior to warming (Figs. **??** - **??**). Portions of Central Europe have experienced more dramatic warming in winter versus summer *(48; 49,* though this varies over time and space). Yet even if warming uniquely occurs in the winter, our results suggest that delays due to decreased chilling only occur at warming above at least 4°C for most sites, though responses vary by species (Fig. 3, **??**). At high warming, predicted declines in sensitivity were due to declines in chilling—photoperiod had comparatively little effect on budburst day of year, even for the photosensitive species *F. sylvatica* (Fig. **??**).

Our predictions leave open the question of what underlies declining sensitivities across Europe, but one possibility is that it may be a statistical artifact of how temperature sensitivities are calculated. Physiologically, budburst is triggered by the accumulation of forcing temperatures during the spring *(15; 50)*. However, researchers today often estimate temperature sensitivities from long-term observational data using a linear regression of annual budburst date versus mean spring temperature, or other aggregated temperature metrics *(e.g., 11)*. This approach has the benefit of yielding an easily interpretable metric—days change per °C—but

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will estimate systematically lower sensitivities given warmer daily temperatures, even with no change in the underlying cues (Fig. ??). We found the declining sensitivities observed in European data are of the same magnitude as those predicted from a statistical artifact (sensitivity declines of 0.8 ± 0.3 days/°C in European data versus 0.9 ± 0.5 days/°C in simulations), and the data also show a related decline in leafout variance that would not be immediately predicted from shifting cues (see Potential statistical artifacts in declines of temperature sensitivity in observational long-term data in the Supplemental Materials and 51, for further details). This statistical artifact is likely not confined to phenological studies; it should apply to any research using a similar days/°C metric to estimate an underlying thermal accumulation model where the thermal sum per day is non-stationary, as is the case with climate change.

Our results unify decades of experimental studies, which have shown the importance of chilling, forcing, and daylength to determining budburst timing, with long-term observational data, where forcing appears to dominate responses to recent warming. We do not find strong evidence for delaying budburst in the near future, and suggest that recent observed declines in temperature sensitivity may be related to statistical artifacts from estimating complex cues from observational data. Instead, our predictions suggest budburst will continue to advance in many well-studied European regions in the future. The most dramatic changes in future spring phenology will come from regions where winter warming causes large changes in chilling, with implications for ecosystem services related to phenology. Thus identifying processes that plants undergo when accumulating chilling versus forcing will be critical for the most accurate forecasts (15; 22).

References

[1] R. K. Campbell, A. I. Sugano, *Botanical Gazette* pp. 290–298 (1975).

[2] O. M. Heide, *Scientia Horticulturae* **115**, 309 (2008).

[3] D. F. B. Flynn, E. M. Wolkovich, *New Phytologist* **219**, 1353 (2018).

[4] O. M. Heide, *Journal of Experimental Botany* **62**, 5397 (2011).

[5] Y. S. H. Fu, *et al.*, *Nature* **526**, 104 (2015).

[6] C. M. Zohner, B. M. Benito, J. C. Svenning, S. S. Renner, *Nature Climate Change* **6**, 1120 (2016).

[7] IPCC, *Climate Change 2014: Impacts, Adaptation, and Vulnerability* (Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 2014).

[8] A. J. Miller-Rushing, R. B. Primack, *Ecology* **89**, 332 (2008).

[9] A. Menzel, *et al.*, *Global Change Biology* **12**, 1969 (2006).

[10] E. E. Cleland, I. Chuine, A. Menzel, H. A. Mooney, M. D. Schwartz, *Trends in Ecology & Evolution* **22**, 357 (2007).

[11] E. M. Wolkovich, *et al.*, *Nature* **485**, 494 (2012).

[12] T. Rutishauser, J. Luterbacher, C. Defila, D. Frank, H. Wanner, *Geophysical Research Letters* **35**, L05703 (2008). Rutishauser, This Luterbacher, Juerg Defila, Claudio Frank, David Wanner, Heinz.

[13] H. Y. Yu, E. Luedeling, J. C. Xu, *Proceedings of the National Academy of Sciences of the United States of America* **107**, 22151 (2010).

[14] X. Wang, *et al.*, *Nature communications* **10**, 2389 (2019).

[15] I. Chuine, *et al.*, *Global Change Biology* **22**, 3444 (2016).

[16] C. A. Harrington, P. J. Gould, *Frontiers in Plant Science* **6**, 120 (2015).

[17] D. Basler, C. Körner, *Tree Physiology* **34**, 377 (2014).

[18] A. Caffarra, A. Donnelly, I. Chuine, M. B. Jones, *Climate Research* **46**, 147 (2011).

[19] A. Caffarra, A. Donnelly, I. Chuine, *Climate Research* **46**, 159 (2011).

[20] H. Fraga, J. G. Pinto, J. A. Santos, *Climatic Change* pp. 1–15 (2019).

[21] O. Heide, *Physiologia Plantarum* **88**, 531 (1993).

[22] R. K. Singh, T. Svystun, B. AlDahmash, A. M. Jönsson, R. P. Bhalerao, *New Phytologist* **213**, 511 (2017).

[23] Y. Vitasse, D. Basler, *European Journal of Forest Research* **132**, 1 (2013).

[24] Y. Vitasse, D. Basler, *Tree physiology* **34**, 174 (2014).

[25] J. Laube, *et al.*, *Global Change Biology* **20**, 170 (2014).

[26] D. Basler, C. Körner, *Agricultural and Forest Meteorology* **165**, 73 (2012).

[27] A. Caffarra, A. Donnelly, *International Journal of Biometeorology* **55**, 711 (2011).

9

[28] R. Ohlemüller, E. S. Gritti, M. T. Sykes, C. D. Thomas, *Global Ecology and Biogeography* **15**, 395 (2006).

[29] J. W. Williams, S. T. Jackson, *Frontiers in Ecology and the Environment* **5**, 475 (2007).

[30] J. W. Williams, S. T. Jackson, J. E. Kutzbacht, *Proceedings of the National Academy of Sciences of the United States of America* **104**, 5738 (2007).

[31] T. Stocker, D. Qin, G. Platner, *Working Group I Contribution to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Summary for Policymakers (IPCC, 2013)* (2013).

[32] Y. Xu, V. Ramanathan, D. G. Victor, Global warming will happen faster than we think (2018).

[33] E. M. Wolkovich, *et al.*, Observed Spring Phenology Responses in Experimental Environments (OS-PREE), doi:10.5063/F1QV3JQR (2019).

[34] E. Richardson, *HortScience* **9**, 331 (1974).

[35] F. Dennis, *HortScience* **38**, 347 (2003).

[36] A. Gelman, J. Hill, *Data analysis using regression and multilevel/hierarchical models* (Cambridge University Press, 2006).

[37] Y. H. Fu, *et al.*, *Global Change Biology* **25**, 1696 (2019).

[38] N. L. Bradley, A. C. Leopold, J. Ross, W. Huffaker, *Proceedings of the National Academy of Sciences* **96**, 9701 (1999).

[39] J. Gauzere, C. Lucas, O. Ronce, H. Davi, I. Chuine, *Ecological Modelling* **441**, 108805 (2019).

[40] O. Heide, A. Prestrud, *Tree Physiology* **25**, 109 (2005).

[41] A. M. Roberts, C. Tansey, R. J. Smithers, A. B. Phillimore, *Global Change Biology* **21**, 2603 (2015).

[42] C. van der Schoot, L. K. Paul, P. L. H. Rinne, *Journal of Experimental Botany* **65**, 1699 (2014).

[43] S. Fishman, A. Erez, G. Couvillon, *Journal of Theoretical Biology* **124**, 473 (1987).

[44] J. H. Weinberger, *et al.*, *Proceedings. American Society for Horticultural Science* (1950), vol. 56, pp. 122–28.

[45] C. A. Polgar, R. B. Primack, E. H. Williams, S. Stichter, C. Hitchcock, *Biological Conservation* **160**, 25 (2013).

[46] Y. Vitasse, *New Phytologist* **198**, 149 (2013).

[47] J. Laube, T. H. Sparks, N. Estrella, A. Menzel, *New Phytologist* **202**, 350 (2014).

[48] C. Li, B. Stevens, J. Marotzke, *Geophysical Research Letters* **42**, 8131 (2015).

[49] R. C. J. Balling, P. J. Michaels, P. C. Knappenberger, *Climate Research* **9**, 175 (1998).

[50] H. Hänninen, *Canadian Journal of Botany* **73**, 183 (1995).

[51] S. Güsewell, R. Furrer, R. Gehrig, B. Pietragalla, *Global Change Biology* **23**, 5189 (2017).
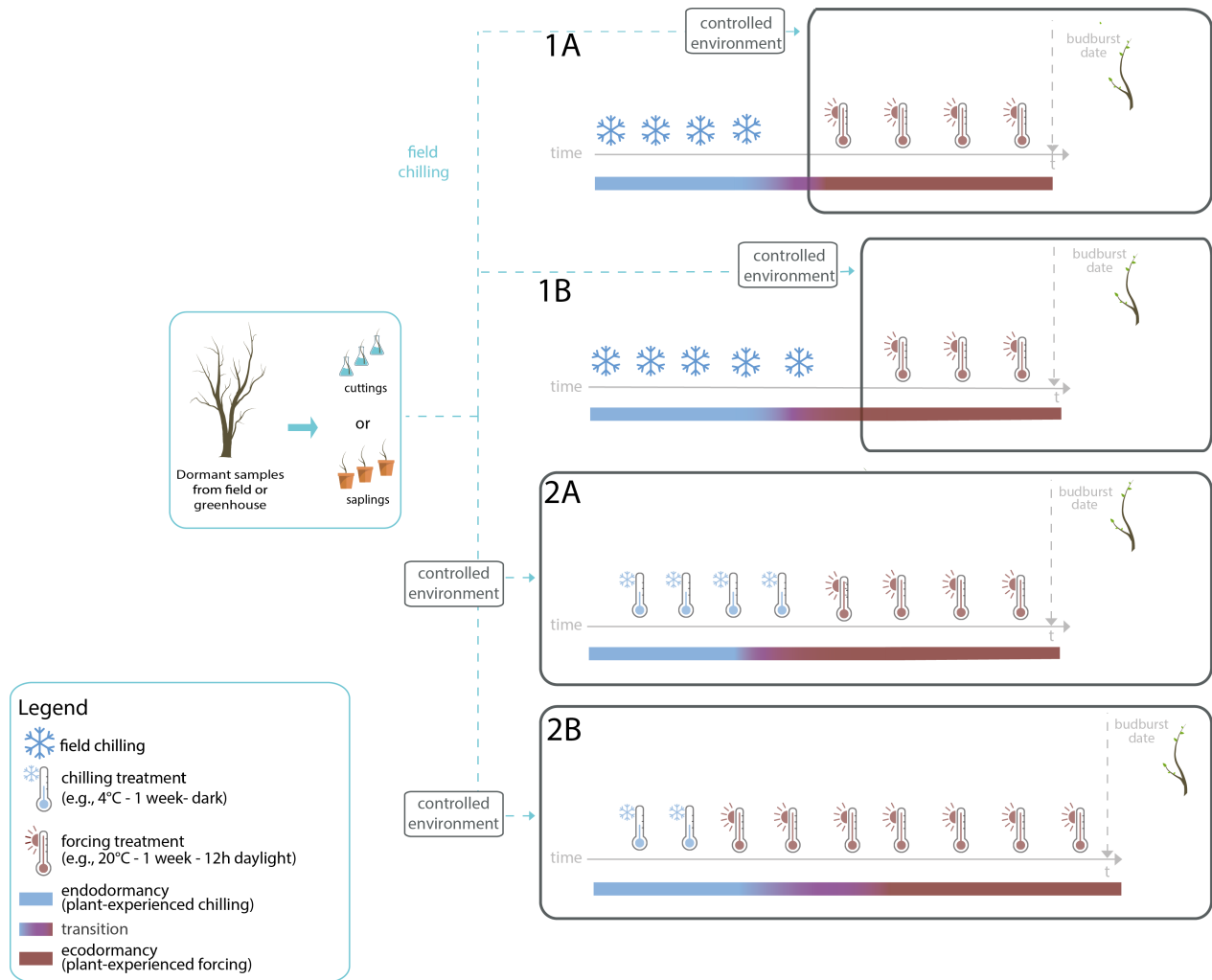
Acknowledgements

Figures

Figure 1: **Controlled environment experiments that test temperature effects on woody plant phenology,** as we synthesize here, generally manipulate photoperiod and temperature cues, which include chilling and forcing. Chilling is manipulated by either using natural chilling in the field (1A-B, in which plant material is collected after different numbers of days in the fall/winter) or experimentally (2A-B, in which plant material is placed in controlled environment chambers set to different chilling temperatures and/or durations). Chilling treatments, which are generally applied first (in field or experimentally), are designed to break plant endodormancy, after which forcing treatments are imposed by moving plant material to warmer temperatures that allow budburst to occur. Ideally, this experimental transition aligns with the physiological shift from endo- to ecodormancy (e.g., 1A, though it could also occur with experimentally applied chilling). A challenge with controlled environment studies is that species-specific chilling requirements are rarely known, so experimental treatments may not always align with what the plant experiences (i.e., physiological shifts in dormancy). Thus, in some cases, chilling treatments may bridge across what plants experience as both chilling and forcing (1B and 2A, where plants transition into ecodormancy before "forcing" treatments are applied), or chilling treatments may end before endodormancy is fully broken (2B). In the studies synthesized here, photoperiod (not shown) is most often manipulated in forcing treatments.
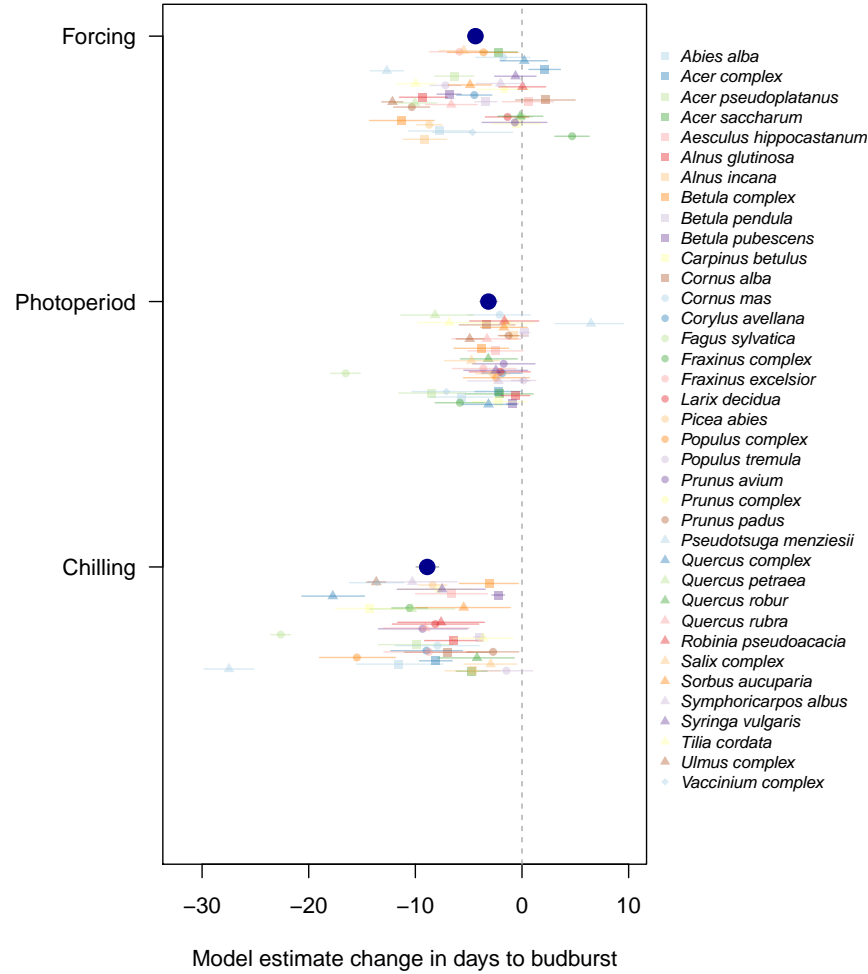
Figure 2: **Estimated effects of chilling, forcing, and photoperiod on budburst timing across 67 (modeled as 36 separate taxa, see *Models* in the Supplemental Materials) in 42 controlled environment studies.** Using standardized units, which allow comparisons across cues, we show that most species (smaller symbols) are responsive to most cues, with chilling being the strongest cue when considering overall estimates across species (larger, dark blue circles). Overall estimates shown here were generally similar to other model formulations, including using data from 203 species, and using different methods for calculating chilling (Figs. **??**, **??**; Tables **??-??**). Lines represent 50% uncertainty intervals (other intervals provided in Tables **??-??**)
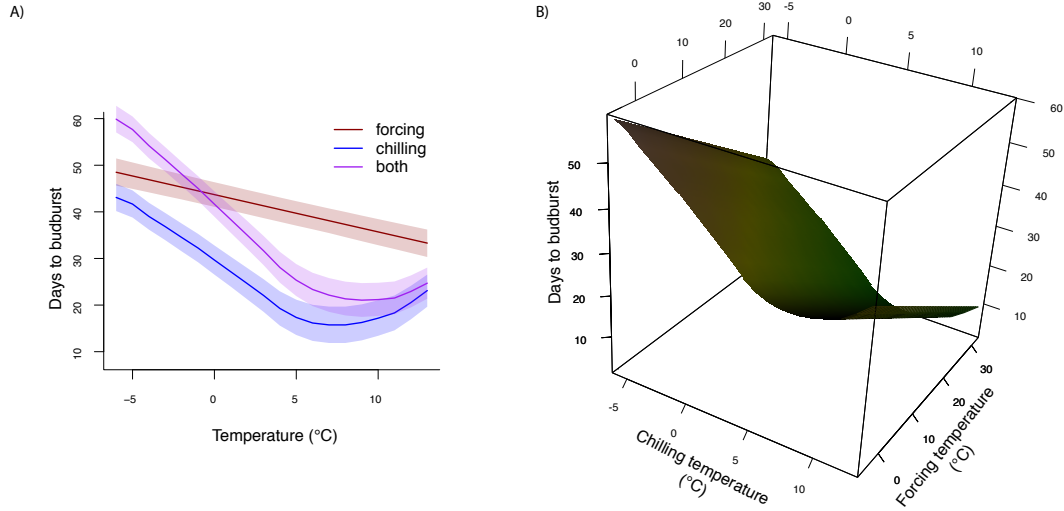
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Figure 3: **Estimates of budburst across a range of forcing temperatures and estimated chilling** (converted to a representative mean temperature, see *Estimating chilling* in Methods and Supplemental Materials) based on overall estimates of chilling and forcing effects (Fig. 2). Note that days to budburst is relative to experimental methods and thus not comparable to day of year in the field, shading (in A) represents 50% uncertainty intervals. Panel A shows the effect of chilling temperature on budburst, with forcing kept at the mean level across all experiments (16°C); the effect of forcing temperature with chilling kept at the mean level across all experiments (1324 chilling units), and the effect of varying both chilling and forcing temperatures simultaneously. Panel B shows all possible combinations of chilling and forcing across the experimental conditions. Maximum advances in budburst occur at intermediate chilling temperatures (*e.g.*, here at mean winter temperatures of 6-7°C) and the highest forcing (here at 32°C). We set photoperiod to eight hours, which is the most common photoperiod treatment in the database.
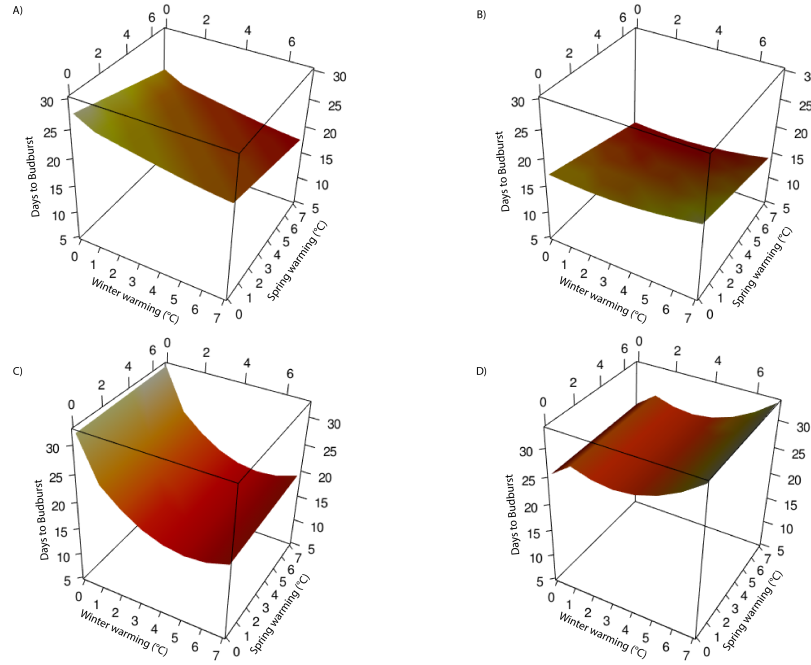
Figure 4: **Implications of warming on budburst timing varies across species and sites**, depending strongly on pre-warming climate conditions related to chilling for each site. Here we show species-level estimates from our model (Fig. 2) for the two most common species in the OSPREE database: *Betula pendula* (A, B) and *Fagus sylvatica* (C, D), for sites that highlight the diversity of possible budburst responses to warming (Fig. **??**, which shows general trends across many sites in Central Europe). In some sites, warming increases total chilling estimates (A, C) leading to greater advances in budburst (compared to forcing alone), whereas warming decreases total chilling estimates in other sites (B, D), leading to smaller advances and, eventually, delays with substantial warming. See Fig. **??** in the Supplemental Materials for a simplified two-dimensional version.