

RH: Interactive cues & phenology

## **Integrating experiments to predict interactive cue effects on spring phenology with warming**

E. M. WOLKOVICH<sup>1,2</sup>, C. J. CHAMBERLAIN<sup>2,3</sup>, D. M. BUONAIUTO<sup>2,3</sup>, A. K. ETTINGER<sup>4</sup>  
& I. MORALES-CASTILLA<sup>5</sup>

<sup>1</sup> *Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, 2424 Main Mall,  
Vancouver, BC V6T 1Z4*

<sup>2</sup> *Arnold Arboretum of Harvard University, 1300 Centre Street, Boston, Massachusetts, 02131, USA*

<sup>3</sup> *Organismic & Evolutionary Biology, Harvard University, 26 Oxford Street, Cambridge, Massachusetts,  
02138, USA*

<sup>4</sup> *The Nature Conservancy, 74 Wall Street, Seattle, Washington USA*

<sup>5</sup> *Global Change Ecology and Evolution Group, Department of Life Sciences, University of Alcalá,  
Alcalá de Henares 28805, Spain*

**Corresponding author:** Lizzie, see <sup>3</sup> above ; E-mail: e.wolkovich@ubc.ca

*Summary:* Climate change has advanced plant phenology globally 4-6 days per °C on average. Such shifts are some of the most reported and predictable biological impacts of rising temperatures. Yet as climate change has marched on, phenological shifts have appeared muted over recent decades—failing to match simple predictions of an advancing spring with continued warming. The main hypothesis for these changing trends is that interactions between spring phenological cues—long-documented in lab environments—are playing a greater role in natural environments due to climate change. Here we argue that accurately linking shifts observed in long-term data to underlying phenological cues is slowed by biases in observational studies and limited integration of lab experiments. We synthesize seven decades of lab experiments to quantify how much phenological cue-space has been studied and how treatments compare to shifts in cues caused by climate change. Most studies focus on one cue, limiting our ability to make accurate predictions, but some well-studied forest species offer opportunities to advance forecasting. We outline how greater integration of controlled environment experiments with long-term data and insights from physiology, alongside a new generation of lab experiments could transform our fundamental understanding of phenology and improve predictions.

*Keywords:* phenology, climate change, spring warming, chilling, forcing, daylength, photoperiod, non-linear responses, leafout, budburst

# 1 Main text

Shifts in spring plant phenology are one of the most reported and predictable changes with climate change. Decades of research have documented advancing budburst, leafout and flowering with climate change (Delpierre *et al.*, 2009; Yu *et al.*, 2010; Ellwood *et al.*, 2013; Jochner *et al.*, 2013; Hereford *et al.*, 2017), especially in temperate systems where long-term records highlight how humans have altered the timing of spring (Schwartz, 1997; Menzel *et al.*, 2006). Recently, however, these advances have appeared to slow (Fu *et al.*, 2015) or even reverse (Yu *et al.*, 2010)—failing to match simple predictions of an advancing spring with continued warming (Ellwood *et al.*, 2013). The main hypothesis for this failure is that spring warming—which most observational studies focus on—is no longer the only environmental cue that matters to predicting responses to warming (Chuine *et al.*, 2016; Gauzere *et al.*, 2019).

Despite the strong focus of spring phenology research on spring temperatures, increasing evidence suggests a more complicated physiology for most plant species (e.g., Zohner *et al.*, 2016; Gauzere *et al.*, 2019; Ettinger *et al.*, 2020). For many species three major cues underlie spring phenology: chilling (cool temperatures, generally occurring in the fall and winter), forcing (warm temperatures, generally occurring in the late winter and early spring), and photoperiod (daylength).

Together, chilling, forcing and photoperiod may produce non-linear responses. Predicting these non-linearities is a common goal in plant phenology research today (Güsewell *et al.*, 2017; Chen *et al.*, 2019), but has been slowed by data gaps and the complexity of spring phenological cues. Non-linear responses can generally occur through two alternative pathways. First, while each cue alone may yield linear responses in the mid-range of values, extremely high or low values can produce threshold responses (Heide, 1993a; Partanen *et al.*, 1998; Singh *et al.*, 2017; Rinne *et al.*, 2018). Such extreme values are uncommon in natural environmental regimes and thus unlikely to explain recent observations of shifts.

Most hypotheses for the observed decline instead invoke the interaction between cues to produce non-linear responses to warming. Yet estimating the effect of interactions from observational data, where cues themselves often covary, may produce spurious results (Ettinger *et al.*, 2020). In contrast, controlled environment (e.g., growth chamber) studies are designed to understand and estimate multiple cues, and can tease out their interactive effects.

Here we argue that controlled environmental studies to estimate cue interactions are critical to robust forecasts of phenological shifts with climate change. Such studies have been conducted for over 70 years, but they have rarely been reviewed and are often poorly integrated into the current phenological literature on climate change (e.g., Fu *et al.*, 2015; Richardson *et al.*, 2018). To address this gap, we synthesize these studies (Fig. S1) to quantify how much of the cue-space (i.e., the possible range of each cue and interactions across cues) has been studied and how treatments compare to shifts in cues caused by climate change. Based on this, we suggest a path forward where advances in physiology, greater integration of controlled environment studies with forecasting, and multi-species modeling can yield more robust predictions.

Given our aim to improve understanding of current trends and forecasts we focus on early vegetative phases (budburst and leafout), which are critical to plant growth and carbon storage, have shifted most with climate change (Cleland *et al.*, 2007), and have been studied the most in experiments (see Supporting Information). Our review concentrates on woody species phenology, where most research on budburst and leafout has occurred. Most of our conclusions and suggested approaches, however, could be adapted to non-woody species and other phenophases with similar underlying interactive cues.

## 1.1 Why observational studies are not enough to robustly estimate cues

The first step towards robust phenological predictions is accurate measurements of chilling, forcing and photoperiod cues. Recently, efforts have focused on estimating these cues from long-term observational data (e.g., Luedeling *et al.*, 2013). Yet observational studies may struggle to accurately estimate any of the three major cues due to two types of statistical issues. First, in most observational data these cues correlated: forcing increases alongside longer photoperiods (Elmendorf & Ettinger, 2020), and changes to chilling and photoperiod cues with warming predict the same response—later spring phenology. Approaches that attempt to disentangle some of these correlations, such as leveraging trends across elevation or latitude, may run afoul of other correlations (Tansey *et al.*, 2017). Second, most observational studies focus on linear models of each cue, often without interactions between cues (Visser & Holleman, 2001; Polgar *et al.*, 2014). Even when interactions are invoked to explain declining responses to warming, they are often not estimated (e.g., Fu *et al.*, 2015).

In contrast to observational studies, controlled environment studies can manipulate each cue independently. By experimentally decoupling cues, they can tease out interactions between them. Perhaps unsurprisingly then, most of our fundamental understanding of these cues comes from controlled environment studies.

Controlled environment studies have shown that chilling, forcing and photoperiod cues together determine the transition from dormancy to growth (usually first observed as budburst) each year for most temperate woody species (van der Schoot *et al.*, 2014; Chuine & Regniere, 2017; Singh *et al.*, 2017; Chang *et al.*, 2021). Chilling and forcing cues are generally understood to be accumulation processes, where plants integrate chilling and forcing experienced over time to meet a threshold value at which they can budburst, leafout or flower (Chuine & Regniere, 2017). In contrast, photoperiod is generally not considered an integral cue, but one evaluated daily. In practice, these two types of processes are often abstracted into the effects of average experienced values (mean temperature or daylength), generally over some temporal window in long-term data (e.g., Wolkovich *et al.*, 2012a; Fu *et al.*, 2015), or by holding temperatures and light regimes more constant in experiments (e.g., Worrall & Mergen, 1967; Heide, 1993a; Skuterud & Dietrichson, 1994).

Climate change is likely altering each of the three cues. Long term data suggests that warming has already translated into shifts in forcing and chilling (Fu *et al.*, 2015; Piao *et al.*, 2017), but such estimates are complicated by our limited understanding of chilling (see ‘The chilling enigma

of over-winter temperatures’ in Supporting Information). Additionally, the relevant photoperiod a plant experiences at critical physiological points may change dramatically with warming (Ettinger *et al.*, 2021).

Researchers hypothesize that these shifts—in forcing, chilling and photoperiod experienced near the time of an event—produce non-linearities in plant responses to warming. While a growing number of observational studies have attempted to document non-linear shifts in phenology due to interactive cues (Fu *et al.*, 2015; Gauzere *et al.*, 2019), understanding exactly when cues cause non-linearities may be more complicated than often presented (Fig. 1), and almost always relies on knowledge gained from controlled environment studies (e.g., Fu *et al.*, 2015; Richardson *et al.*, 2018; Gauzere *et al.*, 2019).

## 1.2 Interactions alone are unlikely to produce non-linearities with warming

Interactions are well-documented in controlled environment studies. Multiple studies now show that the threshold of forcing needed for budburst depends on the sum of chilling over the fall and winter and also the photoperiod experienced in the spring (e.g., Zohner & Renner, 2014; Flynn & Wolkovich, 2018). Higher forcing is generally needed given lower chilling (Fig. 2) and shorter photoperiods (Basler & Körner, 2014; Fu *et al.*, 2019). This yields a sub-additive interaction of forcing x chilling and forcing x photoperiod, where high values of both cues together produce a more muted response than would be expected from examining either cue alone.

These interactions may produce a smaller response, but they do not necessarily produce non-linearities (Fig. 1). Instead most hypotheses for non-linear responses tacitly require both the interaction of cues and covarying shifts in cues with warming to produce non-linearities. For example, if some species have a critical photoperiod for budburst and warming leads to forcing cues being met before the critical threshold—effectively shifting both the forcing and photoperiod cues—then we would expect incomplete or highly delayed budburst (Singh *et al.*, 2017; Rinne *et al.*, 2018). Alternatively, the threshold could be crossed in the opposite direction: in this case, pre-climate change conditions would cause budburst to occur at the extreme values of some cues, but warming could push budburst into values where responses are more linear. This is the mechanism often suggested for declining responses to warming in some temperate trees (Fu *et al.*, 2015; Piao *et al.*, 2017; Gauzere *et al.*, 2019), specifically that plants previously accumulated sufficient chilling such that forcing was the dominant cue—whereas warming has now reduced chilling such that more forcing is needed for budburst (producing an overall muted effect when estimated as change in days per °C, see Fu *et al.*, 2015, for one example). As this example highlights, however, changes in a single cue are unlikely to occur without additional effects on other cues—complicating how well we can understand them in long-term data without robust understanding of the exact cue requirements from other methods.

## 1.3 Forecasting non-linear responses: Do we have the necessary data?

Controlled environment studies can help predict non-linear responses by allowing researchers to examine the effects of one cue with the others held constant, and examine interactive effects—

given the appropriate study design. Such experiments may be especially useful for forecasting if they contain enough variation in treatments to capture precisely where non-linearities occur, and are designed across a range of levels relevant to current versus future conditions (Shen *et al.*, 2015). Indeed, one of the major advantages of experiments is that they allow treatments outside of the historical range of a species' (or region's) climate—an option observational data cannot provide.

To understand how valuable currently available data may be to forecasting, we reviewed controlled environment studies over the last seven decades, quantifying the range of treatments and how they compare to current and future conditions. These studies were rarely conducted for climate change research, and most often done for reasons of fundamental or applied science (e.g., horticulture or forestry). Yet they are some of the best available data for how plants respond to the environment and thus a critical resource for climate change research today.

#### *How studies and their experimental treatments vary globally*

Controlled environment studies have been conducted with 226 woody species across the globe, with the majority of papers reporting research occurring in Europe (54 of 84 papers; and 93 of 136 studies across papers; a study is a unique experiment within a paper), followed by North America (22 papers and 32 studies, Fig. S1, Table S1). Most studies manipulate one cue though studies of two or three cues have occurred in almost every decade (Fig. S3). Across study designs, chilling was the most commonly studied cue (69% of 117 studies that manipulated at least one cue; with roughly half of these studies—47%—using outdoor chilling through multiple field sampling dates), followed by forcing and photoperiod (43% and 40%, respectively).

Studies have covered only a small portion of cue-space, especially for experimentally applied chilling (Fig. 3), with certain levels of cues much more common than others. This suggests we have greater inference at certain cue levels (e.g., 12 hour photoperiods and 20°C forcing), but also more limited understanding beyond them, which could limit forecasting. The levels of cues also varied across latitude with a general trend toward examining more extreme values at higher latitudes (Fig. S4). This trends appears related to latitudinal clines in the environment—higher latitudes experience colder temperatures and longer photoperiods, and see similar shifts in their controlled environment study designs. But this trend also introduces a bias in results, as any comparisons of studies from lower and higher latitudes are also comparing a different range of cues (see 'Trends in experimental treatments over cue levels and space' in Supporting Information).

#### *How studies manipulate cues*

To robustly test for interactions studies must use an experimental design that includes most or all combinations of all levels of cues studied (factorial design). Of the studies manipulating at least one cue, 37% (43 studies) additionally manipulated another cue in a factorial design, with the studies almost evenly split across all possible two-way interactions. Studies examining three cues directly were rare: we identified only three studies examining all three cues at once. Two of these were on *Picea abies* (Worrall & Mergen, 1967; Sogaard *et al.*, 2008), and the other on

*Betula pendula* (Skuterud & Dietrichson, 1994). A slightly larger set of studies (5 studies from 4 papers) examined three cues indirectly—manipulating photoperiod and forcing in controlled environments but equating chilling with sequential removal of tissue from the field—for 11 species (Schnabel & Wample, 1987; Heide, 1993a; Partanen *et al.*, 1998; Basler & Körner, 2014).

*Do we have enough data to estimate interactions?*

The small number of studies examining multiple cues hinders forecasting by limiting our understanding of how—when combined—cues will determine future budburst with continued warming. Because the cues are all known to be interactive, estimates of any one cue are influenced by the level of each other cue. But, unfortunately, current studies often do not report the levels of each cue (see ‘NA’ in Fig. 3), and many studies use field conditions to apply treatments (Fig. 3, and ‘Paths forward’ below).

Of the 136 studies we reviewed, only 72 reported enough information to estimate forcing, chilling and photoperiod treatments (Ettinger *et al.*, 2020). This combined with the uneven sampling of cue-space (Fig. 3) makes it impossible to estimate the effect of interactions between cues with current data (Ettinger *et al.*, 2020). While a few studies estimated interaction terms (e.g., Zohner & Renner, 2014), most studies did not, likely because of the increased effort in study design. In addition to needing enough controlled environments to apply a full-factorial design, the statistical power needed to robustly estimate an interaction is much greater than a simple main effect (such as the effect of forcing or photoperiod alone)—requiring a 16X greater sample size (Gelman *et al.*, 2020).

*How relevant are treatments to current and future conditions?*

The utility of controlled environment studies to forecasting also depends on how relevant treatments are to current and future conditions. Estimating such relevance is difficult as it depends on a species’ geographical range and projections considered. However, a simple analysis of two well studied species, *Fagus sylvatica* and *Betula pendula*, suggests experiments have generally bracketed the range of projected temperatures (Fig. 4), though there is a limited number of chilling studies that directly manipulate chilling temperature (Fig. 3). Indeed, we found no studies with multiple chill temperatures for *Fagus sylvatica*, even though it is one of the most well-studied species (Fig. 4C).

Experimental treatments were generally larger than expected shifts due to climate change. This is not surprising given that certain forcing and photoperiod treatments were extremely common across studies (see above). It also makes sense from an experimental-statistical perspective: if the goal of an experiment is to identify if a cue is present then larger treatment differences should yield larger effect sizes and higher statistical power. Such large shifts, however, may be risky to extrapolate from when predicting effects of warming.

## 1.4 Paths forward

We argue that controlled environment experiments will be critical for accurate predictions of phenology given future warming. How accurate such predictions are will depend on the design

of future experiments, breakthroughs in our physiological understanding of the major cues, and how well these two areas can be integrated with long-term data to improve models.

#### *Improving controlled environment studies*

We expect the most useful future experiments for forecasting will be designed to identify threshold effects, optimal temperatures/photoperiods, and non-linearities from interactive cues (Caffarra *et al.*, 2011). Identifying threshold effects and optimal temperatures or photoperiods generally requires many different levels of a single cue, which can make such experiments difficult to cross with other cues. Yet, where a threshold or optimum occurs is likely dependent on the level of other cues, making studies of interactive cues critical for robust forecasting. To test for non-linearities due to interactive cues, studies must be fully-crossed (i.e., every combination of levels is present in treatments), which can quickly require a large number of controlled environments. However, as experimental results support that all cues are dependent on the level of other cues (Stearns & Olson, 1958; Flynn & Wolkovich, 2018) and long-term data hint at multiple cues, we believe this should be a major research aim.

Controlled environment studies may also be more readily applied to forecasting by exploring more realistic conditions. While identifying thresholds, optima and non-linearities may involve considering informative extremes in levels of cues, most changes in cues due to climate change are and will be on a (relatively) smaller scale than experiments currently study (Fig. 4). While species distribution models generally assume species will remain in the same climatic conditions (Elith & Leathwick, 2009), most evidence suggests species will lag in their spatial responses, meaning shifts in cues in the current range may be important to the fate of trailing edge populations (Bertrand *et al.*, 2011; Lenoir & Svenning, 2015). Thus, researchers should design experiments that include values of cues within the current and projected future species' range, rather than focusing on extreme comparisons only (e.g., 12 versus 24 hour photoperiods). In particular, our findings suggest forcing treatments (e.g., 20 and 25°C) are often much higher than the average temperatures that study species experience near budburst (Fu *et al.*, 2015; Gusewell *et al.*, 2017), and studies often do not consider chilling temperatures.

#### *Incorporating our understanding of physiology into forecasts of phenology*

Even with all the suggested above improvements, controlled environment studies will still be fundamentally limited in their utility for prediction without a deeper understanding of how major phenological cues act physiologically (Bahuguna & Jagadish, 2015). This problem is most acute in our understanding of dormancy, which divides chilling and forcing (Chang *et al.*, 2021). Researchers may use the terms 'chilling' and 'forcing' for their treatments, but they rarely have physiological evidence that these are the actual conditions plants experience (Chuine *et al.*, 2016).

Chilling is generally defined as accumulated cool temperatures in the first phase of dormancy (endodormancy), after which plants enter ecodormancy, when accumulated forcing leads to budburst (Chuine *et al.*, 2016). Measuring endodormancy and its transition into ecodormancy is notoriously difficult (e.g., Junttila & Hänninen, 2012). In practice, most phenology studies use the terms 'chilling' and 'forcing' to mean 'cool temperatures' (either in the fall and winter or ap-



plied in experimental conditions) and ‘warm temperatures’ (either in the spring or applied after sufficient chilling) and generally hope they correspond to endo- and eco-dormancy—without any evidence of this hoped-for correspondence. Some studies use the sequential transfer of cuttings to warm conditions to estimate the transition from endo- to eco-dormancy, with rapid and full budburst (e.g., >90% of buds on a cutting) generally meaning a plant is ecodormant (e.g., Junttila & Hänninen, 2012), but, given that this is labor- and space-intensive, few studies include this method.

Physiologists have long recognized this issue and recent breakthroughs provide new insights into what causes dormancy at the cellular level (van der Schoot *et al.*, 2014). Thus far, research has been on a very limited number of species (Singh *et al.*, 2017; Rinne *et al.*, 2018), making extrapolation to other species difficult. Such results, however, hold promise for a much improved physiological understanding in the near future.

An improved understanding of endodormancy release could revolutionize models of chilling, and in turn, estimates of forcing. Forcing in controlled environment experiments is often defined simply as warm temperatures (or warm temperatures after cool temperatures). Future estimates could more accurately define forcing using only temperatures during ecodormancy, assuming tractable tests of endo- and eco-dormancy and the uptake of such tests in controlled environment studies. With these experiments in hand, researchers could quickly build improved models of chilling, and forcing and—for the first time—provide accurate estimates of how chilling has and will shift with climate change.

*Improving integration of controlled environment and physiological studies with long-term data*  
Integrating long-term observational, physiological and controlled environment studies would help advance our understanding and climate change forecasting. With important exceptions (e.g., Gauzere *et al.*, 2017), studies of long-term observational phenology data have moved forward independently from advances in our physiological understanding and from controlled environment studies. Similarly, controlled environment studies generally do not use long-term data to help interpret results or define treatments.

Studies that have integrated results from both controlled environment experiments and with long-term observations provide a path forward (Caffarra *et al.*, 2011; Satake *et al.*, 2013; Chuine & Regniere, 2017). Such research underlies progress towards model development that relies continuously on a back-and-forth process between developing models based on both long-term data and experiments, then testing predictions with new experiments and newly-available observational data (Satake *et al.*, 2013). This research has been done only for a handful of crop and model organism species, though our synthesis suggests that *Betula pendula* and *Fagus sylvatica* are good candidates for expanding to wild forest species across larger landscapes.

#### *Building species-rich predictions*

Given the efforts and data involved in models for a single species, building up to multi-species predictions may appear daunting, but multi-species models are crucial for accurate forecasts that

can apply to diverse regions and large-scale vegetation models. Long-term data is generally more species-rich than controlled environment studies. For example long-term observational data in the PEP725 and NECTAR databases together have multi-site data on more than 2500 species (Wolkovich *et al.*, 2012b; Templ *et al.*, 2018), while our review of controlled environment studies found most studies focused on only one species, with data on a total of 226 species. Thus, more diverse controlled environment studies may be the current major data limitation. Beyond data, however, new modeling approaches can help integrate current and future data more powerfully.

Bayesian hierarchical models are specifically designed to synthesize across multiple sources of data. With the right information, they can attribute variation across studies to the species studied, the cues and remaining unmeasured variation in studies (i.e., unreported lab and chamber differences may be captured by including a parameter to estimate a ‘study’ effect). Such models are extremely powerful for building species-rich predictions, but—like all models—are more robust with more data. With greater efforts to publish data and improved treatment reporting (i.e., all cue conditions defined, even when not manipulated, Fig. 3) all studies—whether designed to improve models or forecasting, or not—can be included in such models.

## 1.5 Right now: It’s your tomorrow

Research on phenology had been conducted for centuries before anthropogenic climate change caused earlier budburst and leafout across much of the globe (Lamb, 1948; Sparks & Carey, 1995). Decades of controlled environment studies contributed to our fundamental understanding of the drivers of spring plant phenology. Today, climate change requires leveraging these decades and centuries of research for more accurate predictions that can help humans adapt to warming.

We have outlined how researchers could better harness the power of controlled environment experiments to transform our fundamental understanding of phenology and advance forecasting. Controlled environment studies can critically rule out, or support, hypotheses to explain observed discrepancies in long-term data and open up new pathways to use long-term data to understand current trends, helping the field move beyond trying to tease out cues using only long-term data where cues are inherently correlated. While understanding, modeling and predicting interactions among cues and their effects on phenology is challenging, it will yield more accurate predictions—with valuable implications to more realistically assess the effects of climate change on plant biodiversity.

## 2 References

- Bahuguna, R.N. & Jagadish, K.S. (2015) Temperature regulation of plant phenological development. *Environmental and Experimental Botany* **111**, 83–90.
- Basler, D. & Körner, C. (2014) Photoperiod and temperature responses of bud swelling and bud burst in four temperate forest tree species. *Tree Physiology* **34**, 377–388.
- Bertrand, R., Lenoir, J., Piedallu, C., Riofrío-Dillon, G., de Ruffray, P., Vidal, C., Pierrat, J.C. & Gégout, J.C. (2011) Changes in plant community composition lag behind climate warming in lowland forests. *Nature* **479**, 517.
- Caffarra, A., Donnelly, A. & Chuine, I. (2011) Modelling the timing of *Betula pubescens* bud-burst. II. Integrating complex effects of photoperiod into process-based models. *Climate Research* **46**, 159–170.
- Chang, C.Y.Y., Brautigam, K., Huner, N.P.A. & Ensminger, I. (2021) Champions of winter survival: cold acclimation and molecular regulation of cold hardiness in evergreen conifers. *New Phytologist* **229**, 675–691.
- Chen, L., Huang, J.G., Ma, Q.Q., Hanninen, H., Tremblay, F. & Bergeron, Y. (2019) Long-term changes in the impacts of global warming on leaf phenology of four temperate tree species. *Global Change Biology* **25**, 997–1004.
- Chuine, I., Bonhomme, M., Legave, J.M., García de Cortázar-Atauri, I., Charrier, G., Lacointe, A. & Améglio, T. (2016) Can phenological models predict tree phenology accurately in the future? The unrevealed hurdle of endodormancy break. *Global Change Biology* **22**, 3444–3460.
- Chuine, I. & Regniere, J. (2017) Process-based models of phenology for plants and animals. *Annual Review of Ecology, Evolution, and Systematics* **48**, 159–182.
- Cleland, E.E., Chuine, I., Menzel, A., Mooney, H.A. & Schwartz, M.D. (2007) Shifting plant phenology in response to global change. *Trends in Ecology & Evolution* **22**, 357–365.
- Delpierre, N., Dufrene, E., Soudani, K., Ulrich, E., Cecchini, S., Boe, J. & Francois, C. (2009) Modelling interannual and spatial variability of leaf senescence for three deciduous tree species in France. *Agricultural and Forest Meteorology* **149**, 938–948.
- Elith, J. & Leathwick, J.R. (2009) Species distribution models: ecological explanation and prediction across space and time. *Annual review of ecology, evolution, and systematics* **40**, 677–697.
- Ellwood, E.R., Temple, S.A., Primack, R.B., Bradley, N.L. & Davis, C.C. (2013) Record-breaking early flowering in the eastern united states. *Plos One* **8**, ellwood, Elizabeth R. Temple, Stanley A. Primack, Richard B. Bradley, Nina L. Davis, Charles C.

364 Elmendorf, S.C. & Ettinger, A.K. (2020) Is photoperiod a dominant driver of secondary growth  
365 resumption? *Proceedings of the National Academy of Sciences of the United States of America*  
366 **117**, 32861–32864.

367 Ettinger, A., Chamberlain, C., Morales-Castilla, I., Buonaiuto, D., Flynn, D., Savas, T., Samaha,  
368 J. & Wolkovich, E. (2020) Winter temperatures predominate in spring phenological responses  
369 to warming. *Nature Climate Change* pp. 1–6.

370 Ettinger, A.K., Buonaiuto, D.M., Chamberlain, C.J., Morales-Castilla, I. & Wolkovich, E.M.  
371 (2021) Spatial and temporal shifts in photoperiod with climate change. *New Phytologist* **230**,  
372 462–474.

373 Flynn, D.F.B. & Wolkovich, E.M. (2018) Temperature and photoperiod drive spring phenology  
374 across all species in a temperate forest community. *New Phytologist* **219**, 1353–1362.

375 Fu, Y.H., Piao, S., Zhou, X., Geng, X., Hao, F., Vitasse, Y. & Janssens, I.A. (2019) Short  
376 photoperiod reduces the temperature sensitivity of leaf-out in saplings of *Fagus sylvatica* but  
377 not in horse chestnut. *Global Change Biology* **25**, 1696–1703.

378 Fu, Y.S.H., Zhao, H.F., Piao, S.L., Peaucelle, M., Peng, S.S., Zhou, G.Y., Ciais, P., Huang, M.T.,  
379 Menzel, A., Uelas, J.P., Song, Y., Vitasse, Y., Zeng, Z.Z. & Janssens, I.A. (2015) Declining  
380 global warming effects on the phenology of spring leaf unfolding. *Nature* **526**, 104–107.

381 Gauzere, J., Delzon, S., Davi, H., Bonhomme, M., de Cortazar-Atauri, I.G. & Chuine, I. (2017)  
382 Integrating interactive effects of chilling and photoperiod in phenological process-based mod-  
383 els. A case study with two European tree species: *Fagus sylvatica* and *Quercus petraea*. *Agricu-*  
384 *cultural and Forest Meteorology* **244**, 9–20.

385 Gauzere, J., Lucas, C., Ronce, O., Davi, H. & Chuine, I. (2019) Sensitivity analysis of tree phe-  
386 nology models reveals increasing sensitivity of their predictions to winter chilling temperature  
387 and photoperiod with warming climate. *Ecological Modelling* **441**, 108805.

388 Gelman, A., Hill, J. & Vehtari, A. (2020) *Regression and Other Stories*. Cambridge University  
389 Press.

390 Güsewell, S., Furrer, R., Gehrig, R. & Pietragalla, B. (2017) Changes in temperature sensitivity  
391 of spring phenology with recent climate warming in Switzerland are related to shifts of the  
392 pre-season. *Global Change Biology* **23**, 5189–5202.

393 Heide, O. (1993a) Daylength and thermal time responses of budburst during dormancy release  
394 in some northern deciduous trees. *Physiologia Plantarum* **88**, 531–540.

395 Heide, O. (1993b) Dormancy release in beech buds (*Fagus sylvatica*) requires both chilling and  
396 long days. *Physiologia Plantarum* **89**, 187–191.

397 Hereford, J., Schmitt, J. & Ackerly, D.D. (2017) The seasonal climate niche predicts phenology  
398 and distribution of an ephemeral annual plant, *mollugo verticillata*. *Journal of Ecology* **105**,  
399 1323–1334.

- Jochner, S., Caffarra, A. & Menzel, A. (2013) Can spatial data substitute temporal data in phenological modelling? a survey using birch flowering. *Tree Physiology* **33**, 1256–1268.
- Junttila, O. & Hänninen, H. (2012) The minimum temperature for budburst in *Betula* depends on the state of dormancy. *Tree physiology* **32**, 337–345.
- Lamb, R.C. (1948) Effect of temperatures above and below freezing on the breaking of rest in the Latham raspberry. *Proceedings of the American Society for Horticultural Science*, vol. 51, pp. 313–315.
- Lenoir, J. & Svenning, J.C. (2015) Climate-related range shifts—a global multidimensional synthesis and new research directions. *Ecography* **38**, 15–28.
- Luedeling, E., Guo, L., Dai, J., Leslie, C. & Blanke, M.M. (2013) Differential responses of trees to temperature variation during the chilling and forcing phases. *Agricultural and Forest Meteorology* **181**, 33–42.
- Menzel, A., Jakobi, G., Ahas, R., Scheifinger, H. & Estrella, N. (2003) Variations of the climatological growing season (1951–2000) in Germany compared with other countries. *International Journal of Climatology* **23**, 793–812.
- Menzel, A., Sparks, T.H., Estrella, N., Koch, E., Aasa, A., Ahas, R., Alm-Kuebler, K. *et al.* (2006) European phenological response to climate change matches the warming pattern. *Global Change Biology* **12**, 1969–1976.
- Partanen, J., Koski, V. & Hänninen, H. (1998) Effects of photoperiod and temperature on the timing of bud burst in Norway spruce (*Picea abies*). *Tree Physiology* **18**, 811–816.
- Piao, S., Liu, Z., Wang, T., Peng, S., Ciais, P., Huang, M., Ahlstrom, A., Burkhardt, J.F., Chevallier, F., Janssens, I.A. *et al.* (2017) Weakening temperature control on the interannual variations of spring carbon uptake across northern lands. *Nature climate change* **7**, 359.
- Polgar, C.A., Primack, R.B., Dukes, J.S., Schaaf, C., Wang, Z. & Hoeppepner, S.S. (2014) Tree leaf out response to temperature: comparing field observations, remote sensing, and a warming experiment. *International journal of biometeorology* **58**, 1251–1257.
- Richardson, A.D., Hufkens, K., Milliman, T., Aubrecht, D.M., Furze, M.E., Seyednasrollah, B., Krassovski, M.B., Latimer, J.M., Nettles, W.R., Heiderman, R.R., Warren, J.M. & Hanson, P.J. (2018) Ecosystem warming extends vegetation activity but heightens vulnerability to cold temperatures. *Nature* **560**, 368–+.
- Richardson, E. (1974) A model for estimating the completion of rest for ‘Redhaven’ and ‘Elberta’ peach trees. *HortScience* **9**, 331–332.
- Rinne, P.L.H., Paul, L.K. & van der Schoot, C. (2018) Decoupling photo- and thermoperiod by projected climate change perturbs bud development, dormancy establishment and vernalization in the model tree populus. *Bmc Plant Biology* **18**.

- 435 Satake, A., Kawagoe, T., Saburi, Y., Chiba, Y., Sakurai, G. & Kudoh, H. (2013) Forecasting  
436 flowering phenology under climate warming by modelling the regulatory dynamics of flowering-  
437 time genes. *Nature Communications* **4**.
- 438 Savage, J. & Vellend, M. (2015) Elevational shifts, biotic homogenization and time lags in  
439 vegetation change during 40 years of climate warming. *Ecography* **38**, 546–555.
- 440 Schnabel, B.J. & Wample, R.L. (1987) Dormancy and cold hardiness in *Vitis vinifera* L. cv.  
441 White Riesling as influenced by photoperiod and temperature. *American Journal of Enology*  
442 *and Viticulture* **38**, 265–272.
- 443 Schwartz, M.D. (1997) Spring index models: An approach to connecting satellite and surface  
444 phenology. *Phenology in Seasonal Climates I* (eds. H. Lieth & M.D. Schwartz), pp. 22–38,  
445 Backhuys Publisher, Leiden, The Netherlands.
- 446 Shen, M., Piao, S., Dorji, T., Liu, Q., Cong, N., Chen, X., An, S., Wang, S., Wang, T. & Zhang,  
447 G. (2015) Plant phenological responses to climate change on the Tibetan Plateau: research  
448 status and challenges. *National Science Review* **2**, 454–467.
- 449 Singh, R.K., Svystun, T., AlDahmash, B., Jönsson, A.M. & Bhalerao, R.P. (2017) Photoperiod-  
450 and temperature-mediated control of phenology in trees—a molecular perspective. *New Phy-*  
451 *tologist* **213**, 511–524.
- 452 Skuterud, R. & Dietrichson, J. (1994) Budburst in detached birch shoots (*Betula pendula*) of  
453 different varieties winter-stored in darkness at three different temperatures. *Silva Fennica* **28**,  
454 223–224.
- 455 Sjøgaard, G., Johnsen, Ø., Nilsen, J. & Junttila, O. (2008) Climatic control of bud burst in young  
456 seedlings of nine provenances of norway spruce. *Tree Physiology* **28**, 311–320.
- 457 Sønsteby, A. & Heide, O.M. (2014) Chilling requirements of contrasting black currant (*Ribes*  
458 *nigrum* L.) cultivars and the induction of secondary bud dormancy. *Scientia Horticulturae*  
459 **179**, 256–265.
- 460 Sparks, T.H. & Carey, P.D. (1995) The responses of species to climate over two centuries - an  
461 analysis of the Marsham phenological record, 1736–1947. *Journal of Ecology* **83**, 321–329.
- 462 Stearns, F. & Olson, J. (1958) Interactions of photoperiod and temperature affecting seed ger-  
463 mination in *Tsuga canadensis*. *American Journal of Botany* **45**, 53–58.
- 464 Tansey, C.J., Hadfield, J.D. & Phillimore, A.B. (2017) Estimating the ability of plants to plas-  
465 tically track temperature-mediated shifts in the spring phenological optimum. *Global Change*  
466 *Biology* **23**, 3321–3334.
- 467 Templ, B., Koch, E., Bolmgren, K., Ungersböck, M., Paul, A., Scheifinger, H., Rutishauser, T.,  
468 Busto, M., Chmielewski, F.M., Hájková, L., Hodzić, S., Kaspar, F., Pietragalla, B., Romero-  
469 Fresneda, R., Tolvanen, A., Vučetić, V., Zimmermann, K. & Züst, A. (2018) Pan European  
470 Phenological database (PEP725): a single point of access for European data. *International*  
471 *Journal of Biometeorology* **62**, 1109–1113.

- Thielges, B. & Beck, R. (1976) Control of bud break and its inheritance in *Populus deltoides*.  
*Tree Physiology and Yield Improvement* **14**, 253–259.
- van der Schoot, C., Paul, L.K. & Rinne, P.L.H. (2014) The embryonic shoot: a lifeline through  
 winter. *Journal of Experimental Botany* **65**, 1699–1712.
- Visser, M.E. & Holleman, L.J. (2001) Warmer springs disrupt the synchrony of oak and winter  
 moth phenology. *Proceedings of the Royal Society of London. Series B: Biological Sciences*  
**268**, 289–294.
- Wolkovich, E.M., Cook, B.I., Allen, J.M., Crimmins, T.M., Betancourt, J.L., Travers, S.E., Pau,  
 S., Regetz, J., Davies, T.J., Kraft, N.J.B., Ault, T.R., Bolmgren, K., Mazer, S.J., McCabe,  
 G.J., McGill, B.J., Parmesan, C., Salamin, N., Schwartz, M.D. & Cleland, E.E. (2012a)  
 Warming experiments underpredict plant phenological responses to climate change. *Nature*  
**485**, 494–497.
- Wolkovich, E.M., Cook, B.I. & Regetz, J. (2012b) NECTAR: Network of Ecological and Climato-  
 logical Timings Across Regions, <http://knb.ecoinformatics.org/knb/metacat/nceas.988/knb>.
- Worrall, J. & Mergen, F. (1967) Environmental and genetic control of dormancy in *Picea abies*.  
*Physiologia Plantarum* **20**, 733–745.
- Yu, H.Y., Luedeling, E. & Xu, J.C. (2010) Winter and spring warming result in delayed spring  
 phenology on the Tibetan Plateau. *Proceedings of the National Academy of Sciences of the*  
*United States of America* **107**, 22151–22156.
- Zohner, C.M., Benito, B.M., Svenning, J.C. & Renner, S.S. (2016) Day length unlikely to con-  
 strain climate-driven shifts in leaf-out times of northern woody plants. *Nature Climate Change*  
**6**, 1120–1123.
- Zohner, C.M. & Renner, S.S. (2014) Common garden comparison of the leaf-out phenology  
 of woody species from different native climates, combined with herbarium records, forecasts  
 long-term change. *Ecology Letters* **17**, 1016–1025.

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499

500 *Data Availability:* Upon acceptance for publication, data from a systematic literature review  
501 included in the paper will be freely available at KNB ([knb.ecoinformatics.org](http://knb.ecoinformatics.org)).

502

### 503 **3 Figures**



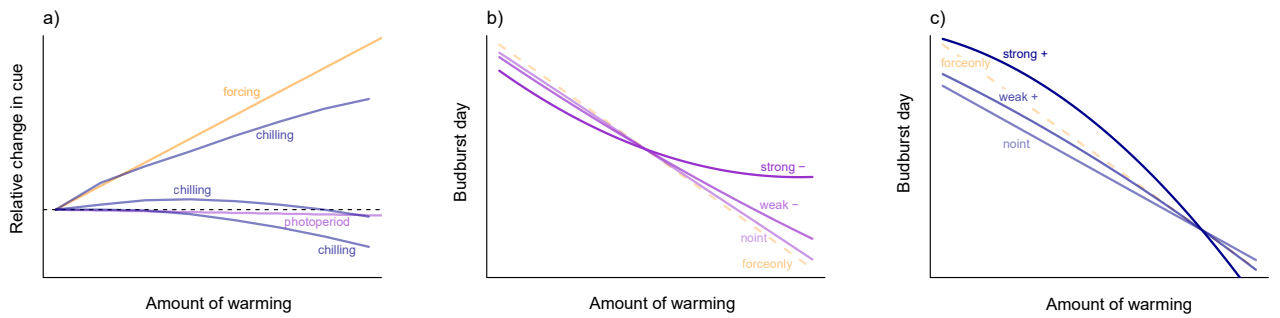


Figure 1: **Nonlinear phenological responses to warming can occur when multiple cues shift in concert with climate change.** Much research focuses on how warming increases forcing, but warming may also alter other cues, including photoperiod experienced near the time of the event, which is expected to shorten, and chilling, which may either increase or decrease (a). Shifts in forcing alongside shifts in a second cue may produce non-linearities due to the interaction between cues (b-c) showing the effect of: forcing-only in yellow, both cues without an interaction in lighter purple/blue, and both cues with an interaction in darker purple/blue). The overall change in budburst day predicted with warming depends on the sign of the second cue (negative in b, positive in c), as well as its strength (weak/strong).

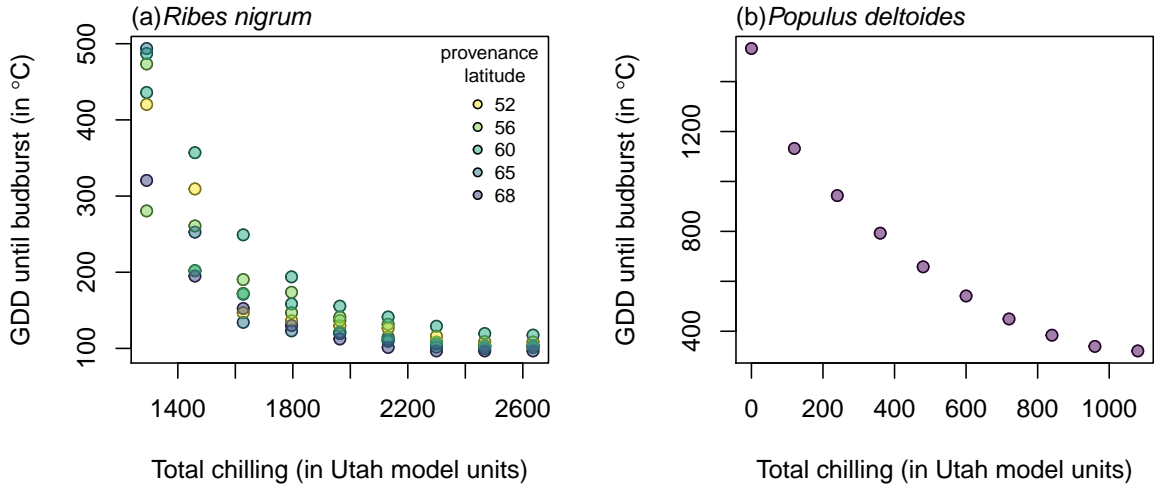


Figure 2: **A common example of how the level of one cue can modify the required level of another cue** comes from experiments finding that the amount of chilling affects the amount of forcing needed for budburst. Here, we show this from experiments in black currant (*Ribes nigrum*) from Sønsteby & Heide (2014) and eastern cottonwood (*Populus deltoides*) from Thielges & Beck (1976). Forcing is estimated as growing degree days (GDD, sum of temperatures  $> 0^{\circ}\text{C}$ ) while chilling is estimated using the Utah Model (see Richardson, 1974).

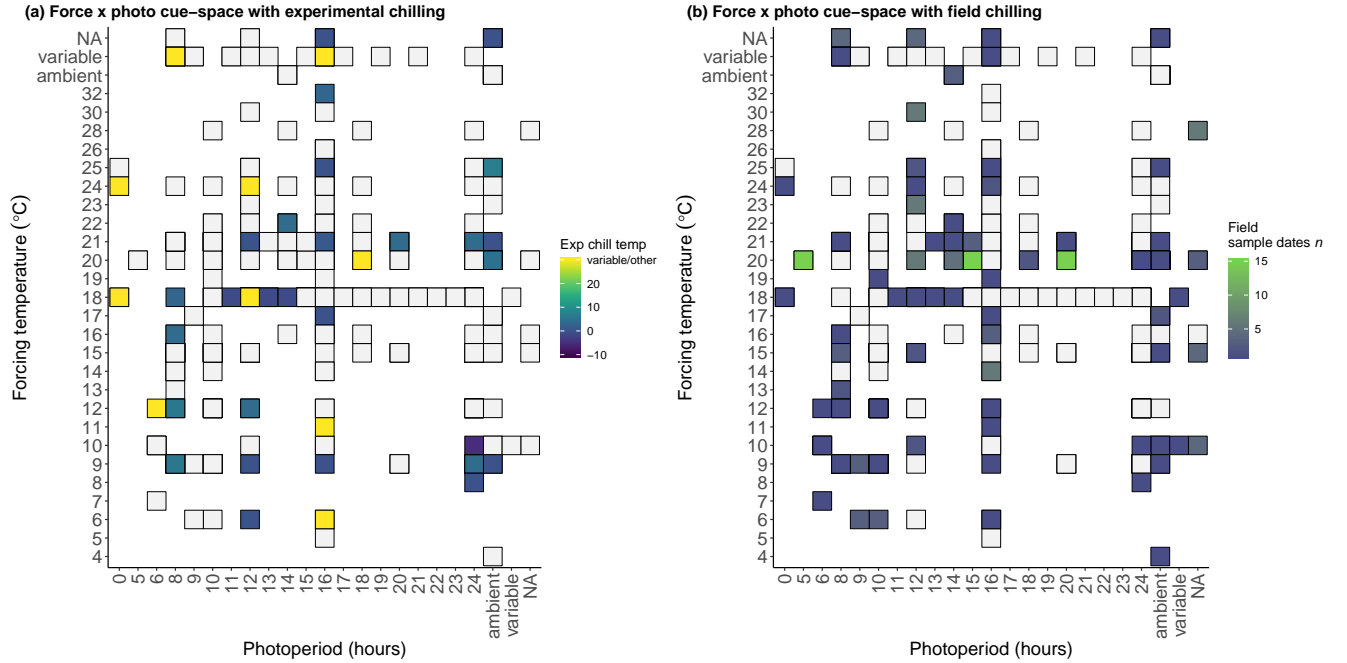


Figure 3: **Studies have examined a range of cue-space** (forcing temperature x photoperiod across two methods to manipulate chilling: a) experimentally or b) using multiple field sampling dates), but a focus on certain levels of cues and limited reporting of all cues makes using these data to estimate non-linearities challenging (see ‘Forecasting non-linear responses: Do we have the necessary data?’). Gray squares indicate a combination of forcing x photoperiod not present for that method of chilling design (includes studies that did not clearly report chilling treatments), while a value of ‘NA’ indicates that we could not estimate a level of a particular cue because it was not clearly reported. Photoperiod treatments were generally only applied (or reported) during forcing conditions.

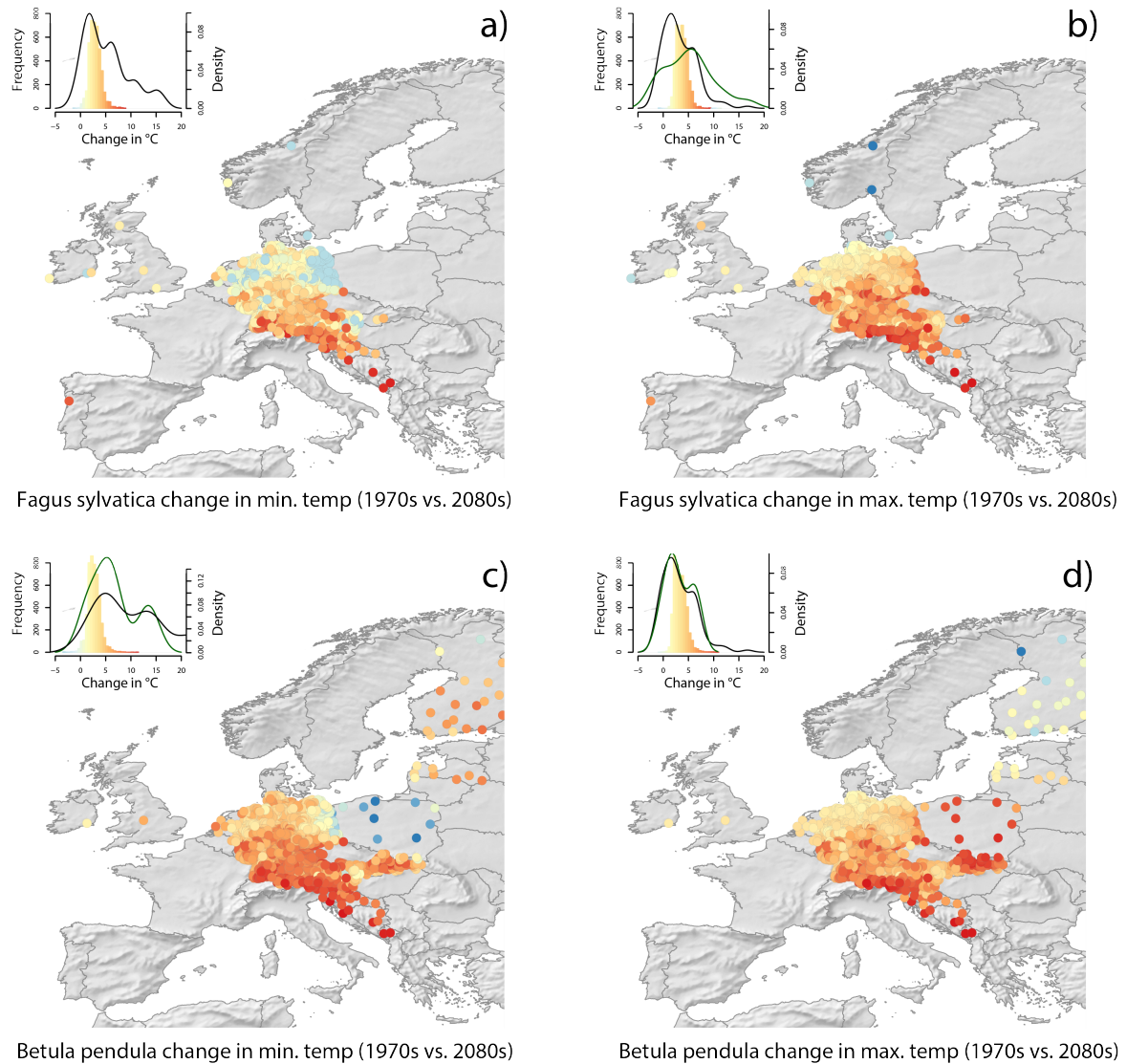


Figure 4: **Findings from controlled environment experiments are relevant for forecasting applications** based on comparison of two well-studied species: *Fagus sylvatica* (a-b) and *Betula pendula*. Points represent a site with spring phenology data (from the PEP725 database, Templ *et al.*, 2018) and show the predicted change (2080s average - 1970s average) in minimum (a,c) and maximum (b,d) temperatures 60 days before leafout. We found that treatment differences (see lines in inlay plots, green lines show the treatments for that exact species, while black lines show across all species in the OSPREE database) generally bracketed predicted changes (histograms in inlay plots show the same predicted changes in temperature represented also by the mapped points), but were on average much larger in magnitude. We show chilling (a,c) and forcing (b,d) differences in treatments across studies (for *Fagus sylvatica* there are no chilling treatments of differing temperatures, so green line is absent in inset of a). For more details, see ‘Comparing experimental treatments to forecasted trends’ in the Supporting Information.