

# 2. FREE ENERGY PERTURBATION (FEP)

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## 1 Free Energy Perturbation (FEP)

Free energy perturbation simulations can be used for theoretical prediction of binding free energies and along with it the elucidation and/or prediction of chemical reaction mechanisms. In this step-by-step tutorial you will be able to compute the relative binding free energy of camphane and camphor to an enzyme which catalyzes the addition of molecular oxygen to nonactivated hydrocarbons at physiological temperature. This oxydizing enzyme is called cytochrome P450 cam [1].

### 1.1 Theoretical background

The Free Energy Perturbation method is just that, a method, it is not, by any means, a theory. The method, in broad lines, allows a travel between states given a forced path. This makes sense since, by definition, the Free Energy of a system is a thermodynamic state function and therefore its value must be independent of the path taken. An approximation is made that a partition function for a perturbed system can be separated from the full partition function of the system as given by equation 3. The original expressions given by Zwanzig [2] to describe the idea of the Free Energy Perturbation (FEP) method as applied to hot gases are given below:

$$\exp(-\beta\bar{A}_N) = Q_N = \frac{1}{N!} \int_v \dots \int_v \times \exp(-\beta V_N) d\tau_1 \dots d\tau_N \quad (1)$$

$$V_N = V_N^{(0)} + V_N^{(1)} \quad (2)$$

$$\exp - \beta(\bar{A}_N - \bar{A}_N^{(0)}) \equiv \exp(-\beta\bar{A}_N^{(1)}) \quad (3)$$

$$\exp(-\beta\bar{A}_N^{(1)}) = \langle \exp(-\beta V_N^{(1)}) \rangle_0 \quad (4)$$

Where  $\beta = 1/\kappa T$ ,  $\kappa$  is Boltzmann's constant,  $T$  is the temperature in Kelvin,  $Q_N$  is the configurational partition function of the system,  $\bar{A}$  is the configurational free energy, and  $V_N$  is the total potential energy of the system.

Notice that the paper is called "High Temperature Equation of State by a Perturbation Method. I. Nonpolar Gases". Why the title specifically states the high temperature case is because it deals with a gas where particles are not interacting with each other and so can be considered to be having fully elastic collisions.

A more modern notation to describe the perturbation method as applied to free energies is given by Pohorille, Jarzynski and Chipot[3] in the following expression:

$$\exp(-\beta\Delta A) = \langle \exp(-\beta\Delta U) \rangle \quad (5)$$

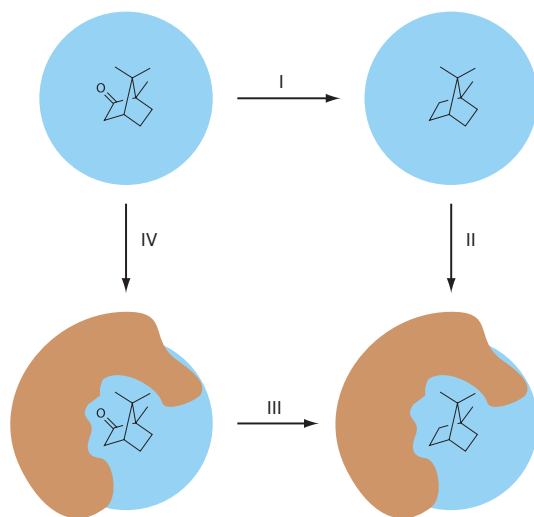


Figure 1: Thermodynamic cycle for ligand binding of camphene (CMA) and camphor (CAM) to the P450cam enzyme. The brown blob denotes the enzyme, and the blue background represents water.

Eq. 4 is derived from the statistical mechanics expression for the Helmholtz free energy,  $A$ , and the configurational partition function integral,  $Z$ .

$$A = -kT \cdot \ln(Z) \quad (6)$$

$$Z = \int e^{-V/(kT)} d\Gamma \quad (7)$$

At constant pressure and volume the change in Helmholtz free energy equals that in Gibbs free energy. The free energy difference between the two different states A and B, that are represented with potential energies  $V_A$  and  $V_B$  respectively, can be rearranged into the eq. 4 as follows:

$$\begin{aligned} \Delta G_{A \rightarrow B} &= -(kT \cdot \ln(Z_B) - kT \cdot \ln(Z_A)) \\ &= -kT \cdot \ln(Z_B/Z_A) \\ &= -kT \cdot \ln\left(\left(\int e^{-V_B/(kT)} d\Gamma\right) / \left(\int e^{-V_A/(kT)} d\Gamma\right)\right) \\ &= -kT \cdot \ln\left(\left(\int e^{-V_B/(kT)} e^{-V_A/(kT)} e^{V_A/(kT)} d\Gamma\right) / \left(\int e^{-V_A/(kT)} d\Gamma\right)\right) \\ &= -kT \cdot \ln \langle e^{-\Delta V_{A \rightarrow B}/(kT)} \rangle_A \end{aligned} \quad (8)$$

FEP is very accurate concerning small perturbations, when the difference between the potential  $V_A$  and  $V_B$  is smaller than 2 kT. The free energy of association of the ligands to a protein is calculated with FEP as a relative difference between the ligands. The absolute binding energy is defined as a free energy associated with moving the ligand in water to a solvated protein, paths II and IV in Figure 1. The relative binding energies are defined as a difference between two simulations where the ligand A is stepwise transformed to ligand B in water and in solvated protein, paths I and III in Figure 1.

## 1.2 FEP simulations using qdyn

Before starting the tutorial change the directory to `protein` or `water` in the FEP folder.

In this tutorial we are going to analyze the perturbation of camphor (CAM) to camphane (CMA) in water and in the P450cam enzyme. P450 enzymes catalyze the hydroxylation of unactivated alkanes, and P450cam specifically catalyzes the hydroxylation of camphor. Although the system investigated here is P450cam the rules in this tutorial are general and can be applied to any system of interest.

A FEP simulation is specified in **Q** with a `name.fep` file where the transformation is defined in detail. Every `name.fep` file is split into a number of sections where we can define atoms, charges, bonds and angles which are changing during the FEP simulation (see Figure 2). Note that a section in **Q** follows the syntax [...].

- Open the `cma_cam.fep` file, in the `protein` or `water` folder, that is used for the CMA to CAM perturbation. Try to locate the different sections that are used in the FEP file. Can you understand how CMA is transformed to CAM?

- What effect do they have on the simulation? (HINT: For the detailed explanation of the different sections consult Appendix A!)

```
[atoms]
...

[FEP]
...

[change_charges]
...

[atom_types]
...

.
.
.
```

Figure 2: FEP file format.

Having a FEP file the perturbation is defined in Q by adding a specifier

```
fep          name.fep
```

in the input files for **qdyn**. Additionally it is necessary to add the section `[lambdas]` where the lambda values are defined,  $\lambda_1$  and  $\lambda_2$ . The lambda values are used to transform the potential  $V_A$  to  $V_B$  in small steps to improve the convergence. The energies are sampled on the potential  $V$ ,

$$V = \lambda_1 V_A + \lambda_2 V_B = \lambda_1 V_A + (1 - \lambda_1) V_B \quad (9)$$

where  $\lambda_1$  varies between 0 and 1. How many lambda steps are used in the simulation depends on difference between  $V_A$  and  $V_B$  and the type of perturbation.

- Open an input file `name.inp`. In what section is the fep file read?

- Find the `[lambdas]` section. How many lambda steps are used in the perturbation and how big is each step? (HINT: Look in several `cma_camN.inp` files, where  $N=0..30$ .)

- How many steps are used in the simulation in each file?
- Also note the specifiers

energy	name.en
energy	25

that are used for saving the energies every 25th step to the file name.en.

### 1.3 Analysis of FEP simulations using qfep

All steps in this tutorial take approximately 2-3 hours running in parallel in eight processor cores in a 2.3 GHz processor. **qfep** is the analysis program which allows you to compute the free energy differences between states using energies which have been stored in the binary files with ".en" extension, that is, files such as `cma_cam30.en`. To run **qfep** you must issue the following command for both the simulation in plain water and in the protein:

```
qfep < qfep.inp > qfep.out
```

The `qfep.inp` file contains all necessary instructions issued to **qfep** and the `qfep.out` contains the full output.

- Open `qfep.inp` and `qfep.out`. Try to understand the different commands. What are they specifying? (HINT: Check out Appendix B!)

- The free energy difference for the perturbation from CMA to CAM can be found at the end of table summarized in Part 1. What is the difference between water and protein?

Water simulation:  $\Delta G_{A \rightarrow B}^w =$  (kcal/mol)

Protein simulation:  $\Delta G_{A \rightarrow B}^p =$  (kcal/mol)

The free energy is calculated as an average of forward and backward calculations at every lambda point. The forward and backward free energy values are also summarized in Part 1 of the `qfep.out` file in columns `sum(dGf)` and `sum(dGr)`. The theoretical error of the FEP simulation is half the value of the difference between forward and backward calculations.

-What is the theoretical error of FEP for water and protein simulations?

Water simulation:  $\Delta G_{A \rightarrow B}^{w,error} =$  (kcal/mol)

Protein simulation:  $\Delta G_{A \rightarrow B}^{p,error} =$  (kcal/mol)

The relative free energy of binding is determined as the difference between the free energies for the protein and water simulations.

Relative binding free energy:

Calculated:  $\Delta \Delta G_{bind,rel}^{calc} = \Delta G_{A \rightarrow B}^p - \Delta G_{A \rightarrow B}^w =$  (kcal/mol)

Experimental:  $\Delta \Delta G_{bind,rel}^{exp} = \Delta G_{A \rightarrow B}^p - \Delta G_{A \rightarrow B}^w = -2.0$  (kcal/mol)

Alternatively the free energy profiles can be plotted and investigated graphically. The program `gnuplot` is used for plotting different graphs.

- Open `gnuplot` by typing `gnuplot` in the shell window and then write `load 'fep.part1.pgp'`. This script plots the free energy as a function of lambda, as summarized in the table in Part 1 of `qfep.out`.

- What is the shape of the curve? What is the spacing between the points? Does it look ok?

Molecular dynamics simulations also gives us an opportunity to investigate the structures as they are propagated through time.

- Type **pymol** and open `cma_cam.pse` which loads `cma_cam0.pdb`, `cma_cam10.pdb` and `cma_cam30.pdb` files in **pymol**. The name .pdb files represent structures at the lambda state (1,0), (0.63,0.37) and (0,1) respectively.

- What is the difference between the different ligand structures?

- What are the interactions between the ligands and the surrounding. (HINT: Check out Tyr87 residue!)

Here ends the FEP part of the lab course!

## A FEP file format

Table 1: FEP file format

[atoms]: Define Q-atoms.

column	description
1	Q-atom number (counting from 1 up).
2	Topology atom number.

[PBC]: For periodic boundary conditions.

keyword	value	comment
switching- _atom	Topology atom number.	Required with periodic boundary conditions.

[FEP]: General perturbation information.

keyword	value	comment
states	Number of FEP/EVB states.	Optional, default 1.
offset	Topology atom number.	Optional, default 0. This number is added to all topology atom numbers given in the FEP file.
offset_residue	Residue/fragment number.	Optional. Set offset to the topology number of the first atom in the given residue minus one.
offset_name	Residue/fragment name.	Optional. Set offset to the topology number of the first atom in the first residue with the given name minus one.
qq_use- _library- _charges	This is a special feature for studying <i>e.g.</i> electrostatic linear response. Set to 'on' to use the library charges from the topology for intra-Q-atom interactions, i. e. change only Q-atom-surrounding electrostatic interactions.	Optional, default off.
softcore- _use_max- _potential	Set to 'on' if the values entered in the [softcore] section are the desired maximum potentials (kcal/mol) at $r = 0$ . Qdyn will then calculate pairwise $\alpha_{ij}$ to be used in equation 10. 'off' means the values are to be used directly in equation 10.	Optional, default off.

[change\_charges]: Redefine charges of Q-atoms.

column	description
1	Q-atom number (referring to numbering in atoms section).
2...	Charge (e) in state 1, state 2, ...

[atom\_types]: Define new atom types for Q-atoms: Standard LJ parameters and parameters for the exponential repulsion potential  $U_{soft} = C_i \cdot C_j e^{-a_i \cdot a_j \cdot r_{i,j}}$ .

1	Name (max 8 characters).
2	Lennard-Jones A parameter ( $\text{kcal}^{\frac{1}{2}} \cdot \text{mol}^{-\frac{1}{2}} \cdot \text{\AA}^{-6}$ ) for geometric combination or $R^*$ ( $\text{kcal} \cdot \text{mol}^{-1} \cdot \text{\AA}^{-12}$ ) for arithmetic combination rule.
3	LJ B parameter ( $\text{kcal}^{\frac{1}{2}} \cdot \text{mol}^{-\frac{1}{2}} \cdot \text{\AA}^{-3}$ ) or $\epsilon$ ( $\text{kcal} \cdot \text{mol}^{-1} \cdot \text{\AA}^{-6}$ ).
4	Soft repulsion force constant $C_i$ ( $\text{kcal}^{\frac{1}{2}} \cdot \text{mol}^{-\frac{1}{2}}$ ) in $U_{soft}$ .
5	Soft repulsion distance dependence parameter $a_i$ ( $\text{\AA}^{-\frac{1}{2}}$ ) in $U_{soft}$ .
6	Lennard-Jones A parameter ( $\text{kcal}^{\frac{1}{2}} \cdot \text{mol}^{-\frac{1}{2}} \cdot \text{\AA}^{-6}$ ) or $R^*$ ( $\text{kcal} \cdot \text{mol}^{-1} \cdot \text{\AA}^{-12}$ ) for 1-4 interactions.
7	LJ B parameter ( $\text{kcal}^{\frac{1}{2}} \cdot \text{mol}^{-\frac{1}{2}} \cdot \text{\AA}^{-3}$ ) or $e$ ( $\text{kcal} \cdot \text{mol}^{-1} \cdot \text{\AA}^{-6}$ ) for 1-4 interactions.
8	Atomic mass (u).

[change\_atoms]: Assign Q-atom types to Q-atoms.

1	Q-atom number.
2...	Q-atom type name in state 1, state 2, ...

Table 1: FEP file format

**[soft\_pairs]:** Define pairs which use soft repulsion.

1	Q-atom number of first atom in pair.
2	Q-atom number of second atom in pair.

**[excluded\_pairs]:** Define pairs to exclude from non-bonded interactions. Note: also non-Q-atoms can be excluded.

1	Topology atom number of first atom in pair.
2	Topology atom number of second atom in pair.
3...	Exclusion effective (1) or not (0) in state 1, state 2, ...

**[el\_scale]:** Define q-atom pairs for scaling of the electrostatic interaction. Can be useful e.g. when highly charged intermediates appear in FEP/EVB. The scale factor applies to all states. Note: only Q-atom pairs can be scaled.

1	q-atom number of first atom in pair
2	q-atom number of second atom in pair
3	electrostatic scale factor (0..1)

**[softcore]:** Define q-atom softcore potentials. The meaning of these entries depends on the value of `softcore_use_max_potential`.

1	q-atom number
2...	Desired potential at $r = 0$ for all of this q-atom's vdW interactions in state 1, state 2, ... or the actual $\alpha$ value used in equation 10. An $\alpha$ of 200 yields vdW potentials at $r = 0$ of 10-50 kcal/mol for heavy atom - heavy atom interactions. Set to 0 if softcore is not desired for this q-atom.

**[monitor\_groups]:** Define atom groups whose non-bonded interactions are to be monitored (printed in the log file).

1...	Topology atom number of first and following atoms in group.
------	---

**[monitor\_group\_pairs]:** Define pairs of `monitor_groups` whose total non-bonded interactions should be calculated.

1	First <code>monitor_group</code> number.
2	Second <code>monitor_group</code> number.

**[bond\_types]:** Define Q-bond types using Morse or harmonic potentials,

$$E_{\text{Morse}} = D_e \left( 1 - e^{-\alpha(r-r_0)} \right)^2 \quad E_{\text{Harmonic}} = \frac{1}{2} k_b (r - r_0)^2.$$

Morse and harmonic potentials can be mixed (but each bond type is either kind). Entries with four values are Morse potentials and entries with three values are harmonic.

	Morse potential	Harmonic potential
1	Q-bond type number (starting with 1).	
2	Morse potential dissociation energy, $D_e$ (kcal·mol <sup>-1</sup> ).	Harmonic force constant $k_b$ (kcal·mol <sup>-1</sup> ·Å <sup>-2</sup> ).
3	Exponential co-efficient $\alpha$ in Morse potential (Å <sup>-2</sup> ).	Equilibrium bond length $r_0$ in harmonic potential (Å).
4	Equilibrium bond length $r_0$ in Morse potential (Å).	

**[change\_bonds]:** Assign Q-bond types. Note: shake constraints for the redefined bonds are removed. The order in which atoms are given is not important.

1	Topology atom number of first atom in bond.
2	Topology atom number of second atom in bond.
3...	Q-bond type number (referring to numbering in <code>bond_types</code> section) or 0 to disable bond in state 1, state 2, ...

**[angle\_types]:** Define Q-angle types.

1	Q-angle type number (starting with 1).
2	Harmonic force constant (kcal·mol <sup>-1</sup> ·rad <sup>-2</sup> ).
3	Equilibrium angle (°).



Table 1: FEP file format

**[change\_angles]:** Assign Q-angle types.

1	Topology atom number of first atom in angle.
2	Topology atom number of middle atom in angle.
3	Topology atom number of third atom in angle.
4...	Q-angle type number (referring to numbering in angle_types section) or 0 to disable angle in state 1, state 2, ...

**[torsion\_types]:** Define Q-torsion types.

1	Q-torsion type number (starting with 1).
2	Force constant = $\frac{1}{2}$ · barrier height (kcal·mol <sup>-1</sup> ).
3	Periodicity (number of maxima per turn).
4	Phase shift (°).

**[change\_torsions]:** Assign Q-torsion types. Note: The order of atoms (1, 2, 3, 4 or 4, 3, 2, 1) is not important.

1	Topology atom number of first atom in torsion.
2	Topology atom number of second atom in torsion.
3	Topology atom number of third atom in torsion.
4	Topology atom number of fourth atom in torsion.
5...	Q-torsion type number (referring to numbering in torsion_types section) or 0 to disable torsion in state 1, state 2, ...

**[improper\_types]:** Define Q-improper types.

1	Q-improper type number (starting with 1).
2	Harmonic force constant (kcal·mol <sup>-1</sup> ·rad <sup>-2</sup> ).
3	Equilibrium angle (°).

**[change\_impropers]:** Assign Q-improper types. Note: The order of atoms (1, 2, 3, 4 or 4, 3, 2, 1) is not important.

1	Topology atom number of first atom in improper.
2	Topology atom number of second atom in improper.
3	Topology atom number of third atom in improper.
4	Topology atom number of fourth atom in improper.
5...	Q-improper type number (referring to numbering in improper_types section) or 0 to disable improper in state 1, state 2, ...

**[angle\_couplings]:** Couple Q-angles to Q-bonds, *i.e.* scale angle energy by the ratio of the actual value of the Morse bond energy to the dissociation energy.

1	Q-angle number (line number within change_angles section).
2	Q-bond number (line number within change_bonds section).

**[torsion\_couplings]:** Couple Q-torsions to Q-bonds.

1	Q-torsion number (line number within change_torsions section).
2	Q-bond number (line number within change_bonds section).

**[improper\_couplings]:** Couple Q-impropers to Q-bonds.

1	Q-improper number (line number within change_impropers section).
2	Q-bond number (line number within change_bonds section).

**[shake\_constraints]:** Define extra shake constraints. The effective constraint distance will be the sum of the distances given for each state, weighted by their *l* values. Note: constraints defined here do not override constraints imposed by setting the shake flag to *on* in the Qdyn input file. To remove a constraint the bond must be redefined as a Q-bond. The order in which atoms are given is not important.

1	Topology atom number of first atom.
2	Topology atom number of second atom.
3...	Constraint distance (Å) in state 1, state 2, ...

Table 1: FEP file format

**[off-diagonals]:** Define off-diagonal elements of the Hamiltonian, represented by  $H_{i,j} = A_{i,j} \cdot e^{-\mu_{i,j} \cdot r_{k,l}}$  where i and j are states and k and l are Q-atoms.

1	State i.
2	State j.
3	Q-atom k.
4	Q-atom l.
5	$A_{i,j}$ (kcal·mol <sup>-1</sup> ).
6	$\mu_{i,j}$ (Å <sup>-1</sup> ).

Softcore equation:

$$V_{vdW}(r) = \frac{A_{ij}}{(r^6 + \alpha_{ij})^2} - \frac{B_{ij}}{r^6 + \alpha_{ij}} \quad \text{or} \quad V_{vdW}(r) = \epsilon \cdot \left( \frac{R_{ij}^{*12}}{(r^6 + \alpha_{ij})^2} - 2 \cdot \frac{R_{ij}^{*6}}{r^6 + \alpha_{ij}} \right) \quad (10)$$

## B qfep input summary

Some of the commands are affecting only analysis of the empirical valance bond (EVB) simulations, while others are common for both FEP and EVB. The ones affecting FEP are marked in bold text and are of concern in FEP simulations.

HINT:

→ specifies the input commands

# denotes the written output.

```
->Number of energy files:
# Number of files = 31
-> No. of states, no. of predefined off-diag elements:
# Number of states = 2
# Number of off-diagonal elements = 0
-> Give kT & no. of pts to skip:
# kT = 0.596
# Number of data points to skip = 80
-> Give number of gap-bins:
# Number of gap-bins = 40
-> Give minimum # pts/bin:
# Minimum number of points per bin= 10
-> Give alpha for state 2:
# Alpha for state 2 = 0.00
-> Number of off-diagonal elements:
# Number of off-diagonal elements = 0
-> linear combination of states defining reaction coord:
# Linear combination co-efficients= 1.00 0.00
name1.en
name2.en
.
.
nameN.en
```

The # Number of files is the total number of files used for the FEP simulation, # Number of data points to skip is the number of points that are discarded as the equilibration at each lambda step and # kT specifies the temperature of the simulation. The energy files name.en are read last in **qfep**.

## References

- [1] Schlichting, I., Berendzen, J., Chu, K., Stock, A. M., Maves, S. A., Benson, D. E., Sweet, R. M., Ringe, D., Petsko, G. A., and Sligar, S. G. (2000) The catalytic pathway of cytochrome P450cam at atomic resolution. *Science*, **287**, 1615–1622.

- [2] Zwanzig, R. W. (1954) High-temperature equation of state by a perturbation method. I. nonpolar gases. *The Journal of Chemical Physics*, **22**, 1420–1426.
- [3] Pohorille, A., Jarzynski, C., and Chipot, C. (2010) Good practices in free-energy calculations. *The Journal of Physical Chemistry B*, **114**, 10235–10253.