# **Sequence Analysis 1**

# A. Concepts, conservation & substitution

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SCIE2100 | BINF6000 | Bioinformatics 1 – Introduction

### Lecture outline

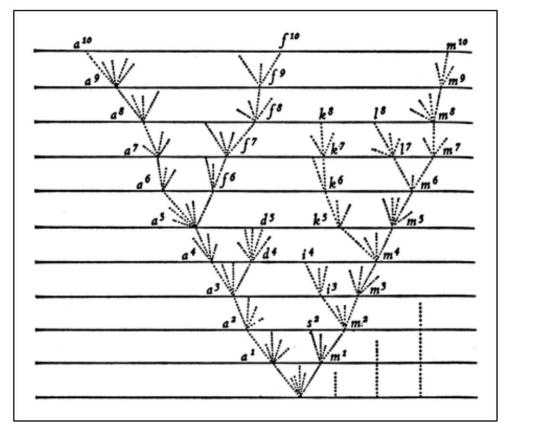
- Concept of homology
  - Homology and sequence similarity
- Sequence change in evolution
  - Random change versus biological evolution
  - Sequence change versus conservation
- Quantifying sequence change
  - Observed versus null probabilistic models
  - Log-odds score
  - Basic principles of PAM and BLOSUM matrices, and their differences

# **Concept of homology**

Organisms related by genealogical descent with modification

Traits (and genes) are passed on from one generation to the next (inheritance)

Charles Darwin (1859). On the Origin of Species



Features derived from a common ancestor are said to be **homologous**. This applies to any feature – morphological or molecular (genes, RNAs, proteins).

time

# Homology and sequence similarity

• Sequence similarity (or sequence identity): a measure of identical residues shared between two sequences, usually in an alignment (where identical/similar residues were aligned); the extent to which two sequences are invariant

Example:

Seq1 ACGTAGCTAGCTACCT

Seq2 ACGTAGCTAGCTAGCT

%identity between

Seq1 & Seq2

**= 19/20 = 95%** 

- High level of sequence similarity usually, but not necessarily, indicates evidence for homology
- Similar sequences may be homologs

# Homology and sequence similarity

 Conservation: unchanged/invariant positions when comparing two sequences. At amino acid level, this can also refer to changes at an amino acid position that preserves the physicochemical property

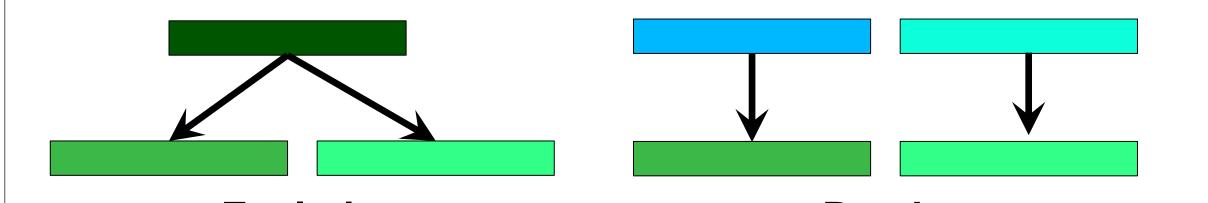
Phenylalanine (**F**)
Valine (**V**) are
hydrophobic

Tyrosine (**Y**) and Histidine (**H**) are polar

Homology is an evolutionary relationship that either exists or not (i.e. it is **all-or-nothing**, there are no "degrees of homology"). We may be able to quantify how confident we are in believing that two molecules/sequences are homologous, but they are nonetheless either homologous or non-homologous.

# Sequence change in evolution

Two alternative explanations to observed sequence similarity:

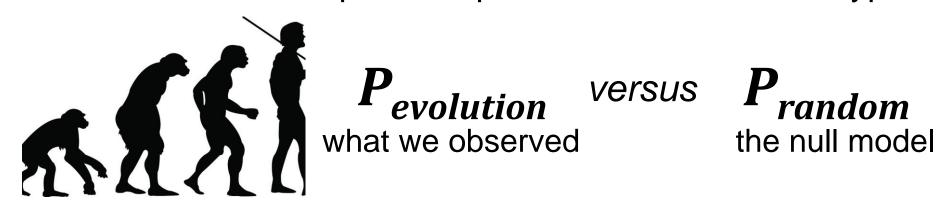


# **Evolution**

similarity is due to shared ancestry

Random similarity is coincidental by pure chance

Compare the probabilities of the two hypotheses:





# Random sequence change

What are the chances of randomly matching 2 basepairs (bp) in a human-size genome (3 x 10<sup>9</sup> bp)?

Total genome size

3,000,000,000 = 187,500,000

$$\frac{187,500,000}{3,000,000,000} \times 100 = 6.25\%$$

Probability of a match to occur at random

number of

possible

matches

# Random sequence change

What are the chances of randomly matching **N** basepairs (bp) in a human-size genome (3 x 10<sup>9</sup> bp)?

Length of match in bp (N)	# possible matches $(3 \times 10^9 / 4^N)$	Probability $(P_{random})$
1	750,000,000	25.0 %
2	187,500,000	6.25 %
10	2,861	9.5 x 10 <sup>-5</sup> %
20	$2.73 \times 10^{-3}$	9.1 x 10 <sup>-11</sup> %
300	7.23 x 10 <sup>-172</sup>	2.4 x 10 <sup>-179</sup> %

### Random change or biological evolution?

Chimpanzee genome size ≈ human genome size ≈ **3Gbp**. Consider a **300bp**-region in chimpanzee and in human:

%identity

 $= 296/300 \times 100\%$ 

**= 98.67%** 

GGCTGTCATCACTTAGACCTCACCCTGTGG AGCCACACCCTAGGGTTGGCCAATCTACTC CCAGGAGCAGGAGCCAGGGCT GGGCATAAAAGTCAGGGCAGAGCCATCTAT TGCTTACATTTGCTTCTGACACAACTGTGT TCACTAGCAACCTCAAACAGACACCATGGT **GCATCTGACTCCTGAGGAGAAGTCTGCCGT** TACTGCCCTGTGGGGCAAGGTGAACGTGGA TGAAGTTGGTGGTGAGGCCCTGGGCAGGTT GGTATCAAGGTTACAAGACAGGTTTAAGGA Example

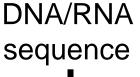
GGCTGTCATCACTTAGACCTCACCCTGTGG AGCCACACCCTAGGGTTGGCCAATCTACTC CCAGGAGCAGGAGCCAGGGCT GGGCATAAAAGTCAGGGCAGAGCCATCTAT TGCTTACATTTGCTTCTGACACAACTGTGT TCACTAGCAACCTCAAACAGACACCATGGT ACCGCTGACTCCTGAGGAGAAGTCTGCCGT TACTGCCCTGTGGGGCAAGGTGAACGTGGA TGAAGTTGGTGGTGAGGCCCTGGGCAGGTT GGTATCAAGGTTACAAGACAGGCTTAAGGA

**P**<sub>random</sub> of 296bp identical match in a 3Gbp genome

 $= 6.17 \times 10^{-177} \%$ 

Do you think human and chimpanzee share a common ancestry?

# Sequence change versus conservation





Protein sequence



Tertiary structure



**Function** 

- Selective pressure for divergence e.g. genetic diversity (to increase viability), e.g. fast-evolving genes
- Substitutions, insertions, deletions, translocations, genetic transfers/exchange, etc.
- Adaptation due to changes in environments (biotic and abiotic stressors)
- Copy/replication errors

- Selective pressure for preservation of critical gene function, protein structure and function, including non-coding sequences and regulatory elements
- Especially true for critical machineries and slow-evolving genes e.g. housekeeping genes, ribosomal RNA genes (phylogenetic markers)

# Quantifying sequence change

Homology and sequence conservation is commonly observed at the protein

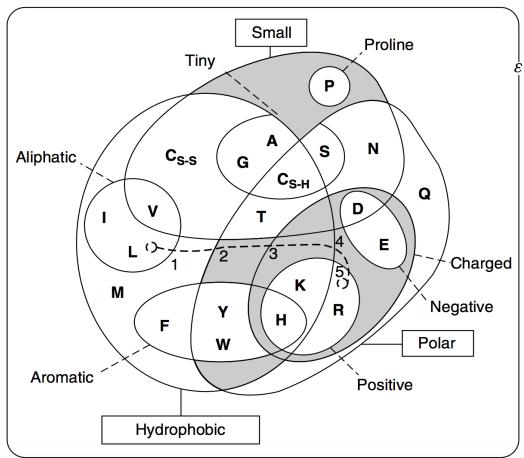
level. Why?

 Codon degeneracy: nearly one-third of the bases in coding regions are under a weak (if any) selection

 Greater information content: 20 amino acids versus 4 bases

Among a set of homologous sequences: **observed substitution frequency** for each amino acid can be used to quantify sequence change

 Commonly weighted based on shared physicochemical properties of amino acids

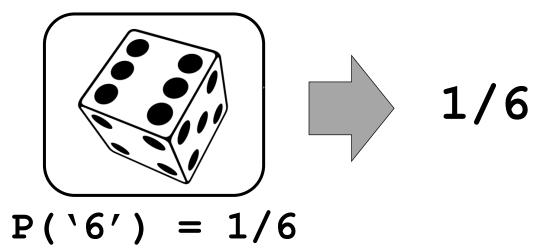


Betts & Russell (2003) Chapter 14. *Bioinformatics for Geneticists*.In: Barnes MR & Cray IC (Eds). John Wiley & Sons. Chapter 13 in the 2nd Edition (QH430 .B375 2007)

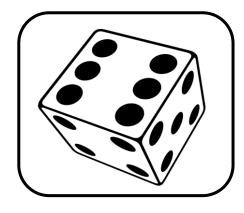
# Probability in an unbiased null model

On a die:

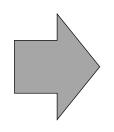
Landing a 6?



Landing a 6 then a 2?







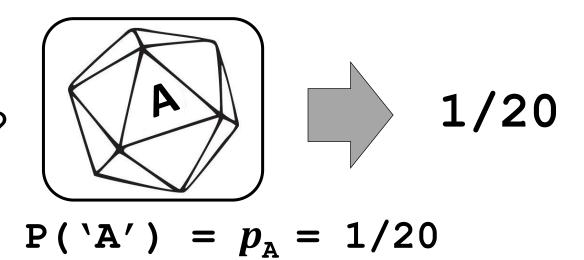
1/36

$$P('6' \land '2') = 1/6 \times 1/6$$
  
= 1/36

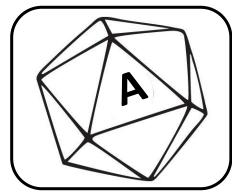
# Probability in an unbiased null model

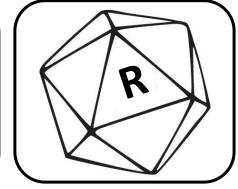
Imagine a 20-face die (each face represents an amino acid)

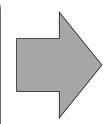
Observing an **A**? (Alanine)



Observing **A** in one but **R** in another?
(Arginine)





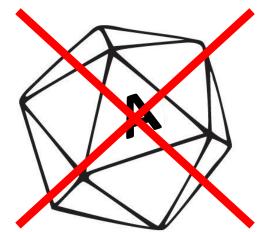


1/400

$$P('A' \land 'R') = p_A \times p_R = 1/20 \times 1/20$$
  
= 1/400

# Null model based on empirical data Amino acid composition (%) in UniProt/TrEMBL (Feb 2021) http://www.ebi.ac.uk/uniprot/TrEMBLstats

He Asp Pro Lys Phe Asn Gln Tyr Met



 $p_{\rm s} = 0.0672$ 

### Amino acid composition (%)

(A) 9.23Gln (Q) 3.77(L) 9.86(S) Ser Ala Leu (R) 5.845.54 Glu (E) Lys (K) 4.89 Arq 6.20 (T)1.30 (N) 3.78Gly (G) 7.35 Met (M) 2.35 (W) Asn (D) 5.48 2.20 (F) 3.88 Tyr 2.87 His (H) Phe Asp (C) 1.24(I)5.53 Pro (P) 4.94 6.91 Cys

# Probability in a little-more-realistic null model

Observing A in a sequence

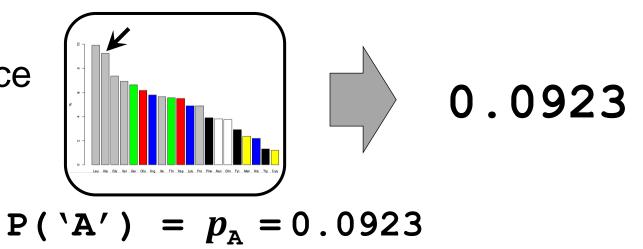
sequence Y ...R...

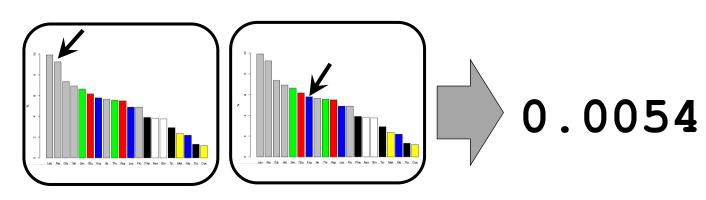
Aligned position:

 $X_i$ : position i in sequence X

 $Y_i$ : position j in sequence Y

Observing **A** at  $X_i$  and **R** at  $Y_j$  simply by chance





$$P(X_i = A' \land Y_j = R') = p_A \times p_R$$
  
= 0.0923 \times 0.0584  
= 0.0054

# Log-odds score for amino acid substitution

 $p_a$ : **prior** probability of observing residue a  $p_b$ : **prior** probability of observing residue b

 $q_{ab}$ : **joint** probability of observing residue a and residue b in the same column

 $\gamma$ : a scaling factor (e.g.  $\gamma = 10$  if  $\log_{10}$  scale is used)

# Log-odds score for a substituted by b $S_{ab} = \gamma \log \left( \frac{q_{ab}}{p_a \cdot p_b} \right)$

 $S_{ab} > 0$  (**positive**): **more** frequent than expected  $(q_{ab} > p_a \times p_b)$  $S_{ab} < 0$  (**negative**): **less** frequent than expected  $(q_{ab} < p_a \times p_b)$ 

Consider the A-R substitution Seq1 YPSVPFSAGP in these two sequences: Seq2 YPVLPFSRGP

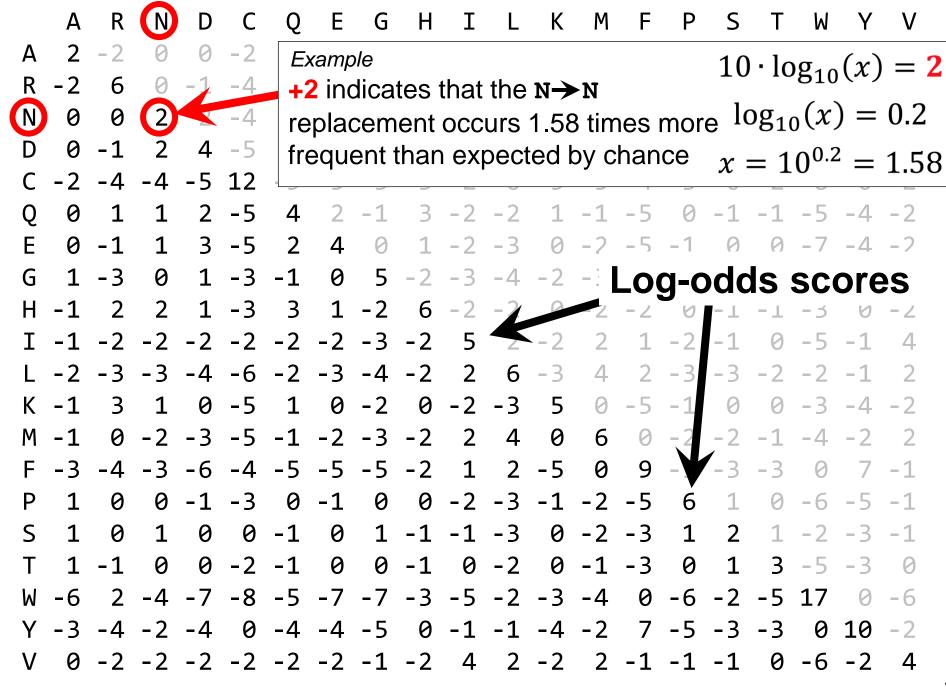
Example

 $P_{observed}$  for  $A \rightarrow R$  substitution =  $q_{AR}$ 

 $P_{null}$  for  $A \rightarrow R$  substitution =  $p_A \times p_R$ 

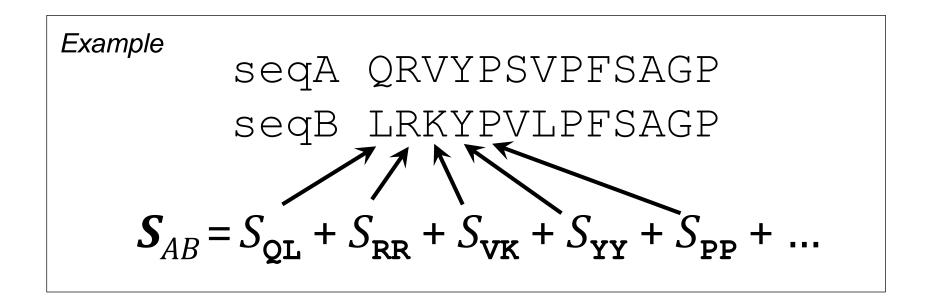
# Substitution matrix

captures the propensity of any amino acids to be substituted by another amino acid due to biological reasons



# Why do we use log-odds score?

- Logarithms are easier to use for a scoring system
- They allow us to sum the scores of aligned residues (rather than multiplying the probabilities for independent mutations)
- The sum of log-odds is equivalent to product of probabilities



# Point Accepted Mutation (PAM) matrices

PAM matrices (Dayhoff, Schwartz & Orcutt, 1978)

based on 1,572 observed mutations in
 71 families of closely related proteins

An **accepted point-mutation** is a single-residue mutation that was incorporated into the protein (and passed to its progeny), thus it:

- (a) did not disrupt the protein function or
- (b) was **beneficial** to the organism (e.g. in evolutionary terms, it increased the fitness of the species)

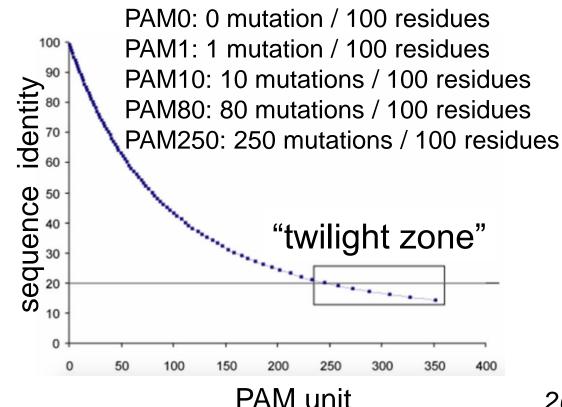


Margaret O. Dayhoff (1925–1982)

### **PAM** units

1 PAM unit: a series of accepted point mutations (and no insertions of deletions) has converted  $S_1$  to  $S_2$  with an average of **one accepted** point-mutation event per 100 amino acids. It measures the rate of divergence, i.e. the evolutionary distance.

- PAM unit between two sequences is not necessarily the same as percent difference in sequence identity
- Single position may undergo > 1 mutation, which could also result in no change observed in the sequence,



### Construction of PAMn substitution matrices

1. for PAM1 matrix, protein sequences with >85% identity are used

- GICGG GICGL Evolutionary tree **ALSGA ALCAA** (parsimony Similar
- 2. count of amino acid replacements are recorded along branches of a phylogenetic tree
  - sequences

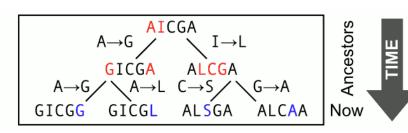
### **Count matrix**

3. transition probability for each pair of amino acids is calculated based on the count matrix and the occurrence of these amino acids in the dataset

### **Transition probability matrix**

- 4. matrices of other PAM units are extrapolated from PAM1 via matrix multiplication:  $PAM2 = (PAM1)^2$ ;  $PAM250 = (PAM1)^{250}$
- 5. probabilities can be transformed into log-odds scores

### **Scoring matrix**



### count matrix

oodiii iiidaan					
	Α	R	N	D	С
Α	9867	2	9	10	3
R	1	9913	1	0	1
N	4	1	9822	36	0
D	6	0	42	9859	0
C	1	1	0	0	9973

PAM1

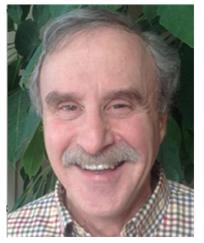
Dropapility matrix						
	Α	R	N	D	С	
Α	13	6	9	9	5	
R	3	17	4	3	2	
N	4	4	6	7	2	PAM250
D	5	4	8	11	1	
С	2	1	1	1	52	-

### scoring matrix

		_				
Α	2		PAM250			
R	-2	6 PAIVI230				
N	0	0	2			
D	0	-1	2	4		
С	-2	-4	-4	-5	12	

# **BLOck SUbstitution Matrix (BLOSUM)**

- Based on clustering of distantly related proteins
- Blocks database consists of >2000 locally aligned (blocks) of conserved regions from >500 groups of distantly related proteins
- Observed amino acid frequencies derived based on aligned blocks (no phylogenetic trees)





Steven & Jorja Henikoff

- In BLOSUMn matrices, sequences with identity > n% are clustered
- Scores are derived from intercluster differences (among sequences sharing < n% identity).

# **BLOSUM: clustering based on %identity**

### Example

AKLGGREAVE AKLIGREAVE DKIGGHPAIE DNIGGQPAIE DKIGGQPAIE EKLGGTTAVD EKLGGTTAMK EKLGGTAAVQ EKLGGQAAVQ YEAIGEELLS

Cluster at % identity ≥ **80** 

AKLGGREAVE AKLIGREAVE

DKIGGHPAIE
DNIGGQPAIE
DKIGGQPAIE

Identity for each possible pairwise sequences within each cluster ≥ 80%

EKLGGTTAVD
EKLGGTTAMK
EKLGGTAAVQ
EKLGGQAAVQ

These four clusters form the basis for BLOSUM80 matrix

YEAIGEELLS

# **BLOSUM:** deriving transition probability

Calculate  $q_{\rm QN}$  for BLOSUM**50** 

Example		• 3 possible pairs of clusters (1-2, 1-3 and 2-3)					
4	ATCKQ	5 amino acid residues in length -					
) <del> </del>	ATCRN	$A$ : total number of aligned pairs = $3 \times 5 = 15$					
		<b>B:</b> total $Q \longrightarrow N$ substitution frequency <b>between</b> each cluster					
	ASCK <b>N</b>	between	$\frac{1}{4}$ is Q in 1 compares to $\frac{1}{2}$ is N in 2 $\binom{1}{3}$ $\binom{1}{4}$ $\binom{3}{4}$ $\binom{1}{4}$				
	SSCRN	1-2	$\frac{1}{4}$ is <b>Q</b> in <b>1</b> compares to $\frac{1}{2}$ is <b>N</b> in <b>2</b> $\left(\frac{1}{4} \times \frac{1}{2}\right) + \left(\frac{3}{4} \times \frac{1}{2}\right) = \frac{4}{8} = \frac{1}{2}$				
2	SDCE <b>Q</b> SECE <b>N</b>		1/2 is Q in 2 compares to 0 is N in 3 1/2 is N in 2 compares to 1/1 is Q in 3				
3	TECRQ	between 1-3	<sup>1</sup> / <sub>4</sub> is Q in 1 compares to 0 is N in 3 <sup>3</sup> / <sub>4</sub> is N in 1 compares to 1/1 is Q in 3				
	clustered at %identity ≥ 5		<b>B</b> Total = $\frac{1}{2} + \frac{1}{2} + \frac{3}{4} = \frac{4+4+6}{8} = \frac{14}{8}$				

$$q_{QN} = \frac{B}{A} = \frac{14}{8} \div 15 = 0.1167$$

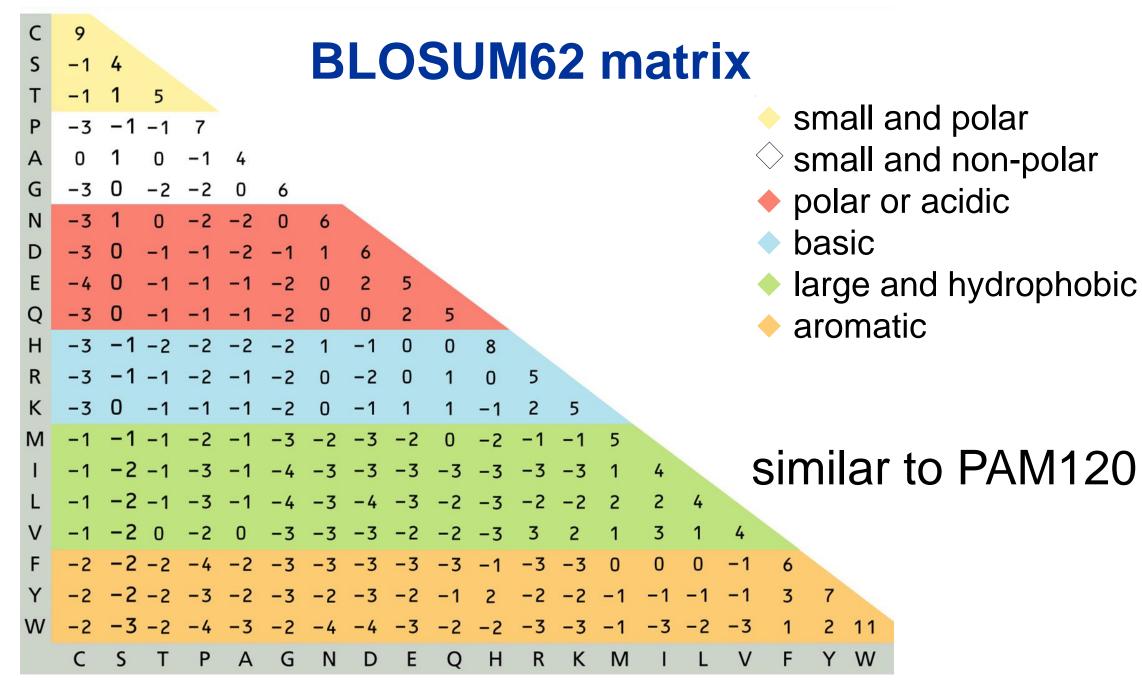
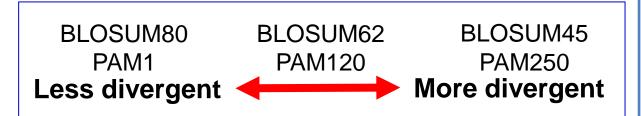


Figure 4.4 Textbook (pg. 83)

### **PAM** matrices

- Uses closely related proteins
- Based on an explicit evolutionary model (i.e. replacements counted on the branches of a phylogenetic tree)
- mutations observed throughout a global alignment
- All mutations are counted the same
- Higher PAM units denote larger evolutionary distance



### **BLOSUM** matrices

- Uses evolutionarily divergent proteins
- Based on an implicit model of evolution (no trees)
- Based only on highly conserved regions in series of local alignments without gaps
- Uses groups of sequences within which not all mutations are counted the same
- Larger numbers in the BLOSUM matrix naming scheme denote higher sequence similarity (and thus smaller evolutionary distance)

These matrices could be too simplistic. Further developments result in more-realistic substitution models based on more-extensive protein data, e.g. **JTT** and **WAG**.

### Reflection

What is homology, and how does it relate to shared sequence similarity?

Biologically, why would DNA/proteins sequences be conserved, or different from one another?

How do we quantify and model sequence change?

Why do we prefer log-odds score to probability values?

How can we model substitutions based on empirical data?

What are PAM and BLOSUM matrices, and how do they differ?