## Assignment 3: Data Exploration

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#### **OVERVIEW**

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

#### **Directions**

- 1. Rename this file <FirstLast>\_A03\_DataExploration.Rmd (replacing <FirstLast> with your first and last name).
- 2. Change "Student Name" on line 3 (above) with your name.
- 3. Work through the steps, **creating code and output** that fulfill each instruction.
- 4. Be sure to **answer the questions** in this assignment document.
- 5. When you have completed the assignment, **Knit** the text and code into a single PDF file.
- 6. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

The completed exercise is due on Sept 30th.

### Set up your R session

1. Check your working directory, load necessary packages (tidyverse), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX\_Neonicotinoids\_Insects\_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON\_NIWO\_Litter\_massdata\_2018-08\_raw.csv). Name these datasets "Neonics" and "Litter", respectively. Be sure to include the subcommand to read strings in as factors.

#### getwd()

```
## [1] "/Users/danleizou/EDA-Fall2022"
```

```
## [1] "/Users/danleizou/EDA-Fall2022"

#creating datasets
Neonics <- read.csv("./Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv", stringsAsFactors = TRUE)
Litter <- read.csv("./Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv", stringsAsFactors = TRUE)</pre>
```

## Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: We might be interested to see how certain species of insects react to neonicotinoids. This data would help keep track of how effective the insecticides are against different species. Some may be killed by the usage of such insecticides, while others may end up adapting to live amidst the usage or even become immune to them.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Wood litter and debris that fall to the forest ground add nutrients to the soil, and provide cover for other animals that live in the forest. The amount of wood litter and debris may also be indicative of the overall health of the forest and the trees in it.

4. How is litter and woody debris sampled as part of the NEON network? Read the NEON\_Litterfall\_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. Litter and fine woody debris samplign is executed at terrestrial NEON sites that contain woody vegetation over 2m tall. 2. Litter and fine woody debris are collected from elevated and ground traps, respectively. 3. In sites with > 50% aerial cover of woody vegetation >2m in height, placement of litter traps is random.

### Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
#checking dimensions of Neonics dataset
dim(Neonics)
```

## [1] 4623 30

6. Using the summary function on the "Effect" column, determine the most common effects that are studied. Why might these effects specifically be of interest?

# #checking summary of Effect column in dataset summary(Neonics\$Effect)

##	Accumulation	Avoidance	Behavior	Biochemistry
##	12	102	360	11
##	Cell(s)	Development	Enzyme(s)	Feeding behavior
##	9	136	62	255
##	Genetics	Growth	Histology	Hormone(s)
##	82	38	5	1
##	Immunological	Intoxication	Morphology	Mortality
##	16	12	22	1493
##	Physiology	Population	Reproduction	
##	7	1803	197	

Answer: Population and Mortality are the most common effects studied. These might be of interest in order to keep track of how the population of each species changes over time as the insecticide is introduced and throughout its future usage.

7. Using the summary function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.

## #checking most commonly studied species summary(Neonics\$Species.Common.Name)

##	Honey Bee	Parasitic Wasp
##	667	285
##	Buff Tailed Bumblebee	Carniolan Honey Bee
##	183	152
##	Bumble Bee	Italian Honeybee
##	140	113
##	Japanese Beetle	Asian Lady Beetle
##	94	76
##	Euonymus Scale	Wireworm
##	75	69
##	European Dark Bee	Minute Pirate Bug
##	66	62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
## ##	57	51
##	Erythrina Gall Wasp 49	Beetle Order 47
##	<del></del> -	<del>-</del> '
	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
## ##	<del>-</del> '	Proff to iled Promblehee
##	True Bug Order 45	Buff-tailed Bumblebee 39
##	45 Aphid Family	Cabbage Looper
##	Aprilu Family	Cabbage Looper
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23

##	Ladybird Beetle	Mason Bee
##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	Tooth marked Fungua Bootle
##	Potato Leafhopper 20	Tooth-necked Fungus Beetle 20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order
##	17	17
##	Egg Parasitoid	Insect Class
##	17	17
##	Moth And Butterfly Order	Oystershell Scale Parasitoid
##	17 Hemlock Woolly Adelgid Lady Beetle	17 Hemlock Wooly Adelgid
##	16	nemiock wooly kdeigid
##	Mite	Onion Thrip
##	16	16
##	Western Flower Thrips	Corn Earworm
##	15	14
##	Green Peach Aphid	House Fly
##	14	14
##	Ox Beetle	Red Scale Parasite
##	14	14
##	Spined Soldier Bug	Armoured Scale Family
##	14	13
##	Diamondback Moth	Eulophid Wasp
##	13	13
##	Monarch Butterfly	Predatory Bug
## ##	Yallar Farran Magazita	13 Braconid Parasitoid
##	Yellow Fever Mosquito	braconid Parasitoid 12
##	Common Thrip	Eastern Subterranean Termite
##	12	12
##	Jassid	Mite Order
##	12	12
##	Pea Aphid	Pond Wolf Spider
##	12	12
##	Spotless Ladybird Beetle	Glasshouse Potato Wasp
##	11	10
##	Lacewing	Southern House Mosquito
##	10	10

```
## Two Spotted Lady Beetle Ant Family
## 10 9
## Apple Maggot (Other)
## 9
```

Answer: The six most common species studied are the Honey Bee, Parasitic Wasp, Buff Tailed Bumblebee, Carniolan Honey Bee, Bumbe Bee, and Italian Honeybee. These are all pollinator insects, with 5 of them being bee species.

8. Concentrations are always a numeric value. What is the class of Conc.1..Author. in the dataset, and why is it not numeric?

```
#checking Conc.1..Author class
class(Neonics$Conc.1..Author.)

## [1] "factor"

## ["factor"]
```

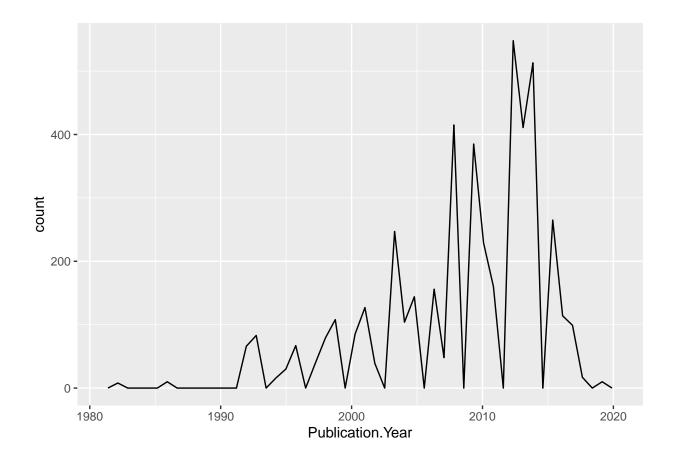
Answer: The class of Conc.1..Author. is a character class because the data is categorical, not numeric.

## Explore your data graphically (Neonics)

9. Using geom\_freqpoly, generate a plot of the number of studies conducted by publication year.

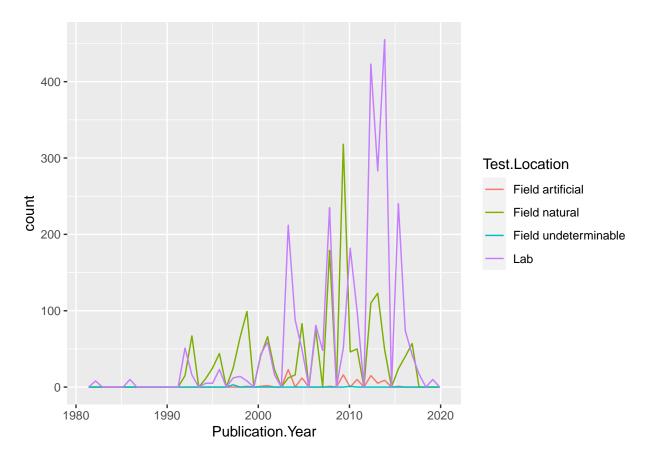
```
#creating line plot by pub year
library(ggplot2)

ggplot(Neonics) +
  geom_freqpoly(aes(x = Publication.Year), bins = 50)
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
#adding color to line plot by pub year
ggplot(Neonics) +
geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins = 50)
```

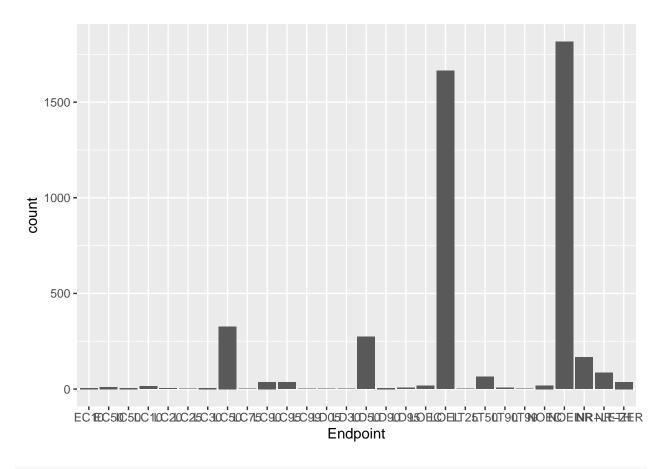


Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are labs, peaking at its highest around 2013/2014. The next most common test location was a natural field around 2007/2008. The most common test location does change over time, and it has since decreased drastically as we reached 2020.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX\_CodeAppendix for more information.

```
#creatign bar graph of endpoint counts
ggplot(Neonics, aes(x = Endpoint)) +
  geom_bar()
```



which.max(table(Neonics\$Endpoint))

## NOEL ## 25

##NOEL ##25

Answer: NOEL and 25 were the two most common endpoints. They are defined by

## Explore your data (Litter)

12. Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the unique function, determine which dates litter was sampled in August 2018.

class(Litter\$collectDate) #checking class of collectDate

## [1] "factor"

## [1] "factor" - is not recognized as a date

Litter\$collectDate <- as.Date(Litter\$collectDate) #changing it to date class class(Litter\$collectDate) #confirming new class of collectDate

```
## [1] "Date" - now recognized as a date
unique(Litter$collectDate) #checking whihc dates litter was sampled in Aug 2018
## [1] "2018-08-02" "2018-08-30"
## [1] "2018-08-02" "2018-08-30"
 13. Using the unique function, determine how many plots were sampled at Niwot Ridge. How is the
    information obtained from unique different from that obtained from summary?
unique(Litter$plotID) #checking how many plots sampled at Niwot Ridge with 'unique'
   [1] NIWO 061 NIWO 064 NIWO 067 NIWO 040 NIWO 041 NIWO 063 NIWO 047 NIWO 051
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
## 1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051 NIWO_058
## [10] NIWO_046 NIWO_062 NIWO_057
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 ... NIWO_067
summary(Litter$plotID) #comparing it against 'summary' results
## NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 NIWO_061
         20
                  19
                           18
                                     15
                                              14
                                                                16
                                                                          17
## NIWO_062 NIWO_063 NIWO_064 NIWO_067
         14
                  14
                           16
## [1] NIWO 061 NIWO 064 NIWO 067 NIWO 040 NIWO 041 NIWO 063 NIWO 047 NIWO 051 NIWO 058
```

## [1] "Date"

## [10] NIWO\_046 NIWO\_062 NIWO\_057

19

17

18

15

##

20

## NIWO\_064 NIWO\_067 16

> Answer: Both functions showed that 12 plots were sampled at Niwot Ridge. The summary function shows us how many tests were done at each of the 12 plots, giving us an overview of the whole dataset. The unique function only lays out what the 12 plots sampled are, without the count of how many tests were done at each plot.

## 12 Levels: NIWO\_040 NIWO\_041 NIWO\_046 NIWO\_047 NIWO\_051 NIWO\_057 NIWO\_058 ... NIWO\_067 ## NIWO 040 NIWO 041 NIWO 046 NIWO 047 NIWO 051 NIWO 057 NIWO 058 NIWO 061 NIWO 062 NIWO 063

14

8

16

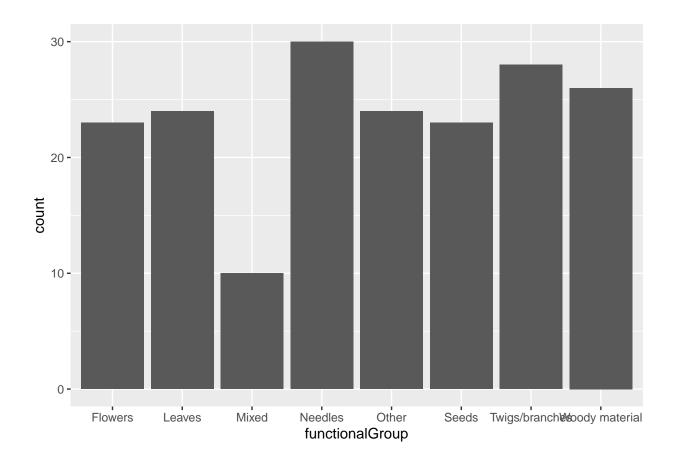
17

14

14

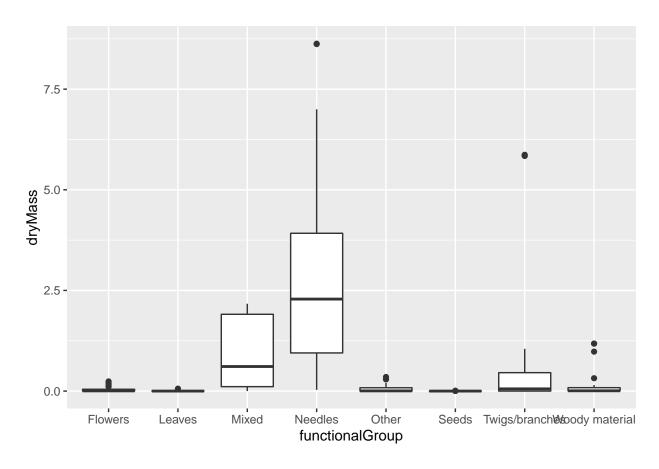
14. Create a bar graph of functional Group counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
#creating bar graph to show functional group counts
ggplot(Litter, aes(x = functionalGroup)) +
  geom_bar()
```



15. Using geom\_boxplot and geom\_violin, create a boxplot and a violin plot of dryMass by functional-Group.

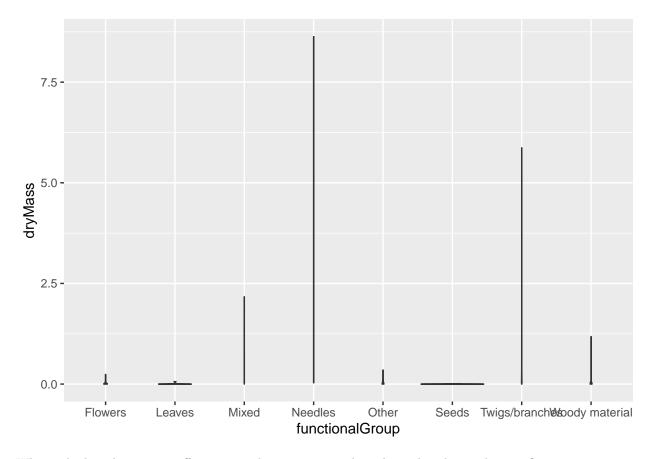
```
#creating boxplot of the relation between dry mass and functional group
ggplot(Litter) +
  geom_boxplot(aes(x = functionalGroup, y = dryMass))
```



```
## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values

## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values

## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: The boxplot is easier to read, and provides a clearer picture of the data. It shows details like outliers and the median of the data. The violin plot only shows the complete range of the data, with no additional details about where all the points fall within that range.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles tend to have the highest biomass at these sites, with a greater range and maximum than other types of litter.