https://doi.org/10.1093/icb/icac098

Society for Integrative and Comparative Biology

### **GENOMICS PAPER**

## Population Genomic Time Series Data of a Natural Population Suggests Adaptive Tracking of Fluctuating Environmental Changes

Markus Pfenninger\*†‡, and Quentin Foucault\*†

\*Department Molecular Ecology, Senckenberg Biodiversity and Climate Research Centre, Senckenberganlage 25, 60325 Frankfurt am Main, Germany; † Institute for Molecular and Organismic Evolution, Johannes Gutenberg University, Johann-Joachim-Becher-Weg 7, 55128 Mainz, Germany; † LOEWE Centre for Translational Biodiversity Genomics, Senckenberg Biodiversity and Climate Research Centre, Senckenberganlage 25, 60325 Frankfurt am Main, Germany

<sup>1</sup>E-mail: Markus.Pfenninger@senckenberg.de

Synopsis Natural populations are constantly exposed to fluctuating environmental changes that negatively affect their fitness in unpredictable ways. While theoretical models show the possibility of counteracting these environmental changes through rapid evolutionary adaptations, there have been few empirical studies demonstrating such adaptive tracking in natural populations.

Here, we analyzed environmental data, fitness-related phenotyping and genomic time-series data sampled over 3 years from a natural *Chironomus riparius* (Diptera, Insecta) population to address this question. We show that the population's environment varied significantly on the time scale of the sampling in many selectively relevant dimensions, independently of each other. Similarly, phenotypic fitness components evolved significantly on the same temporal scale (mean 0.32 Haldanes), likewise independent from each other. The allele frequencies of 367,446 SNPs across the genome showed evidence of positive selection. Using temporal correlation of spatially coherent allele frequency changes revealed 35,574 haplotypes with more than one selected SNP. The mean selection coefficient for these haplotypes was 0.30 (s.d. = 0.68). The frequency changes of these haplotypes clustered in 46 different temporal patterns, indicating concerted, independent evolution of many polygenic traits. Nine of these patterns were strongly correlated with measured environmental variables. Enrichment analysis of affected genes suggested the implication of a wide variety of biological processes.

Thus, our results suggest overall that the natural population of *C. riparius* tracks environmental change through rapid polygenic adaptation in many independent dimensions. This is further evidence that natural selection is pervasive at the genomic level and that evolutionary and ecological time scales may not differ at all, at least in some organisms.

#### Introduction

Most natural environments consist of an almost infinite number of aspects and parameters that are constantly changing in space and time (Kingsolver et al. 2012). Organisms in such habitats therefore, potentially have to deal with a fluctuating selective regime (Bell 2010) where the direction of selection frequently changes (Siepielski et al. 2009). The question is whether and to what extent natural populations rapidly and constantly adapt to the fluctuations in their environment in a process termed adaptive tracking (Vander Wal et al. 2013). The answer is crucial for our understanding of the relative roles of adaptation vs. phenotypic plas-

ticity (King and Hadfield 2019), eco-evolutionary processes (Rudman et al. 2018), the maintenance of genetic variation (Barton and Keightley 2002) and the process of balancing selection in natural populations (Abdul-Rahman et al. 2021).

Theoretical work has shown that rapid adaptation, a prerequisite for adaptive tracking, is possible almost in real time in natural populations (Messer and Petrov 2013; Botero et al. 2015; Matuszewski et al. 2015), in particular if polygenic traits are the target of selection (Jain and Stephan 2017; Barghi et al. 2020). However, despite the importance of the question whether and to what extent selective tracking plays a role in

nature, empirical studies are rare. There are a few examples of selective tracking of the fluctuating environment for phenotypic traits (Grant and Grant 1989; Marrot et al. 2017; de Villemereuil et al. 2020). In particular, the selectively driven beak variability of Galapagos finches in response to different weather conditions in different years, is the classic example of very rapid evolutionary adaptation to a variable environment (Boag and Grant 1981). On the molecular level, there are several demonstrations of rapid selectively driven changes (Yang et al. 2016; Margres et al. 2017; Bitter et al. 2019; Rennison et al. 2019; Zong et al. 2021). Seasonally selected polymorphisms with correlated allele frequency trajectories were also observed in natural populations of a dipteran species, Drosophila melanogaster (Bergland et al. 2014; Croze et al. 2017). However, studies bringing together the observed temporal fitness differences among different phenotypes with the underlying molecular variants are scarce. In a complex seminatural experiment (Rudman et al. 2021), could demonstrate adaptive tracking on both the phenotypic and genomic level. However, it has been stressed that realworld examples are needed to understand how common adaptive tracking is in natural populations (Hendry 2019).

In a recent article, Pfenninger et al. (2022) could show with population genomic time series data and experiments that a natural population of the non-biting midge Chironomus riparius (Meigen, 1803) selectively tracked a short-term weather event. Moreover, long term genomic time series data suggested that the spatially unlinked alleles involved in the investigated polygenic trait under selection changed temporally in a correlated fashion over a longer period (Pfenninger et al. 2022). This raised the question whether there are more spatially unlinked haplotypes in the genome that change over time in correlated fashion, thus indicating independent responses to the respectively same selective forces. Or, in other words, does this species generally adaptively track the environmental changes in its habitat? To demonstrate adaptive tracking, it is necessary to show that fitness-related phenotypic and selection driven genomic changes happen on the same time scale as selectively relevant environmental changes.

Here, we used environmental data, fitness-related phenotyping, and population genomic time-series data of the same natural *C. riparius* population to tackle this question. *Chironomus riparius* is a multivoltine species with up to 15 generations per year in Europe (Oppold et al. 2016). Therefore, the different generations are subjected to widely varying environmental conditions. Accordingly, extensive research on temperature and photoperiod has shown that several traits can

and do adapt locally (Waldvogel et al. 2018), and temporally among seasons (Foucault et al. 2018; Doria et al. 2022a) also on the genomic level. But also other factors are known to act as selection pressures on this species (e.g., organic load, Kraak et al. (2000), conductivity, (Pfenninger and Nowak 2008), nitrogen, (Nemec et al. 2012), temperature, (Nemec et al. 2013), and anthropogenic substances, (Nowak et al. 2009). The high effective and demographic population size (>1,000,000, Waldvogel et al. (2018)) and the very high number of offspring per breeding pair (400-800) allows for rapid adaptation (Pfenninger and Foucault 2020). A range of genomic resources and parameters are available (Oppold and Pfenninger 2017; Schmidt et al. 2020) and established routines for evolutionary experiments in the laboratory exist (Foucault et al. 2019b), which makes the species a suitable object to test the hypothesis that adaptive tracking takes place in natural populations. Here, we addressed the following questions:

- Did known selection pressures change over the periods monitored in the natural population, thus providing a fluctuating selective regime?
- Is there fitness-related phenotypic evolution on the same time scale in the population?
- Can we find evidence for selection-driven allele frequency changes in the genome?
- Are there spatially unlinked selected haplotypes whose allele frequencies evolve in a temporally correlated fashion, indicative of polygenic adaptation?
- If so, how many of such selected trajectory clusters can we identify?
- Are the frequency changes of the haplotypes in these clusters correlated with recorded environmental changes?

## Material and methods

#### Sampling

In February 2016, >500 living red *thummi*-type chironomid larvae were sampled with a sieve at a single site situated in a small river (Hasselbach, Hessen, Germany, 50.167562°N, 9.083542°E) following the protocol of Foucault et al. (2019b). The specimen's specific identity was ascertained by DNA-barcoding of a mitochondrial (COI) and a nuclear locus (L44). Eighty thus identified *C. riparius* were pooled and subjected to PoolSeq sequencing (see below). The sampling was likewise repeated in September 2016, assuming that the two annual samples thus were subjected to the selective extremes of the winter and summer season, respectively. This sampling scheme was repeated for the two following years, 2017 and 2018, resulting in a time

series of six population pools (NP0–NP5). From February to September, the population produced about seven generations, in the period from autumn to early spring about three. The entire sampling period thus covered about 27 generations (Supplementary Fig. S1). We used additional *C. riparius* individuals from the field, sampled at the same time as the individuals for genomic analysis except for September 2018, to perform standardized life cycle fitness tests at three temperatures (14°C, 20°C, and 26°C).

#### **Environmental parameters**

Mean monthly water parameters for the period 2009– 2018 were obtained from the Hasselbach water treatment plant of the Abwasserverband Freigericht. The values were continuously measured automatically in the river past the outflow, less than 50 m from the sampling site. The following parameters were recorded: water temperature [°C], conductivity [mS/cm], oxygen, biological oxygen demand (BOD), chemical oxygen demand (COD), NH<sub>4</sub>, NO<sub>3</sub>, NO<sub>2</sub>, N<sub>tot</sub>, and PO<sub>4</sub>, all (mg/L). To assess the similarity of the entirety of the parameters over time, the monthly means were summarized in a seasonal mean for the period before sampling, that is, from March to September and from October to February. These values were subjected to a Principal Components Analysis (PCA). Temporal autocorrelation was calculated in Past 4 (Hammer et al. 2001).

#### Phenotypic evolution

We used an established life cycle fitness test to assess three fitness component phenotypes of the natural population under standard conditions in a common garden experiment. The experiment was carried out at 14°C, which corresponds to the annual average water temperature at the sampling site. A fraction of the sampled larvae from each sampling date was divided into five replicates (30 individuals each) and brought to hatch and reproduce. To distinguish phenotypic evolution from phenotypic plasticity, the resulting F1 generation was monitored for the parameters developmental rate (measured as emergence mean time 50 EmT50), fecundity (viable eggs per female), mortality (proportion of emerging midges), and a resulting overall fitness parameter (population growth rate per day). The assessed phenotypes are known to have a complex genetic basis (Doria et al. 2022a). We tested with an One-way ANOVA for significant changes in mean trait values, using the time points of sampling as categories. A detailed description of the life cycle fitness test can be found in (Foucault et al. 2019b).

We calculated rates of phenotypic evolution among sampling dates for each trait in Haldanes (*h*) as

$$h = \frac{\frac{(x_1 - x_2)}{s_p}}{g},$$

where  $x_n$  representS the mean trait value at the time point n,  $s_p$  the pooled standard deviation, and g the number of generations between the two time points (Haldane 1949).

#### Population genomic analyses of pooled samples

DNA was extracted for each pool from the field or the experiment using a blood and tissue extraction kit on pooled samples of 80 larval head capsules, respectively. Integrity and quality of extracted DNA was controlled using electrophoresis, and the DNA concentration for each samples measured with a fluorimeter.

Whole genome pool-sequencing was carried out with 250bp paired end reads. Reads were trimmed using the wrapper tool autotrim (Waldvogel et al. 2018) that integrates trimmomatic (Bolger et al. 2014) for trimming and fastQC (Andrews 2010) for quality control. The trimmed reads were then mapped on the latest C. riparius reference genome (Schmidt et al. 2020) using the BWA mem algorithm (Li and Durbin 2009). Low quality reads were subsequently filtered and SNPs were initially called using samtools(Li et al. 2009). The pipelines Popoolation and Popoolation2 (Kofler et al. 2011a, 2011b) were used to call SNPs, remove indels, and to estimate genetic diversity as Watterson's theta each pool. Allele frequencies for all SNPs with coverage between  $15 \times$  and  $70 \times$  were estimated with the R library PoolSeq (Taus et al. 2017).

## Identification of allele frequency trajectories with signs of selection

Selected SNP loci were identified by allele-frequency changes (AFC) larger than expected by neutral drift. First, Fisher's exact tests for the AFCs among all consecutive time points (NP0 > NP1, etc.) and among the first and last sampling were calculated. This test identifies AFC larger than expected by chance, but does not account for genetic drift expected. Therefore, neutral simulations were used to compute false discovery rate *q*-values < 0.001 with parameters (number of SNPs, starting allele frequencies in the ancestral population, sequence coverage, and number of generations) matching those of the respective sample. As effective population size, we used 15,000, which constitutes a very conservative estimate (see Waldvogel et al. 2018). All calculations and simulations were performed with the R-library poolSeq (Taus et al. 2017). A SNP locus was

considered showing signs of selection if at least one sampling interval or between the first and the last sample period had an AFC significantly larger than expected. The allele frequency trajectory, that is, the vector of allele frequencies of such putatively selected SNP site over time was recorded.

## Inference of genome-wide level of linkage disequilibrium

The expected length of segregating haplotypes depends on the recombination rate and their age. The former can be approximated by an estimate of linkage disequilibrium LD (Feder et al. 2012). We estimated the extent of LD and its decay using the maximum likelihood method of (Feder et al. 2012) on one of the pools (NP0). This method exploits the haplotype information contained in (paired) reads and has been shown to yield accurate estimates if mean LD decay to background levels is shorter than the read length (Feder et al. 2012).

## Clustering of spatially coherent SNPs into haplotypes

Most SNPs showing larger than expected AFC are hitchhiking with the actually selected site in selected haplotypes. The extent of such haplotypes was determined following the rationale of Franssen et al. (2016) that physically linked variants following the same evolutionary trajectory should show correlated allele frequencies through time. However, contrary to an experimental evolution situation with a constant selection pressure, it could not be expected that the observed AFC were always in the same direction. Since this is a prerequisite for the software used in Franssen et al. (2016), we had to develop our own algorithm. Our algorithm considered consecutively all temporal trajectories of significant SNPs along a scaffold. A SNP was added to a haplotype if the absolute correlation coefficient of its temporal trajectory to any other trajectory of the SNPs already in the haplotype equalled or exceeded a given threshold (0.75). This was repeated until the next SNP position did not meet the criterion. The number of SNPs in the haplotype, its start and end positions, and the mean allele frequency were then recorded and a new haplotype was started at the next position considered. The analysis was carried out with a custom Python script.

#### Validation of haplotypes

Inferred haplotypes were validated with 30 whole genome resequenced individuals ( $25 \times$  coverage, 150 bp paired end reads) from all sampling periods. The reads from these individuals were quality checked, mapped against the reference genome as described above and

the genotypes called with the GATK pipeline (Van der Auwera et al. 2013). Consensus sequences of 100 randomly chosen haplotypes with more than two SNPs, including one neighboring haplotype were extracted from the individual VCF-files, aligned and phased with FastPhase in DNAsp (Rozas et al. 2017). The resulting phased alignments were checked for linkage of the previously identified target haplotype's typical alleles and linkage equilibrium to the neighboring haplotypes in the same software.

## Calculation of selection coefficients for haplotypes

The selection coefficient s was calculated from the change in mean allele frequencies between samplings for all selected haplotypes by the formula given in (Kimura 1970). Assuming codominance (i.e., h = 0.5), the selection coefficient s can be calculated as

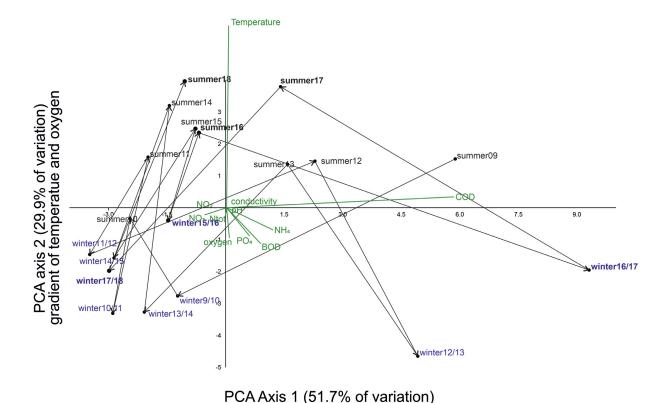
$$s = 2 * \left( \ln \left( \frac{p_t}{1 - p_t} \right) - \ln \left( \frac{p_0}{1 - p_0} \right) \right) * t^{-1},$$

where  $p_0$  is the allele frequency of the rising allele in the ancestral population and  $p_t$  its frequency in generation t.

# Hierarchical clustering of mean haplotype trajectories

For each haplotype, we calculated the mean allele frequency trajectory (MHT) over the allele frequencies of all constituting SNPs. The matrix of absolute pairwise correlation coefficients among all MHT was transformed into a distance matrix (1–correlation coefficient), hierarchically clustered with UPGMA and the resulting dendrogram plotted. The number of clusters considered was determined by applying a clustering threshold of 0.25, corresponding to a minimum similarity of 0.75 of all MHT within a cluster.

To determine this threshold, sensibly balancing between arbitrary lumping of actually different trajectories and over-splitting of trajectories differing rather by stochastic fluctuations, we applied a randomization approach. From the empirical AFC distribution of all SNPs in the analyzed set (i.e., at least one significant AFC among consecutive sampling periods or between the first and last sampling period, see above), we randomly drew 1000 times the empirically found number of MHTs. Starting with a distance threshold of 0.05, we iteratively increased the threshold by 0.05. In each iteration, the trajectories of the 1000 random MHTs were hierarchically clustered as described above and the number of resulting clusters recorded. The simulated distribution was then compared to the empirically observed number of clusters, applying the same thresh-



**Fig. 1** PCA on seasonal physico-chemical water parameters in the natural population for the decade from 2009–2018. Shown are the first two axes, accounting for 82.2% of environmental variation. Periods after which phenotypic and genomic assessments of the population were performed are indicated in bold and with larger dots. Winter and summer samples are shown in different colours. The temporal trajectory of the environmental conditions are indicated by arrows connecting consecutive periods.

gradient of organic water pollution

old. The smallest distance threshold for which the observed number of clusters was smaller than the 0.05 quantile of the randomly simulated distribution was deemed appropriate for the analysis. This was the case for a distance threshold of 0.25; that is, the trajectories within a cluster showed a correlation of at least 0.75. The analysis was performed with a custom Python script.

## Correlation of environmental parameters with haplotype trajectories

The seasonal mean environmental parameters (see above) before the first and the respective periods between two sampling times were correlated to the means of all identified MHT patterns. To avoid problems with multiple testing and lacking statistical power (Sjölander and Vansteelandt 2019), we employed a Bayesian implementation of a test on correlation with default values (Bååth 2014). We report only correlations with convincing posterior evidence (>95% posterior credibility) for positive, respectively negative associations.

#### **GO-term enrichment analysis**

The annotated gene overlapping with a haplotype was identified as a putative selection target. The resulting gene lists for all member haplotypes of each MHT pattern were tested for over-representation of GO terms in the category "biological function" in the R-library TopGO with a Fisher's exact test (Alexa and Rahnenführer 2016).

#### **Results**

#### Fluctuating enviroment

Measured physico-chemical water parameters varied substantially through time (Fig. 1, Supplementary Fig. S2). Comparison with a broken stick model suggested that the first two Principal Component axes were meaningful. Axis 1 (51.8% of variation) of the PCA was mainly determined by parameters describing organic water pollution (chemical oxygen demand, NH<sub>4</sub>, biological oxygen demand, and conductivity), axis 2 (21.7%) reflected mostly seasonal changes (temperature, oxygen, and PO<sub>4</sub>, Fig. 1). The measured environmental regime never returned close to any previous

state. This was mirrored in the poor temporal autocorrelation of the parameters, except for the regular seasonal temperature changes (Supplementary Fig. S3).

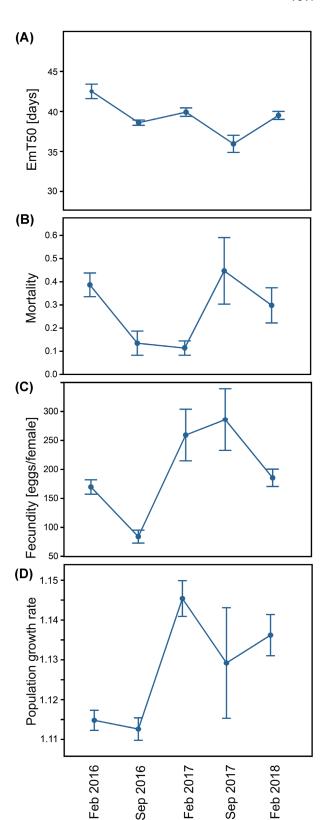
#### Strong phenotypic evolution

Exposed to the same standard conditions in a common garden setting, the F1 generation derived from the natural population at different points in time showed signs of strong and rapid phenotypic evolution, with idiosyncratic patterns across traits. The EmT50 value varied strongly and significantly with the seasons (F =11.53, d.f. = 4,  $P = 5 \times 10^{-5}$ ) between 35.9 and 42.5 days, with an overlaid overall decreasing trend (Fig. 2A). Mean evolutionary rate for the trait between sampling dates was 0.29 Haldanes (min = 0.07, max = 0.42) with a reversal of direction between each sampling date (Fig. 2A). Mortality varied also significantly, albeit marginally among sampling dates (F = 3.25, d.f. = 4, P = 0.03). Between 11 and 45% of the midges did not hatch in the laboratory environment (Fig. 2B). The rate of evolution was 0.22 (min = 0.07, max = 0.31) with two changes of direction (Fig. 1B). The strongest phenotypic evolution was observed for fecundity (Fig. 2C). The mean fecundity varied significantly between 84 and 286 offspring per female (F = 6.07, d.f. = 4, P = 0.002). The evolutionary rate of change for the trait was on average 0.45 (min = 0.20, max = 0.96) standard deviations per generation. The direction of the evolution changed twice during the period of observation (Fig. 2C). The resulting population fitness remained always positive (range = 1.112-1.145), but varied significantly among sampling dates (F = 3.84, d.f. = 4, P = 0.02, Fig. 1D).

# Large parts of the genome showed evidence of selective evolution

The genome-wide mean estimate of distance-dependent linkage disequilibrium (LD) dropped to a mean  $r^2$  of 0.5 within 10 base pairs and to the genome-wide background level of about 0.2 within 120 bp (Supplementary Fig. S5). Consequently, the expected haplotype structure was extremely short.

Allele frequency estimates for all sampling dates were obtained for 22,693,348 SNP positions in the genome. Neutral drift simulations, taking observed sampling variance of the poolSeq approach into account, showed that the chosen selection detection threshold of 2.0 -  $\log_{10}$ p for an allele frequency change (AFC) between consecutive sampling times or between the first and the last sampling implied a false discovery rate of q < 0.001. This corresponded to an AFC of at least 0.15. Under the condition of at least one selected AFC between any two consecutive sampling times or the first and the last sam-



**Fig. 2** Development of phenotypic traits and population fitness over time in the natural population. (**A**) Developmental time measured as EmT50. (**B**) Mortality measured as proportion of emerged individuals. (**C**) Fecundity as viable eggs per female. (**D**) Population growth rate per day as integrative measure of population fitness.

pling time, identified a set of 367,446 SNPs with signs of selection.

Clustering spatially coherent SNPs with signs of selection into haplotypes resulted in 281,153 haplotypes of which 35,574 consisted of more than a single SNP. Of the 100 haplotypes randomly chosen for validation, 98 showed the expected linkage between the marker SNPs of the haplotype (Supplementary Fig. S6). In two cases, both with more than two haplotype marker SNPs, one of the marginal SNPs was only slightly linked (r < 0.6). All haplotypes checked were distinct from their respective spatial neighbors. Only the 35,574 haplotypes with more than a single SNP were retained for subsequent analyses. In these, the average number of SNPs per haplotype was 3.30 +/- 2.64 (mean +/- s.d.). The mean length of inferred haplotypes was 893.1 +/- 1790.3 bp. Neighboring haplotypes on the same scaffold were on average 3466.0 + / -4293.5 bp apart. Of all 13,490 annotated genes in the genome, 5793 (= 42.9%) harbored at least one selected haplotype, some more.

The mean selection coefficient over all retained haplotypes was 0.30 with a standard deviation of 0.68. The latter is equivalent to the intensity of selection over time, and its much larger value relative to the mean indicated large changes in magnitude and frequent reversals in the direction of selection (Bell 2010).

#### Clustering of haplotype trajectories

To avoid using an arbitrary clustering threshold, we used a Monte-Carlo simulation approach. Clustering of simulated random MHTs, drawn from the observed AFC distribution, indicated that a clustering threshold of 0.75 yielded informative clusters without undue lumping (Supplementary Fig. S7). Applying this threshold, the 35,574 MHTs were grouped into 46 distinct cluster (Fig. 3). The number of MHT cluster members was highly different. The largest cluster (MHT16) contained 17,914 haplotype trajectories, the smallest merely three (MHT28), the mean was 816.8 (Supplementary Table S1).

# Correlation of haplotype trajectories to environmental parameter changes and GO analysis

Nine MHT cluster patterns were highly correlated to measured water parameters in the period before sampling. Three MHTs were correlated to changes in NO<sub>3</sub>. One MHTs followed variation on Biological Oxygen Demand (BOD) closely. Two other MHTs were associated with NO<sub>2</sub>, MHT3 was correlated to PO<sub>4</sub>, MHT 38 to temperature, while the strongest association was found for MHT 34 and oxygen (Fig. 4). For 44 of the 46 MHT clusters, at least one significantly enriched GO

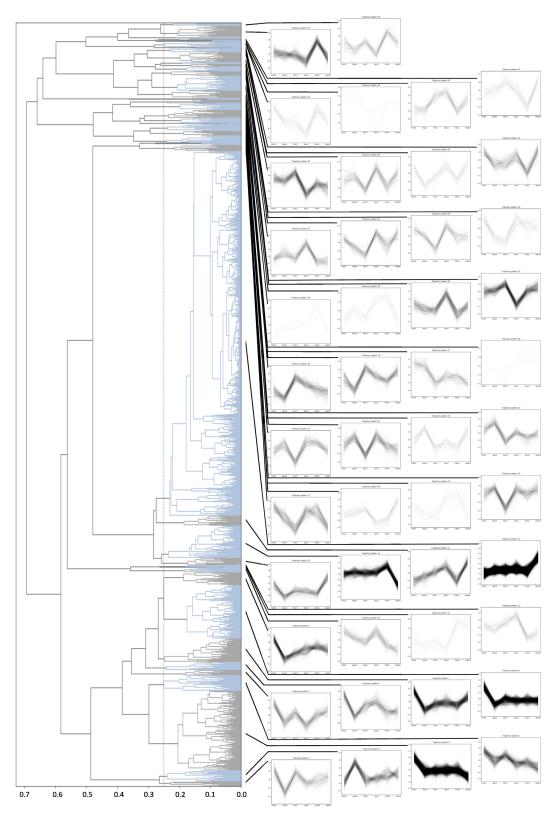
term was found (Supplementary Table S2). Most enriched terms for the individual MHT clusters were on a very high hierarchical level. Consequently, they were rather uninformative or affected only a small minority of the genes with overlap to a selected haplotype. However, some hinted at a specific biological process or supported the observed association of the MHT cluster to a measured physico-chemical variable.

The enriched terms for MHT4 (GO terms 0051704 multiorganism process, 0006952 defense response) suggested reaction to a biological interaction. The largest cluster MHT16 contained hundreds of genes involved in osmoregulation, reflected in the enrichment for GO terms 0055085 transmembrane transport and 0006811 ion transport. Despite affecting only about 20 genes, MHT36 showed an enrichment of terms associated with porphyrin-containing compound biosynthetic process (0006779) and pigment biosynthetic process (046148). Genes affected by MHT40 haplotypes were enriched for signal transduction processes (0007186 G proteincoupled receptor signaling pathway), perhaps in conjunction with hormones (1901362 organic cyclic compound biosynthetic process, 0008202 steroid metabolic process). MHT3 was enriched for term (0006811 ion transport). MHT39, with calcium (transmembrane) transport (terms 0006813 and 0071805), which is co-annotated to nitric-oxide synthase inhibitor activity. There was an enrichment of the term 0005975 carbohydrate metabolic process in MHT21. MHT22 was enriched for 0007688 sensory perception of smell and 0006979 response to oxidative stress.

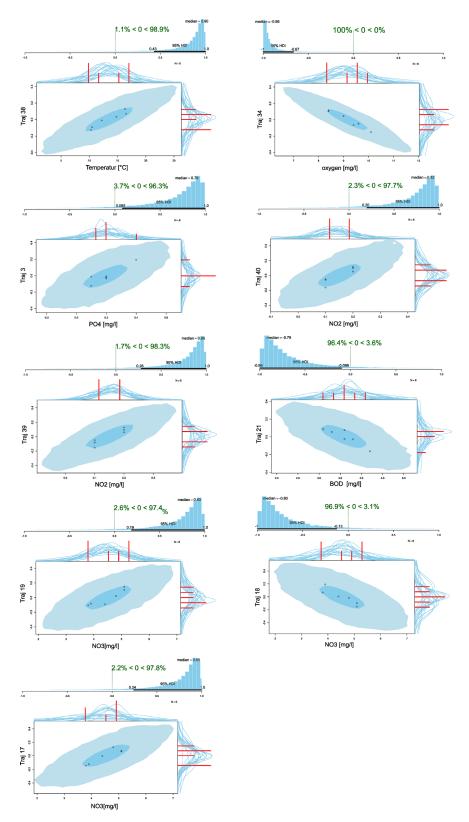
#### **Discussion**

## Complex and unpredictable multidimensional fluctuating selective regime

The largely uncorrelated temporal changes as well as the observed range of the measured physico-chemical water parameters suggested that the local environment presented a highly fluctuating, multidimensional selective regime for *C. riparius*. For most of these parameters, the fitness relevance and thus their selective potential for the species has already been shown (Ristola et al. 1999; Kraak et al. 2000; Pfenninger and Nowak 2008; Nemec et al. 2012; 2013). Even though some parameters showed seasonal changes, these were either overlain by long-term trends or were minor in comparison to the observed changes that did not seem to follow any regular pattern. In consequence, the selective regime as a whole defined by this physico-chemical parameter set was at no point in time identical or even similar to any other period surveyed. Moreover, only a small subset of the potentially selectively relevant biotic and abiotic parameters was measured. For example,



**Fig. 3** Temporal dynamics and complexity of allele frequencies trajectories with signs of selection. The left side of the panel shows a UPGMA-dendrogram on the pairwise distance (I—correlation coefficients) of all 35,574 haplotype MHTs with at least one significant AFC. The hatched vertical line presents the inferred clustering threshold of 0.25. The right side of the panel shows the MHTs for each cluster. For better visibility and comparability, the individual MHTs in each cluster were polarized into the same direction and normalized by substracting the respective mean.



**Fig. 4** Correlations between measured environmental parameters and MHTs. Shown are only associations with a posterior credibility > 95%. Please note that the direction of the correlation depends on the arbitrary polarization of the alleles in the allele frequency trajectory.

predation (Correia et al. 2005) and other intra- and interspecific ecological interactions that are known to play an important role for the fitness of *C. riparius* (van de Bund and Davids 1993; Hooper et al. 2003) were not taken into account. It is therefore reasonable to assume that an unknown, but a much larger number of selective factors are simultaneously, yet in strength and direction more or less independently, acting on this population. The local habitat is therefore probably conforming to the multi-scale theory of the environment (Bell 2010).

## Continuous phenotypic evolution with frequent changes in direction

The measured phenotypes of the natural population varied significantly among sampling dates and inversed the direction of change repeatedly and independently across traits. To distinguish between phenotypic plasticity and adaptation, we measured the traits in the F1 generation under standard laboratory conditions, which may be viewed as too few to exclude imprinting effects (Glastad et al. 2019). The laboratory conditions, however, are themselves not selectively neutral for field populations (Pfenninger and Foucault 2020). Given the propensity for rapid adaptation in the laboratory, particularly in the first few generations when confronted with a new stressor (e.g., Doria et al. 2022a, 2022b), it was necessary to balance between these two pitfalls.

We observed strong evolutionary responses, with per generation rates that were in range of those observed for insects in laboratory experiments (Hoffmann and Ross 2018) or in the semi-natural setting of Rudman et al. (2021). Because there were frequent changes in the direction of change, the chosen test interval was crucial; as in the latter study, the choice of another time scale could have suggested no phenotypic change at all

Most importantly for the diagnosis of adaptive tracking was that phenotypic evolution occurred on the same temporal scale as the environmental change. However, we did not test for a direct link between any environmental parameter and a phenotypic trait, because (i) the measured traits are rather highly inclusive fitness components, thus integrating the influence of many lower level traits, and (ii) transient, short term extreme events were not recorded by the long-term averages, but are known to exert important selective pressure (Pfenninger et al. 2022). All measured traits have strong links to fitness, which also varied over the monitored period. This concurred to selective dynamic patterns generally known from natural populations (Kingsolver et al. 2012).

## Rapid and pervasive selective genomic evolution

The analysis of population genomic time series showed that tens of thousands loci showed signs of at least one selection-driven change in allele frequency across sampling periods, corresponding to <30 generations. Rapid and substantial changes in allele frequencies were observed throughout the genome, with many changes of the direction of selection (Fig. 3). As the chosen approach accounted for drift and sampling variance (Taus et al. 2017), and immigration was unlikely to change allele frequencies noticeably (Foucault et al. 2019a), the observed significant changes are likely due to selection alone. This supports the increasing observations of rapid selective change in natural populations (e.g., Torda et al. 2017; Dayan et al. 2019). Even though highly likely, the observed signatures of molecular selection are, however, no definite proof of adaptive evolution. The observation that the standard deviation of the selection coefficients, an equivalent to selection intensity, was much larger than the respective mean (0.68 vs. 0.30) proved substantial changes in magnitude and frequent reversals in the direction of selection. The acting selection regime, thus corresponded to the multi-scale model of fluctuating selection advocated by Bell (2010).

The propensity to rapid adaptation with simultaneous preservation of genetic variation in C. riparius is thus probably due to an interplay of genomic, life cycle, and demographic traits of the species. On the genomic level, the low observed mean LD resulted in a highly fragmented haplotype structure, comparable in scale to other insects like *Drosophila* (Feder et al. 2012). A short range haplotype structure is a precondition for the observed short distance between haplotypes responding to selection. Such a short range LD structure is due to the recombination rate, which in turn has been shown to evolve under the influence of fluctuating seasonal selection (Aggarwal et al. 2021). Since only haplotypes with more than a single SNP were considered, this may even present an underestimate, because ancient selected polymorphisms may very well be completely separated from their background by recombination (Pfenninger et al. 2022). In some instances, even different parts of the same gene followed different evolutionary trajectories. The observed fine-scaled selective response to independently fluctuating environmental parameters requires that many different recombinational variants are permanently present in the population. This may be one reason for the gross overproduction of offspring in the species (Foucault et al. 2019b). A large offspring number implies a respectively large number of independent recombinational events, creating a wide variety of new genotypes from which to select. Together with the large, fluctuating demographic and effective population size, the preconditions for rapid, fine-scaled and variable evolutionary reaction to fluctuating changes in the selective regime are therefore given.

## The number of temporal evolutionary trajectories is limited but dynamic

In comparison to the huge number of potentially independent loci in terms of physical linkage reacting to selection, the number of statistically distinguishable evolutionary trajectories was quite limited (46). This was not necessarily expected, since in polygenic traits, not all contributing loci need necessarily follow the temporal trajectory of the selective pressure closely (unpublished results). Despite expected trade-offs and pleiotropy between loci (Li et al. 2019), this suggested nevertheless that rather the number of detectable selection pressures was limited than the capacity of the population to follow them. That the majority of genes was not measurably affected by selection suggested unused adaptive potential, but could also be due to small effect size loci that did not reach statistical significance in the present analysis.

While the order of magnitude of estimated effectively acting selection pressures is certainly correct, the exact value may not be. On the one hand, the statistically derived clustering threshold was possibly too rigid and therefore distinguished too many clusters, as indicated by the correlation of some very similar MHT clusters (MHT21, MHT22 and MHT17, MHT18, MHT19) to the same environmental variables (Fig. 4). On the other hand, in longer time series other selection pressures could have elicited additional statistically detectable MHT or split co-varying but distinctive selection pressures into distinguishable haplotype trajectories, thus increasing the estimate of acting selective agents. A hint in this direction was the increase from 36 distinguishable MHTs in a preliminary analysis after five temporal samplings (data not shown) to 46 after six. It was therefore possible that the number of selection pressures the population can effectively follow independently within few generations may be finally limited, but could be different than estimated here. However, general conclusions will require testing the framework presented here with simulation studies and application to more populations over more generations and different species.

Changes in intensity and direction of MHTs were observed frequently, thus corroborating the results gained from phenotypic traits. The number of haplotypes that showed a continuous increase or decline in allele frequency over the monitored period were relatively limited (MHT4, 724 loci), suggesting that adaptation pro-

ceeds mainly by short-time-scale tracking of varying selection pressures and not by slow fixation of alleles.

As expected from studies on *Drosophila* (Bergland et al. 2014), MHT patterns reflecting seasonal variation were identified. MHT38 highly correlated to temperature and MHT40 showed respective patterns, accounting for about 1.2% of all selected haplotypes. This is in the range found for seasonal fluctuations of allele frequencies in *Drosophila* (Bergland et al. 2014; Machado et al. 2021). However, compared to the high variability of the remaining MHT patterns, the seasonal variations in temperature, precipitation, resource availability etc. were surprisingly of relatively minor importance.

## Environmental associations and functional gene enrichments of MHTs

The number of MHTs in each cluster was highly different, ranging from a few to many thousands. On average, many independent loci were therefore involved in the same adaptive response, which is typical for polygenic traits (Jain and Stephan 2017). However, enrichment analyses in the GO category Biological Process for genes overlapping with haplotypes from respective MHT clusters rarely yielded clearly interpretable patterns. In some instances, hypotheses on the acting selection pressures could be gained from the enrichment analyses. In particular, MHT4 suggested a biological interaction, perhaps in defence to parasites. Immune genes adaptively tracking the abundance of potential pathogens over seasons were also found in wild D. melanogaster populations (Behrman et al. 2018). Another interesting set of enriched genes associated with porphyrin-containing pigment metabolism was found in MHT36. The most important pigments in Chironomids are hemoglobins, giving the characteristic red colour to these midges (also called "bloodworms"). They play an important role in hypoxia/anoxia (Grazioli et al. 2016) and are negatively influenced in their function by nitrite (Ha and Choi 2008). It was thus not surprising that the highest correlations of this MHT were to oxygen (r = 0.74) and  $NO_2$  (r = 0.78), respectively. Less than a quarter of the MHT patterns were highly (>0.9) correlated with the few measured environmental parameters, indicating that only a small proportion of selectively relevant parameters were measured. Some correlations suggestively matched the inferred functions from the GO enrichment analysis. MHT39, enriched for genes involved in *nitric-oxide synthase inhibitor activity*, was highly correlated to NO2, underlining the importance of nitrogen metabolism for the species (Pfenninger and Nowak 2008). Another match was found for MHT21, strongly associated with biological oxygen demand, which reflects the availability of organic matter in the stream,

an important food source for the midges (Ristola et al. 1999). Among the GO terms enriched in this MHT, *carbohydrate metabolism* was found, insinuating an adaptive reaction to the food availability.

#### Adaptive tracking of environmental changes

The observed substantial rapid phenotypic and selectively driven genomic evolution with changing signs on the same time scale as putatively selectively relevant environmental variation strongly suggested that the natural *C. riparius* population studied here adaptively tracked their environment. This implies the existence of abundant, unlinked genetic variation under fluctuating selection. Indication that adaptive tracking is indeed beneficial for the species comes from the observation that genetically variable populations are much less impaired in population fitness and thus demography compared to artificially impoverished populations when exposed to stressors in multigeneration experiments (Vogt et al. 2007; Oetken et al. 2009).

In classical population genetic predictions, adaptive tracking as observed here is viewed as unlikely to evolve, because fluctuating selection should rapidly eliminate genetic variation (Hedrick 2006). Several recent models; however, have called these predictions into question. It was shown that in a multivoltine species, adaptive tracking is more likely to evolve than phenotypic plasticity or bet-hedging (Botero et al. 2015). Dominance shifts of adaptive alleles (Wittmann et al. 2017) and boom-bust cycles (Bertram and Masel 2019) can additionally contribute to the maintenance of genetic variation and are probably common in species satisfying these conditions (Kain et al. 2015).

Species following such an evolutionary regime should be able to cope also with long term trends as imposed by global change. The next logical step will be to increase the temporal sampling and analyse the spatial dimension. This promises to obtain statistically reliable associations whose functional causalities may then be experimentally validated (but see Pfenninger and Foucault 2020).

#### Conclusion

The present study suggested that natural populations can adaptively track the environmental changes in their habitat. It thus confirms findings of a recent experimental study on *Drosophila* (Rudman et al. 2021), indicating the relevance of the process for real world populations. Pending further support from additional taxa, these studies provide support for the idea that evolutionary and ecological time scales may not differ at all (Hairston Jr et al. 2005). In addition, the rapid and pervasive action of natural selection on the genomic level

challenges long-standing population genetic paradigms (Messer et al. 2016; Kern and Hahn 2018).

Beyond these important scientific questions, the presented framework opens the possibility to use population genomic time series for applied functional environmental monitoring (Pfenninger and Bálint 2022). While currently rather the presence or absence of species is used to evaluate the state of environments, the monitoring of selective changes associated with certain anthropogenic or environmental factors could serve as an early warning system before any community change would be visible. Such knowledge may soon be of crucial importance to direct biodiversity conservation and mitigation strategies in a rapidly changing world.

#### **Author contributions**

MP: conceptualization, methodology development, software programming, validation, formal analysis, investigation, resources, data curation, writing—original DrMHT preparation, writing—review and editing preparation, visualization preparation, supervision, project administration management, and funding acquisition. QF: investigation and writing—review and editing preparation.

#### **Acknowledgments**

We thank Andreas Wieser for help with sampling and the Abwasserverband Freigericht for generously providing the environmental Hasselbach data.

#### **Funding**

The work was supported by the German Research Foundation [grant number PF390/8-1].

#### Supplementary Data

Supplementary Data available at *ICB* online.

#### **Conflicts of interest**

The authors declare the absence of any conflict of interest.

#### **Data availability**

Sequencing data are publicly available on ENA (project ERP115516, samples ERS4040036-ERS4040041). Core scripts and sync-files were made available on DRYAD (doi.org/10.5061/dryad.fxpnvx0nd).

#### References

- Abdul-Rahman F, Tranchina D, Gresham D. 2021. Fluctuating environments maintain genetic diversity through neutral fitness effects and balancing selection. Mol Biol Evol 38:4362–75.
- Aggarwal DD, Rybnikov S, Sapielkin S, Rashkovetsky E, Frenkel Z, Singh M, Michalak P,Korol AB. 2021. Seasonal changes in recombination characteristics in a natural population of Drosophila melanogaster. Heredity 127:278–87.
- Alexa A, Rahnenführer J. 2016. Gene set enrichment analysis with topGO. R package version 2.24. 0. https://bioconductor.org/packages/release/bioc/html/topGO.html, last accessed date: December 2020.
- Bååth R. 2014. Bayesian first aid: a package that implements Bayesian alternatives to the classical\*. test functions in R. Proc of useR 2014:2.
- Barghi N, Hermisson J, Schlötterer C. 2020. Polygenic adaptation: a unifying framework to understand positive selection. Nat Rev Genet 21:769–81.
- Barton NH, Keightley PD. 2002. Understanding quantitative genetic variation. Nat Rev Genet 3:11–21.
- Behrman EL, Howick VM, Kapun M, Staubach F, Bergland AO, Petrov DA, Lazzaro BP, Schmidt PS.2018. Rapid seasonal evolution in innate immunity of wild *Drosophila melanogaster*. Proc R Soc B Biol Sci 285:20172599.
- Bell G. 2010. Fluctuating selection: the perpetual renewal of adaptation in variable environments. Philos Trans Royal Soc B Biol Sci 365:87–97.
- Bergland AO, Behrman EL, O'Brien KR, Schmidt PS, Petrov DA. 2014. Genomic evidence of rapid and stable adaptive oscillations over seasonal time scales in *Drosophila*. PLos Genet 10:e1004775.
- Bertram J, Masel J. 2019. Different mechanisms drive the maintenance of polymorphism at loci subject to strong versus weak fluctuating selection. Evolution 73:883–96.
- Bitter MC, Kapsenberg L, Gattuso J-P, Pfister CA. 2019. Standing genetic variation fuels rapid adaptation to ocean acidification. Nat Commun 10:1–10.
- Boag PT, Grant PR. 1981. Intense natural selection in a population of Darwin's finches (Geospizinae) in the Galapagos. Science 214:82–85.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–20.
- Botero CA, Weissing FJ, Wright J, Rubenstein DR. 2015. Evolutionary tipping points in the capacity to adapt to environmental change. Proc Natl Acad Sci 112:184–89.
- Bund van de WJ, Davids C. 1993. Complex relations between freshwater macro-and meiobenthos: interactions between *Chironomus riparius* and *Chydorus piger*. Freshw Biol 29:1–6.
- Correia AM, Bandeira N, Anastácio PM. 2005. Predator–prey interactions of *Procambarus clarkii* with aquatic macroinvertebrates in single and multiple prey systems. Acta Oecol 28:337–43.
- Croze M, Wollstein A, Božičević V, Živković D, Stephan W, Hutter S. 2017. A genome-wide scan for genes under balancing selection in *Drosophila melanogaster*. BMC Evol Biol 17:1–12.
- Dayan DI, Du X, Baris TZ, Wagner DN, Crawford DL, Oleksiak MF. 2019. Population genomics of rapid evolution in natural

- populations: polygenic selection in response to power station thermal effluents. BMC Evol Biol 19:1–20.
- Doria HB, Caliendo C, Gerber S, Pfenninger M. 2022. Photoperiod is an important seasonal selection factor in *Chironomus riparius* (Diptera: Chironomidae). Biol J Linn Soc 135: 277–90.
- Doria HB, Hannappel P, Pfenninger M. 2022. Whole genome sequencing and RNA-seq evaluation allowed to detect Cd adaptation footprint in *Chironomus riparius*. Sci Total Environ 819:152843.
- Feder AF, Petrov DA, Bergland AO. 2012. LDx: estimation of linkage disequilibrium from high-throughput pooled resequencing data. PLoS One 7:e48588.
- Foucault Q, Wieser A, Heumann-Kiesler C, Diogo J, Cocchiararo B, Nowak C, Waldvogel A-M. 2019. An experimental assessment of reproductive isolation and its consequences for seasonal hybridization dynamics. Biol J Linn Soc 126:327–37.
- Foucault Q, Wieser A, Waldvogel A-M, Feldmeyer B, Pfenninger M. 2018. Rapid adaptation to high temperatures in *Chironomus riparius*. Ecol Evol 8:12780–89.
- Foucault Q, Wieser A, Waldvogel A-M, Pfenninger M. 2019. Establishing laboratory cultures and performing ecological and evolutionary experiments with the emerging model species *Chironomus riparius*. J Appl Entomol 143:584–92.
- Franssen SU, Barton NH, Schlötterer C. 2016. Reconstruction of haplotype-blocks selected during experimental evolution. Mol Biol Evol 34:174–84.
- Glastad KM, Hunt BG, Goodisman MAD. 2019. Epigenetics in insects: genome regulation and the generation of phenotypic diversity. Annu Rev Entomol 64:185–203.
- Grant BR, Grant PR. 1989. Natural selection in a population of Darwin's finches. Am Nat 133:377–93.
- Grazioli V, Rossaro B, Parenti P, Giacchini R, Lencioni V. 2016. Hypoxia and anoxia effects on alcohol dehydrogenase activity and hemoglobin content in *Chironomus riparius* Meigen, 1804. J Limnol 75:347–54.
- Ha M-H, Choi J. 2008. Effects of environmental contaminants on hemoglobin of larvae of aquatic midge, *Chironomus riparius* (Diptera: Chironomidae): A potential biomarker for ecotoxicity monitoring. Chemosphere 71:1928–36.
- Haldane JBS. 1949. Suggestions as to quantitative measurement of rates of evolution. Evolution 3:51–56.
- Hammer Ø, Harper DAT, Ryan PD. 2001. PAST-Palaeontological statistics. Palaeontol Electronica 4:1–9.
- Hedrick PW. 2006. Genetic polymorphism in heterogeneous environments: the age of genomics. Annu Rev Ecol Evol Syst 37:67–93.
- Hendry AP. 2019. A critique for eco-evolutionary dynamics. Funct Ecol 33:84–94.
- Hoffmann AA, Ross PA. 2018. Rates and Patterns of Laboratory Adaptation in (Mostly) Insects. J Econ Entomol 111:501–9.
- Hooper HL, Sibly RM, Hutchinson TH, Maund SJ. 2003. The influence of larval density, food availability and habitat longevity on the life history and population growth rate of the midge *Chironomus riparius*. Oikos 102:515–24.
- Jain K, Stephan W. 2017. Modes of rapid polygenic adaptation. Mol Biol Evol 34:3169–75.
- Jr Hairston, Ellner SP, Geber MA, Yoshida T, Fox JA. 2005. Rapid evolution and the convergence of ecological and evolutionary time. Ecol Lett 8:1114–27.

- Kain JS, Zhang S, Akhund-Zade J, Samuel AD, Klein M, Bivort de BL. 2015. Variability in thermal and phototactic preferences in Drosophila may reflect an adaptive bet-hedging strategy. Evolution 69:3171–85.
- Kern AD, Hahn MW. 2018. The neutral theory in light of natural selection. Mol Biol Evol 35:1366–71.
- Kimura M. Crow JF. 1970. Introduction to population genetics theory. New York: Harper & Row Publishers.
- King JG, Hadfield JD. 2019. The evolution of phenotypic plasticity when environments fluctuate in time and space. Evol Lett 3:15–27.
- Kingsolver JG, Diamond SE, Siepielski AM, Carlson SM. 2012. Synthetic analyses of phenotypic selection in natural populations: lessons, limitations and future directions. Evol Ecol 26:1101–18.
- Kofler R, Orozco-terWengel P, De Maio N, Pandey RV, Nolte V, Futschik A, Kosiol C, Schlotterer C. 2011a. PoPoolation: a toolbox for population genetic analysis of next generation sequencing data from pooled individuals. PLoS One 6:e15925.
- Kofler R, Pandey RV, Schlötterer C. 2011. PoPoolation2: identifying differentiation between populations using sequencing of pooled DNA samples (Pool-Seq). Bioinformatics 27:3435–36.
- Kraak MH, Stuijfzand SC, Admiraal W. 2000. Interaction between organic matter and toxicants in polluted river water: beneficial and inhibiting effects on the midge *Chironomus riparius*. In: Hoffrichter O. LAte 20<sup>th</sup> century research on chironomidae. Aachen: Shaker Verlag. p.507–11,
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler transform. Bioinformatics 25: 1754–60.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N,Marth G,Abecasis G,Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The sequence alignment/map format and SAMtools. Bioinformatics 25:2078–79.
- Li Y, Petrov DA, Sherlock G. 2019. Single nucleotide mapping of trait space reveals Pareto fronts that constrain adaptation. Nat Ecol Evol 3:1539–51.
- Machado HE, Bergland AO, Taylor R, Tilk S, Behrman E, Dyer K, Fabian DK, Flatt T, Gonzalez J, Karasov TL et al.. 2021. Broad geographic sampling reveals the shared basis and environmental correlates of seasonal adaptation in Drosophila. E-life 10:e67577.
- Margres MJ, Wray KP, Hassinger ATB, Ward MJ, McGivern JJ, Moriarty Lemmon E, Lemmon AR, Rokyta DR. 2017. Quantity, not quality: rapid adaptation in a polygenic trait proceeded exclusively through expression differentiation. Mol Biol Evol 34:3099–3110.
- Marrot P, Garant D, Charmantier A. 2017. Multiple extreme climatic events strengthen selection for earlier breeding in a wild passerine. Philos Trans R Soc B Biol Sci 372:20160372.
- Matuszewski S, Hermisson J, Kopp M. 2015. Catch me if you can: adaptation from standing genetic variation to a moving phenotypic optimum. Genetics 200:1255–74.
- Messer PW, Ellner SP, Hairston NG. 2016. Can population genetics adapt to rapid evolution? Trends Genet 32:408–18.
- Messer PW, Petrov DA. 2013. Population genomics of rapid adaptation by soft selective sweeps. Trends Ecol Evol 28: 659–69.
- Nemec S, Heß M, Nowak C, Pfenninger M. 2012. Experimental evidence for niche segregation in a sister species pair of non-biting midges. Hydrobiologia 691:203–12.

- Nemec S, Patel S, Nowak C, Pfenninger M. 2013. Evolutionary determinants of population differences in population growth rate x habitat temperature interactions in *Chironomus riparius*. Oecologia 172:585–94.
- Nowak C, Vogt C, Pfenninger M, Schwenk K, Oehlmann J, Streit B 2009. Rapid genetic erosion in pollutant-exposed experimental chironomid populations. Environ Pollut. 157:881–6.
- Oetken M, Jagodzinski LS, Vogt C, Jochum A, Oehlmann J. 2009. Combined effects of chemical and temperature stress on *Chironomus riparius* populations with differing genetic variability. J Environ Sci Health A 44:955–62.
- Oppold AM, Pedrosa JAM, Bálint M, Diogo JB, Ilkova J, Pestana JLT, Pfenninger M. 2016. Support for the evolutionary speed hypothesis from intraspecific population genetic data in the non-biting midge *Chironomus riparius*. Proc R Soc B Biol Sci 283:20152413.
- Pfenninger M, Bálint M. 2022. On the use of population genomic time series for environmental monitoring. Am J Bot 109:497–
- Pfenninger M, Foucault Q, Waldvogel A-M, Feldmeyer B. 2022. Selective effects of a short transient environmental fluctuation on a natural population (preprint). bioRxiv. https://doi.org/10.1101/2022.02.10.479864
- Pfenninger M, Foucault Q. 2020. Genomic processes underlying rapid adaptation of a natural Chironomus riparius population to unintendedly applied experimental selection pressures. Mol Ecol 29:536–48.
- Pfenninger M, Nowak C. 2008. Reproductive isolation and ecological niche partition among larvae of the morphologically cryptic sister species *Chironomus riparius* and *C. piger*. PLoS One 3:e2157.
- Rennison DJ, Rudman SM, Schluter D. 2019. Genetics of adaptation: Experimental test of a biotic mechanism driving divergence in traits and genes. Evol Lett 3:513–20.
- Ristola T, Pellinen J, Ruokolainen M, Kostamo A, Kukkonen JV. 1999. Effect of sediment type, feeding level, and larval density on growth and development of a midge (*Chironomus riparius*). Environ Toxicol Chem 18:756–64.
- Rozas J, Ferrer-Mata A, Sánchez-DelBarrio JC, Guirao-Rico S, Librado P, Ramos-Onsins SE, Sanchez-Gracia A et al. 2017. DnaSP 6: DNA sequence polymorphism analysis of large data sets. Mol Biol Evol 34:3299–3302.
- Rudman SM, Barbour MA, Csilléry K, Gienapp P, Guillaume F, Hairston NG, Jr, Hendry AP, LaskyJR, Rafajlovic M, Rasanen K et al. 2018. What genomic data can reveal about ecoevolutionary dynamics. Nat Ecol Evol 2:9–15.
- Rudman SM, Greenblum SI, Rajpurohit S, Betancourt NJ, Hanna J, Tilk S, Yokoyama T, Petrov DA, Schmidt P. 2021. . Science. 375, DOI: 10.1126/science.abj7484 .
- Schmidt H, Hellmann SL, Waldvogel A-M, Feldmeyer B, Hankeln T, Pfenninger M. 2020. A high-quality genome assembly from short and long reads for the non-biting midge *Chirono*mus riparius (Diptera). G3 (Bethesda) 10:1151–57.
- Siepielski AM, DiBattista JD, Carlson SM. 2009. It's about time: the temporal dynamics of phenotypic selection in the wild. Ecol Lett 12:1261–76.
- Sjölander A, Vansteelandt S. 2019. Frequentist versus Bayesian approaches to multiple testing. Eur J Epidemiol 34:809–21.
- Taus T, Futschik A, Schlötterer C. 2017. Quantifying selection with pool-seq time series data. Mol Biol Evol 34:3023–34.

- Torda G, Donelson JM, Aranda M, Barshis DJ, Bay L, Berumen ML, Bourne DG, Cantin N, Foret S, Matz M et al. 2017. Rapid adaptive responses to climate change in corals. Nat Clim Chang 7:627–36.
- Van der Auwera GA, Carneiro MO, Hartl C, Poplin R, Del Angel G, Levy-Moonshine A, Jordan T, Shakir K, Roazen D, Thibault J et al. 2013. From FastQ data to high-confidence variant calls: the genome analysis toolkit best practices pipeline. Curr Protoc Bioinformatics 43:11–10.
- Vander Wal E, Garant D, Festa-Bianchet M, Pelletier F. 2013. Evolutionary rescue in vertebrates: evidence, applications and uncertainty. Philos Trans R Soc B Biol Sci 368:20120090.
- Villemereuil de P, Charmantier A, Arlt D, Bize P, Brekke P, Brouwer L, Cockburn A,Gamelon M, Hamel S, Hegelbach J et al. 2020. Fluctuating optimum and temporally variable selection on breeding date in birds and mammals. Proc Natl Acad Sci 117:31969–78.
- Vogt C, Pupp A, Nowak C, Jagodzinski LS, Baumann J, Jost D, Oetken M, Oehlmann J. 2007. Interaction between genetic di-

- versity and temperature stress on life-cycle parameters and genetic variability in midge *Chironomus riparius* populations. Climate Research 33:207–14.
- Waldvogel AM, Wieser A, Schell T, Patel S, Schmidt H, Hankeln T, Feldmeyer B, Pfenninger M. 2018. The genomic footprint of climate adaptation in *Chironomus riparius*. Mol Ecol 27:1439–56.
- Wittmann MJ, Bergland AO, Feldman MW, Schmidt PS, Petrov DA. 2017. Seasonally fluctuating selection can maintain polymorphism at many loci via segregation lift. Proc Natl Acad Sci 114:E9932–E9941.
- Yang JI, Li W-R, Lv F-H, He S-G, Tian S-L, Peng W-F, Sun Y-W, Zhao Y-X, Tu X-L, Zhang M et al. 2016. Whole-genome sequencing of native sheep provides insights into rapid adaptations to extreme environments. Mol Biol Evol 33: 2576–92.
- Zong S-B, Li Y-L, Liu J-X. 2021. Genomic architecture of rapid parallel adaptation to fresh water in a wild fish. Mol Biol Evol 38:1317–29.