

Vegetation structure drives mosquito community composition in UK's largest managed lowland wetland

Daniel Smith^{1,2*†}, Stefanie Schäfer^{1†}, Nick Golding³,
Miles Nunn³, Steven White¹, Amanda Callaghan², Bethan Purse¹

¹*Department, UK Centre for Ecology and Hydrology, MacLean
Building, Wallingford, OX10 8BB, UK.

²School of Biological Sciences, University of Reading, Whiteknights,
Reading, RG6 2AJ, UK.

³Department, Organization, Street, City, 610101, State, Country.

*Corresponding author(s). E-mail(s): dansmi@ceh.ac.uk;

Contributing authors: smsc@ceh.ac.uk;

nick.golding.research@gmail.com; miles.nunn@akaritx.com;

smwhit@ceh.ac.uk; a.callaghan@reading.ac.uk; beth@ceh.ac.uk;

[†]These authors contributed equally to this work.

Abstract

Purpose: The rising burden of mosquito-borne diseases in Europe extends beyond urban areas, encompassing rural and semi-urban regions near managed and natural wetlands. This is evident in outbreaks of Usutu and West Nile viruses. While wetland management policies focus on biodiversity and ecosystem services, few studies explore the impact on mosquito vectors.

Methods: Our research addresses this gap, examining juvenile mosquito and aquatic predator communities in 67 ditch sites within a south England coastal marsh, subjected to different wetland management tiers. Using joint distribution models, we analyse how mosquito communities respond to abiotic and biotic factors influenced by wetland management.

Results: Of the 12 mosquito species identified, *Culiseta annulata* (Usutu virus vector) and *Culex pipiens* (Usutu and West Nile virus vector) constitute 47% of 6825 larval mosquitoes. Abundant predators include Coleoptera (water beetles) adults, Corixidae (water boatmen), and Zygoptera (Damselfly) larvae. Models reveal that "Tier 3" management sites (higher winter water levels, lower

agricultural intensity) associated with shade and less floating vegetation are preferred by specific mosquito species. All mosquito species, except *Anopheles maculipennis s.l.*, are negatively impacted by potential predators. *Culiseta annulata* shows positive associations with shaded and turbid water, contrary to Corixidae preferences.

Conclusion: In conclusion, Tier 3 areas managed for biodiversity, characterised by higher seasonal water levels and reduced livestock grazing intensity, provide favourable habitats for key mosquito species, known vectors of arboviruses like Usutu and West Nile. Our findings emphasise the impact of biodiversity-focused wetland management, altering mosquito breeding site vegetation to enhance vector suitability. Further exploration of these trade-offs is crucial for comprehending the broader implications of wetland management.

Keywords: Mosquito, Wetlands, Management, Disease, Community Ecology

1 Background

The burden and risk of mosquito-borne diseases (MBDs) is increasing across Europe, not only in urban areas, driven by invasive *Aedes* mosquitoes (e.g., Dengue, Chikungunya, Zika, [Medlock et al \(2012\)](#)), but also by native species in more rural or peri-urban areas, at the interfaces between human habitation, agriculture, and natural ecosystems (e.g., West Nile virus, Usutu, Sindbis, [Buckley et al \(2003\)](#); [Cheng et al \(2018\)](#)). These changes in risk are attributed to multiple interacting global drivers including climate change cite:([Brugueras et al, 2020](#)), increased trade and travel ([Benelli et al, 2020](#); [Kilpatrick, 2011](#)), and land use change, including agricultural intensification and urbanisation ([Gottdenker et al, 2014](#); [Meyer Steiger et al, 2016](#); [Townroe and Callaghan, 2014](#)). At local scales, altered human behaviour through uptake of activities in areas with long standing mosquito presence can be driver of MBD risk, through increased potential contact time between people and competent vectors ([Lambin et al, 2010](#)). Man-made habitat modification that leads to shifts in abundance and species composition of mosquito populations can also alter the interaction dynamics between mosquitoes, humans, and animal reservoir hosts, increasing the relative risk of zoonotic disease spillover ([Meyer Steiger et al, 2016](#)).

In parallel, there is an increased policy focus on managing natural ecosystems such as wetlands to maximise the provision of ecosystem services and enhance biodiversity [Acreman et al \(2011\)](#); [Defra \(2023b\)](#); [Gibbs \(2000\)](#). Within the UK, for example, government policies and payment schemes to landowners encourage the creation, restoration, and management of existing wetlands to increase biodiversity and foster local and regional flood resilience programs ([Defra, 2023a,b,c](#)). Water is a requirement for mosquito breeding and so there is an urgent need to understand how policy-driven changes in wetlands impact mosquito communities, as well as their interactions with animal and human hosts, and how this trades off with disease transmission risk ([Dale and Knight, 2008](#); [Martinou et al, 2020](#); [Medlock and Vaux, 2015a](#); [Hanford et al, 2020](#)).

There is growing evidence globally that wetland management for biodiversity can affect mosquito communities (Rey et al, 2012), not only by changing aquatic breeding site characteristics and vegetation but also via impacts on mosquito predators (Griffin and Knight, 2012; Saha et al, 2012), and that this can lead to public health co-benefits or dis-benefits depending on local context. Some studies have found that mosquito density increases after wetland construction and management (Schäfer et al, 2004; Jiannino and Walton, 2004), but if implemented correctly, wetland management schemes that create diverse and permanent wetland habitats can decrease mosquito populations by simultaneously decreasing habitats suitable for larval mosquitoes while increasing those suitable for known mosquito predators (James-Pirri et al, 2009; Rochlin et al, 2012b; Rose, 2001). Dynamically altering wetland water levels during the mosquito breeding seasons, including complete drying of water bodies during peak breeding months can lead to desiccation of mosquito larvae and prove an adequate control method, but these strategies can negatively impact other aquatic flora and fauna of the wetland (Russell, 1999). In Australia, draining and re-filling of urban wetlands to manage an invasive fish species, led to increased abundance of mosquito species compared to undrained urban wetlands (Hanford et al, 2020). In some contexts, integrated management for biodiversity and reduced public health risks and nuisance biting from mosquitoes has been possible. Aptly named Integrated Marsh Management Schemes employed in salt marshes in the USA, combine tidal flow restoration and vegetation management favouring fish and wildlife biodiversity, with management of open water surfaces (Open Marsh Water Management) to enhance habitats for larvivorous fish predators (Rochlin et al, 2012a).

In Europe and the UK, there is a notable dearth of data regarding the influence of wetland management on mosquito communities, encompassing both nuisance biters and potential disease vectors (Hawkes et al, 2020). Existing research is limited in its examination of the potential trade-offs between conservation-oriented management practices aimed at preserving biodiversity and their implications for public, animal, and wildlife health (Martinou et al, 2020). This knowledge gap is increasingly pressing for Europe, particularly considering the reported 3,500 human cases of West Nile fever, resulting in 379 deaths, between 2010 and 2018. Furthermore, the heightened circulation of the Usutu virus, associated with mosquitoes in and around wetlands, adds urgency to the need for a comprehensive understanding of the impacts of wetland management on mosquito communities (Ferraguti et al, 2016). Specifically, in Southern England, where Usutu virus is currently impacting blackbird populations (Folly et al, 2022), and given the proximity to West Nile virus circulation in Germany and the Netherlands (Bakonyi and Haussig, 2020), there is an increased likelihood of further incursions of mosquito-borne pathogens in the short to medium term. Studies conducted in UK fenlands have explored the links between wetland management and mosquito abundance, and revealed that emergent vegetation and sediment build-up can lead to warmer waters and increased densities of Culicine mosquito species, while drainage of water levels can decrease Culicine abundance but create a more suitable habitat for *Anopheles maculipennis s.l.*, a species complex known for its nuisance biting behaviour (Medlock and Vaux, 2015b).

139 Combining empirical mosquito surveys with statistical spatial modelling of abiotic
140 and biotic drivers of mosquito community composition across wetland management
141 gradients may lead to a more detailed understanding of impacts of wetland man-
142 agement on candidate vector species and biting risks. Utilising such an approach in
143 marshes in the east of England (North Kent Marshes), [Golding et al \(2015\)](#) found
144 that ditch shrimp (*Palaemon varians*) and fish predators reduced the prevalence
145 of mosquito larvae, namely of *An. maculipennis s.l.* (a species complex thought to
146 include minor and historical malaria vectors) and *Culex modestus* (a bridge vector for
147 WNV) and suggested that habitat management for these species could both increase
148 biodiversity and reduce mosquito numbers.

149 Species distribution models have been applied at national, sub-national and local
150 scales to study the impacts of wetland changes on individual mosquito vector species,
151 but these ignore important species community interactions. However, community mod-
152 elling approaches such as joint species distribution models ([Golding and Harris, 2015](#);
153 [Ovaskainen et al, 2017](#); [Pichler and Hartig, 2021](#)) might offer great advantages. These
154 models can help identify shared responses to environmental conditions ([Poggiato et al,](#)
155 [2021](#)) and account for potential biotic interactions such as competition and preda-
156 tion. These interactions can strongly influence mosquito population dynamics and
157 persistence ([Beketov and Liess, 2007](#); [Braks et al, 2004](#); [Saha et al, 2012](#)) and will
158 likely modulate individual vector species responses to wetland changes ([Rey et al,](#)
159 [2012](#)). This study applies community joint modelling methods to sampled larval and
160 adult mosquito population data in a UK wetland that has been subject to manage-
161 ment changes under agri-environmental schemes, where water levels, livestock grazing
162 pressure and mechanical interventions are differentially managed, with the following
163 objectives:

- 164 1. To understand the role of abiotic (physico-chemical water parameters, ditch
165 morphology, vegetation structure) and biotic factors (predator communities) in
166 determining larval mosquito community composition
- 167 2. To determine whether wetland management changes under recent agri-
168 environmental schemes are likely to have increased the larval abundance and
169 diversity of key UK mosquito vectors of important mosquito-borne viruses.

171 2 Methods

172 2.1 Study site

173 The Somerset Levels and Moors (SLM), the largest remaining lowland wet grassland
174 in the UK, spanning 650 km² in the southwest of England, holds unique ecological
175 significance. Designated under the European Commission Habitats Directive and the
176 UK Biodiversity Action Plan, it serves as an exemplary coastal grazing marsh habitat
177 (JNCC, 1994). This area plays a crucial role in providing essential ecosystem services
178 to local communities and tourists, boasting high biodiversity with a notable presence
179 of wading and migratory birds year-round ([Acreman et al, 2011](#)).
180
181
182
183
184

The SLM's history is marked by periodic winter inundation over the past 10,000 years, contributing to the development of fertile peat soils and rich biodiversity. However, human activities, such as drainage and ditching for seasonal grazing pastures, began as early as the 9th century and intensified in the mid-20th century, reaching a peak with peat extraction and agricultural practices. Recognizing its environmental sensitivity, the SLM received designation as an Environmentally Sensitive Area (ESA) in 1987. Subsequently, agri-environmental schemes were implemented to support farmers in adopting management practices beneficial to biodiversity and flood management (Morris et al, 2008). This led to the transformation of arable land back into wet grassland.

Operating in a tiered system, these agri-environment schemes prescribe different measures, including encouraging lower grazing pressure, minimizing mechanical intervention in fields and surrounding ditches, and maintaining higher minimum water levels, especially in winter (Tier 3 versus Tiers 1 and 2). We anticipate that these tiered prescriptions will influence shading, vegetation structure, ditch morphology, and the presence of macro-invertebrate predators. Consequently, these conditions are expected to impact mosquito species. Our study aims to sample and investigate the key drivers of mosquito community composition in Tier 3 versus Tier 1 sites across the SLM, shedding light on the ecological dynamics influenced by wetland management practices.

2.2 Ecological Survey

We randomly selected 17 ditch locations across two management regimes, eight in Tier 1 management and 9 in Tier 3 of the SLM (Fig 1). At each site, we selected four sampling sites (ditches) within a 500m radius of the location, often part of an interconnected ditch system. We surveyed these sampling sites using a standard dipping protocol across three time points: spring (May), summer (June/July) and autumn (August/September) for three years, from 2009 to 2011. We set up six dip-points, for which we took GPS locations, at each sampling site along the ditch from one to six meters, randomly determined by the throw of a dice. During each visit, we took a complete submersion dip sample from both water-body margins as well as the centre of the ditch using a 1-liter volume mosquito dipper at each of the dip points.

We recorded the abundance of mosquito larvae and pupae, and that of potential mosquito predator groups at each dip point. Aquatic macroinvertebrate species were identified in situ to order and suborder, where possible, using (Dobson et al, 2012). Mosquito larvae and pupae were preserved in 70% ethanol and identified to species or species complex level in the laboratory using the morphological keys by (Becker et al, 2010; Schaffner et al, 2001; Cranston, 1987; Snow and Terzi, 1990). During each visit, details of bankside, emergent and floating channel vegetation were recorded (Cope et al, 2009; Jermy et al, 1995). Plants within and at the edges of the ditch were identified to genus or species level, and their percentage cover and height estimated. Vegetation height and percentage cover values were averaged across species in three groups based on their functional impact, bank, emergent, and floating vegetation, since these vegetation structures are likely to have differential impacts on habitat suitability across mosquito species (Table 1). We measured the physicochemical characteristics

of the ditch at each sampling site, assessing ditch width, and area of the ditch shaded (a proxy for habitat openness) as well as pH, temperature, turbidity, and salinity of the water. Average values for the covariates listed in Table 1 were summarised across the six dip-points per ditch site in each season.

235

236 2.3 Statistical Analysis

237

We used a joint multivariate hierarchical generalized mixed linear model approach, to account for the interdependency of species responses to the environment and species responses to each other in the ecosystem, by modelling all species simultaneously and accounting for each species responses to measured and unmeasured environmental covariates through latent variable factors (Wilkinson et al, 2019). We fitted our model using the R package Hierarchical Modelling of Species Communities (HMSC; Ovaskainen et al (2017); Ovaskainen and Abrego (2020)) framework, to explore how biotic and abiotic interactions drive mosquito larval distribution across the SLM.

246

The multispecies generalised linear latent variable model (with probit link function) was fitted to the presence-absence data for 4 mosquitoes and 8 predator groups obtained from our 320 sampling sites with abiotic covariates on a linear scale (Table 1). We excluded any species that occurred fewer than ten times to increase statistical stability (Ovaskainen and Abrego, 2020), leading to the exclusion of one mosquito species and four predator groups (see results). To account for potential spatial biases in the sampling data, we generated a distance matrix, calculated from the average coordinates across the six dip points that make up each sampling site, to represent the spatial scales between each sampling unit as a spatially structured random effect (F. Dormann et al, 2007). We considered the impact of temporal effects on sampling methods by including a nested random effect for both year and time point. In this case we consider the height and areal cover of different plant functional groups bank, emergent, and floating vegetation) as an abiotic driver as we expect them to function as a regulator of population fitness through shielding of predation or similar processes (Saha et al, 2009).

260

The model was fitted using four Monte-Carlo Markov Chains (MCMC) with a transient period of 5000 samples and target of 1000 samples per chain using a thinning rate of 1000 for a total of 4 million MCMC post-transient in samples in total. Parameter convergence was measured using Gelman and Rubens potential scale reduction factor (PSRF, Gelman and Rubin (1992)). We used five-fold cross validation to validate model performance, comparing predictive and explanatory values of Tjur's R2 and the area under curve (AUC) statistic for each species (Lobo et al, 2008; Tjur, 2009). We examined the importance of different sets of covariates in our model by partitioning the variation explained by fitting of partial models (Borcard et al, 1992; Ovaskainen and Abrego, 2020). We originally aimed to construct abundance models that included the same covariates, as we hypothesised that biotic interactions would have a greater impact on species abundance than species presence. However, due to the high complexity of the model, it was deemed computationally infeasible to achieve an acceptable fit and run-times exceeded one month without reasonable convergence for all species (Howard et al, 2014).

275

276

Table 1 Effect of Environmental Variables on Mosquito Abundance: Impact and expected ecological implications of key environmental factors on mosquito populations, including vegetation cover, water characteristics, and habitat structure.

Variable	Description	Impact on Mosquito Abundance
Floating Vegetation Cover (%)	The percentage of the water surface covered by floating plants or plants with significant leaf coverage.	Expected negative impact on mosquito population density. Dense floating aquatic plants, like <i>Lemna spp.</i> , can inhibit mosquito larvae and pupae reaching the water surface for air Cuthbert et al (2020) ; Eid et al (1992) .
Bank Height (cm)	The height of vegetation along the ditch margins and banks.	Increased structural complexity may provide favorable microhabitats for adult mosquitoes and increase availability of sheltered resting places Sauer et al (2021) .
Bank Vegetation Cover (%)	The percentage of banks covered by plant matter.	Cover would be expected to provide similar benefits as height.
Emergent Vegetation Height (cm)	The height of vegetation emerging vertically from the waterbody.	Emergent vegetation may provide increased shelter for mosquito females ovipositing on the water surface and improve larval and pupal survival through predator avoidance Saha et al (2009) .
Emergent Vegetation Cover (%)	The percentage area of emergent vegetation.	Cover would be expected to provide similar benefits as height.
Shaded Water (%)	Percentage of water surface shaded.	Expected positive impact on mosquito species with a preference for heavily vegetated or cool breeding sites Hawkes et al (2020) .
Width (cm)	Width of the waterbody.	Wider waterbodies may be more favorable for vertebrate predators, negatively impacting mosquito density Sunahara et al (2002) . Some species, like <i>An. maculipennis s.l.</i> , prefer more open habitats Hawkes et al (2020) .
Water Temp (°C)	Water temperature at sampling.	Larval development time is shorter, and survival is better at moderately higher temperatures Bayoh and Lindsay (2004) ; Mereta et al (2013) ; Rydzanicz and Lone (2003) .
Dissolved O ₂ (ppm)	Concentration of dissolved oxygen in the water.	Uncertain impact as many UK species are tolerant to a broad range of dissolved oxygen levels.
pH	Water acidity/basicity at sampling.	Most mosquitoes prefer neutral pH levels for optimum growth and are tolerant to moderate fluctuations Emidi et al (2017) .
Turbidity	Water clarity, indication of water flow.	Turbid waters are expected to increase mosquito larvae survival since predator efficiency is reduced Cano-Rocabayera et al (2020) ; Utne-Palm (2002) .

277
278
279
280
281
282
283
284
285
286
287
288
289
290
291
292
293
294
295
296
297
298
299
300
301
302
303
304
305
306
307
308
309
310
311
312
313
314
315
316
317
318
319
320
321
322

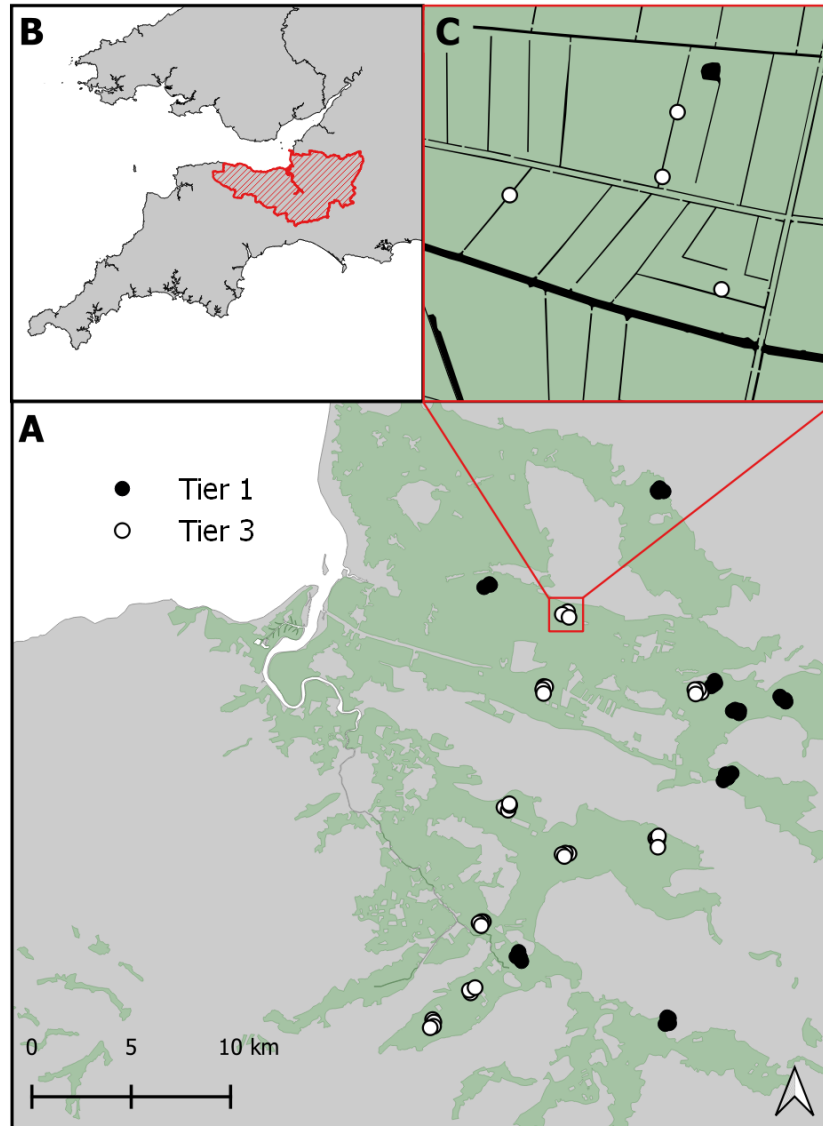


Fig. 1 **A** Map of Somerset Levels and Moors study site. Extent of coastal grazing marsh in green with Tier 1 and Tier 3 locations with sampling sites superimposed (black and white circles respectively). **B** Location of the study site (red hatching) in South England. **C** Inset frame showing detailed hierarchical spatial sampling design for each sampling site (circles) in which four ditches were sampled within a 500m square radius for each location.

To understand if management tiers influence potential abiotic drivers of mosquito populations, we estimated the marginal effect of management tier on each covariate measured in our sampling procedure (Table 1). We modelled each covariate separately against management level as a categorical factor, with both a random effect for site and a nested random effect of season within year to account for temporal differences in covariate distribution. Bayesian multivariate models were built in the probabilistic programming language Stan using the BRMS package in R (Carpenter et al, 2017; Bürkner, 2017). Covariates measured on a percentage scale (metrics of vegetation cover and shaded area) used a Zero-One Inflated Beta response distribution. Bank and emergent vegetation height used lognormal hurdle mixed response distributions to account for over-dispersion and the influence of zero values. All other covariates used a student-t distribution for robust estimation of parameter values. Significance was measured across the 95% CI using mean equal tailed intervals of the posterior distribution.

3 Results

3.1 Differences in environmental conditions between management tiers

Metrics of ditch vegetation structure differed significantly between sites subject to Tier 1 versus Tier 3 management, whilst physicochemical properties of the waterbody and ditch structure parameters did not (Table 2, Table S1). Though water-bodies were on average 8cm wider in Tier 3 managed areas, this difference was not statistically significant (95% CI [-23.94, 6.31]). There was no measurable difference in turbidity (95% CI [-0.15, 0.18]) or salinity (95% CI [-0.12, 0.15]) between the management tiers, and pH values were on average -0.3 lower in Tier 3 areas, but this was also non-significant (95% CI [0.03, 0.65]).

Bank vegetation was more likely to be present (95% CI [-0.29, -0.10], Table S1), and when present it was significantly taller, by 25cm on average (95% CI [-55.78, -4.24]), in Tier 3 ditches than Tier 1 ditches, but we found no differences in the levels of bank-side vegetation cover between tiers (Mean = 0.01, 95% CI [-0.05, 0.07]). Similarly, we found that emergent channel vegetation was 29% more likely to be present in Tier 3 areas (95% CI [-0.39, -0.18]), and when emergent vegetation was present it was 5cm taller on average than in Tier 3 areas than Tier 1 areas (95% CI [-12.39, -0.34]). There was no measurable difference in the probability of floating vegetation cover being 0% (95% CI [-0.15, 0.06]) or 100% (95% CI [-0.1, 0.04]) between tiers, but on average there was 10% less floating vegetation cover in Tier 3 areas than in Tier 1 areas and this was significant (95% CI [0.01, 0.19]). The amount of shaded area of the channel did not vary significantly between tiers (Mean = 0.12 95% CI [-0.01, 0.27]), but the probability of a waterbody being completely shaded was 48% higher in Tier 3 areas than Tier 1 (95% CI [-0.8, -0.2]), and the probability of a waterbody having no shade was 10% more likely in Tier 3 areas (95% CI [0.03, 0.016]).

Table 2 Differences in environmental variables between the Tier 1 (T1) and Tier 3 (T3) wetland management regimes. Table shows the marginal effect of Management Tier for each environmental covariate from pairwise posterior distribution contrasts of T1-T3 values. PD (Probability of Direction) estimates above 97.5 are deemed significant and highlighted in bold. MPE (Mean Parameter Estimates) with Lower MPE_{Low} and Upper MPE_{High} estimates represent the equal tailed 95% CI estimate across the model's posterior distribution. Full parameter estimates for each model covariate are given in Table S1.

Model Covariate	Effect of Tier 3	PD (%)	MPE	MPE _{Low}	MPE _{High}
Salinity	-	57.40	0.01	-0.12	0.15
Emergent Vegetation Height (Height _{Emerg})	Taller Emergent Vegetation	98.41	-5.30	-12.39	-0.34
Dissolved Oxygen (DO ²)	-	89.17	6.59	-4.10	17.40
pH	-	96.53	0.30	-0.03	0.65
Turbidity	-	59.00	0.02	-0.15	0.18
Floating Vegetation Cover (Cover _{Float})	Less Floating Vegetation Cover	98.33	0.09	0.01	0.19
Bankside Vegetation Cover (Cover _{Bank})	-	76.62	0.01	-0.05	0.07
Emergent Vegetation Cover (Cover _{Emerg})	-	70.88	-0.01	-0.04	0.02
Water Temperature	-	88.33	-0.82	-2.24	0.59
Shaded	-	96.37	0.12	-0.01	0.27
Ditch Width	-	87.67	-8.58	-23.94	6.31
Bank Vegetation Height (Height _{Bank})	Taller Bank Vegetation	99.62	-25.28	-55.78	-4.24

3.2 Abundance and prevalence of sample mosquito and predator taxa

We recorded twelve different aquatic macroinvertebrates taxa in the SLM of which five were mosquitoes (Table 3). We identified 6896 mosquito larvae in total. *Culiseta annulata* (n = 3250, 47.13%) and *Culex pipiens* (n = 3248, 47.10%) made up the highest proportion of these larvae, followed by *Anopheles claviger* (n = 292, 4.23%) and *Anopheles maculipennis* s.l. (n = 105, 1.52%). *Anopheles maculipennis* s.l. were most prevalent, occurring in 13% of the sample sites, followed by *Cs. annulata* (12%), *An. claviger* (11%), *Cx. pipiens* (10%), and lastly *Aedes (Ochlerotatus) caspius* which was present in just a single sampling site (< 0.1%). Because of the low abundance and low prevalence, *Ae. caspius* was omitted from the subsequent analysis.

We identified eight potential predator taxa that were prevalent across at least 10 sites to be included in this statistical analysis (Table 3). Of these taxa, adult *Coleoptera* (water beetles) were most prevalent, being present in the most sampling units (27%, n = 308) predator individuals recorded), followed by Corixidae (water boatmen) which were also the most abundant predator species (26%, n = 647), Zygoptera larvae (damselflies, 25%, n = 349) and Coleoptera larvae (19%, n = 139). The other four taxa had a much lower prevalence and abundance across all sampling units, including Gammaridae (*Palaemon varians* or common ditch shrimp, 8%, n = 103), Anisoptera larvae (dragonflies, 5%, n = 31), *Ilyocoris cimicoides* (Saucer bugs, 3%, n = 19) and *Nepa cinerea* (Water scorpions, 3%, n = 11).

Table 3 Relative prevalence (rate of occurrence across all sites) and total (and proportional) abundance of mosquito and predator taxa across sampled sites among sampled individuals across study sites.

Taxon	Prevalence (%)	Abundance (Total)	Mean Abundance per Sample Site
<i>Anopheles maculipennis</i>	13	105	2.44 ± 2.22
<i>Anopheles claviger</i>	11	292	8.11 ± 11.84
<i>Culex pipiens</i>	10	3248	101.50 ± 244.43
<i>Culiseta annulata</i>	13	3250	81.25 ± 160.62
Corixidae	26	647	7.70 ± 20.89
Coleoptera larvae	19	139	2.24 ± 1.70
Coleoptera adults	27	308	3.58 ± 3.30
Zygoptera larvae	26	349	4.20 ± 5.55
Anisoptera larvae	5	31	1.82 ± 1.42
<i>Ilyocoris cimicoides</i>	3	19	1.90 ± 2.18
<i>Nepa cinerea</i>	3	11	1.10 ± 0.32
Gammaridae	8	103	4.29 ± 4.65

3.3 Overall accuracy of community models and partitioning of variance between key sets of drivers

Parameter convergence of the HMSC model was satisfactory with all chains generating sufficient effective samples and PSRF values (Fig S2). Explanatory AUC values (for the training dataset) were high for all mosquito species (0.86-0.99) and predictive

AUC values (from the cross-validation) were reasonable (0.75-0.89). Explanatory AUC values were similarly high for potential predator taxa, but predictive AUC values were much lower for some of the less abundant taxa (*Nepa cinerea* = 0.4, *Ilyocoris cimicoides* = 0.55, *Anisoptera* larvae = 0.55, *Coleoptera* larvae = 0.57). All other predator taxa had adequate predictive AUC values above 0.69 (Table 4).

Metrics of variance explained for the training dataset were higher for Culicine species (*Cx. pipiens* s.l. Tjur's R^2 = 0.47 ; *Cs. annulata* Tjur's R^2 = 0.55) than Anopheline species (*An. maculipennis* s.l. Tjur's R^2 = 0.12; *An. claviger* (Tjur's R^2 = 0.23). When examining the importance of different sets of covariates, for mosquito species, we found that spatiotemporal effects accounted for on average 43% (SD 29%) of all variation explained by the models (Fig 2, Table S3). Tjur's R^2 values for predator taxa were much lower than for the mosquito species, except for Corixidae (Tjur's R^2 = 0.27) and Zygoptera (Tjur's R^2 = 0.28) larvae.

Random effects accounted for substantial variation in Culicine species and low amounts of variation for Anopheline species (Fig 2). For the Anopheline species, a higher proportion of variance was explained by chemical and channel structure covariates than for Culicine species. Temporal effects of year and season explained less variation in presence of mosquito species compared to the predator taxa, and little in Anopheline species (Table S3)

Table 4 Accuracy with which community models explained and predicted the distributions of mosquito and predator taxa including Under the Curve (AUC) and Tjur's R^2 values for explanation and prediction.

Taxa	Explanatory	Predictive		
	AUC	R ²	AUC	R ²
<i>Anopheles maculipennis</i> s.l.	0.86	0.12	0.75	0.06
<i>Anopheles claviger</i>	0.91	0.23	0.84	0.14
<i>Culex pipiens</i> s.l.	0.98	0.40	0.84	0.22
<i>Culiseta annulata</i>	0.99	0.53	0.89	0.38
Corixidae	0.88	0.27	0.82	0.21
Coleoptera larvae	0.78	0.08	0.57	0.02
Coleoptera	0.81	0.15	0.69	0.08
Zygoptera larvae	0.89	0.27	0.80	0.19
Anisoptera larvae	0.86	0.03	0.55	0.00
<i>Ilyocoris cimicoides</i>	0.87	0.03	0.50	0.00
<i>Nepa cinerea</i>	0.99	0.04	0.40	-0.01
Gammaridae	0.87	0.14	0.76	0.08

3.4 Larval mosquito responses to environmental drivers

Culex pipiens was significantly positively associated with bank vegetation cover (Mean = 0.03, 90% CI[0.01,0.04]), negatively associated with bank vegetation height (Mean = -0.006, 90% CI[-0.017,-0.001]), and negatively associated with floating vegetation cover (Mean = -0.01 90% CI[-0.018, -0.001]) (Fig 3). *Culiseta annulata* was significantly positively associated with more bankside vegetation cover (Mean = 0.02, 90% CI[0.001,0.035]) and high turbidity areas (Mean =1.41, 90% CI[0.28,2.65]). *Anopheles*

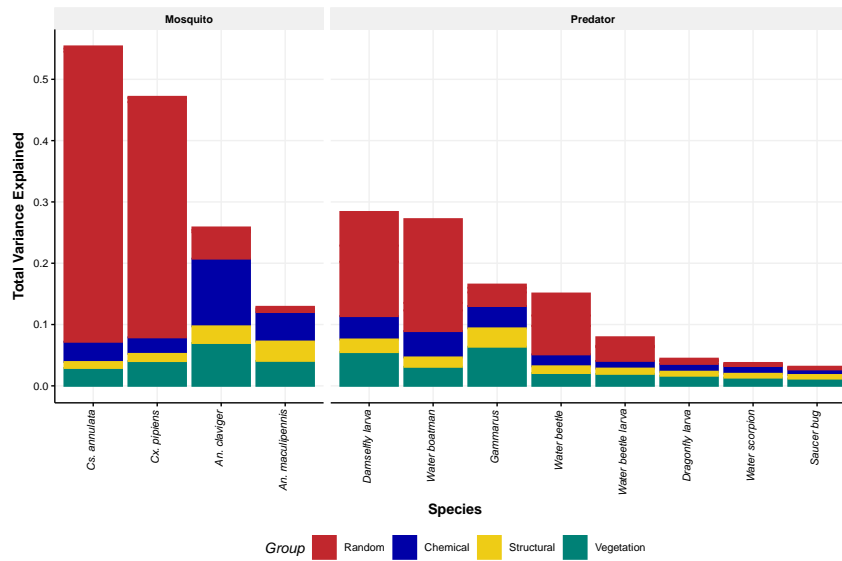


Fig. 2 Variance partitioning and total variance explained by each component for larval mosquitoes and predator species prevalent in the study side. Random effects are the variance explained by year season and the spatial component of the model summed for each species. The “chemical” category includes the physicochemical covariates pH, dissolved oxygen, salinity, turbidity and water temperature, the “structural” category includes the width and relative shadiness of the water body, and the “vegetation” category includes all vegetation metrics; floating, emergent and bank, as covariates in the model. Detailed breakdown of the variance explained by the random effects is provided in Table S3.

maculipennis s.l. showed strong preference for habitats with little shade (Mean = -1.13, 90% CI[-2.20,-0.13]) and higher levels of emergent vegetation (Mean = 0.015 90% CI[0.004,0.026]) (Fig 3). *Anopheles claviger* exhibited a strong preference for shaded habitats (Mean = 1.23, 90% CI[0.34,2.17]), and ditches with little floating vegetation cover (Mean = -0.015 90% CI[-0.025,-0.006]) (Fig 3).

Several potential predator taxa were also significantly correlated with an array of physicochemical and vegetation drivers (Fig 3), but we only interpret these further for those predatory taxa for which a larger percentage of variance in occurrence was explained by the model, namely water boatmen (*Corixidae* spp) and damselfly larvae (*Zygoptera*, Fig 2). The probability of occurrence of water boatmen was significantly negatively associated with lower shading of waterbodies (Mean = -0.96, 90% CI[-1.74,-0.23]). The probability of occurrence of damsel fly (*Zygoptera* spp) larvae was significantly positively impacted by higher levels of floating (Mean = 0.009, 90% CI[0.003,0.015]) and height of bank vegetation (Mean = 0.005, 90% CI[0.001,0.009], Fig 3).

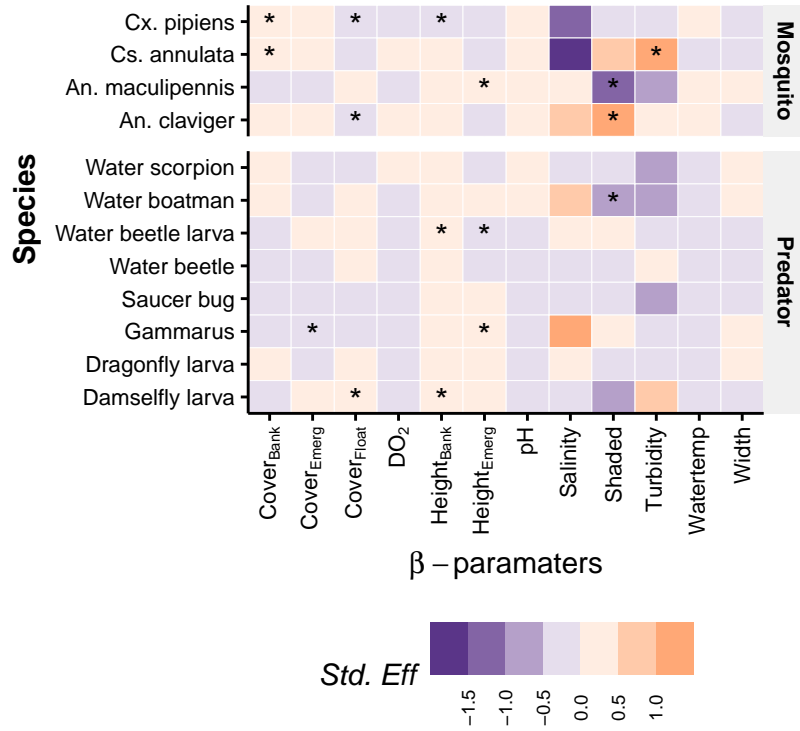


Fig. 3 Species responses to covariates by standardised coefficient effect size. “*” Indicates significance of parameter and cells that are blank indicate where the effect was not significant i.e., the 90% credible interval bridged zero. Height_{Emerg} = Emergent Vegetation height, Height_{Bank} = Emergent Vegetation height, Cover_{Float} = Floating Vegetation cover, Cover_{Bank} = Bank Vegetation height, Cover_{Emerg} = Emergent Vegetation cover.

3.5 Residual association between species

We found significant positive residual species associations between all mosquito species except *An. maculipennis* s.l. after accounting for environmental responses in the HMSC community model (Fig 4). Additionally, we found that all species of mosquito except *An. maculipennis* s.l. show significant negative associations with potential predator taxa including water beetle larvae and adults (Coleoptera), and damselfly (Zygoptera) larvae, water boatmen (Corixidae) and Gammaridae. Saucer bugs (*Ilyocoris*), dragonfly (Anisoptera) larvae and water scorpions (*Nepa cinerea*) do not show any significant associations with any other species. All other predator taxa show significant positive associations with one another (Fig 4).

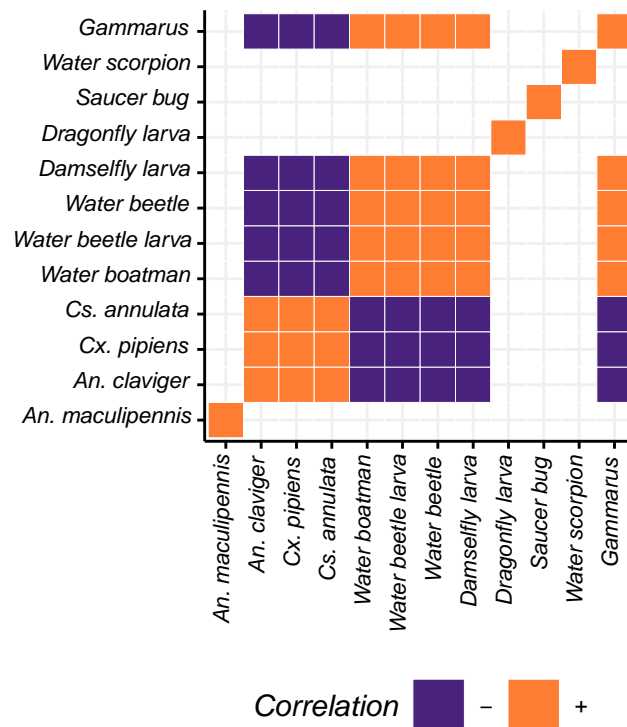


Fig. 4 Significant species residual correlations drawn from the Ω model parameter for each species in Hmsc. Coloured regions show which species meet the 95% significance threshold and are expected to be found together more than can be explained by model covariates alone. Correlations that don't meet this significance threshold are blank.

4 Discussion

4.1 Vegetation structure as a key driver of mosquito communities including potential vectors.

Increased water levels in Tier 3 areas have been previously shown to favour the establishment of wetland meadow plant species which increase the diversity and quality of vegetation in these areas compared to Tier 1 areas (Acreman et al, 2011). Our study supports this, with Tier 3 areas leading to significant increases in emergent and bank-side vegetation height, increasing the structural complexity of vegetation compared to Tier 1 areas (Table 2).

Areas such as wetlands and marshes tend to harbour a wide variety of mosquito species, due to the presence of a variety of suitable water bodies for oviposition, and

691 aquatic plants that provide shelter, food, and protection from predators, as well as a
 692 diverse set of host species from which to draw bloodmeals (Medlock et al, 2005; Becker
 693 et al, 2010). Adult mosquitoes benefit from structural complexity of vegetation, this
 694 complexity includes features such as diverse plant species, varied plant heights, and
 695 the presence of multiple layers of vegetation, all of which contribute to the creation
 696 of sheltered, shaded, and protected spaces. Such complexity in vegetation structure
 697 provides suitable conditions for adult mosquitoes, offering refuge from direct sunlight,
 698 wind, and other environmental elements, thereby enhancing their overall habitat suit-
 699 ability. that provides shelter, shade and protection from the sun and wind (Becker
 700 et al, 2010). Juvenile mosquitoes may also perceive similar benefits from the underwa-
 701 ter structures of algae and plant roots as refuges from predators (Collins et al, 2019).
 702 A review by Rey et al (2012) found that wetlands with high vegetative complexity
 703 had a greater diversity of mosquito species compared to wetlands with low vegetative
 704 complexity. Consistent with these prior studies, we found that occurrence of three of
 705 the key mosquito species in the study area (*An. claviger* and *An. maculipennis s.l.*,
 706 and to a lesser extent, the *Cx. pipiens* complex) was favoured by more complex chan-
 707 nel vegetation structure characteristic of Tier 3 management (increased height and
 708 cover of emergent and bankside vegetation, Table 2). Consistent with the associations
 709 described by Hawkes et al (2020). *An. maculipennis* s.l. showed significant preference
 710 for less shaded environments suggesting a preference for open style habitats, while *An.*
 711 *claviger* showed a preference for heavily shaded habitats (Fig 3). *Cx. pipiens* and *An.*
 712 *claviger*, both of which can cause significant biting nuisances, are likely to find Tier
 713 3 areas more favourable because of their preference for little floating vegetation (Fig
 714 3, Table 2). Floating vegetation can provide a physical barrier between mosquito and
 715 oviposition site as well as larvae and air, dissuading oviposition in these areas (Eid
 716 et al, 1992). Yet, previous studies have found positive associations between floating
 717 vegetation cover and mosquito species presence, suggesting the impacts of this factor
 718 on mosquito larvae is complex and context-dependent (Cuthbert et al, 2020; Golding
 719 et al, 2015).

720 Except for the association of turbid water with *Cs. annulata* presence, no signif-
 721 icant effects of physicochemical characteristics of the water on mosquito occurrence
 722 were found (Fig 3). This aligns with prior knowledge that Culicine species, *Cx. pipi-*
 723 *ens* and *Cs. annulata* utilise a breadth of oviposition sites, including drainage ditches,
 724 artificial containers, and small stagnant waters, that vary widely in water parameters
 725 (Hawkes et al., 2020). We found that physicochemical factors had a larger contribu-
 726 tion to variance explained for the Anopheline species. *Anopheles maculipennis s.l.*, and
 727 *An. claviger*, at 11% and 6% respectively, suggesting more restricted oviposition site
 728 preferences. The SLM system is an interconnected network of ditches that covers an
 729 area over several hundred square kilometres, leading to relatively homogeneous water
 730 chemistry across our study area. This means that the range of conditions experienced
 731 by our sampled species might not be large enough to elucidate any meaningful differ-
 732 ences in water parameter preferences (and indeed the Tier management regimes did
 733 not differ significantly in physico-chemical conditions).

734
 735
 736

4.2 Biotic drivers of larval mosquitoes

Consistent with prior studies of mosquito community composition at landscape level, we found that biotic interactions may affect the distribution of mosquitos across a wetland environment (Golding et al, 2015). Many of the potential predator taxa such as dragonfly and damselfly larvae are frequently observed as effective larval mosquito predators in other contexts and indeed some such as dragonfly larvae have been investigated for biological control of mosquitoes (Medlock and Snow, 2008; Onyeka, 1983; Saha et al., 2012). Water beetles and water boatmen have also been implicated in mosquito larval predation, but their relative predation pressure is thought to be linked to the vulnerability of mosquito larvae (Jeffries, 1988; Medlock and Snow, 2008).

As described above, vegetation structure in and around waterbodies affects the availability of refugia from predators and consequently the effectiveness of predator avoidance strategies of immature mosquitoes (Saha et al, 2009). Environments with complex underwater vegetation limit the space for predators and mosquito larvae to interact and reduces overall predator efficiency (Saha et al, 2009; Sunahara et al, 2002). The higher cover and height of emergent vegetation detected in Tier 3 areas could provide complex vegetation structure both above and below the water level, providing shady refugia that improve predator avoidance in these sites.

It's crucial to recognise that the species interactions deduced from residual correlations in joint occurrence models are not as dependable as direct observations of predator-prey interactions. Instead, these inferred interactions may be indicative of unmeasured factors such as shared or non-shared environmental preferences between species (Poggiato et al, 2021). In essence, while joint occurrence models provide valuable insights, caution should be exercised in attributing the correlations solely to direct predator-prey interactions, as other environmental factors might contribute to the observed patterns (Zurell et al, 2018). For example, though some mosquito species were found to be negatively correlated with Gammaridae we suspect this may reflect different preferences for unmeasured environmental conditions, since the UK Gammarus species (*Palaemon varians*) is omnivorous and occupies different depths of the waterbodies to mosquito larvae, leading to limited potential predation opportunities (P. Scarlett, *pers comm*, June 2023).

The community models exhibited relatively lower performance for predator species when compared to mosquito species. This implies that to comprehensively grasp how wetland management may influence predator effects on mosquito populations in this context, additional and more detailed data on predators, with improved taxonomic resolution, could be valuable. It's worth considering that while this suggestion is important, it may not be the exclusive solution, and other approaches could also contribute to a more nuanced understanding. Prior studies seem to suggest that management plans targeting biodiversity, such as that seen in Tier 3, will have a positive impact on abundance of key predator taxa such as fish (Chandra et al, 2008; Griffin and Knight, 2012). Increased predator abundance would provide a potential control agent for mosquito populations, but few studies have shown this in the field, and none in the UK (Griffin and Knight, 2012; Medlock and Snow, 2008; Saha et al, 2012). Our study indicates that water beetle larvae and adults, dragonfly, and damselfly nymphs, and water boatmen may be key predator taxa that play a role in regulating mosquito

populations within lowland wet grasslands, and that these roles should be investigated further to fully understand trade-offs between biodiversity management and mosquito biting risk.

5 Conclusion

We have shown here how management schemes directed at increasing the biodiversity of grazed wetlands, could increase the suitability of those habitats for immatures of some key mosquito vectors and nuisance biters, encouraging diverse vegetation structure in and around water bodies, that may reduce their vulnerability to predators. However, thinning or removal of vegetation is not a viable strategy to control mosquito populations, being at odds with the targets of wetland management strategies. Vegetation removal impinges upon important wetland ecosystem functions by decreasing biodiversity, lowering water quality and reducing flood resilience of an area (Acreman et al, 2011; Rochlin et al, 2012a).

Furthermore, to interpret disease risk given future incursions of viruses such as West Nile virus, Sindbis virus or Usutu virus into the UK, it would be necessary to understand how these impacts of wetland management on juvenile mosquito populations cascade through into impacts on the ratio of adult vectors to susceptible hosts (a key parameter in disease transmission (Smith et al, 2004), by sampling adult vectors, hosts and their interactions (e.g., via blood meal analysis) across wetland gradients into areas of human habitation (Hanford et al, 2020). This would provide the evidence-base for co-development of integrated mosquito management and risk awareness strategies among cross-sectoral stakeholders that would minimise risk of exposure while aligning with environmental wetland management goals (Martinou et al, 2020). Given the diverse and growing mosquito-borne pathogen threats to people living in and around wetland ecosystems, and the diverse assemblages of potential mosquito vector species involved, the combination of joint models with empirical surveys provides an effective way of inferring the complex ecological interactions that will underpin the trade-offs between disease risk and wetland management.

Supplementary information. If your article has accompanying supplementary file/s please state so here.

Acknowledgements. Acknowledgements are not compulsory. Where included they should be brief. Grant or contribution numbers may be acknowledged.

Declarations

5.1 Funding

This research was supported by the QMEE CDT, funded by NERC grant number NE/P012345/1.

5.2 Conflicts of Interest

The authors report no conflicts of interest or competing interests.

5.3 Ethics approval and consent	829
Not applicable.	830
	831
5.4 Consent for publication	832
Not applicable.	833
	834
5.5 Data availability	835
Data and code are available on GitHub/Dryad (https://github.com/dansmith/Smithetal2024) <i>To be committed to the repository shortly and archived when accepted</i>	836
	837
	838
	839
	840
	841
5.6 Materials availability	842
Not applicable.	843
	844
5.7 Code availability	845
Data and code are available on GitHub/Dryad (XXXXX - Will upload and DOI once accepted)	846
	847
	848
	849
5.8 Author contribution	850
The study was conceived by Daniel Smith, Stefanie Schäfer, Bethan Purse, and Miles Nunn. Field data collection and processing were carried out by Stefanie Schäfer, Miles Nunn, Bethan Purse, Nick Golding. Daniel Smith and Bethan Purse performed modelling and statistical analyses. Daniel Smith, Stefanie Schäfer, and Bethan Purse all contributed in writing the manuscript. The manuscript received valuable feedback and comments from Amanda Callaghan and Steven White.	851
	852
	853
	854
	855
	856
	857
	858
References	859
	860
Acreman MC, Harding RJ, Lloyd C, et al (2011) Trade-off in ecosystem services of the Somerset Levels and Moors wetlands. <i>Hydrological Sciences Journal</i> 56(8):1543–1565. https://doi.org/10.1080/02626667.2011.629783	861
	862
	863
	864
Bakonyi T, Haussig JM (2020) West Nile virus keeps on moving up in Europe. <i>Eurosurveillance</i> 25(46). https://doi.org/10.2807/1560-7917.ES.2020.25.46.2001938	865
	866
	867
Bayoh MN, Lindsay SW (2004) Temperature-related duration of aquatic stages of the Afrotropical malaria vector mosquito <i>Anopheles gambiae</i> in the laboratory. <i>Medical and Veterinary Entomology</i> 18(2):174–179. https://doi.org/10.1111/j.0269-283X.2004.00495.x	868
	869
	870
	871
Becker N, Petric D, Zgomba M, et al (2010) <i>Mosquitoes and Their Control</i> . Springer, Berlin, Heidelberg, https://doi.org/10.1007/978-3-540-92874-4	872
	873
	874

875 Beketov MA, Liess M (2007) Predation risk perception and food scarcity induce alter-
876 ations of life-cycle traits of the mosquito *Culex pipiens*. *Ecological Entomology*
877 32(4):405–410. <https://doi.org/10.1111/j.1365-2311.2007.00889.x>
878
879 Benelli G, Wilke ABB, Beier JC (2020) *Aedes albopictus* (Asian Tiger Mosquito).
880 *Trends in Parasitology* 36(11):942–943. <https://doi.org/10.1016/j.pt.2020.01.001>
881
882 Borcard D, Legendre P, Drapeau P (1992) Partialling out the Spatial Component of
883 Ecological Variation. *Ecology* 73(3):1045–1055. <https://doi.org/10.2307/1940179>
884
885 Braks MAH, Honório NA, Lounibos LP, et al (2004) Interspecific Competition Between
886 Two Invasive Species of Container Mosquitoes, *Aedes aegypti* and *Aedes albopictus*
887 (Diptera: Culicidae), in Brazil. *Annals of the Entomological Society of America*
888 97(1):130–139. [https://doi.org/10.1603/0013-8746\(2004\)097\[0130:ICBTIS\]2.0.CO;2](https://doi.org/10.1603/0013-8746(2004)097[0130:ICBTIS]2.0.CO;2)
889
890 Bruguera S, Fernández-Martínez B, Martínez-de la Puente J, et al (2020) Environ-
891 mental drivers, climate change and emergent diseases transmitted by mosquitoes
892 and their vectors in southern Europe: A systematic review. *Environmental Research*
893 191:110038. <https://doi.org/10.1016/j.envres.2020.110038>
894
895 Buckley A, Dawson A, Moss SR, et al (2003) Serological evidence of West Nile virus,
896 Usutu virus and Sindbis virus infection of birds in the UK. *The Journal of General*
897 *Virology* 84(Pt 10):2807–2817. <https://doi.org/10.1099/vir.0.19341-0>
898
899 Bürkner PC (2017) **Brms** : An *R* Package for Bayesian Multilevel Models Using *Stan*.
900 *Journal of Statistical Software* 80(1). <https://doi.org/10.18637/jss.v080.i01>
901
902 Cano-Rocabayera O, Vargas-Amengual S, Aranda C, et al (2020) Mosquito larvae
903 consumption in turbid waters: The role of the type of turbidity and the larval
904 stage in native and invasive fish. *Hydrobiologia* 847(5):1371–1381. <https://doi.org/10.1007/s10750-020-04195-0>
905
906 Carpenter B, Gelman A, Hoffman MD, et al (2017) *Stan* : A Probabilistic Program-
907 ming Language. *Journal of Statistical Software* 76(1). <https://doi.org/10.18637/jss.v076.i01>
908
909 Chandra G, Bhattacharjee I, Chatterjee SN, et al (2008) Mosquito control by
910 larvivorous fish. *The Indian Journal of Medical Research* 127(1):13–27
911
912 Cheng Y, Tjaden NB, Jaeschke A, et al (2018) Evaluating the risk for Usutu virus
913 circulation in Europe: Comparison of environmental niche models and epidemiologi-
914 cal models. *International Journal of Health Geographics* 17(1):35. <https://doi.org/10.1186/s12942-018-0155-7>
915
916
917 Collins CM, Bonds JAS, Quinlan MM, et al (2019) Effects of the removal or reduction
918 in density of the malaria mosquito, *ANOPHELES GAMBIAE s.l.*, on interacting
919 predators and competitors in local ecosystems. *Medical and Veterinary Entomology*
920

33(1):1–15. https://doi.org/10.1111/mve.12327	921
Cope T, Gray A, Tebbs MC (2009) Grasses of the British Isles. No. 13 in B.S.B.I. Handbook, Botanical Society of the British Isles, London	922
Cranston PS (ed) (1987) Keys to the Adults, Male Hypopygia, Fourth-Instar Larvae, and Pupae of the British Mosquitoes (Culicidae): With Notes on Their Ecology and Medical Importance. No. no. 48 in Scientific Publication / Freshwater Biological Association, Freshwater Biological Association, Ambleside, Cumbria	923
Cuthbert RN, Vong GYW, Paolacci S, et al (2020) Aquatic plant extracts and coverage mediate larval mosquito survivorship and development. Biological Control 145:104263. https://doi.org/10.1016/j.biocontrol.2020.104263	924
Dale PER, Knight JM (2008) Wetlands and mosquitoes: A review. Wetlands Ecology and Management 16(4):255–276. https://doi.org/10.1007/s11273-008-9098-2	925
Defra (2023a) 25 Year Environment Plan. https://www.gov.uk/government/publications/25-year-environment-plan	926
Defra (2023b) Countryside Stewardship: Get funding to protect and improve the land you manage. https://www.gov.uk/guidance/countryside-stewardship-get-funding-to-protect-and-improve-the-land-you-manage	927
Defra (2023c) Third National Adaptation Programme (NAP3). https://www.gov.uk/government/publications/third-national-adaptation-programme-nap3	928
Dobson M, Pawley S, Fletcher M, et al (2012) Guide to Freshwater Invertebrates: No. 68, 1st edn. Freshwater Biological Assn., Ambleside, Cumbria	929
Eid MAA, Kandil MAE, Moursy EB, et al (1992) Effect of the duck-weed, <i>Lemna minor</i> vegetations on the mosquito, <i>Culex pipiens pipiens</i> . International Journal of Tropical Insect Science 13(03):357–361. https://doi.org/10.1017/S174275840001362X	930
Emidi B, Kisinza WN, Mmbando BP, et al (2017) Effect of physicochemical parameters on Anopheles and Culex mosquito larvae abundance in different breeding sites in a rural setting of Muheza, Tanzania. Parasites & Vectors 10(1):304. https://doi.org/10.1186/s13071-017-2238-x	931
F. Dormann C, M. McPherson J, B. Araújo M, et al (2007) Methods to account for spatial autocorrelation in the analysis of species distributional data: A review. Ecography 30(5):609–628. https://doi.org/10.1111/j.2007.0906-7590.05171.x	932
Ferraguti M, Martínez-de la Puente J, Roiz D, et al (2016) Effects of landscape anthropization on mosquito community composition and abundance. Scientific	933

967 Reports 6(1):29002. <https://doi.org/10.1038/srep29002>
968
969 Folly AJ, Sewgobind S, Hernández-Triana LM, et al (2022) Evidence for overwintering
970 and autochthonous transmission of Usutu virus to wild birds following its redetection
971 in the United Kingdom. *Transboundary and Emerging Diseases* 69(6):3684–3692.
972 <https://doi.org/10.1111/tbed.14738>
973
974 Gelman A, Rubin DB (1992) Inference from iterative simulation using multiple
975 sequences. *Statistical science* 7(4):457–472
976
977 Gibbs JP (2000) Wetland Loss and Biodiversity Conservation. *Conservation Biology*
978 14(1):314–317. <https://doi.org/10.1046/j.1523-1739.2000.98608.x>
979
980 Golding N, Harris DJ (2015) BayesComm: Bayesian community ecology analysis. R
981 package version 01-2
982
983 Golding N, Nunn MA, Purse BV (2015) Identifying biotic interactions which drive
984 the spatial distribution of a mosquito community. *Parasites & Vectors* 8(1):367.
985 <https://doi.org/10.1186/s13071-015-0915-1>
986
987 Gottdenker NL, Streicker DG, Faust CL, et al (2014) Anthropogenic Land Use Change
988 and Infectious Diseases: A Review of the Evidence. *EcoHealth* 11(4):619–632. <https://doi.org/10.1007/s10393-014-0941-z>
989
990 Griffin LF, Knight JM (2012) A review of the role of fish as biological control agents of
991 disease vector mosquitoes in mangrove forests: Reducing human health risks while
992 reducing environmental risk. *Wetlands Ecology and Management* 20(3):243–252.
993 <https://doi.org/10.1007/s11273-012-9248-4>
994
995 Hanford JK, Webb CE, Hochuli DF (2020) Management of urban wetlands for con-
996 servation can reduce aquatic biodiversity and increase mosquito risk. *Journal of*
997 *Applied Ecology* 57(4):794–805. <https://doi.org/10.1111/1365-2664.13576>
998
999 Hawkes F, Medlock JM, Vaux AG, et al (2020) Wetland mosquito survey handbook:
1000 Assessing suitability of British wetlands for mosquitoes
1001
1002 Howard C, Stephens PA, Pearce-Higgins JW, et al (2014) Improving species distribu-
1003 tion models: The value of data on abundance. *Methods in Ecology and Evolution*
1004 5(6):506–513. <https://doi.org/10.1111/2041-210X.12184>
1005
1006 James-Pirri MJ, Ginsberg HS, Erwin RM, et al (2009) Effects of Open Marsh Water
1007 Management on Numbers of Larval Salt Marsh Mosquitoes. *Journal of Medical*
1008 *Entomology* 46(6):1392–1399. <https://doi.org/10.1603/033.046.0620>
1009
1010 Jeffries M (1988) Individual vulnerability to predation: The effect of alternative prey
1011 types. *Freshwater Biology* 19(1):49–56. <https://doi.org/10.1111/j.1365-2427.1988.tb00326.x>
1012

Jermy AC, Tutin TG, Jermy AC, et al (1995) Sedges of the British Isles, 2nd edn. No. 1 in B.S.B.I. Handbook, Botanical Society of the British Isles, London	1013 1014 1015
Giannino JA, Walton WE (2004) Evaluation of vegetation management strategies for controlling mosquitoes in a southern California constructed wetland. Journal of the American Mosquito Control Association 20(1):18–26	1016 1017 1018 1019
Kilpatrick AM (2011) Globalization, Land Use, and the Invasion of West Nile Virus. Science 334(6054):323–327. https://doi.org/10.1126/science.1201010	1020 1021 1022
Lambin EF, Tran A, Vanwambeke SO, et al (2010) Pathogenic landscapes: Interactions between land, people, disease vectors, and their animal hosts. International Journal of Health Geographics 9(1):54. https://doi.org/10.1186/1476-072X-9-54	1023 1024 1025 1026
Lobo JM, Jiménez-Valverde A, Real R (2008) AUC: A misleading measure of the performance of predictive distribution models. Global Ecology and Biogeography 17(2):145–151. https://doi.org/10.1111/j.1466-8238.2007.00358.x	1027 1028 1029
Martinou AF, Schäfer SM, Bueno Mari R, et al (2020) A call to arms: Setting the framework for a code of practice for mosquito management in European wetlands. Journal of Applied Ecology 57(6):1012–1019. https://doi.org/10.1111/1365-2664.13631	1030 1031 1032 1033 1034
Medlock JM, Snow KR (2008) Natural predators and parasites of British mosquitoes – a review. European Mosquito Bulletin p 11	1035 1036 1037
Medlock JM, Vaux AG (2015a) Impacts of the creation, expansion and management of English wetlands on mosquito presence and abundance – developing strategies for future disease mitigation. Parasites & Vectors 8(1):142. https://doi.org/10.1186/s13071-015-0751-3	1038 1039 1040 1041 1042
Medlock JM, Vaux AG (2015b) Seasonal dynamics and habitat specificity of mosquitoes in an English wetland: Implications for UK wetland management and restoration. Journal of Vector Ecology 40(1):90–106. https://doi.org/10.1111/jvec.12137	1043 1044 1045 1046 1047
Medlock JM, Snow KR, Leach S (2005) Potential transmission of West Nile virus in the British Isles: An ecological review of candidate mosquito bridge vectors. Medical and Veterinary Entomology 19(1):2–21. https://doi.org/10.1111/j.0269-283X.2005.00547.x	1048 1049 1050 1051 1052
Medlock JM, Hansford KM, Schaffner F, et al (2012) A Review of the Invasive Mosquitoes in Europe: Ecology, Public Health Risks, and Control Options. Vector Borne and Zoonotic Diseases 12(6):435–447. https://doi.org/10.1089/vbz.2011.0814	1053 1054 1055 1056 1057 1058

1059 Mereta ST, Yewhalaw D, Boets P, et al (2013) Physico-chemical and biological
1060 characterization of anopheline mosquito larval habitats (Diptera: Culicidae): Impli-
1061 cations for malaria control. *Parasites & Vectors* 6(1):320. [https://doi.org/10.1186/](https://doi.org/10.1186/1756-3305-6-320)
1062 [1756-3305-6-320](https://doi.org/10.1186/1756-3305-6-320)
1063
1064 Meyer Steiger DB, Ritchie SA, Laurance SGW (2016) Mosquito communities and
1065 disease risk influenced by land use change and seasonality in the Australian tropics.
1066 *Parasites & Vectors* 9(1):387. <https://doi.org/10.1186/s13071-016-1675-2>
1067
1068 Morris J, Bailey AP, Lawson CS, et al (2008) The economic dimensions of integrating
1069 flood management and agri-environment through washland creation: A case from
1070 Somerset, England. *Journal of Environmental Management* 88(2):372–381. [https:](https://doi.org/10.1016/j.jenvman.2007.03.023)
1071 [//doi.org/10.1016/j.jenvman.2007.03.023](https://doi.org/10.1016/j.jenvman.2007.03.023)
1072
1073 Ovaskainen O, Abrego N (2020) Joint Species Distribution Modelling: With Appli-
1074 cations in R. *Ecology, Biodiversity and Conservation*, Cambridge University Press,
1075 Cambridge, <https://doi.org/10.1017/9781108591720>
1076
1077 Ovaskainen O, Tikhonov G, Norberg A, et al (2017) How to make more out of commu-
1078 nity data? A conceptual framework and its implementation as models and software.
1079 *Ecology Letters* 20(5):561–576. <https://doi.org/10.1111/ele.12757>
1080
1081 Pichler M, Hartig F (2021) A new joint species distribution model for faster and
1082 more accurate inference of species associations from big community data. *Methods*
1083 *in Ecology and Evolution* 12(11):2159–2173. [https://doi.org/10.1111/2041-210X.](https://doi.org/10.1111/2041-210X.13687)
1084 [13687](https://doi.org/10.1111/2041-210X.13687)
1085
1086 Poggiato G, Münkemüller T, Bystrova D, et al (2021) On the Interpretations of Joint
1087 Modeling in Community Ecology. *Trends in Ecology & Evolution* 36(5):391–401.
1088 <https://doi.org/10.1016/j.tree.2021.01.002>
1089
1090 Rey JR, Walton WE, Wolfe RJ, et al (2012) North American Wetlands and Mosquito
1091 Control. *International Journal of Environmental Research and Public Health*
1092 9(12):4537–4605. <https://doi.org/10.3390/ijerph9124537>
1093
1094 Rochlin I, James-Pirri MJ, Adamowicz SC, et al (2012a) The Effects of Integrated
1095 Marsh Management (IMM) on Salt Marsh Vegetation, Nekton, and Birds. *Estuaries*
1096 *and Coasts* 35(3):727–742. <https://doi.org/10.1007/s12237-011-9468-5>
1097
1098 Rochlin I, James-Pirri MJ, Adamowicz SC, et al (2012b) Integrated Marsh Manage-
1099 ment (IMM): A new perspective on mosquito control and best management practices
1100 for salt marsh restoration. *Wetlands Ecology and Management* 20(3):219–232.
1101 <https://doi.org/10.1007/s11273-012-9251-9>
1102
1103 Rose R (2001) Pesticides and Public Health: Integrated Methods of Mosquito Manage-
1104 ment. *Emerging Infectious Diseases* 7(1):17–23. [https://doi.org/10.3201/eid0701.](https://doi.org/10.3201/eid0701.010103)
[010103](https://doi.org/10.3201/eid0701.010103)

Russell RC (1999) Constructed wetlands and mosquitoes: Health hazards and management options—An Australian perspective. <i>Ecological Engineering</i> 12(1):107–124. https://doi.org/10.1016/S0925-8574(98)00057-3	1105 1106 1107 1108
Rydzanicz K, Lone E (2003) Species composition and seasonal dynamics of mosquito larvae in the Wrocław, Poland area. <i>Journal of Vector Ecology</i> 28(2):255–266	1109 1110 1111
Saha N, Aditya G, Saha GK (2009) Habitat complexity reduces prey vulnerability: An experimental analysis using aquatic insect predators and immature dipteran prey. <i>Journal of Asia-Pacific Entomology</i> 12(4):233–239. https://doi.org/10.1016/j.aspen.2009.06.005	1112 1113 1114 1115
Saha N, Aditya G, Banerjee S, et al (2012) Predation potential of odonates on mosquito larvae: Implications for biological control. <i>Biological Control</i> 63(1):1–8. https://doi.org/10.1016/j.biocontrol.2012.05.004	1116 1117 1118 1119
Sauer FG, Grave J, Lühken R, et al (2021) Habitat and microclimate affect the resting site selection of mosquitoes. <i>Medical and Veterinary Entomology</i> 35(3):379–388. https://doi.org/10.1111/mve.12506	1120 1121 1122 1123
Schäfer ML, Lundström JO, Pfeffer M, et al (2004) Biological diversity versus risk for mosquito nuisance and disease transmission in constructed wetlands in southern Sweden. <i>Medical and Veterinary Entomology</i> 18(3):256–267. https://doi.org/10.1111/j.0269-283X.2004.00504.x	1124 1125 1126 1127 1128
Schaffner F, Angel G, Geoffroy B, et al (2001) The Mosquitoes of Europe. An Identification and Training Programme.	1129 1130 1131
Smith DL, Dushoff J, McKenzie FE (2004) The Risk of a Mosquito-Borne Infection in a Heterogeneous Environment. <i>PLoS Biology</i> 2(11):e368. https://doi.org/10.1371/journal.pbio.0020368	1132 1133 1134 1135
Snow KR, Terzi AJE (1990) Mosquitoes. No. 14 in <i>Naturalists' Handbooks</i> , Richmond, Slough	1136 1137 1138
Sunahara T, Ishizaka K, Mogi M (2002) Habitat size: A factor determining the opportunity for encounters between mosquito larvae and aquatic predators. <i>Journal of vector ecology : journal of the Society for Vector Ecology</i>	1139 1140 1141
Tjur T (2009) Coefficients of Determination in Logistic Regression Models—A New Proposal: The Coefficient of Discrimination. <i>The American Statistician</i> 63(4):366–372. https://doi.org/10.1198/tast.2009.08210	1142 1143 1144 1145
Townroe S, Callaghan A (2014) British Container Breeding Mosquitoes: The Impact of Urbanisation and Climate Change on Community Composition and Phenology. <i>PLOS ONE</i> 9(4):e95325. https://doi.org/10.1371/journal.pone.0095325	1146 1147 1148 1149 1150

1151 Utne-Palm A (2002) Visual feeding of fish in a turbid environment: Physical and
 1152 behavioural aspects. *Marine and Freshwater Behaviour and Physiology* 35(1-2):111–
 1153 128. <https://doi.org/10.1080/10236240290025644>
 1154
 1155 Wilkinson DP, Golding N, Guillera-Arroita G, et al (2019) A comparison of joint
 1156 species distribution models for presence-absence data. *Methods in Ecology and*
 1157 *Evolution* 10(2):198–211. <https://doi.org/10.1111/2041-210X.13106>
 1158
 1159 Zurell D, Pollock LJ, Thuiller W (2018) Do joint species distribution models
 1160 reliably detect interspecific interactions from co-occurrence data in homogenous
 1161 environments? *Ecography* 41(11):1812–1819. <https://doi.org/10.1111/ecog.03315>
 1162
 1163
 1164
 1165
 1166
 1167
 1168
 1169
 1170
 1171
 1172
 1173
 1174
 1175
 1176
 1177
 1178
 1179
 1180
 1181
 1182
 1183
 1184
 1185
 1186
 1187
 1188
 1189
 1190
 1191
 1192
 1193
 1194
 1195
 1196