

Final Project

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Math 189
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Application Problems

1. a)

```
library(ISLR2)
head(Carseats)
```

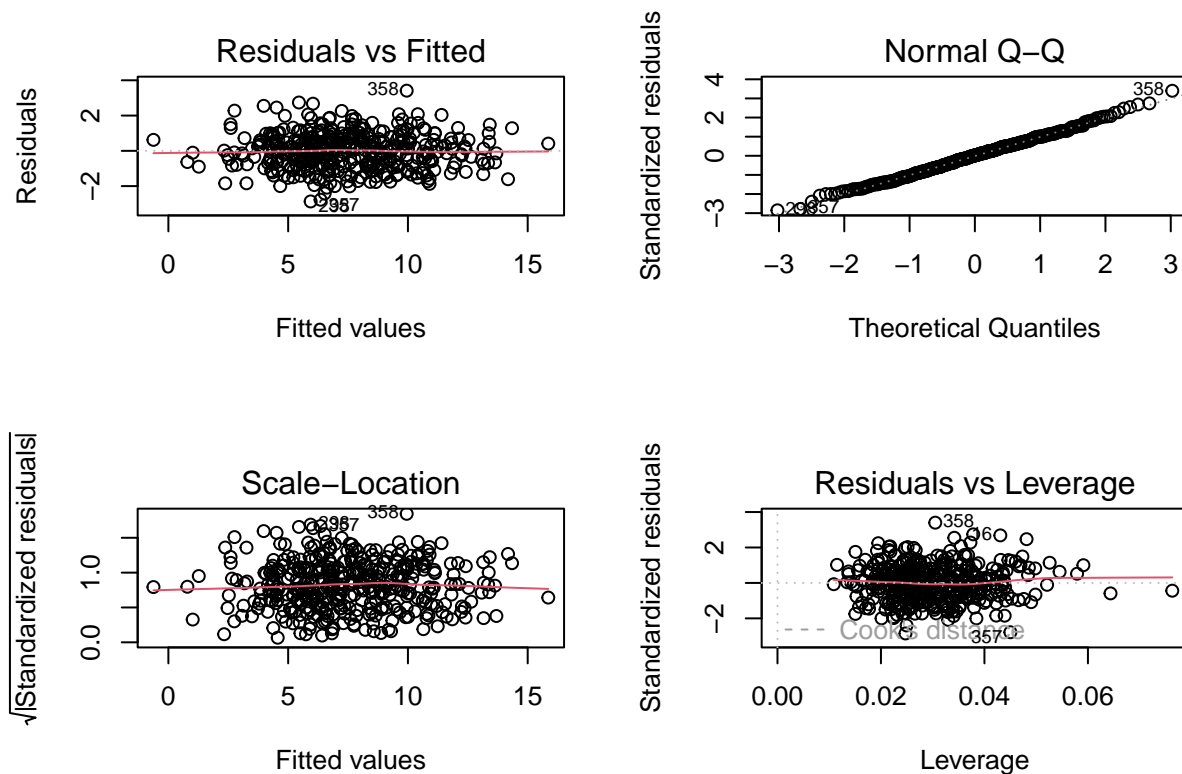
```
##   Sales CompPrice Income Advertising Population Price ShelveLoc Age Education
## 1  9.50      138     73         11         276   120        Bad   42         17
## 2 11.22      111     48         16         260    83        Good  65         10
## 3 10.06      113     35         10         269    80       Medium 59         12
## 4  7.40      117    100          4         466    97       Medium 55         14
## 5  4.15      141     64          3         340   128        Bad   38         13
## 6 10.81      124    113         13         501    72        Bad   78         16
##   Urban  US
## 1   Yes Yes
## 2   Yes Yes
## 3   Yes Yes
## 4   Yes Yes
## 5   Yes  No
## 6   No  Yes
```

```
model <- lm(Sales ~., data = Carseats)
model
```

```
##
## Call:
## lm(formula = Sales ~ ., data = Carseats)
##
## Coefficients:
##   (Intercept)      CompPrice          Income      Advertising
##    5.6606231    0.0928153    0.0158028    0.1230951
##   Population          Price ShelveLocGood ShelveLocMedium
##    0.0002079   -0.0953579    4.8501827    1.9567148
##           Age      Education      UrbanYes           USYes
##   -0.0460452   -0.0211018    0.1228864   -0.1840928
```

1. b)

```
par(mfrow = c(2,2))
plot(model)
```



From the plots above, we see that the model is not violating any assumptions such as linearity or normality. The linear model should be appropriate.

1. c)

```
summary(model)
```

```
##
## Call:
## lm(formula = Sales ~ ., data = Carseats)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -2.8692 -0.6908  0.0211  0.6636  3.4115
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)   5.6606231   0.6034487   9.380  < 2e-16 ***
## CompPrice     0.0928153   0.0041477  22.378  < 2e-16 ***
## Income        0.0158028   0.0018451   8.565 2.58e-16 ***
## Advertising   0.1230951   0.0111237  11.066  < 2e-16 ***
## Population    0.0002079   0.0003705   0.561   0.575
## Price        -0.0953579   0.0026711 -35.700  < 2e-16 ***
## ShelfLocGood  4.8501827   0.1531100  31.678  < 2e-16 ***
## ShelfLocMedium 1.9567148   0.1261056  15.516  < 2e-16 ***
```

```
## Age          -0.0460452  0.0031817 -14.472 < 2e-16 ***
## Education    -0.0211018  0.0197205  -1.070   0.285
## UrbanYes     0.1228864  0.1129761   1.088   0.277
## USYes        -0.1840928  0.1498423  -1.229   0.220
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 1.019 on 388 degrees of freedom
## Multiple R-squared:  0.8734, Adjusted R-squared:  0.8698
## F-statistic: 243.4 on 11 and 388 DF,  p-value: < 2.2e-16
```

The null hypothesis is that the coefficients for CompPrice and Income are equal to zero. The alternative hypothesis is that the coefficients for CompPrice and Income are not equal to zero. The test statistic is the t-test statistic which has a normal distribution. An appropriate significance level is 0.05.

From the table above, we see that the p-value for CompPrice and Income are both below the significance level and we reject the null hypothesis.

2. a)

```
sample <- sample.int(n = nrow(Carseats), size = floor(0.8*nrow(Carseats)), replace = F)
train = Carseats[sample,]
nrow(train)
```

```
## [1] 320
```

```
test = Carseats[-sample,]
nrow(test)
```

```
## [1] 80
```

The proportions for the train/test split are 80/20.

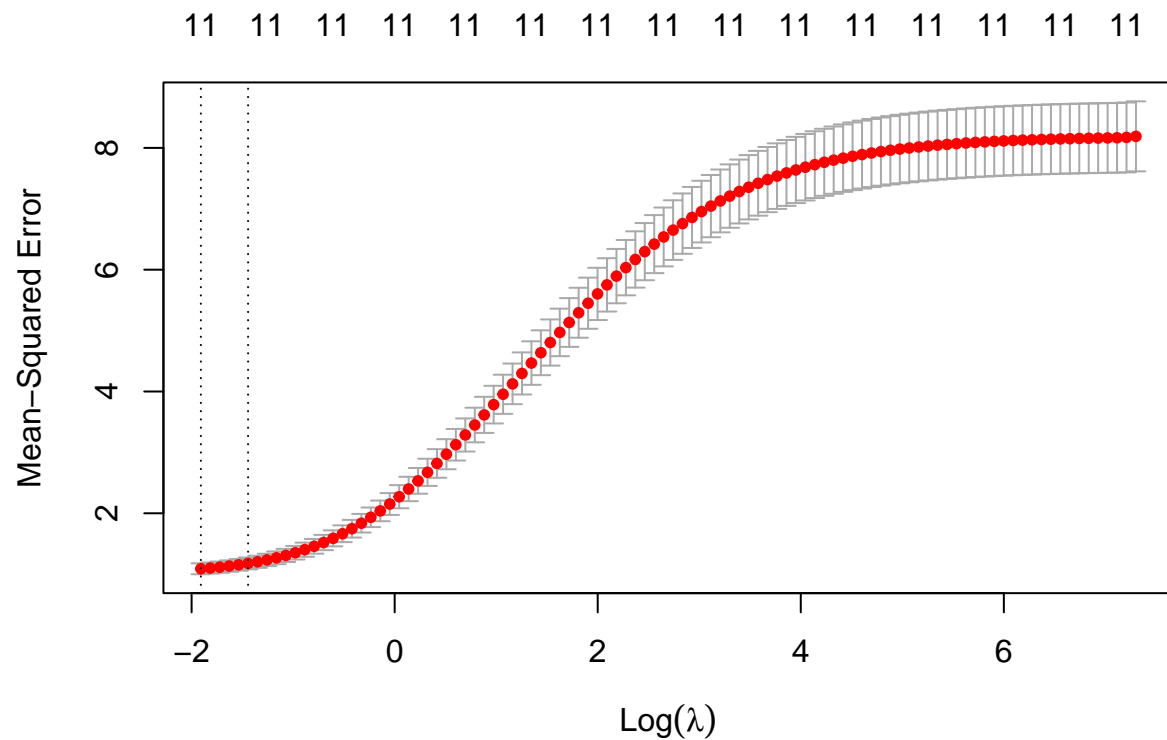
2. b)

```
library(glmnet)
```

```
## Loading required package: Matrix
```

```
## Loaded glmnet 4.1-7
```

```
x <- model.matrix(Sales ~ ., train)[, -1]
y <- train$Sales
set.seed(1)
cv.out <- cv.glmnet(x, y, alpha = 0)
plot(cv.out)
```



```
bestlam <- cv.out$lambda.min
bestlam
```

```
## [1] 0.1482363
```

```
coef(cv.out)
```

```
## 12 x 1 sparse Matrix of class "dgCMatrix"
##              s1
## (Intercept)  6.8501486799
## CompPrice    0.0721633471
## Income       0.0149284924
## Advertising  0.1020216646
## Population   0.0005230278
## Price        -0.0810258502
## ShelveLocGood 4.1239547228
## ShelveLocMedium 1.2942413609
## Age         -0.0408033573
## Education    -0.0212549890
## UrbanYes     0.0604243760
## USYes        0.0379632895
```

2. c)

```
x <- model.matrix(Sales ~ ., test)[-1]
data <- data.frame(pred = predict(cv.out, s = bestlam, newx = x), actual = test$Sales)
head(data)
```

```
##           s1 actual
## 2  12.140019  11.22
## 5   6.277755   4.15
## 6   9.818029  10.81
## 7   6.109526   6.63
## 20  7.498210   8.73
## 21  6.522399   6.41
```

```
sqrt(mean((data$actual - data$s1)^2))
```

```
## [1] 1.230363
```

2. d)

```
library(randomForest)
```

```
## randomForest 4.7-1.1
```

```
## Type rfNews() to see new features/changes/bug fixes.
```

```
library(Metrics)
set.seed(1)
rf <- randomForest(Sales ~ ., data = train, mtry = 10, ntree = 25, importance = TRUE)
rf
```

```
##
## Call:
## randomForest(formula = Sales ~ ., data = train, mtry = 10, ntree = 25,      importance = TRUE)
##           Type of random forest: regression
##           Number of trees: 25
## No. of variables tried at each split: 10
##
##           Mean of squared residuals: 2.841933
##           % Var explained: 64.99
```

```
importance(rf)
```

```
##           %IncMSE IncNodePurity
## CompPrice    9.0611605    241.371408
## Income       2.0756773    139.993412
## Advertising  5.6093425    211.799066
## Population   0.6957289     77.634353
## Price       13.6514033    815.261017
## ShelfLoc     16.3394765    740.199884
## Age          3.6688059    202.656870
## Education    3.6404284     71.252993
## Urban       -0.1668479      7.668564
## US           0.2569514     18.480809
```

```
rmse(test$Sales, predict(rf,test))
```

```
## [1] 1.858639
```

2. e) A marketing team may prefer the ridge regression model in (b) because it has a lower RMSE. Another marketing team may prefer the random forest model because it considers price as being important while the ridge regression model does not.
3. a)

```
set.seed(1)
X <- rt(200, 15)
summary(X)
```

```
##      Min.   1st Qu.   Median     Mean   3rd Qu.    Max.
## -2.268942 -0.665461 -0.008478  0.065234  0.759181  3.230585
```

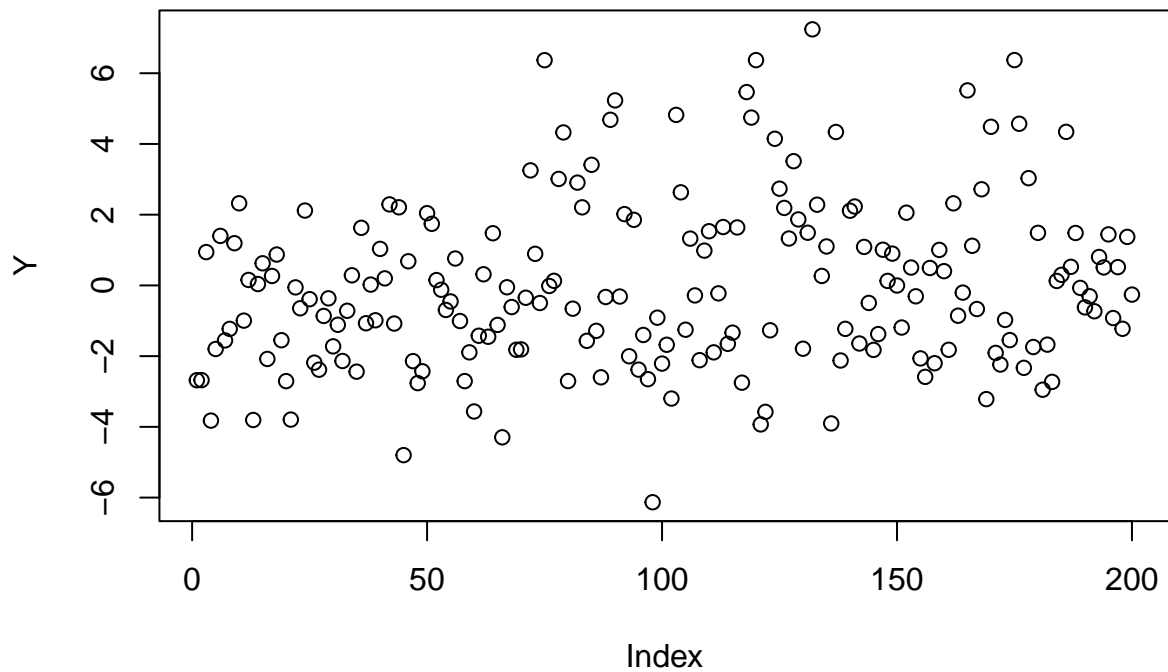
3. b)

```
noise <- rt(200, 5)
summary(noise)
```

```
##      Min.   1st Qu.   Median     Mean   3rd Qu.    Max.
## -4.675369 -0.815115 -0.010529  0.007735  0.756192  4.353930
```

3. c)

```
Y = 5 + 2 * sin(X) - 7 * ( exp(2 * cos(X)) / (1 + exp(2 * cos(X))) ) + noise
plot(Y)
```



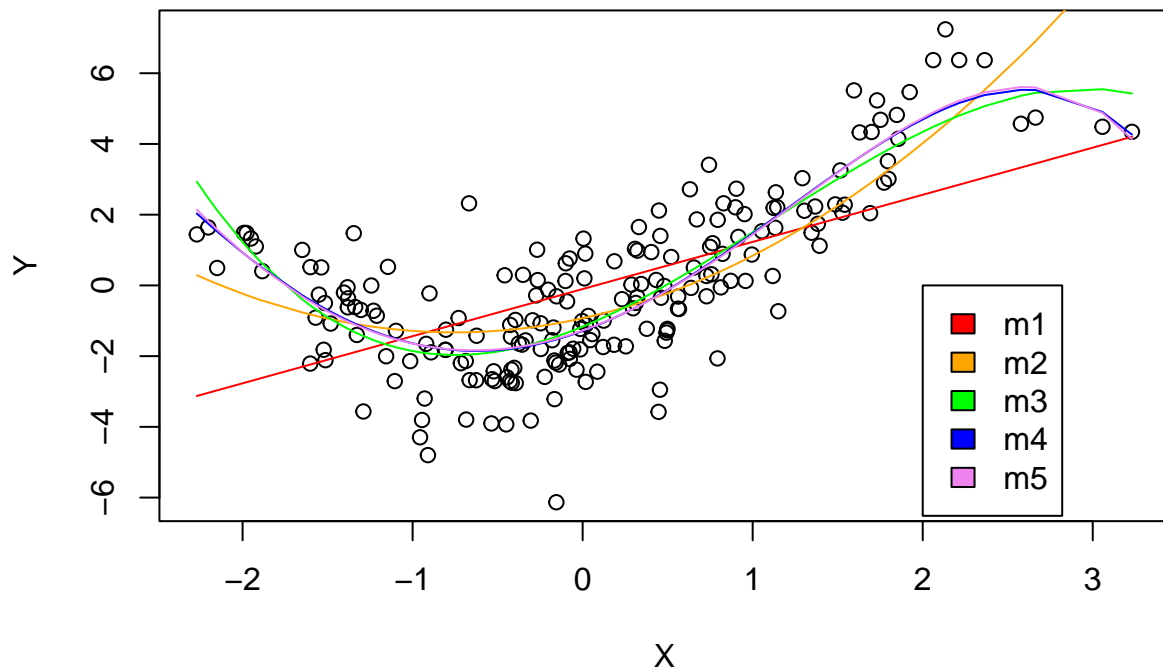
```
summary(Y)
```

```
##      Min.   1st Qu.   Median     Mean  3rd Qu.     Max.
## -6.13140 -1.72791 -0.30768 -0.01605  1.41048  7.24075
```

3. d)

```
df <- data.frame(Y,X)

plot(X, Y)
color <- c("red","orange","green","blue","violet")
for (index in c(0:4))
{
  m <- lm(Y ~ poly(X, index + 1, raw = TRUE), data = df)
  c <- color[index + 1]
  x<-sort(X)
  y<-m$fitted.values[order(X)]
  lines(x, y, col=c)
}
legend(2, y= 0, paste0("m", 1:5), fill=color)
```



3. e) I prefer the model with X to the order of 2 or 3. They neither under-fitted like the linear $m1$ nor over-fitted like $m4$ and $m5$, which barely differ from each other.

f)

```
m2 <- lm(Y ~ poly(X, 2, raw = TRUE), data = df)
predict(m2, newdata = data.frame(X=c(1)), interval = 'confidence')
```

```
##          fit      lwr      upr
## 1 0.8402292 0.5580371 1.122421
```

We are 90% confident that Y is between $[0.5580371, 1.122421]$ when $X = 1$.

3. g)

```
library(boot)
fun <- function(data, idx)
{
  d <- data[idx, ]
  m2 <- lm(Y ~ poly(X, 2, raw = TRUE), data = d)
  predict(m2, newdata = data.frame(X=c(1)))
}
bootstrap <- boot(df, fun, R = 1000)
boot.ci(boot.out = bootstrap, type = c("norm"))
```



```
## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
## Based on 1000 bootstrap replicates
##
## CALL :
## boot.ci(boot.out = bootstrap, type = c("norm"))
##
## Intervals :
## Level      Normal
## 95%      ( 0.5359,  1.1099 )
## Calculations and Intervals on Original Scale
```

We are 90% confident that Y is between [0.5359, 1.1099] when $X = 1$.

4. a)

```
data(College)
head(College)
```

```
##               Private Apps Accept Enroll Top10perc Top25perc
## Abilene Christian University    Yes 1660   1232    721         23         52
## Adelphi University              Yes 2186   1924    512         16         29
## Adrian College                  Yes 1428   1097    336         22         50
## Agnes Scott College              Yes  417    349    137         60         89
## Alaska Pacific University        Yes  193    146     55         16         44
## Albertson College                Yes  587    479    158         38         62
##               F.Undergrad P.Undergrad Outstate Room.Board Books
## Abilene Christian University      2885         537    7440      3300    450
## Adelphi University                2683        1227   12280      6450    750
## Adrian College                    1036          99   11250      3750    400
## Agnes Scott College                510          63   12960      5450    450
## Alaska Pacific University          249         869    7560      4120    800
## Albertson College                  678          41   13500      3335    500
##               Personal PhD Terminal S.F.Ratio perc.alumni Expend
## Abilene Christian University      2200    70      78     18.1         12    7041
## Adelphi University                1500    29      30     12.2         16   10527
## Adrian College                    1165    53      66     12.9         30    8735
## Agnes Scott College                875    92      97      7.7         37   19016
## Alaska Pacific University          1500    76      72     11.9          2   10922
## Albertson College                  675    67      73      9.4         11    9727
##               Grad.Rate
## Abilene Christian University      60
## Adelphi University                56
## Adrian College                    54
## Agnes Scott College                59
## Alaska Pacific University          15
## Albertson College                  55
```

```
set.seed(1)
sample <- sample.int(n = nrow(College), size = floor(0.8*nrow(College)), replace = F)
train = College[sample,]
nrow(train)
```

```
## [1] 621
```

```
test = College[-sample,]  
nrow(test)
```

```
## [1] 156
```

4. b)

```
logreg <- glm(Private ~ ., train,family="binomial")  
logreg
```

```
##  
## Call: glm(formula = Private ~ ., family = "binomial", data = train)  
##  
## Coefficients:  
## (Intercept)      Apps      Accept      Enroll      Top10perc      Top25perc  
##  7.607e-02   -4.834e-04   7.605e-04   6.249e-04   1.576e-03   4.751e-03  
## F.Undergrad P.Undergrad      Outstate      Room.Board      Books      Personal  
## -7.738e-04   1.876e-04   6.896e-04   2.002e-05   1.519e-03  -7.255e-05  
##      PhD      Terminal      S.F.Ratio      perc.alumni      Expend      Grad.Rate  
## -5.717e-02  -3.445e-02  -5.199e-02   4.593e-02   2.056e-04   1.652e-02  
##  
## Degrees of Freedom: 620 Total (i.e. Null);  603 Residual  
## Null Deviance:      712.9  
## Residual Deviance: 186.5      AIC: 222.5
```

The statistic of Top10perc is the percentage of new students being from the top 10% of high school classes. The coefficient for Top10perc can be understood as how important this statistic is as a factor of a college being public or private. Currently, it seems the percentage of new students from the top 10% of high school classes is not an important factor in whether a college is public or private.

4. c)

```
prob <- predict(logreg,newdata = test, type = "response")  
predicted <- ifelse(prob > 0.5, "Yes", "No")  
1 - mean(predicted == test$Private)
```

```
## [1] 0.05769231
```

4. d)

```
library(MASS)
```

```
##  
## Attaching package: 'MASS'  
  
## The following object is masked from 'package:ISLR2':  
##  
## Boston
```

```
lda_model = lda(Private ~ ., train)
predicted <- predict(lda_model,newdata = test)$class
1 - mean(predicted == test$Private)
```

```
## [1] 0.03846154
```

4. e)

```
qda_model = qda(Private ~ ., train)
predicted <- predict(qda_model,newdata = test)$class
1 - mean(predicted == test$Private)
```

```
## [1] 0.07692308
```

4. f)

```
library(e1071)
svm_model <- svm(Private ~ ., train)
predicted <- predict(svm_model,newdata = test)
1 - mean(predicted == test$Private)
```

```
## [1] 0.05128205
```

4. g) I picked the LDA model because it has the lowest test error.

5. a)

```
library(MultBiplotR)
```

```
##
## Attaching package: 'MultBiplotR'

## The following object is masked from 'package:MASS':
##
##      ginv

## The following object is masked from 'package:boot':
##
##      logit
```

```
data(Protein)
head(Protein)
```

```
##           Comunist Region Red_Meat White_Meat Eggs Milk Fish Cereal Starch
## Albania      Yes  South    10.1      1.4  0.5  8.9  0.2  42.3   0.6
## Austria      No   Center     8.9      14.0  4.3 19.9  2.1  28.0   3.6
## Belgium      No   Center    13.5      9.3  4.1 17.5  4.5  26.6   5.7
## Bulgaria     Yes  South     7.8      6.0  1.6  8.3  1.2  56.7   1.1
## Czechoslovakia Yes  Center     9.7     11.4  2.8 12.5  2.0  34.3   5.0
```

```
## Denmark          No North    10.6      10.8  3.7 25.0  9.9   21.9    4.8
##                  Nuts Fruits_Vegetables
## Albania          5.5          1.7
## Austria          1.3          4.3
## Belgium          2.1          4.0
## Bulgaria         3.7          4.2
## Czechoslovakia   1.1          4.0
## Denmark          0.7          2.4
```

```
p <- subset(Protein, select = -c(Comunist,Region))
pca = prcomp(p, scale. = TRUE, rank. =5)
summary(pca)
```

```
## Importance of first k=5 (out of 9) components:
##              PC1    PC2    PC3    PC4    PC5
## Standard deviation    2.0016 1.2787 1.0620 0.9771 0.68106
## Proportion of Variance 0.4452 0.1817 0.1253 0.1061 0.05154
## Cumulative Proportion 0.4452 0.6268 0.7521 0.8582 0.90976
```

5. b)

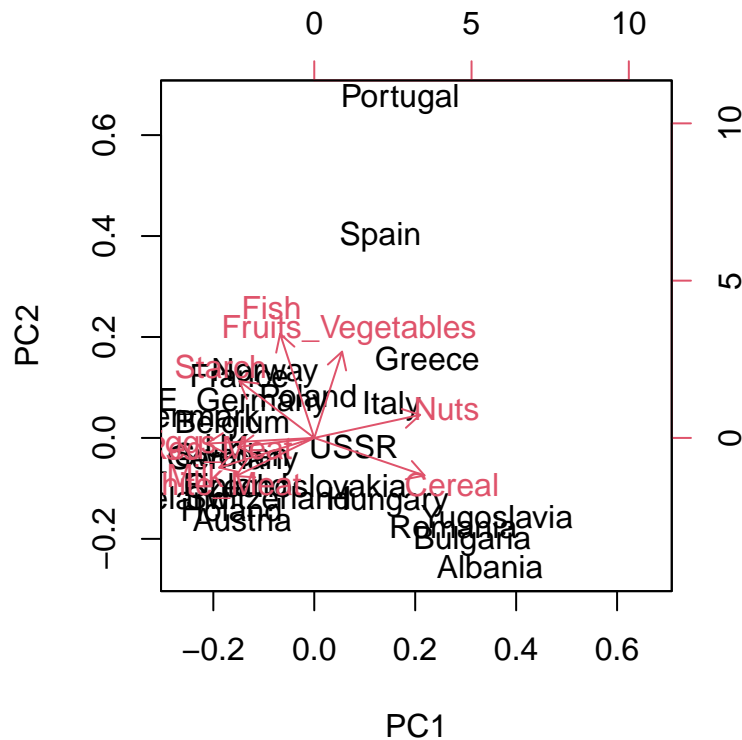
```
pca
```

```
## Standard deviations (1, ..., p=9):
## [1] 2.0016087 1.2786710 1.0620355 0.9770691 0.6810568 0.5702026 0.5211586
## [8] 0.3410160 0.3148204
##
## Rotation (n x k) = (9 x 5):
##              PC1          PC2          PC3          PC4          PC5
## Red_Meat      -0.3026094 -0.05625165 -0.29757957 -0.646476536  0.32216008
## White_Meat    -0.3105562 -0.23685334  0.62389724  0.036992271 -0.30016494
## Eggs          -0.4266785 -0.03533576  0.18152828 -0.313163873  0.07911048
## Milk          -0.3777273 -0.18458877 -0.38565773  0.003318279 -0.20041361
## Fish          -0.1356499  0.64681970 -0.32127431  0.215955001 -0.29003065
## Cereal         0.4377434 -0.23348508  0.09591750  0.006204117  0.23816783
## Starch        -0.2972477  0.35282564  0.24297503  0.336684733  0.73597332
## Nuts           0.4203344  0.14331056 -0.05438778 -0.330287545  0.15053689
## Fruits_Vegetables 0.1104199  0.53619004  0.40755612 -0.462055746 -0.23351666
```

The first principle component has negative associations with non-fish meat and starch, while also having large positive associations with cereal and nuts. The second principle component has large positive associations with fish as well as fruits and vegetables. These two components measure different dietary habits.

5. c)

```
biplot(pca)
```



Based on the plot above, milk is most positively correlated with white meat, most negatively correlated with nuts, and uncorrelated with fish and fruits.

5. d)

```
reg <- Protein[Protein$Region == 'North' | Protein$Region == "Center",]
subset(reg, select = Region)
```

```
##          Region
## Austria    Center
## Belgium    Center
## Czechoslovakia Center
## Denmark     North
## E_Germany   Center
## Finland     North
## France      Center
## Hungary     Center
## Ireland     Center
## Holand      Center
## Norway      North
## Poland      Center
## Sweden      North
## Switzerland Center
## UK          Center
## USSR        Center
## W_Germany   Center
```

```
summary(pca)
```

```
## Importance of first k=5 (out of 9) components:
##              PC1      PC2      PC3      PC4      PC5
## Standard deviation    2.0016 1.2787 1.0620 0.9771 0.68106
## Proportion of Variance 0.4452 0.1817 0.1253 0.1061 0.05154
## Cumulative Proportion 0.4452 0.6268 0.7521 0.8582 0.90976
```

Countries in the north and central regions are grouped close together in the biplot. However, some countries in the north region such as Denmark and Norway are located higher on PC2 compared to countries in the center region. This suggests that countries in the north region have higher consumption of fish and fruits/vegetables.

Conceptual Problems

6. For linear regression, bootstrapping can help validate the model and its confidence intervals. For random forest, bagging uses bootstrapping to reduce variability, which is more helpful.
7. FEWR and FDR are about the rates of type I errors, which are false positives. Correcting for FEWR and FDR may decrease type I errors, but it will also increase type II errors, which are false negatives. This should not be done if the cost of false negatives is higher than the cost of false positives, such as covid test results.
8. Assumptions such as linearity and normality need to be checked because they affect the accuracy of the model. For example, if there is a pattern in the residual plot for a linear model, then it might not have a linear relationship and the model should not be used for inference or prediction.