Darach Miller

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Education

PhD Biology, New York University, New York, NY, USA Genomics and Systems Biology track, dissertation adviser David Gresham

B.S. Genetics, University of California at Davis, Davis, CA, USA

Minor in Quantitative Biology and Bioinformatics

September 2012

- May 2018

September 2007

- December 2011

Research experience

Doctoral research, Gresham lab, New York University

mRNA stability changes can be used to effect rapid changes in gene expression in *Saccharomyces cerevisiae* during environmental changes. I used transcript dynamics modeling to infer candidate mRNAs that are destabilized during a nitrogen upshift, and adapted 4-thiouracil labelling methods to demonstrate that the destabilization occurs using a pulse-chase qPCR experiment. I then scaled the method up to use RNAseq and made some analytical advances to demonstrate that the destabilization is more general than the Nitrogen-Catabolite-Repression regulon. To determine the *trans* genetic factors of this, I redesigned a Fluorescent In-Situ Hybridization protocol to measure mRNA FISH with flow cytometry, then redesigned barcode-sequencing protocols to more robustly handle low-input fixed cell samples with a more advanced error-correcting amplicon design, then combined these approaches with Sort-Seq modeling to estimate mRNA abundance dynamics for 3,300 mutants in a pooled library. This assay confirmed the role of previously known mRNA synthesis control, the central role of the Lsm1-7p/Pat1p complex in eukaryotic mRNA degradation, and pointed towards translation initiation events mediated by elements in the 5' UTR as contributing to the mRNA destabilization.

Summer 2013 - current

Summer internship, Baliga Lab, Institute for Systems Biology

I worked to develop a high-throughput forward-genetic transposon-sequencing screen for *Halobacterium salinarum*. My contribution was improving the efficiency of haloarchaea transformation, so I adapted lipofection reagents from mammalian cell-culture systems to improve transformation efficiency, and characterized the efficiency of the *in-vitro* transposition reaction.

Undergraduate research, Facciotti Lab, University of California at Davis

Haloarchaea express genes that facilitate competition via secreted anti-microbial compounds. I designed and carried out a 60x60 all vs. all inhibition screen via spot assays, to characterize the network of pair-wise interactions. I then used BLAST on their recently completed genome assemblies to identify homologs that were associated with profiles of inhibitory effects.

Fall 2010 - Spring 2012

Summer 2011

Training

CSHL Yeast Genetics and Genomics Course

Three-week intensive course on the methods and practices of yeast genetics, instructed by Brown, Dunham, and Gartenberg.

The Art & Craft of Teaching

Semester-long course on the theory and practice of university teaching, especially focused on engaging students in active learning and using assessment effectively.

Fall 2015

Summer 2015

Awards

NYU Departmental Kopac Teaching Award 2013-2014 Annual teaching award, here for work on graduate-level Applied Genomics course NYU Departmental Kopac Service Award 2014-2015 For making a strong contribution to the department's academic environment, in organization of peer-to-peer skills development workshops Publications Systematic identification of factors mediating accelerated mRNA 2018 degradation in response to changes in environmental nitrogen PLoS Genetics, doi.org/10.1371/journal.pgen.1007406 Darach Miller. Nathan Brandt. David Gresham An upshift in growth upon nitrogen re-feeding is associated with destabilization of a functionally enriched set of mRNA. I measured the extent of this using label-chase and RNA sequencing to track mRNA stability across the upshift, and composed mRNA FISH with FACS and barcodesequencing to estimate the dynamics of GAP1 mRNA repression for 3,230 mutant strains of yeast. Guided by this, we find that modulators of translation initiation and a deletion of the 5' UTR of GAP1 are associated with defective abundance dynamics. Steady-state and dynamic gene expression programs in response to 2016 variation in environmental nitrogen Molecular Biology of the Cell, doi.org/10.1091/mbc.E14-05-1013 Edoardo Airoldi, Darach Miller, Rodoniki Athanasiadou, Nathan Brandt, Farah Abdul-Rahman, Benjamin Neymotin, Tatsu Hashimoto, Tayebeh Bahmani, David Gresham We explored the transcriptome responses of budding yeast to changes in environmental nitrogen. I used mRNA dynamics modeling and 4-thiouracil pulse-chase labeling with qPCR to identify and confirm some mRNA subject to destabilization upon a re-feeding of nitrogen. Growth Rate-Dependent Global Amplification of Gene Expression Preprint, revising Pre-print on bioRxiv, doi.org/10.1101/044735 for resubmission Niki Athanasiadou, Benjamin Neymotin, Nathan Brandt, Darach Miller, Daniel Tranchina, David Gresham We explored the scaling of the yeast transcriptome with growth rate, using a novel statistical normalization method for using spike-ins with RNAseq. To confirm these results, I designed, optimized, and used a fluorescent poly-dT probe flow cytometry assay as orthogonal confirmation. Presentations Accelerated mRNA Degradation Contributes to Gene Expression Remodeling During a August 2017 Nitrogen Upshift Selected talk @ 2017 International Conference of Yeast Genetics and Molecular Biology High-throughput Genetics of mRNA Dynamics using FISH, FACS, and Sequencing August 2017 Selected talk @ New York Area Meeting in Quantitative Biology Genetic factors controlling accelerated mRNA degradation during a nitrogen upshift July 2016 (video) Selected talk @ GSA's TAGC16 in Orlando Screening and Investigating Interactions Between Haloarchaeal Species April 2011 Talk @ UCDavis Undergraduate Research Conference

Posters

mRNA stability regulation accelerates functional reprogramming of the yeast August 2017 transcriptome during a nitrogen upshift CSHL Eukaryotic mRNA processing meeting, ICYGMB 2017 meeting Accelerated mRNA degradation contributes to gene expression remodeling during a September 2015 nitrogen upshift ICYGMB annual meeting. Teaching Quantitative Methods in Human Genetics Spring 2015, Undergraduate-level course Spring 2016 Designed and taught introductory R to explore examples of relevant statistical methods Applied Genomics: Intro. to Bioinf. & Network Modeling Fall 2013 Graduate-level course Designing and running practice in the analysis of genomics datasets using techniques such as network clustering and transcript dynamics modeling. Principles of Biology Fall 2012 Undergraduate-level course Coordinate exercises in benchwork and data analysis of basic molecular biology and genetics. Mentoring Daniel Pham, Undergraduate at North Park University Summer 2017 Mentored as part of 2017 Summer Undergraduate Research Program Finishing undergraduate studies Fall 2016 Alex Ferrena, Undergraduate at New York University Former student, mentored his transition into bioinformatics Starting MS Bioinformatics program at Columbia Stephen Nyarko, Student at Stuyvesant High School Fall 2015, Mentored design of a yeast genetic screen for a science competition Summer 2016 Starting undergraduate studies at Harvard Other experience Social media coordinator, Yeast Course, Cold Spring Harbor Laboratory 2017 Working to build a social media presence for the the CSHL Yeast Course. Building an alumni list and connecting alumni across years to help attendees and boost recruitment of new attendees from outside the community. Research technician, van Winkle Lab, UC Davis 2008 - 2009 Microscopy sample preparation (mouse dissection via microtome). Bike mechanic/instructor, Davis Bike Collective 2007 - 2012 Volunteer mechanic/instructor and shift supervisor at community bike shop. Lots of teaching and reconciling bicycle theory with the extensive natural variation in design and function. Never met a seatpost I couldn't extract.