

SL

pH Titration

Microscopy

Chloride and pH calibration in Hek cells

Materials

Ensure the following materials are available before starting the procedure:

- Transfection kit: Qiagen Effectene
- Plasmid DNA: Midi or mini transfection-grade

Procedure

- Day 1 - Cell Seeding
 - a. Seed 25,000 HEK cells per well in a Labtec chamber with 0.5 mL of culture medium.
- Day 2: Transfection (6-hour protocol)
 - a. Mix 0.3 µg of DNA with 50 µL EC buffer in a clean microcentrifuge tube.
 - b. Add 2.4 µL of Enhancer, vortex gently, and incubate for 5 minutes at room temperature.
 - c. Add 4 µL of Effectene, mix thoroughly, and incubate for 10 minutes at room temperature.
 - d. Add the transfection mixture to the cells dropwise and incubate for 6 hours.

| | 1 well | ... wells |
|-------------|--------|-----------|
| Plasmid DNA | 0.3 µg | |
| Enhancer | 2.4 µL | |
| EC buffer | 50 µL | |
| Effectene | 4 µL | |

- Days 3 & 4: Chloride and pH Calibration
 - a. Remove medium from one well carefully.

- b. Add 1 mL of the desired buffer containing ionophores (prepared fresh).
- c. Incubate the cells for 2 minutes, then change the buffer. Repeat this process 4 times.
- d. Begin imaging cells for chloride or pH calibration after buffer changes.
- e. For wells that are not being imaged immediately, add a drop of 50 mM Tris (pH 7.3) to maintain the cells.

Notes

- Ensure that buffers with ionophores are freshly prepared and kept at appropriate pH levels.
- Imaging should be performed promptly after the buffer changes to ensure accuracy in pH and chloride calibration.