SL pH Titration Microscopy

## Chloride and pH calibration in Hek cells

## **Materials**

Ensure the following materials are available before starting the procedure:

- Transfection kit: Qiagen Effectene
- Plasmid DNA: Midi or mini transfection-grade

## Procedure

- Day 1 Cell Seeding
  - a. Seed 25,000 HEK cells per well in a Labtec chamber with 0.5 mL of culture medium.
- Day 2: Transfection (6-hour protocol)
  - a. Mix  $0.3 \mu g$  of DNA with  $50 \mu L$  EC buffer in a clean microcentrifuge tube.
  - b. Add 2.4  $\mu L$  of Enhancer, vortex gently, and incubate for 5 minutes at room temperature.
  - c. Add  $4\,\mu\text{L}$  of Effectene, mix thoroughly, and incubate for 10 minutes at room temperature.
  - d. Add the transfection mixture to the cells dropwise and incubate for 6 hours.

	1 well	wells
Plasmid DNA	0.3 µg	
Enhancer	2.4 µL	
EC buffer	50 μL	
Effectene	4 μL	

- Days 3 & 4: Chloride and pH Calibration
  - a. Remove medium from one well carefully.

- b. Add 1 mL of the desired buffer containing ionophores (prepared fresh).
- c. Incubate the cells for 2 minutes, then change the buffer. Repeat this process 4 times.
- d. Begin imaging cells for chloride or pH calibration after buffer changes.
- e. For wells that are not being imaged immediately, add a drop of 50 mM Tris (pH 7.3) to maintain the cells.

## Notes

- Ensure that buffers with ionophores are freshly prepared and kept at appropriate pH levels.
- Imaging should be performed promptly after the buffer changes to ensure accuracy in pH and chloride calibration.