

A microscopic image of neurons, showing a large cell body with several branching processes. The image is in grayscale and has a soft, slightly blurred appearance. The text 'Using NEURON to model cells' is overlaid in a large, blue, sans-serif font, and 'Tutorial' is overlaid in a smaller, blue, sans-serif font below it.

Using NEURON to model cells

Tutorial

A dark blue, stylized logo consisting of a series of connected dots and lines, resembling a neural network or a brain structure, located in the bottom left corner of the slide.

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Blue Brain Project

Tutorial adapted from Oren Amsalem, Yoni Leibner (Idan Segev Lab) and András Ecker

Introduction

The [NEURON Simulation Environment](#) is designed for modeling individual neurons and networks of neurons.

It is particularly well-suited to explore problems which are closely linked to experimental data.

NEURON was built and is maintained by [Ted Carnevale](#) and [Michael Hines](#) at Yale. The Blue Brain Project is also heavily involved with the development of the software

[Neuron](#) is written in C & C++, the interface with the simulator is with HOC, but a Python wrapper was build and now commonly used instead of HOC (NEURON as a python package). In addition User-defined mechanisms such as voltage- and ligand-gated ion channels are used in order to expand NEURON (NMODL files, need to be compiled).

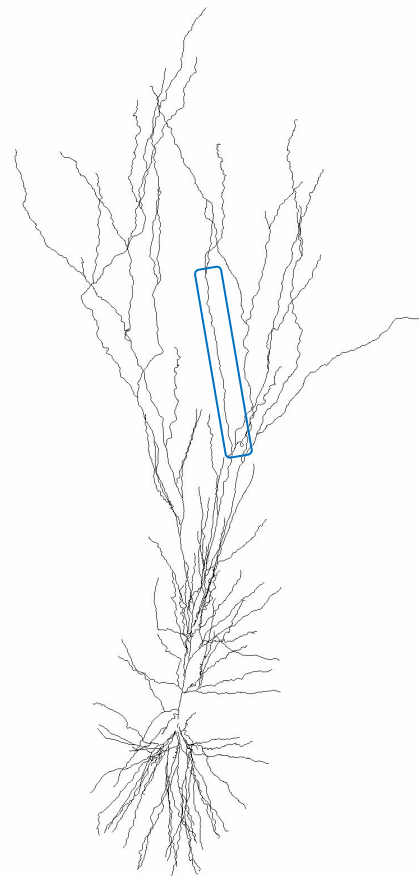
In this presentation we will show the following examples:

- Single compartment model
- Ball and Stick model and the replication of a published result (Gidon et al.)
- Multicompartmental L5 Pyramidal cell and two published results (Hay et al., Doron et al.)

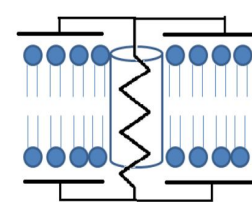
Introduction

Basic concepts of Neuron models:

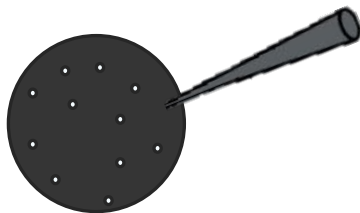
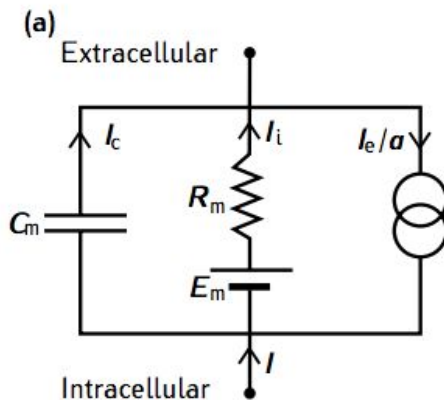
- **Model** - a Neuron object, it can have soma, basal dendrites (dend) and apical dendrite (apic).
- **Section** - more of a morphological definition, a section is usually a dendrite between 2 bifurcation points.
- **Segment** - one RC circuit that resemble (approximate) the computation in a piece of membrane.
- **Clamps** - an object that represent an experimental electrode.
- **Synapses** - object that resemble a single synapse (we will see it later, and how it's activated in the simulation).
- **Distributed Process**: generally ion channels spread on the membrane
- **Point Process** - a more general idea, it's an object that seats on the segment (RC circuit), and simulate clamps, synapses, etc.



Single compartment neuron - single RC circuit



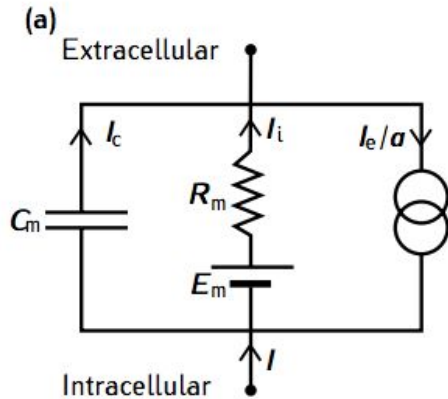
Neuron Cell Membrane
Equivalent Circuit



```
from neuron import h, gui
# create model
soma = h.Section(name="soma")
soma.L = 10 # length  $\mu\text{m}$ 
soma.diam = 10 # diameter  $\mu\text{m}$ 
soma.insert('pas'). # add passive properties
soma.g_pas = 1/10000 # set the specific membrane
                    # resistance to 10000 ohm*cm2
```

```
# current current clamp
stim = h.IClamp(soma(0.5))
stim.delay = 20. # start of the current
                # injection (ms)
stim.dur = 100 # duration (ms)
stim.amp = 0.01 # amplitude (nA)
```

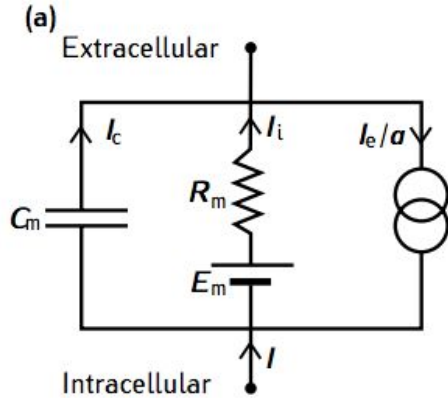
Single compartment neuron - recording



```
# record injected current soma voltage (and time)
soma_v = h.Vector()
soma_v.record(soma(0.5)._ref_v)
stim_current = h.Vector()
stim_current.record(stim._ref_i)
t = h.Vector()
t.record(h._ref_t)

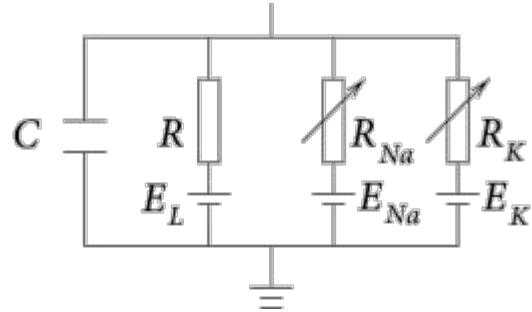
# run simulation
h.tstop = 220 # simulation time (ms)
h.dt = 0.025.
h.v_init = -70. # initial voltage (mV)
h.run()
```

Single compartment neuron - synapse



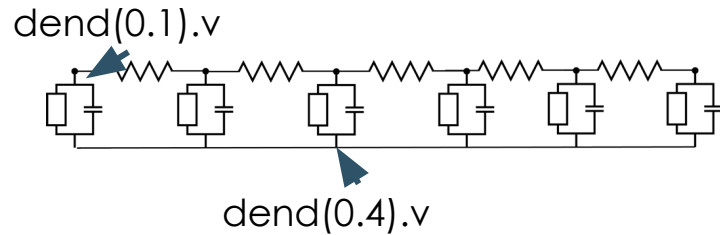
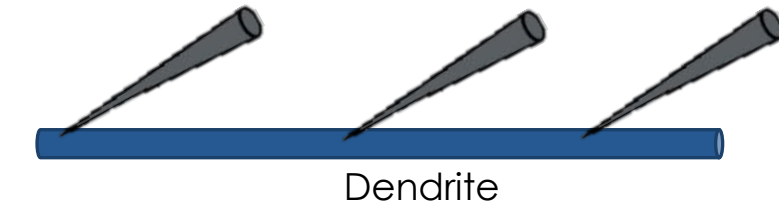
```
# add a synapse
synapse = h.Exp2Syn(soma(0.5))
synapse.tau1 = 0.3 # rise time constant
synapse.tau2 = 1.8 # decay time constant
stim = h.NetStim()
stim.number = 1
stim.noise = 0 # no noise
stim.interval = 1
net_con = h.NetCon(stim, synapse)
net_con.weight[0] = 0.0004 # the maximal
                             conductance
                             of the synapse
```

Single compartment neuron – active conductances

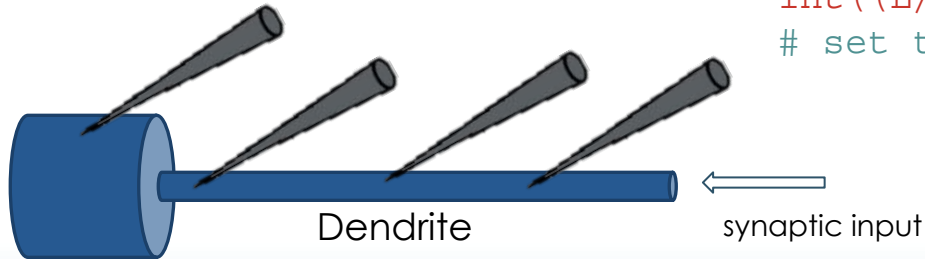


```
from neuron import h, gui
# create model
Soma = h.Section(name="soma")
soma.L = 10 # length  $\mu\text{m}$ 
soma.diam = 10 # diameter  $\mu\text{m}$ 
soma.insert("pas"). # add passive properties
soma.g_pas = 1/10000 # set the specific membrane
                    # resistance to 10000 ohm*cm2
soma.insert("kv") # add potassium channel
                # (from a mod file)
soma.gbar_kv = 2000 # set the potassium conductance
soma.insert("na") # add sodium channel
                # (from a mod file)
soma.gbar_na = 8000 # set the sodium conductance
h.celsius = 30. # set temperature
```

Moving towards the cable model



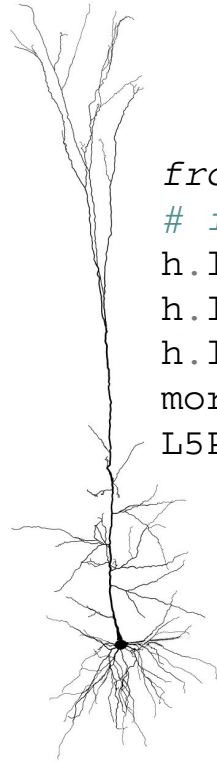
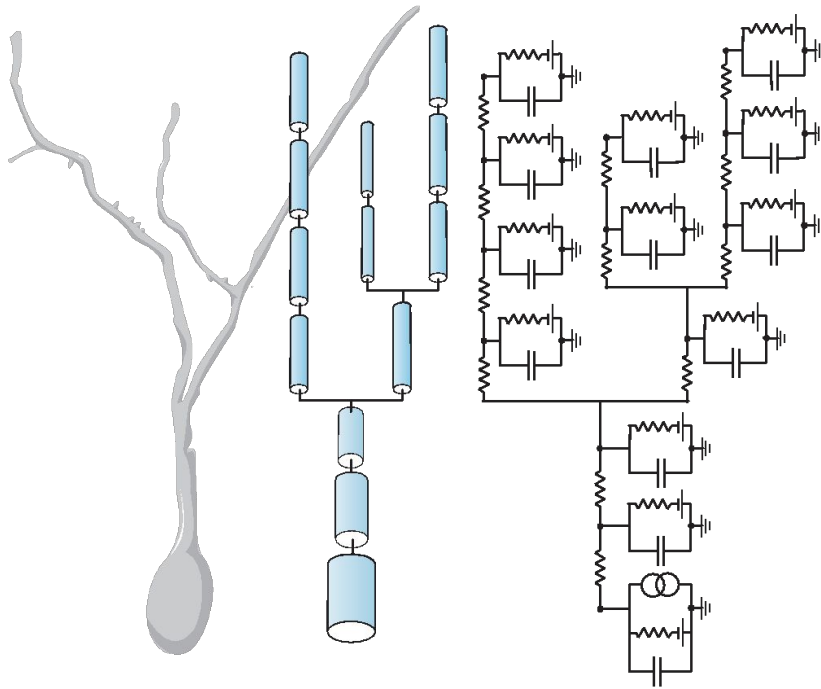
```
# create dendrite
dend = h.Section(name="dend")
dend.L = 500 # um
dend.diam = 1 # um
dend.Ra = 100 # ohm*cm
dend.insert("pas")
dend.g_pas = 1/10000
dend.connect(soma, 1, 0). # connect the end of
the dendrite to the beginning of the soma
h"forall { nseg =
int((L/(0.1*lambda_f(100))+0.9)/2)*2 + 1 }"
# set the number of segments
```



[2_soma_dend_step_current.ipynb](#)
[3_soma_dend_synapse.ipynb](#)

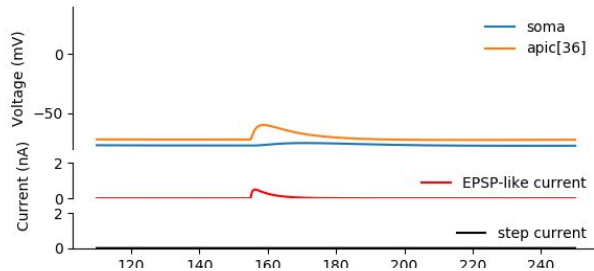
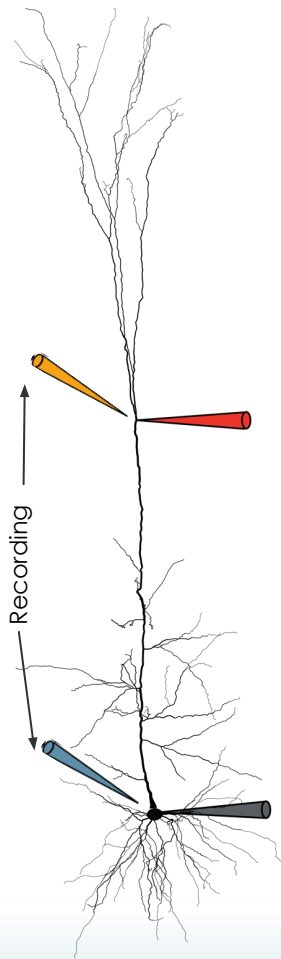
Note: d_lambda rule – [NEURON Book chapter 4](#)

Layer 5b Pyramidal Cell



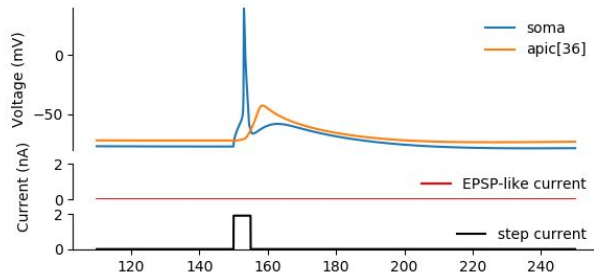
```
from neuron import h, gui
# instantiating cell
h.load_file("import3d.hoc")
h.load_file("models/L5PCbiophys3.hoc")
h.load_file("models/L5PCtemplate.hoc")
morph_fname = "morphologies/cell1.asc"
L5PC = h.L5PCtemplate(morph_fname)
```

4_L5_PC_Hay_et_al.ipynb



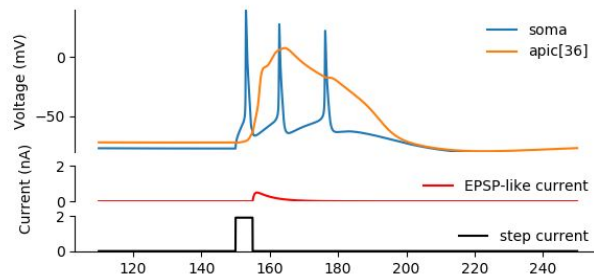
Only EPSP-like current

```
syn.imax = 0.5
stim.amp = 0
h.run();
plot_result(...)
```



Only small step current

```
syn.imax = 0
stim.amp = 1.9
h.run();
plot_result(...)
```

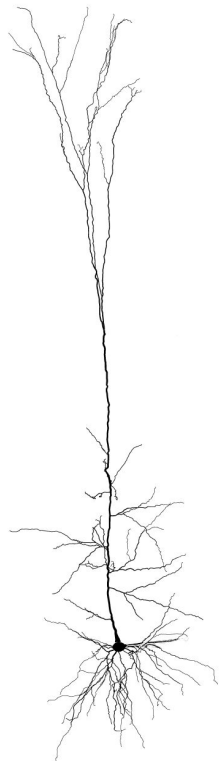


both currents

```
syn.imax = 0.5
stim.amp = 1.9
h.run()
plot_result(...);
```

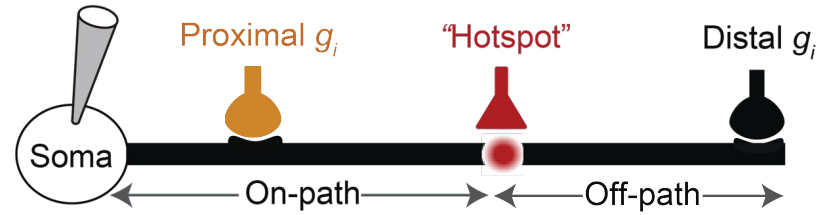
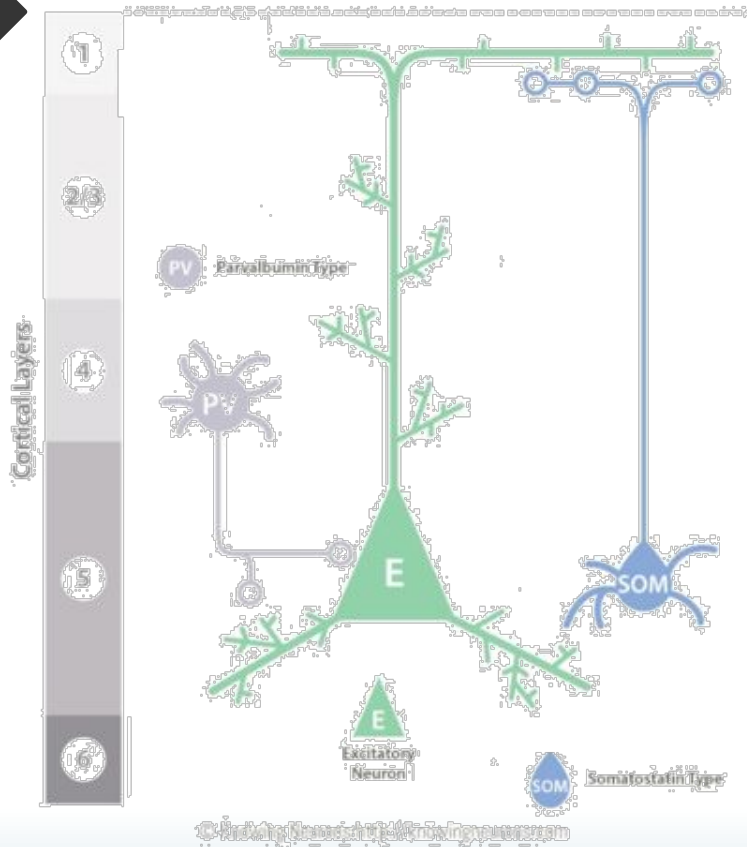
5_calcium_spike_Hay_et_al.ipynb

Layer 5b Pyramidal Cell



```
# place a single synapse
syn_loc = rng_pos.repick()
apical_exc_syns.append(h.ProbAMPANMDA_EMS(sec(syn_loc)))
# create random stim. times
apical_exc_netstims.append(h.NetStim())
apical_exc_netstims[-1].number = t_stop / 1000 * exc_freq
apical_exc_netstims[-1].interval = 1000 / exc_freq # ms
apical_exc_netstims[-1].noise = 1 # make it Poisson like
# connect stimulator to the synapse
apical_exc_netconns.append(h.NetCon(apical_exc_netstims[-1]
                                     ,
                                     apical_exc_syns[-1]))
apical_exc_netconns[-1].weight[0] = gsyn_exc
```

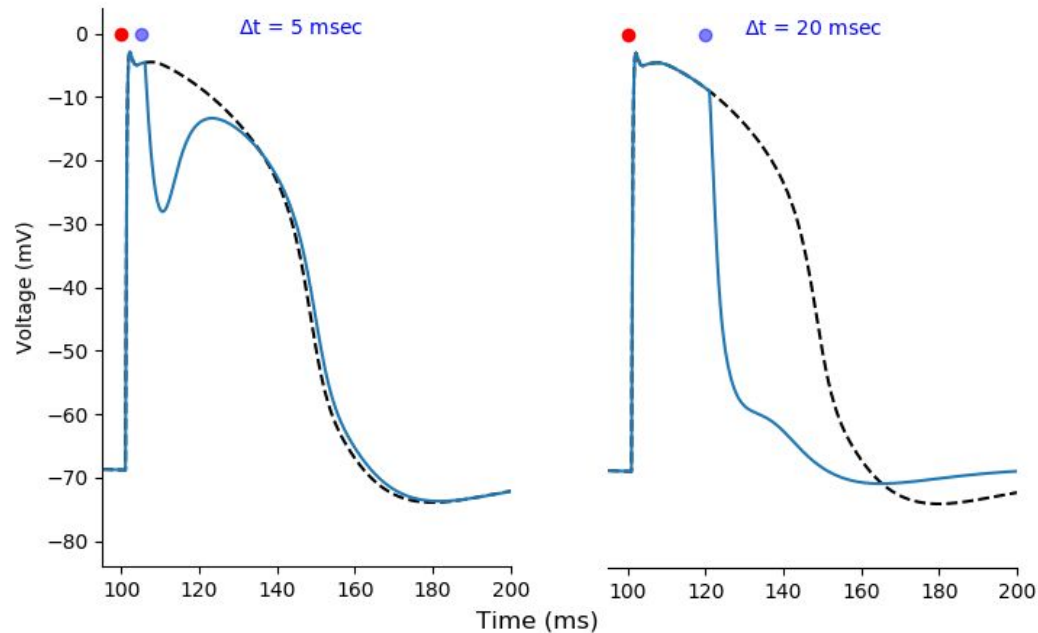
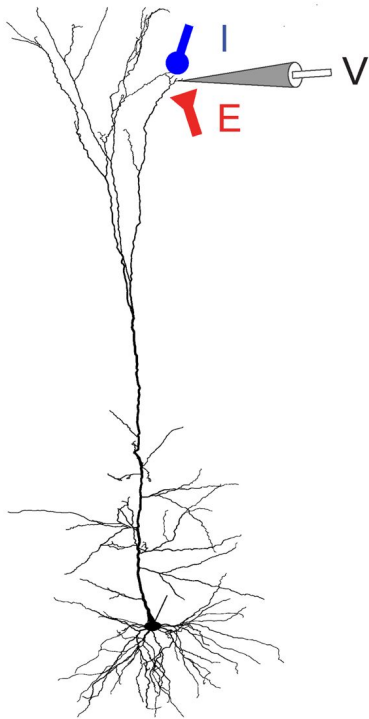
Inhibition location



[+1_inhibition_location_Gidon_et_al.ipynb](#)

"Principles Governing the Operation of Synaptic Inhibition in Dendrite" Albert Gidon, Idan Segev. Neuron, 2012

Timed inhibition



[+2_timed_inhibition_Doron_et_al.ipynb](#)

“Timed Synaptic Inhibition Shapes NMDA Spikes, Influencing Local Dendritic Processing and Global I/O Properties of Cortical Neurons” Michael Doron, Giuseppe Chindemi, Eilif Muller, Henry Markram, Idan Segev. Cell Reports, 2017

Useful links

- [NEURON](#): ([GitHub](#))
 - Documentation: [Python Doc](#), [HOC Doc](#), [ReadTheDocs](#)
 - More [scripting basics](#), [NEURON-Python tutorials](#), [courses](#) and [material](#)
 - Queries: [FAQs](#), [NEURON Forum](#) and [GitHub issues](#)
- Modeling repositories
 - [ModelDB](#) (published models)
 - [NeuroMorpho](#) (morphologies)
 - [Channelpedia](#) and [ICGenealogy](#) (ion channels)
 - [NeuroElectro](#) (neuron parameters)
- Others
 - [Blue Brain Tools](#) (for [subcellular](#), [cellular](#), [circuit](#)-level models among others)
 - [Allen Institute Data and Tools](#)
 - [NetPyNE](#) (modelling networks of neurons)
 - [CoreNEURON](#) (faster NEURON execution)
 - Free edx MOOCs: [Simulation Neuroscience](#), [Simulating a Hippocampus Microcircuit](#)

Useful NEURON Tools and Tips

- [NEURON-Python scripting Basics](#)
- [Units used in NEURON](#)
- [NEURON Tips](#) (mainly HOC)
- Specifying temperature in NEURON [h.celsius](#)
- Don't forget to [compile](#) new mod files before running NEURON
- [Randomness in NEURON models](#)
- NEURON [Sections](#) overview

Blue Brain Project Software

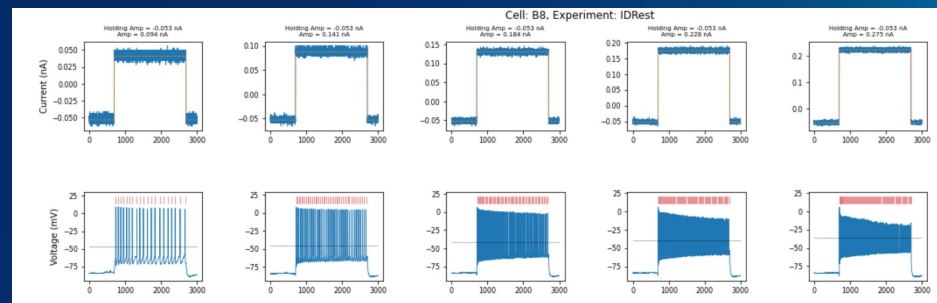
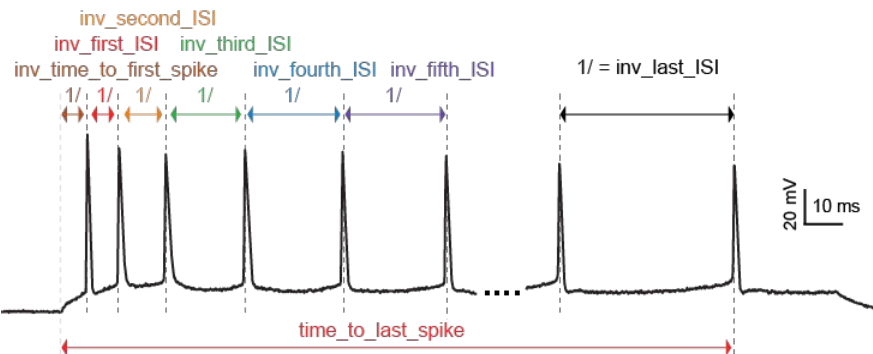
<https://github.com/BlueBrain>

eFEL

Electrophys Feature Extraction Library

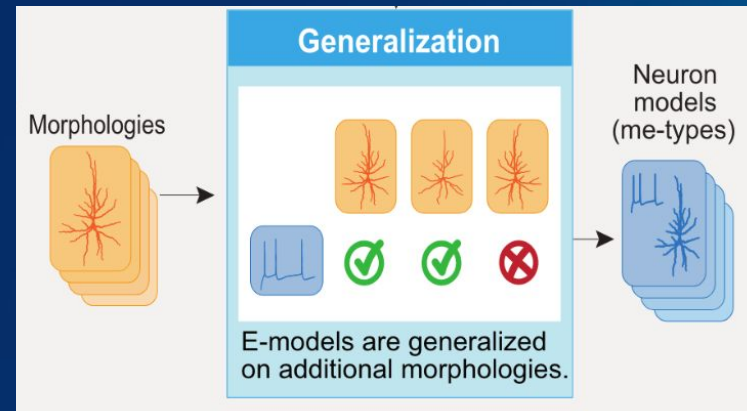
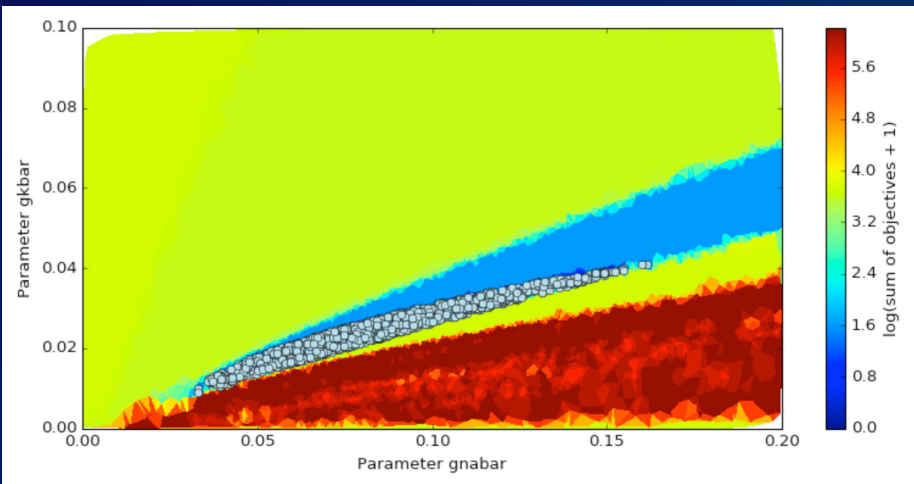
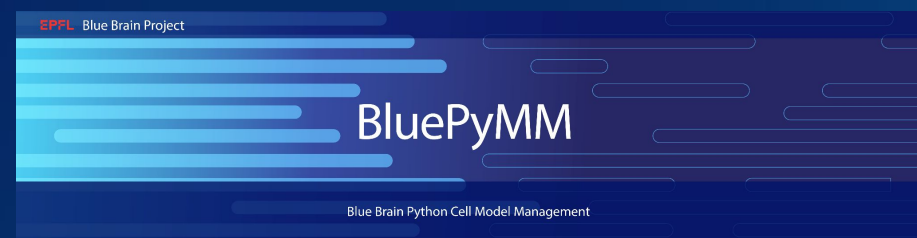
BluePyEfe

Blue Brain Python E-feature extraction



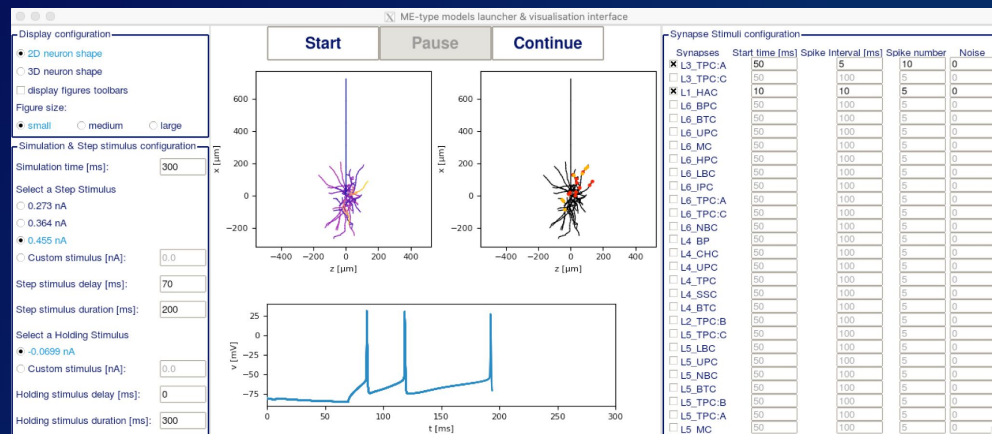
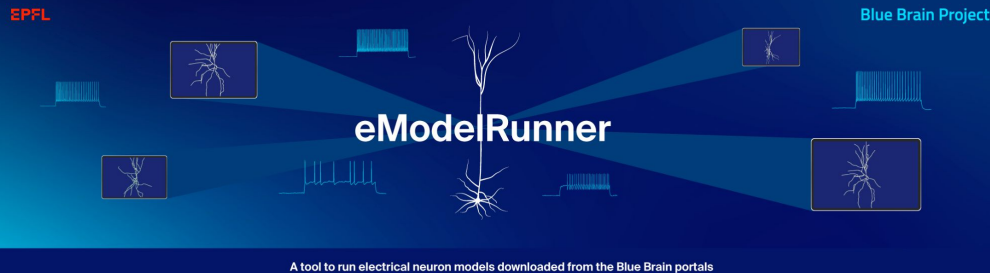
github.com/BlueBrain/eFEL

github.com/BlueBrain/BluePyEfe



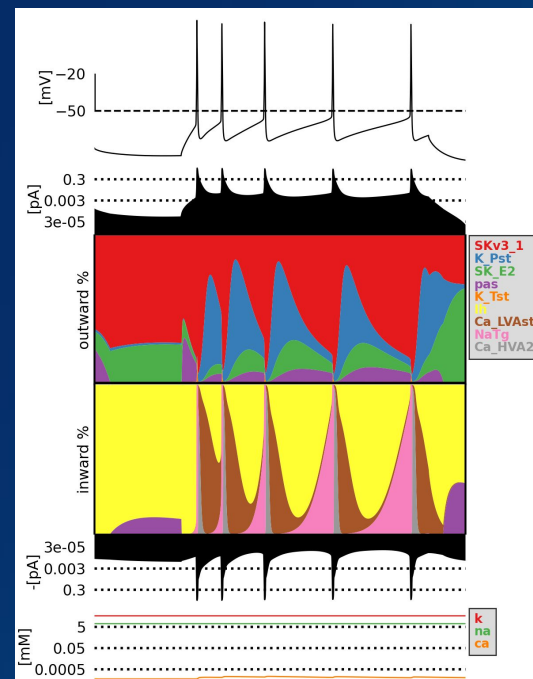
github.com/BlueBrain/BluePyOpt

github.com/BlueBrain/BluePyMM



github.com/BlueBrain/EModelRunner

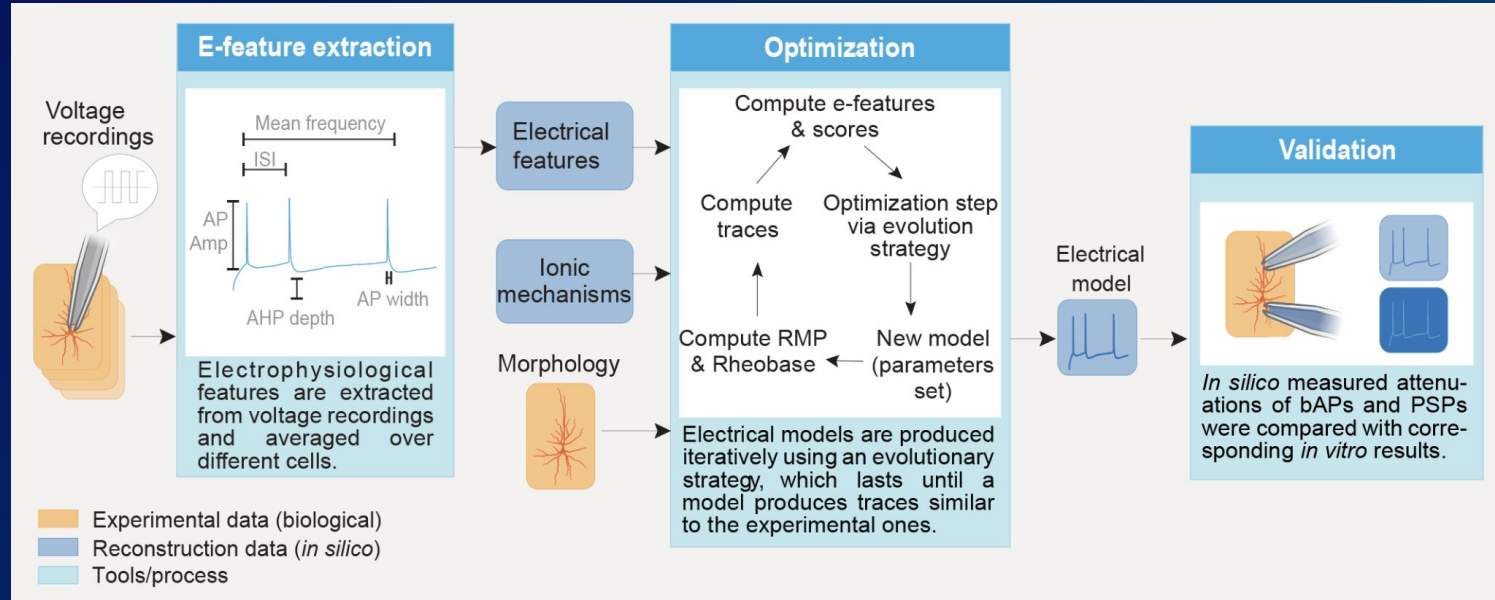
Currentscape



github.com/BlueBrain/Currentscape

BluePyEModel

- Unified detailed single cell model building software pipeline
- Uses eFEL, BluePyEfe, BluePyOpt and Currentscape

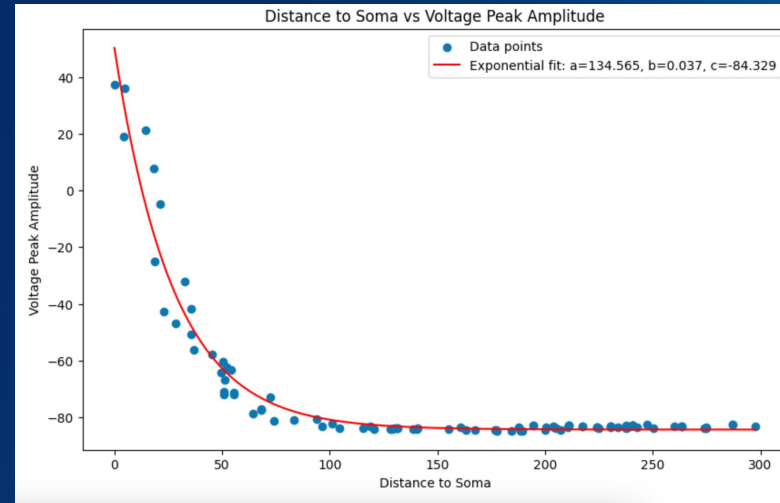
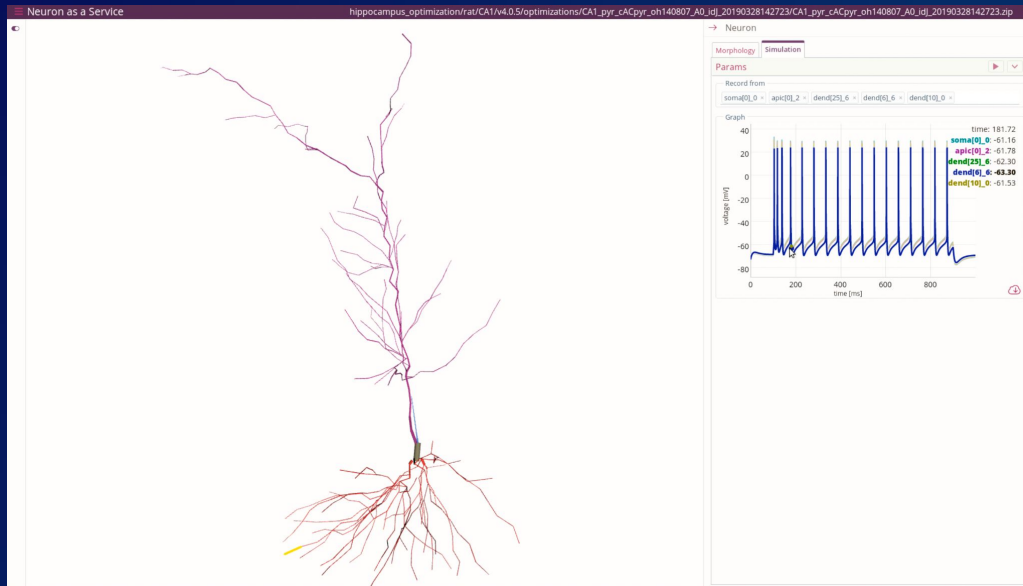


github.com/BlueBrain/BluePyEModel

BlueNaaS-SingleCell

Provides Web UI to access NEURON simulator in order to load the single cell models and run model simulations

BlueCellulab



github.com/BlueBrain/BlueNaaS-SingleCell

github.com/BlueBrain/BlueCelluLab

A graphic with a dark blue background. On the left, there are several horizontal blue bars of varying lengths. On the right, there is a grid of white lines forming a 3D cube-like structure. Inside and around this grid are small blue dots and lines, suggesting a network or data flow. The text "Single-Cell E-Model Suite" is written in white, bold, sans-serif font across the middle of the graphic.

Single-Cell E-Model Suite

- **BluePyEModel** — pipeline for building single cell e-models
- **eFEL** — Electrophys Feature Extraction Library
- **BluePyEfe** — E-feature extraction
- **BluePyOpt** — Parameter optimisation Library
- **BlueCelluLab** — Perform simulations on a single cell or group of cells
- **Currentscape** — Easily plot and visualize currents and ionic concentration in electrical neuron models
- **BluePyMM** — Model management for circuit building
- **EModelRunner** — Runs cells from stand-alone packages
- **BlueNaaS-SingleCell** — Interacts with single cell models through a web application

Thank You!