

ADS502 Group Project

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```
threshold <- 0.5
drug_induced_training_data <- drug_induced_training_data[, colMeans(is.na(drug_induced_training_data)) < threshold]

get_mode <- function(x) {
  uniq_vals <- unique(x)
  uniq_vals[which.max(tabulate(match(x, uniq_vals)))]
}
categorical_cols <- sapply(drug_induced_training_data, is.character)
drug_induced_training_data[categorical_cols] <- lapply(
  drug_induced_training_data[categorical_cols],
  function(x) {
    x[is.na(x)] <- get_mode(x)
    return(x)
  })
write.csv(drug_induced_training_data, "DIA_trainingset_RDKit_descriptors.csv")
```

Data Cleaning

```
summary(drug_induced_training_data[, 1:6])
```

Descriptive Statistics

| ## | Label | SMILES | BalabanJ | BertzCT |
|----|----------------|------------------|---------------|----------------|
| ## | Min. :0.0000 | Length:477 | Min. :0.986 | Min. : 8.0 |
| ## | 1st Qu.:0.0000 | Class :character | 1st Qu.:1.679 | 1st Qu.: 493.3 |
| ## | Median :0.0000 | Mode :character | Median :1.964 | Median : 712.4 |
| ## | Mean :0.2474 | | Mean :2.143 | Mean : 738.6 |
| ## | 3rd Qu.:0.0000 | | 3rd Qu.:2.419 | 3rd Qu.: 943.2 |
| ## | Max. :1.0000 | | Max. :5.083 | Max. :2430.9 |
| ## | Chi0 | Chi0n | | |
| ## | Min. : 3.414 | Min. : 1.725 | | |
| ## | 1st Qu.:13.405 | 1st Qu.:10.391 | | |
| ## | Median :17.646 | Median :14.184 | | |
| ## | Mean :18.130 | Mean :14.372 | | |
| ## | 3rd Qu.:22.052 | 3rd Qu.:17.730 | | |
| ## | Max. :50.120 | Max. :38.475 | | |

```
drug_data <- select(drug_induced_training_data, where(is.numeric))
drug_stats <- describe(drug_data)
head(drug_stats, 10)
```

```
##      vars    n  mean    sd median trimmed   mad  min    max   range skew
## Label      1 477   0.25   0.43   0.00   0.19   0.00 0.00    1.00    1.00  1.17
## BalabanJ    2 477   2.14   0.71   1.96   2.05   0.52 0.99    5.08    4.10  1.49
## BertzCT     3 477 738.63 392.97 712.42 720.01 336.93 8.00 2430.93 2422.93 0.79
## Chi0        4 477  18.13   7.25  17.65  17.71   6.41 3.41   50.12   46.71  0.84
## Chi0n       5 477  14.37   6.09  14.18  14.05   5.50 1.73   38.48   36.75  0.81
## Chi0v       6 477  14.89   6.13  14.68  14.58   5.52 1.73   39.84   38.11  0.77
## Chi1        7 477  11.87   4.83  11.77  11.67   4.29 1.73   31.52   29.78  0.66
## Chi1n       8 477   8.43   3.76   8.35   8.26   3.54 0.61   23.17   22.55  0.65
## Chi1v       9 477   8.96   3.81   9.00   8.79   3.39 0.61   24.44   23.83  0.62
## Chi2n      10 477   6.67   3.28   6.36   6.48   3.04 0.25   19.35   19.10  0.75
##      kurtosis    se
## Label      -0.64 0.02
## BalabanJ    2.66 0.03
## BertzCT     1.74 17.99
## Chi0        1.73 0.33
## Chi0n       1.77 0.28
## Chi0v       1.62 0.28
## Chi1        1.38 0.22
## Chi1n       1.18 0.17
## Chi1v       1.13 0.17
## Chi2n       1.05 0.15
```

```
data_quality_report <- data.frame(
  Variable = names(drug_induced_training_data),
  Type = sapply(drug_induced_training_data, class),
  Missing = sapply(drug_induced_training_data, function(x) sum(is.na(x))),
  Complete = sapply(drug_induced_training_data, function(x) sum(!is.na(x))),
  Unique = sapply(drug_induced_training_data, function(x) length(unique(x)))
)
knitr::kable(head(data_quality_report, 40), caption = "Drug Induced Data Quality Report (Preview)")
```

Data Quality Report

Table 1: Drug Induced Data Quality Report (Preview)

| | Variable | Type | Missing | Complete | Unique |
|----------|----------|-----------|---------|----------|--------|
| Label | Label | integer | 0 | 477 | 2 |
| SMILES | SMILES | character | 0 | 477 | 477 |
| BalabanJ | BalabanJ | numeric | 0 | 477 | 406 |
| BertzCT | BertzCT | numeric | 0 | 477 | 465 |
| Chi0 | Chi0 | numeric | 0 | 477 | 382 |
| Chi0n | Chi0n | numeric | 0 | 477 | 460 |
| Chi0v | Chi0v | numeric | 0 | 477 | 460 |
| Chi1 | Chi1 | numeric | 0 | 477 | 423 |

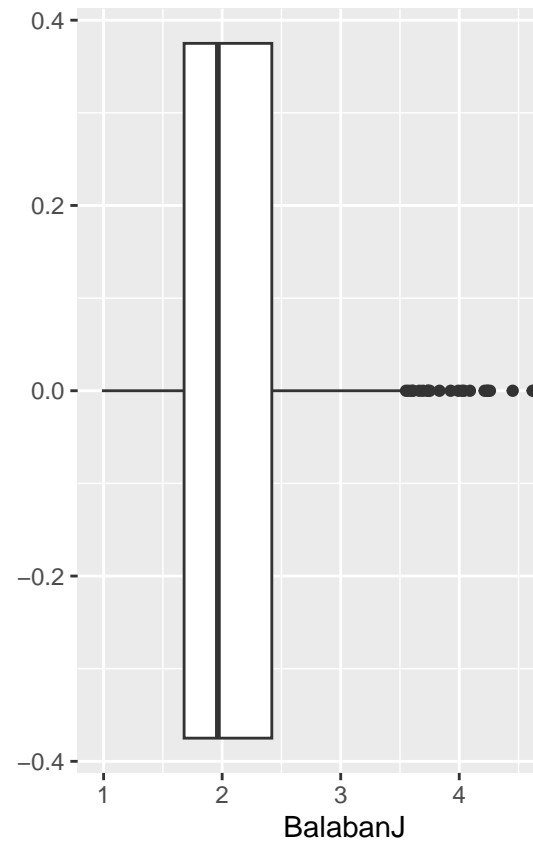
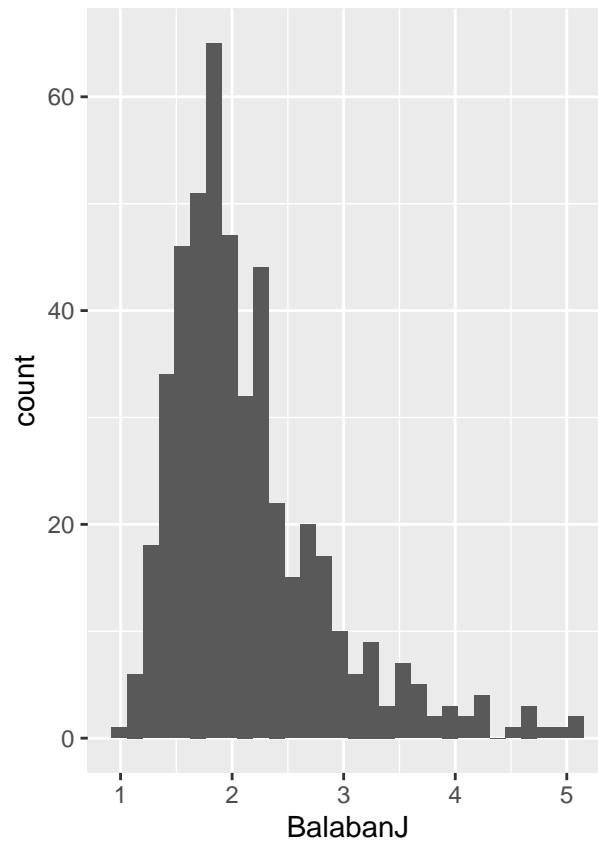
| | Variable | Type | Missing | Complete | Unique |
|---------------------|---------------------|---------|---------|----------|--------|
| Chi1n | Chi1n | numeric | 0 | 477 | 461 |
| Chi1v | Chi1v | numeric | 0 | 477 | 466 |
| Chi2n | Chi2n | numeric | 0 | 477 | 458 |
| Chi2v | Chi2v | numeric | 0 | 477 | 462 |
| Chi3n | Chi3n | numeric | 0 | 477 | 456 |
| Chi3v | Chi3v | numeric | 0 | 477 | 457 |
| Chi4n | Chi4n | numeric | 0 | 477 | 450 |
| Chi4v | Chi4v | numeric | 0 | 477 | 447 |
| EState_VSA1 | EState_VSA1 | numeric | 0 | 477 | 211 |
| EState_VSA10 | EState_VSA10 | numeric | 0 | 477 | 108 |
| EState_VSA11 | EState_VSA11 | numeric | 0 | 477 | 5 |
| EState_VSA2 | EState_VSA2 | numeric | 0 | 477 | 279 |
| EState_VSA3 | EState_VSA3 | numeric | 0 | 477 | 233 |
| EState_VSA4 | EState_VSA4 | numeric | 0 | 477 | 244 |
| EState_VSA5 | EState_VSA5 | numeric | 0 | 477 | 183 |
| EState_VSA6 | EState_VSA6 | numeric | 0 | 477 | 131 |
| EState_VSA7 | EState_VSA7 | numeric | 0 | 477 | 122 |
| EState_VSA8 | EState_VSA8 | numeric | 0 | 477 | 217 |
| EState_VSA9 | EState_VSA9 | numeric | 0 | 477 | 134 |
| ExactMolWt | ExactMolWt | numeric | 0 | 477 | 460 |
| FractionCSP3 | FractionCSP3 | numeric | 0 | 477 | 175 |
| HallKierAlpha | HallKierAlpha | numeric | 0 | 477 | 247 |
| HeavyAtomCount | HeavyAtomCount | integer | 0 | 477 | 53 |
| HeavyAtomMolWt | HeavyAtomMolWt | numeric | 0 | 477 | 420 |
| Ipc | Ipc | numeric | 0 | 477 | 453 |
| Kappa1 | Kappa1 | numeric | 0 | 477 | 450 |
| Kappa2 | Kappa2 | numeric | 0 | 477 | 459 |
| Kappa3 | Kappa3 | numeric | 0 | 477 | 448 |
| LabuteASA | LabuteASA | numeric | 0 | 477 | 466 |
| MaxAbsEStateIndex | MaxAbsEStateIndex | numeric | 0 | 477 | 452 |
| MaxAbsPartialCharge | MaxAbsPartialCharge | numeric | 0 | 477 | 166 |
| MaxEStateIndex | MaxEStateIndex | numeric | 0 | 477 | 452 |

```

hist <- ggplot(drug_induced_training_data, aes(x=BalabanJ)) + geom_histogram(bins = 30)
boxplot <- ggplot(drug_induced_training_data, aes(x=BalabanJ)) + geom_boxplot()

grid.arrange(hist, boxplot, ncol = 2)

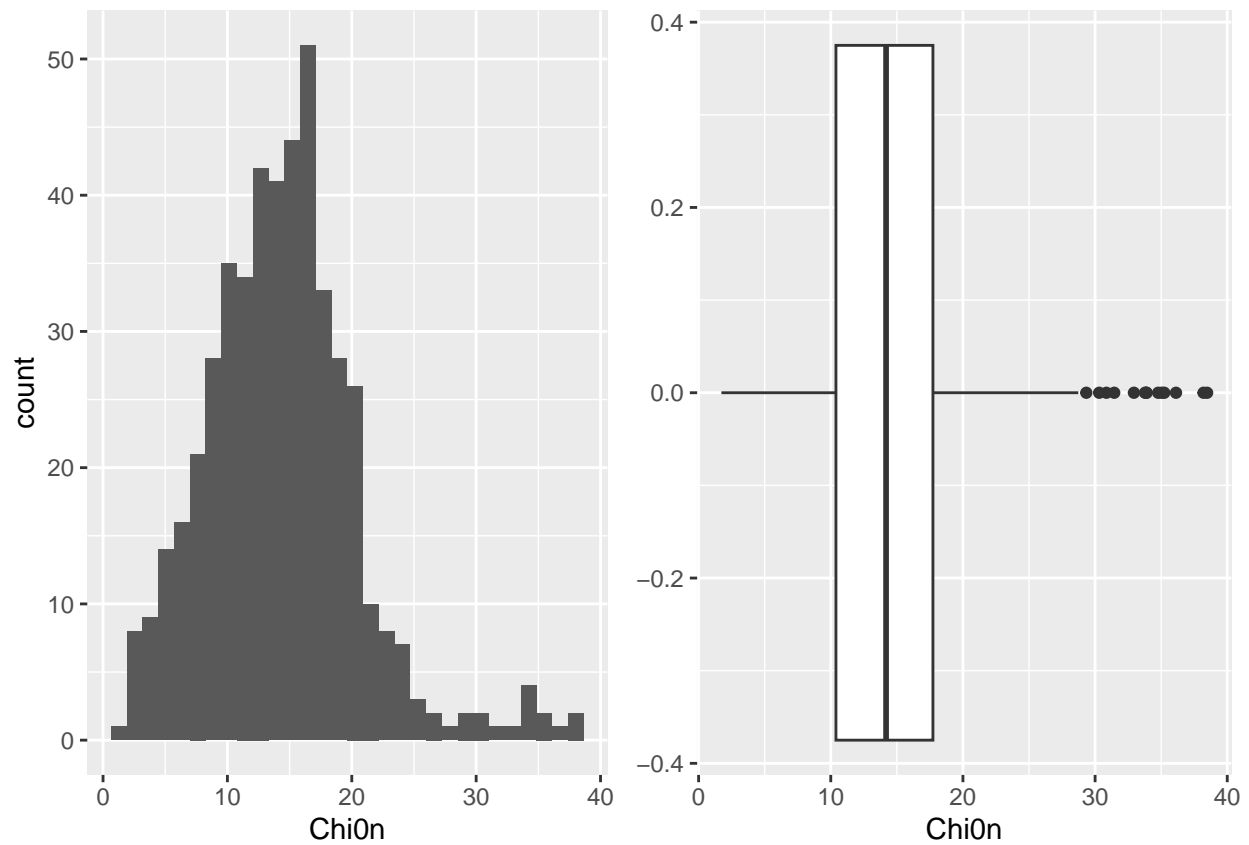
```



Univariate Analysis

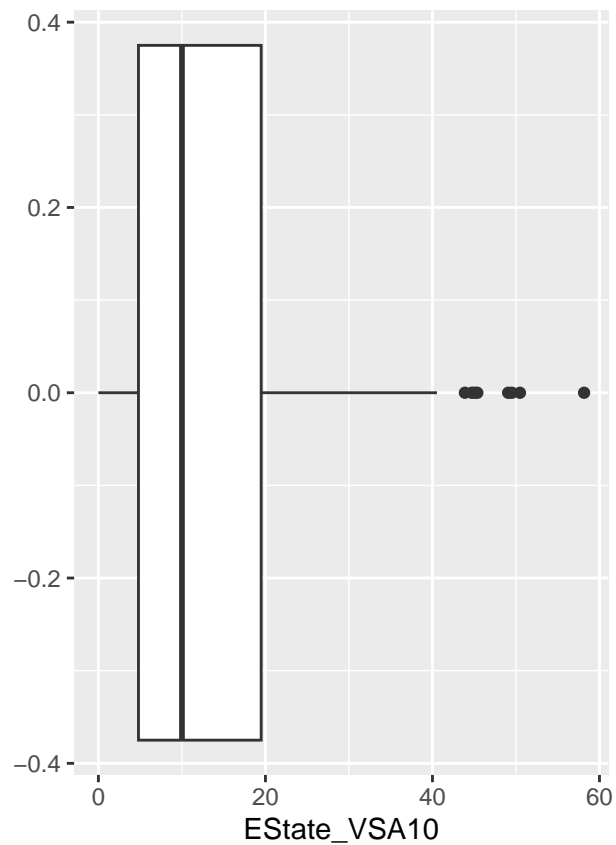
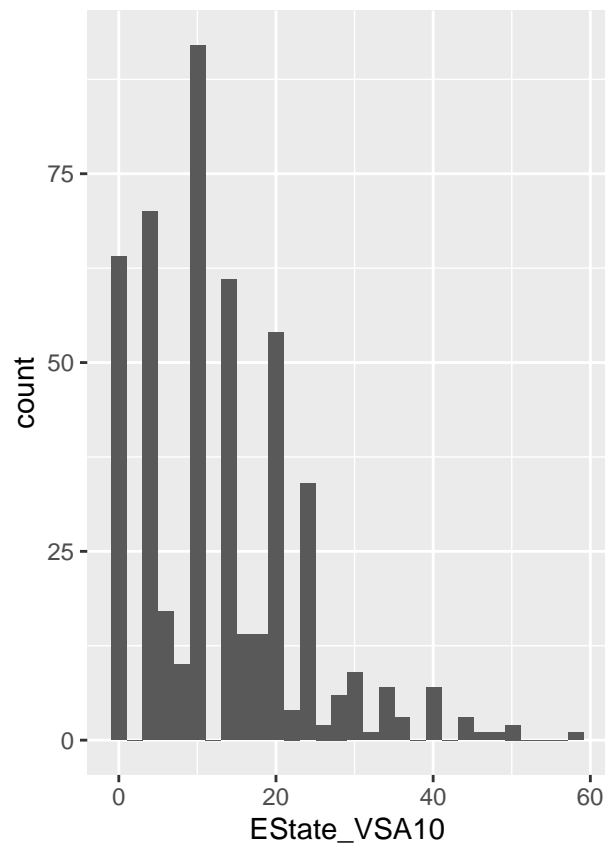
```
hist_2 <- ggplot(drug_induced_training_data, aes(x=Chi0n)) + geom_histogram(bins = 30)
boxplot_2 <- ggplot(drug_induced_training_data, aes(x=Chi0n)) + geom_boxplot()

grid.arrange(hist_2, boxplot_2, ncol = 2)
```



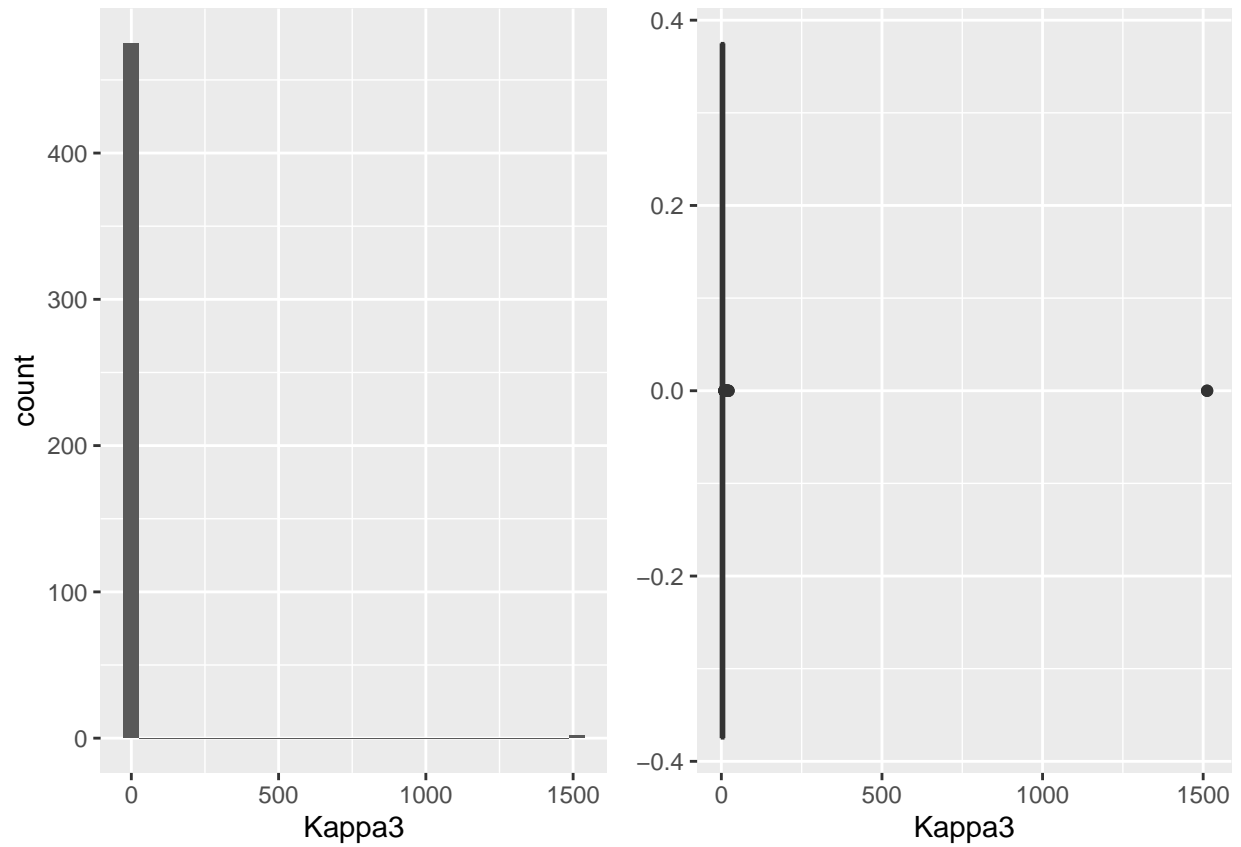
```
hist_3 <- ggplot(drug_induced_training_data, aes(x=EState_VSA10)) + geom_histogram(bins = 30)
boxplot_3 <- ggplot(drug_induced_training_data, aes(x=EState_VSA10)) + geom_boxplot()

grid.arrange(hist_3, boxplot_3, ncol = 2)
```



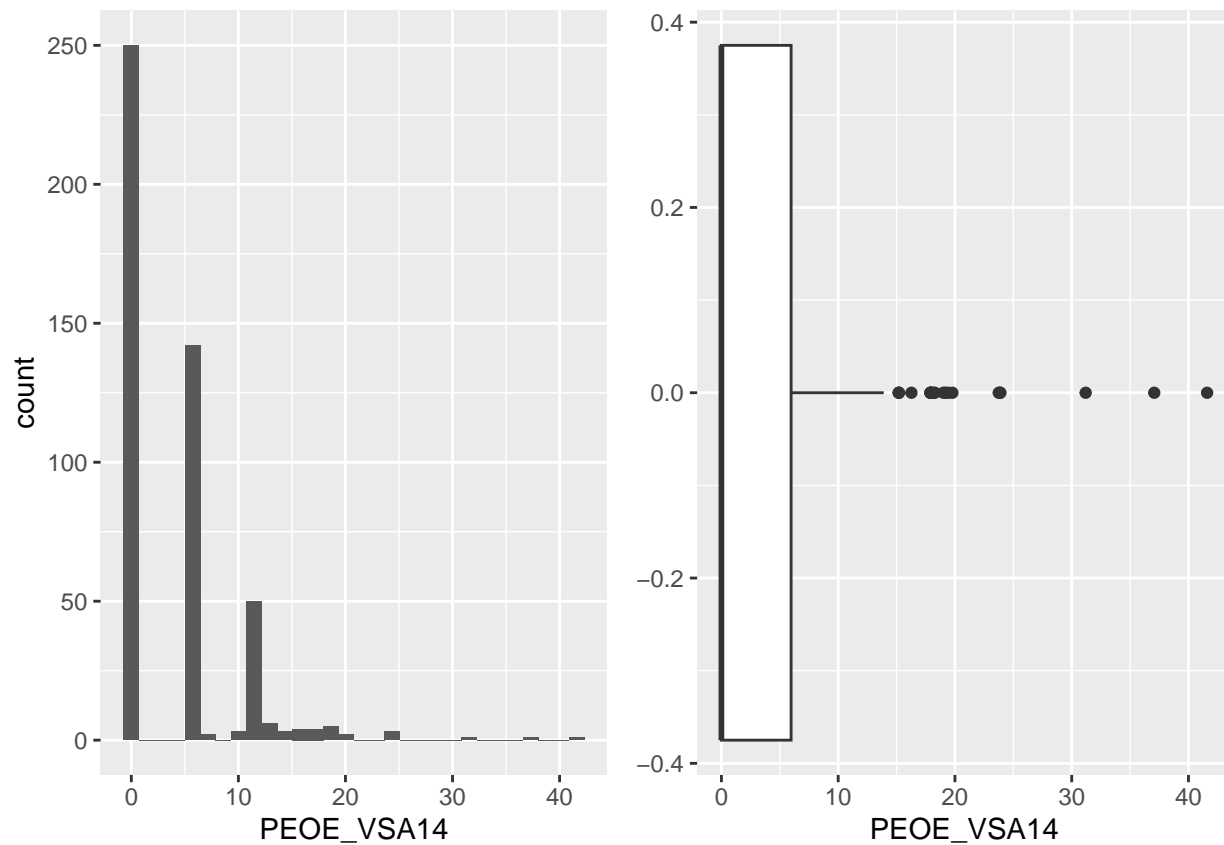
```
hist_4 <- ggplot(drug_induced_training_data, aes(x=Kappa3)) + geom_histogram(bins = 30)
boxplot_4 <- ggplot(drug_induced_training_data, aes(x=Kappa3)) + geom_boxplot()

grid.arrange(hist_4, boxplot_4, ncol = 2)
```



```
hist_5 <- ggplot(drug_induced_training_data, aes(x=PEOE_VSA14)) + geom_histogram(bins = 30)
boxplot_5 <- ggplot(drug_induced_training_data, aes(x=PEOE_VSA14)) + geom_boxplot()

grid.arrange(hist_5, boxplot_5, ncol = 2)
```

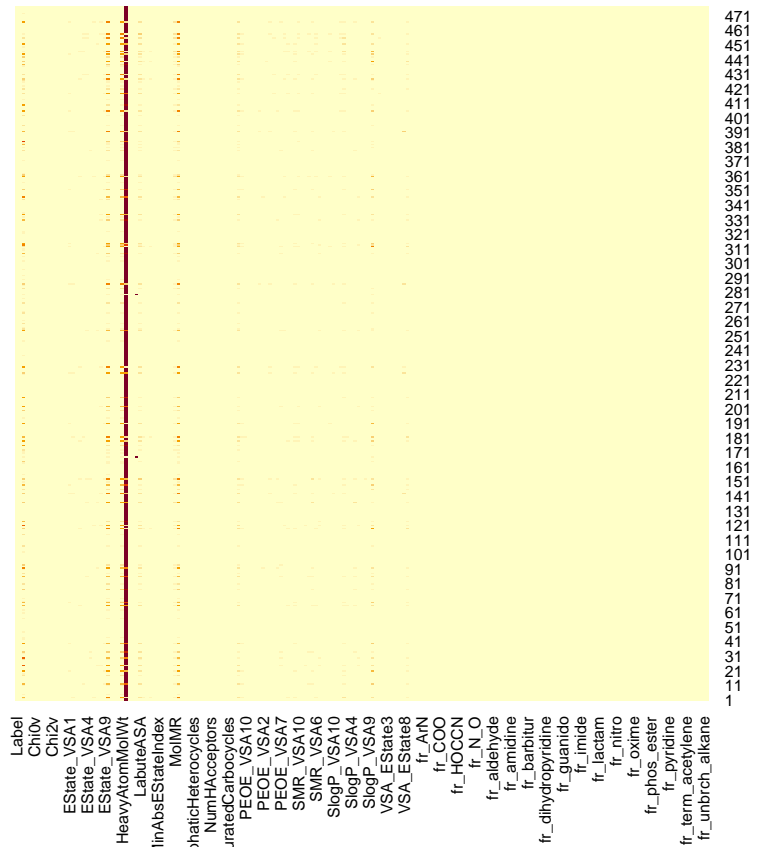


Features are right skewed. Few outliers that do not need to be removed.

```
numeric_cols_indexes <- sapply(drug_induced_training_data, is.numeric)
numeric_features <- drug_induced_training_data[numeric_cols_indexes]

data <- as.matrix(numeric_features)

heatmap(data, Colv = NA, Rowv = NA, scale = "row")
```

Multivariate Analysis

```
print("Pre-balance class cardinality")
```

Class Imbalance

```
## [1] "Pre-balance class cardinality"
```

```
table(drug_induced_training_data$Label)
```

```
##
##    0    1
## 359 118
```

```
# Random Undersampling
balanced_0 <- sample_n(filter(drug_induced_training_data, Label == 0), 118)
filtered_1 <- filter(drug_induced_training_data, Label == 1)
bal_drug_induced_training_data <- rbind(balanced_0, filtered_1)

print("Post-balance class cardinality")
```

```
## [1] "Post-balance class cardinality"
```

```
table(bal_drug_induced_training_data$Label)
```

```
##
##    0    1
## 118 118
```

```
# Drop SMILES feature
bal_drug_induced_training_data <- bal_drug_induced_training_data[, colnames(bal_drug_induced_training_data)

for (col_name in colnames(bal_drug_induced_training_data)){
# For binary responses
  if (length(unique(bal_drug_induced_training_data[[col_name]])) == 2){
    bal_drug_induced_training_data[[col_name]] <- as.factor(bal_drug_induced_training_data[[col_name]])
  }

  # For standarization of numeric responses
  if (is.numeric(bal_drug_induced_training_data[[col_name]])){
    if( max(bal_drug_induced_training_data[[col_name]]) != min(bal_drug_induced_training_data[[col_name]]){
      bal_drug_induced_training_data[[col_name]] <- (bal_drug_induced_training_data[[col_name]] - min(bal_drug_induced_training_data[[col_name]])) / (max(bal_drug_induced_training_data[[col_name]]) - min(bal_drug_induced_training_data[[col_name]]))
    }
    else{
      bal_drug_induced_training_data[[col_name]] <- 0
    }
  }
}
# Final Dataset
# bal_drug_induced_training_data
```

Pre-Model Data Preparation

```
# Missing step --> numeric_features need to be obtained from bal_drug_induced_training_data

model_all <- lm(Label ~ ., data=numeric_features) # with all the independent variables in the dataframe
# summary(model_all)
coeff <- coefficients(model_all)

# To remove correlated where coefficient = NaN -> Correlation
na_coeff_names <- names(coeff)[is.na(coeff)]
na_coeff_names
```

Feature Selection

```
## [1] "MaxEStateIndex"      "NumAliphaticRings"    "NumAromaticRings"
## [4] "NumRadicalElectrons" "NumSaturatedRings"    "RingCount"
## [7] "SMR_VSA8"           "SlogP_VSA9"           "VSA_EState1"
## [10] "VSA_EState2"         "VSA_EState3"          "VSA_EState4"
```

| | | | |
|---------|---------------|-------------------|-------------------|
| ## [13] | "VSA_EState5" | "VSA_EState6" | "VSA_EState7" |
| ## [16] | "fr_COO" | "fr_Nhpyrrole" | "fr_azide" |
| ## [19] | "fr_barbitur" | "fr_benzene" | "fr_diazo" |
| ## [22] | "fr_isocyan" | "fr_isothiocyane" | "fr_prisulfonamd" |
| ## [25] | "fr_thiocyan" | | |