

Macroevolutionary Rates of Species Interactions: Approximate Bayesian Inference from Cophylogenies

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Abstract

Understanding the macroevolutionary dynamics of species interactions such as parasitisms, commensalisms, and mutualisms is an important goal in evolutionary ecology. To this end, statistical inference from extant cophylogenetic systems holds immense potential. However, such inference cannot yet handle the speciation-extinction dynamics that occur simultaneously in the host and symbiont clades on the same timescale. Here we present an Approximate Bayesian Computation (ABC) approach that, while taking into account host and symbiont extinction, infers rates of four types of speciation from a cophylogenetic system: (i) host speciation, (ii) symbiont speciation without host-switching, (iii) symbiont speciation with host-switching, and (iv) cospeciation. The new ABC approach relies on a novel design of summary statistics combining both size-based (i.e., tree sizes) and size-free summary statistics (i.e., the normalized distribution of Branch Length Differences - BLenD) of the cophylogeny. Convergence analyses show that the combined design of summary statistics outperforms size-based or size-free summary statistics alone – achieving satisfactory accuracy in detecting rate heterogeneity between the four types of speciation. Our ABC approach allows the user to infer the predominant mode of speciation within a given cophylogenetic system. The approach is demonstrated with an application to a cophylogenetic dataset of commensalism, in which beetles of one genus mimic those of another. In this system, we identify host speciation as the predominant process with the fastest rate (in events per unit time) among all four types of speciation (i.e., 4.3–5.1 times faster than the median among all four types of speciation). Understanding how and why different types of speciation may predominate in different cophylogenetic systems can have implications for various areas in ecology and evolution such as host conservatism, trait-driven diversification, pathogen spillover risk, and parasite extinction risk. This new approach highlights the need and potential for future efforts to compile time-calibrated cophylogenies.

Key words: Phylogenetic comparative methods, Likelihood-free inference, Simulation-based inference, Birth-death processes, Diversification rates, Coevolution, Co-diversification, Symbiosis

1 Introduction

Biologists have long sought to understand the relationship between macroevolution and ecological interactions. Early examples include how Darwin explains the diversity of life as a result of natural selection in his seminal book, using examples where natural selection occurs as a result of organisms' "struggle for life" against their competitors and natural enemies (Darwin, 1859). A recent comprehensive synthesis has consolidated the macroevolution of species interactions as a rapidly-evolving subfield of ecology and evolution with deep historical roots (see Hembry and Weber 2020 for an overview). Several other syntheses have separately examined the effects of species interactions on macroevolution, reaching varying conclusions about whether generalities exist in how species interactions influence clade diversification (Jablonski, 2008; Harmon et al., 2019; Zeng and Wiens, 2021; Kaur and Pennell, 2023). Another body of research has focused on the effects of macroevolution on species interaction networks. Such examples include how macroevolutionary stability shapes the intricate architecture of species interaction networks (Burin et al., 2021) and statistical inference of the evolutionary history of species interaction networks (Braga et al., 2020, 2021). Recent years have also seen significant advances in theoretical work that, by modeling how speciation arises from mutation in a (meta)community with

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ongoing species interactions, bridges the microevolution-macroevolution gap and provides mechanistic insights into the evolution of ecological communities (Aguilée et al., 2018; Coelho and Rangel, 2018; Maliet et al., 2020; Chaparro-Pedraza et al., 2022; Pontarp et al., 2024; Zeng and Hembry, 2024).

In particular, the joint macroevolutionary history of organisms that interact in pairs (i.e., bipartite interactions) has attracted much attention over the decades. Examples of bipartite interactions include those between plants and pollinators, plants and seed dispersers, hosts and parasites, and between plants and herbivores (see Bronstein 2015 for an overview). Coevolution can arise when reciprocal evolutionary change happens between interacting parties, driving or impeding the diversification of the interacting clades (Yoder and Nuismer, 2010; Hembry et al., 2014). Coevolution can leave detectable signals in the cophylogenetic patterns of bipartite species interactions, such as those formed by hosts and their mutualistic, commensal, or parasitic symbionts. Significant effort has been devoted to studying topological congruence between phylogenies of interacting organisms as an indicator of cospeciation or codiversification (e.g., Legendre et al. 2002; de Vienne et al. 2007; Hoyal Cuthill and Charleston 2012; Hayward et al. 2021; Perez-Lamarque and Morlon 2024; reviewed in Nieberding and Olivieri 2007; de Vienne et al. 2013; Martínez-Aquino 2016; Dismukes et al. 2022). Recent cophylogenetic research has forayed into linking cophylogenetic patterns with their underlying eco-evolutionary processes (Blasco-Costa et al., 2021). Another area of active research is cophylogenetic reconstruction, aiming to reconstruct the joint evolutionary history of interacting clades by finding a parsimonious series of events that give rise to the observed cophylogenetic patterns (e.g., Merkle et al. 2010; Baudet et al. 2015; Sinaimeri et al. 2023; reviewed in Charleston and Libeskind-Hadas 2014).

However, the study of the macroevolutionary rates of cophylogenetic systems remains in its infancy. Macroevolutionary rates can be defined, for single clades, as speciation and extinction rates that can be inferred from phylogenetic systems (Stadler, 2013; Harmon et al., 2014). Here, we define macroevolutionary rates of species interactions (i.e., of interacting clades) as speciation and extinction rates that can be inferred from cophylogenetic systems. Although much recent work has been focused on the advancement of phylogenetic comparative methods for estimating the speciation, extinction, or net diversification rates of single clades (Garamszegi, 2014; Revell and Harmon, 2022; Morlon et al., 2024), single-clade-based phylogenetic methods cannot be readily used for cophylogenetic systems. Specifically, although these single-clade-based methods allow diversification rates to be inferred separately for host and symbiont clades in a given cophylogenetic system, these rate estimates cannot provide a full picture of the prevalence of different types of speciation in the cophylogeny (Charleston and Perkins, 2006; Charleston and Libeskind-Hadas, 2014). Motivated by a desire to better understand parasitic symbionts (i.e., pathogens), Alcalá et al. (2017) represents an important advance in estimating macroevolutionary rates of species interactions from cophylogenetic systems. This approach focuses on estimating the rate of cospeciation and the probability of host switching in the symbiont clade. However, this approach has two major disadvantages when the entire cophylogenetic system, as opposed to the symbiont clade alone, is of interest: (i) it does not allow host speciation and symbiont speciation to occur on the same timescale; (ii) it does not take into account host extinction, in which symbionts relying on the extinct host goes extinct as well. A promising, more recent approach can already tease apart the relative contributions of different macroevolutionary processes (cospeciation, host switching, pollinator speciation, and pollinator extinction) in generating cophylogenetic patterns (Satler et al., 2019). However, this method suffers the same problem of inconsistent timescales between host and symbiont diversification, resulting in zero rate estimates for host switching and cospeciation in some cases. Fortunately, recent advances have allowed simulating cophylogenetic systems as a result of constant-rate birth-death processes, simultaneously incorporating speciation and extinction in both the host and symbiont clades (Dismukes and Heath, 2021). This new model has paved the way for developing new statistical methods for estimating macroevolutionary rates of bipartite species interactions.

Approximate Bayesian Computation (ABC) has been shown to be a useful tool for estimating macroevolutionary rates of species interactions from cophylogenetic systems (Alcalá et al., 2017). ABC is a class of simulation-based inference that has proven especially suitable for models whose likelihoods are impossible or difficult to obtain (Marin et al., 2012; Sunnåker et al., 2013). Initially developed by geneticists, ABC has been increasingly used in ecology and evolution (Pantel and Becks, 2023). In the most basic form of ABC, real data is compared to numerous simulations generated using different parameter samples, and parameters samples that generate simulations similar enough to the real data constitute the approximate posterior distribution of that parameter; the key to successful ABC is designing informative summary statistics that are informative and low in dimensionality (Sunnåker et al., 2013). Conceptual frameworks and standard practices have been established for using approximate Bayesian computation for both parameter estimation and model selection in ecology and evolution (Csilléry et al., 2010, 2012; Janzen et al., 2015; Pontarp et al., 2019; Pantel and Becks, 2023). As for estimating macroevolutionary rates of species interactions from cophylogenetic systems, the existing ABC approach shows promising potential but also suffers significant difficulty in implementation (Alcalá et al., 2017). Specifically, the implementation of this approach requires highly customized representation of co-

102 phylogenies, a multitude of network summary statistics, dimension reduction techniques, and machine learning
 103 techniques. An easy-to-implement ABC approach to estimating macroevolutionary rates of species interactions
 104 from cophylogenies is yet to be developed.

105 Here, we develop a new ABC approach that, while taking into account host and symbiont extinction,
 106 infers the rates of four types of speciation processes from an extant cophylogeny: (i) host speciation, (ii) symbiont
 107 speciation without host-switching, (iii) symbiont speciation with host-switching, and (iv) cospeciation. This
 108 ABC approach is easy to implement thanks to our newly designed summary statistics, which take into account
 109 properties of the cophylogeny that are both related to or irrespective of the sizes of the host and symbiont trees
 110 (i.e., size-based and size-free summary statistics). We show that the performance of the parameter inference
 111 remains reasonably good despite the “curse of dimensionality”, process stochasticity of speciation and extinction,
 112 and partial information.

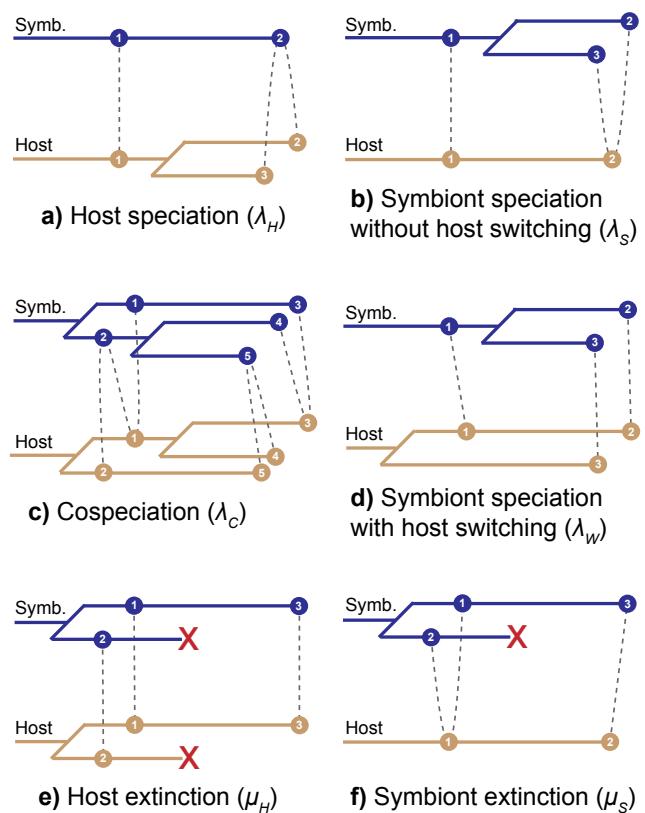


Figure 1: Speciation and extinction events giving rise to cophylogenetic patterns. Macroevolutionary rates of species interactions considered in this study (denoted by Greek letters in parentheses) are defined as, for each type of event, the number of events per unit time. (a) Host speciation (λ_H): both descendant hosts retain association with the original symbiont. (b) Symbiont speciation without host switching (λ_S): both descendant symbionts retain association with the original host. (c) Cospeciation (λ_C): a host and one of its symbionts, randomly selected, undergoes speciation simultaneously (host 1 into hosts 3 and 4; symbiont 2 into symbionts 4 and 5). Each descendant host forms association with one of the descendant symbionts (host 3 with symbiont 4; host 4 with symbiont 5). The ancestral host's and symbiont's remaining associations are randomly sorted among the descendants. (d) Symbiont speciation with host switching (λ_W): same as (b), but here one of the descendant symbionts switches to a new, randomly selected host. (e) Host extinction (μ_H): note that the symbiont(s) relying on this host goes extinct as well. (f) Symbiont extinction (μ_S). These event definitions are identical to those in the simulation tool, except that the rate of symbiont speciation with host switching (λ_W) is denoted by χ in the original study (Dismukes and Heath, 2021).

113 2 Methods

114 In brief, we follow these two steps to estimate the speciation rates of a target cophylogeny: (i) we simulate
 115 cophylogenies following a *birth-death model for co-diversification*. (ii) In an *ABC framework*, the simulated
 116 phylogenies are compared to the target cophylogeny to obtain estimates of the speciation rates. Notably,

the comparison between the simulated and target cophylogenies is decided by *summary statistics*. In our *performance evaluation*, we perform this two-step ABC procedure, using cophylogenies simulated with known parameter values as the target cophylogeny. In our *application to empirical data*, we perform the same two-step ABC procedure using a cophylogeny compiled from empirical data as the target cophylogeny. Below, we provide details of each of these components.

2.1 A Birth-Death Model for Co-diversification

We use the `sim_cophyBD` function from the R package `treeducken` (ver. 1.1.0; Dismukes and Heath 2021) for simulating cophylogenies following a birth–death model. The model in `treeducken` allows the simulation of host and symbiont co-diversification. The model considers six types of speciation or extinction events (Fig. 1): host sepspeciation (λ_H), symbiont speciation without host switching (λ_S), cospeciation (λ_C), symbiont speciation with host switching(λ_W), host extinction (μ_H), and symbiont extinction (μ_S). Each type of event is controlled by a rate parameter that represents the number of events per unit time following a Poisson process. For example, a rate of 1 means one event occurs per unit time in the cophylogenetic system.

Because this approach is designed to infer macroevolutionary rates of species interactions from extant cophylogenetic systems, the extinction rate must not exceed the speciation rate for the host and symbiont clades. We further parameterize the model by defining relative extinction rates ϵ_H (for the host clade) and ϵ_S (for the symbiont clade) as follows,

$$\epsilon_H = \frac{\mu_H}{\lambda_H + \lambda_C}, \quad \epsilon_S = \frac{\mu_S}{\lambda_S + \lambda_C + \lambda_W}$$

where both ϵ_H and ϵ_S range between 0 and 1. In the ABC framework (detailed below), we assume that ϵ_H and ϵ_S are known and aim to estimate the four speciation rates ($\lambda_H, \lambda_S, \lambda_C, \lambda_W$).

2.2 ABC Framework

Speciation rates ($\lambda_H, \lambda_S, \lambda_C, \lambda_W$) in a cophylogenetic system can be collectively seen as a multivariate parameter θ , which is treated as a random variable in Bayesian parameter inference. The posterior probability of θ given observed data D can be, in theory, obtained by

$$P(\theta|D) = \frac{P(D|\theta)P(\theta)}{\int P(D|\theta)P(\theta)d\theta}$$

where $P(\theta)$ is known as the prior distribution and $P(D|\theta)$ as the likelihood function (i.e., the probability of observing the cophylogeny D given the parameter θ). However, when D is a cophylogeny and θ is the parameter in the birth-death model, the likelihood function is difficult to obtain, making Approximate Bayesian Computation (ABC) an especially useful approach for parameter estimation (Alcalá et al., 2017). An ABC approach does not explicitly compute the likelihood but instead uses simulations to approximate the posterior distribution. Here, we use the basic form of a rejection-based ABC approach (Csilléry et al., 2012) following these steps. First, cophylogenies are simulated with the birth-death model (see the previous section in Methods) using samples of θ drawn from the prior distribution. Then, each sample from the prior distribution is accepted if the distance between the resulting simulated cophylogeny and D is below a given threshold. Alternatively, a sample is rejected if the distance is above the threshold. The distances are determined by summary statistics (see the following section in Methods). The threshold is decided by the tolerance rate, which determines the proportion of simulations with the smallest distance to accept. Eventually, accepted samples of θ constitute the approximate posterior distribution.

2.3 Summary Statistics

The key to successful ABC parameter inference is summary statistics that decide the distance between a simulation and the observed data (Sumnåker et al., 2013). A cophylogenetic system (e.g., Fig. 2a) consists of two phylogenies and the network formed by the tips of the two trees. In theory, one could use indices of phylogenies and networks as the summary statistics for ABC. Indices for phylogenies include phylogenetic diversity

(Clarke and Warwick, 2001), the gamma statistic (Pybus and Harvey, 2000) and Sackin index measuring tree shape (Sackin, 1972). Indices for networks include global properties such as nestedness, modularity, connectance (Guimaraes Jr, 2020), and meso-scale properties such as motif frequencies (Simmons et al., 2019). Indices concerning both the phylogeny and network components of a cophylogeny include the mantel correlation, a measure of phylogenetic conservatism of bipartite species association (Maliet et al., 2020). However, single indices tend not to be informative enough as summary statistics for phylogenetic systems (Janzen et al., 2015; Janzen and Etienne, 2024). On the other hand, when we combine multiple of these indices in our preliminary exploratory analyses, parameter convergence is generally difficult. This is unsurprising because the high dimensionality of summary statistics can introduce the “curse of dimensionality”, that is, the match between a simulation and the observed data gets significantly more unlikely when the dimensionality of summary statistics increases (Sunnåker et al., 2013). The difficulty in convergence in our preliminary analyses also agrees with the previous study on estimating macroevolutionary rates of species interactions from cophylogenies, where accurate parameter inference based on numerous indices is difficult without dimension reduction and machine learning techniques (Alcalá et al., 2017). Because of these reasons, we do not choose index-based summary statistics for our approach.

As opposed to index-based summary statistics, normalized curves have been proposed as an alternative type of summary statistics for ABC. Specifically, a pioneering study has used a normalized lineage-through-time curve (nLTT), which is irrespective of the size or length of the phylogenetic tree, to effectively estimate constant speciation rates from single phylogenies while taking into account constant-rate lineage extinction (Janzen et al., 2015). The nLTT summary statistics are informative, computationally efficient, and not obviously affected by the “curse of dimensionality”. Inspired by this work, here we adapt the use of normalized curves as summary statistics for cophylogenetic systems, the details of which are given below.

2.3.1 BLenD Curve

The BLenD (Branch Length Difference) curve is a normalized curve irrespective of the sizes of the cophylogeny of interest. For any association (between a host and a symbiont) in the cophylogeny, it is possible to obtain

$$\delta = \frac{l_H - l_S}{L}$$

where l_H is the branch length of the host tip, and l_S is the branch length of the symbiont tip, and L is the length (or height) of the cophylogeny from its root to tips (Fig. 2). We refer to δ ($-1 < \delta < 1$) as the normalized Branch Length Difference (BLenD). $\hat{f}(\delta)$, the density function of δ , can then be estimated from all observed values of δ in the cophylogeny ($\delta_1, \delta_2, \dots, \delta_n$, where n is the total number of associations; each association is shown as a line with dotted ends in Fig. 2). This estimation is implemented as kernel density estimation using the function `density(kernel = "gaussian")` in the `stats` package in R (ver. 4.4.0; R Core Team 2024). We hereafter refer to $\hat{f}(\delta)$ as the BLenD curve.

Now, let us imagine that we have two cophylogenies instead of one, which we will call Cophylogeny A and Cophylogeny B. Thus, the distance between Cophylogeny A and Cophylogeny B can be defined as:

$$d_{BLenD}(A, B) = \int_{-1}^1 \left| \hat{f}_A(\delta) - \hat{f}_B(\delta) \right| d\delta$$

where $\hat{f}_A(\delta)$ and $\hat{f}_B(\delta)$ are the BLenD curves for Cophylogenies A and B, respectively. $d_{BLenD}(A, B)$ is equal to area between the BLenD curves of Cophylogenies A and B (shaded area in Fig. 2). Similarly to Janzen et al. (2015), this definition of distance achieves a natural weighing of the contribution of all the $\hat{f}(\delta)$ -differences between Cophylogenies A and B; also similarly, the BLenD curve is implemented as a vector of summary statistics. In effect, $d_{BLenD}(A, B)$ is equal to zero only when the two BLenD curves compared are identical.

2.3.2 Tree Sizes

In addition to the newly designed the BLenD curve, we consider the sizes of the two trees (the numbers of tips in the host and symbiont trees) in the cophylogeny as summary statistics because speciation and extinction rates will almost certainly affect the numbers of tips in the host and symbiont phylogenies. The distance between Cophylogenies A and B, based on the tree sizes of their *host* phylogenies, can be defined as:

$$d_{host}(A, B) = |N_{A, host} - N_{B, host}|$$

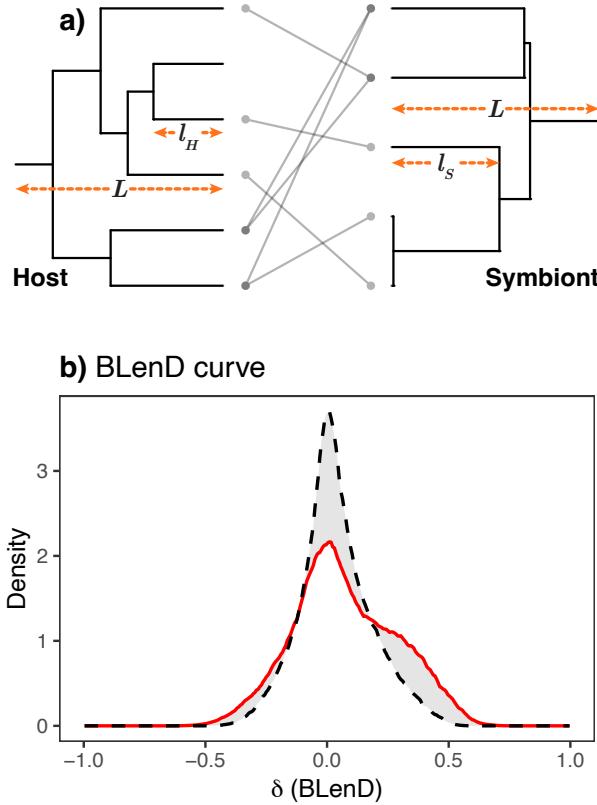


Figure 2: The Branch Length Difference (BLenD) curve as summary statistics for approximate Bayesian computation. **(a)** For each association (line with dotted ends) in the cophylogeny, we measure three lengths: the branch length of the host tip l_H , the branch length of the symbiont tip l_S , and the total length of the cophylogeny L (i.e., assumed to be the same for the host and symbiont phylogenies). Then, we obtain $\delta = (l_H - l_S)/L$ for every association in the cophylogeny. This allows us to estimate the density of δ in the entire cophylogeny (i.e., the BLenD curve) **(b)** Cophylogenies A (solid line) and B (dashed line) with different speciation rates have different BLenD curves. In both Cophylogenies A and B, $\epsilon_H = \epsilon_S = 0.3$. In Cophylogeny A, $\lambda_H = 0.9$, $\lambda_S = 0.3$, $\lambda_C = 0.4$, $\lambda_W = 1.7$; in Cophylogeny B, $\lambda_H = 1.1$, $\lambda_S = 1.2$, $\lambda_C = 0.7$, $\lambda_W = 0.3$. For each curve, the number of replicates is 500 and the median density across all replicates is used.

202 where $N_{A, \text{host}}$ and $N_{B, \text{host}}$ are the sizes of the host phylogenies in Cophylogenies A and B, respectively.
 203 Similarly, the distance between Cophylogenies A and B, based on the tree sizes of their *symbiont* phylogenies,
 204 can be defined as:

$$d_{\text{symb}}(A, B) = |N_{A, \text{symb}} - N_{B, \text{symb}}|$$

205 where $N_{A, \text{symb}}$ and $N_{B, \text{symb}}$ are the sizes of the symbiont phylogenies in Cophylogenies A and B, respectively.
 206 Tree sizes are log-transformed before being used to calculate tree-size-based distances.

207 2.3.3 Combining Summary Statistics

208 Combining different summary statistics in general increases the informativeness of summary statistics for ABC
 209 (Sunnåker et al., 2013), so here we test whether BLenD curve and tree sizes combined can achieve better inference
 210 results than either of them alone. When the BLenD curve and tree sizes are used in tandem, we standardize the
 211 summary statistics by dividing them by their standard deviation to ensure that the BLenD curve and each of
 212 the two tree size statistics have comparable variation among simulations. The distance between Cophylogenies
 213 A and B is computed as the sum of the BLenD-based and tree-size-based distances.

214 2.4 Performance Evaluation

215 We are particularly interested in testing this ABC approach's performance with cophylogenies with rate hetero-
 216 geneity between the four types of speciation ($\lambda_H, \lambda_S, \lambda_C, \lambda_W$). We focus our testing on the most basic form of

rate heterogeneity, that is, one type of speciation occurs at a higher rate than the other three types. This most basic form of rate heterogeneity includes four possibilities: $\lambda_H > \lambda_S = \lambda_C = \lambda_W$, $\lambda_S > \lambda_H = \lambda_C = \lambda_W$, $\lambda_C > \lambda_H = \lambda_S = \lambda_W$, or $\lambda_W > \lambda_H = \lambda_S = \lambda_C$. They correspond to situations where only one type of speciation predominates in the co-phylogenetic system. To test the accuracy of the parameter estimator, we perform convergence analyses to test whether the approximate posterior distribution approaches the true parameter values when tolerance rate approaches zero. We document both the parameter estimates and their residuals (i.e., estimates minus the true value).

Besides the parameter inference problem, we also evaluate how correctly our ABC approach can *categorically* detect a specific type of speciation that occurs at a higher rate than the other three types. A correct detection is considered to have been achieved when the type of speciation with the highest true rate also has the highest median rate estimate in the approximate posterior distribution. We evaluate how the detection correctness of this ABC approach compares to a random guess. To generate simulated cophylogenies for five combinations of ϵ_H and ϵ_S (0/0, 0.3/0.3, 0.7/0, 0/0.7, 0.7/0.7 for ϵ_H / ϵ_S), we run 50000 simulations each containing 100 replicates and take the median BLenD density and tree sizes across all replicates.

2.5 Application to Empirical Data

The ABC approach is applicable to cophylogenetic datasets where the host and symbiont phylogenies are both time-calibrated and have the same stem age. This requirement is important because the timescale has to be held consistent for all six of the speciation/extinction processes (Fig. 1). One empirical dataset that meets this requirement is a dataset of Batesian mimicry compiled by Van Dam et al. (2024), where beetles in the genus *Doliops* mimic beetles in the genus *Pachyrhynchus*, both native to the Philippines. This mimicry can be seen as a case of commensalism, which is defined as a type of interaction where one party in the interaction receives a benefit while the other party receives neither a cost nor a benefit (Bronstein, 2015). Particularly in this case, the mimics (*Doliops*) receive a protective benefit while the models (*Pachyrhynchus*) are largely unaffected. Host-symbiont models have been fitted to this dataset in the original study (Van Dam et al., 2024), with mimics considered as symbionts and models as hosts. The stem age for both *Pachyrhynchus* and *Doliops* has been geologically calibrated for the maximum age of the Philippines and estimated to be 25-30 Myr in the original study. We use 27.5 Myr, the midpoint of this range, as the total length of both phylogenies (distance from the root to tips). We used the `phylo.tracer` function from the `phylotree` package (Revell, 2020) to extract the two beetle phylogenies from Figure 3 of the original study. We then code the association between species manually based on Figure 2 of the original study. For the beetle cophylogeny to be comparable with cophylogenies simulated with treeducken, we rescale the beetle cophylogeny such that the total length of the beetle cophylogeny is equal to the total length of the simulated cophylogenies (i.e., 27.5 Myr on the realistic timescale is convertible to 2 unit time on the `treeducken` timescale as defined in Dismukes and Heath 2021).

For this empirical cophylogenetic dataset, we estimate the four speciation rates ($\lambda_H, \lambda_S, \lambda_C, \lambda_W$) using the same ABC framework (see a previous section in Methods) and the same sets of simulated cophylogenies that we use for performance evaluation (see a section in Methods). As a necessary step when applying ABC to real data (Pantel and Becks, 2023), we also perform posterior predictive checks of model fit to see how well the simulated cophylogenies that are accepted in the ABC framework (i.e., those that contribute to the approximate posterior distribution of the parameter estimates) resemble the beetle cophylogeny .

3 Results

3.1 Performance

3.1.1 Rate Estimation

Here we focus on how different choices of summary statistics compare in terms of convergence. Good convergence is considered to have been achieved when both the precision (i.e., how narrow the approximate posterior distribution is) and accuracy (i.e., how close the approximate posterior distribution is to the true value) of parameter estimation are high. Our convergence analyses reveal that, across all combinations of relative extinction rate ϵ_H and ϵ_S that we consider, parameter estimates generally converge to the true values of the four lambdas ($\lambda_H, \lambda_S, \lambda_C, \lambda_W$) best when a combination of tree sizes and the BLenD curve are used as summary statistics

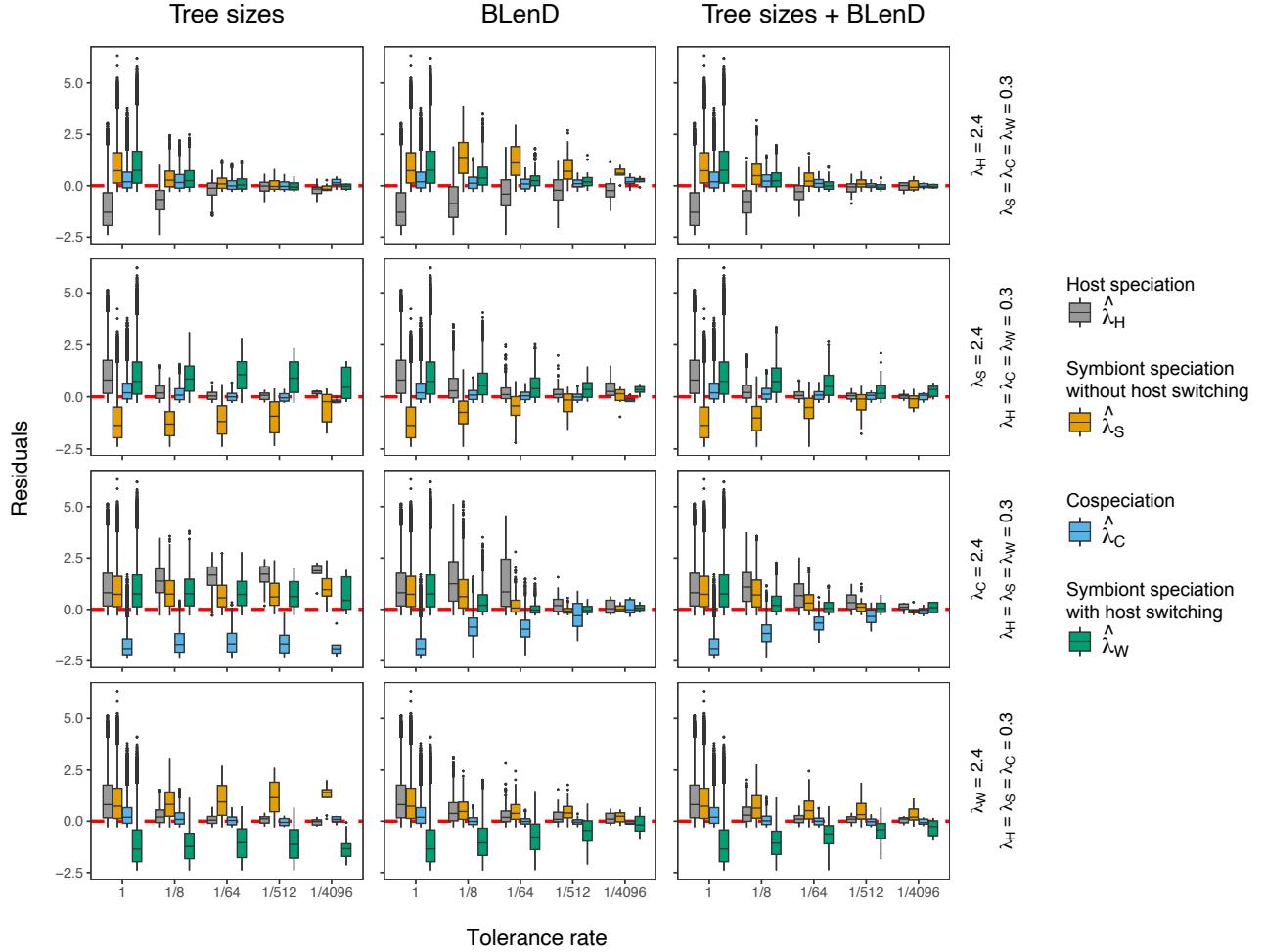


Figure 3: Convergence plots of the speciation rate estimates ($\hat{\lambda}_H, \hat{\lambda}_S, \hat{\lambda}_C, \hat{\lambda}_W$) when $\epsilon_H = \epsilon_S = 0.3$. Residuals are calculated as estimates minus the true values ($\lambda_H, \lambda_S, \lambda_C, \lambda_W$). The estimates converge well if the residuals approach zero as tolerance rate approaches zero. Simulated cophylogenies whose true parameters are known (vertical text on the right) are used as observed data in the ABC framework. Each observed cophylogeny contains 500 replicates; the median BLenD density and tree sizes are taken across all replicates before being used as observations in the ABC. Convergence plots under other assumptions of ϵ_H and ϵ_S are presented in Figs. S1-S4.

Table 1: Performance of speciation rate estimation and speciation rate heterogeneity detection under different assumptions of relative extinctions rates (ϵ_H and ϵ_S). Four rate parameters are considered: λ_H - host speciation; λ_S - host speciation without host switching; λ_C - cospeciation; λ_W - symbiont speciation with host switching.

Relative extinction rates	True speciation rates ^a				Rate estimates ^b				Rate heterogeneity detection		
	ϵ_H	ϵ_S	λ_H	λ_S	λ_C	λ_W	$\hat{\lambda}_H$	$\hat{\lambda}_S$	$\hat{\lambda}_C$	$\hat{\lambda}_W$	Correctness ^c
0	0	2.4	0.3	0.3	0.3	2.19 (0.12)	0.41 (0.18)	0.28 (0.12)	0.14 (0.1)	96%	3.832
0	0	0.3	2.4	0.3	0.3	0.26 (0.16)	2.3 (0.16)	0.3 (0.15)	0.21 (0.14)	72%	2.88
0	0	0.3	0.3	2.4	0.3	0.42 (0.19)	0.22 (0.12)	2.17 (0.15)	0.12 (0.09)	94%	3.752
0	0	0.3	0.3	0.3	2.4	0.22 (0.12)	0.75 (0.77)	0.34 (0.18)	1.74 (0.67)	62%	2.48
0.3	0.3	2.4	0.3	0.3	0.3	2.35 (0.25)	0.33 (0.3)	0.3 (0.12)	0.28 (0.09)	96%	3.856
0.3	0.3	0.3	2.4	0.3	0.3	0.33 (0.17)	2.2 (0.4)	0.3 (0.18)	0.58 (0.31)	47%	1.896
0.3	0.3	0.3	0.3	2.4	0.3	0.41 (0.2)	0.24 (0.2)	2.32 (0.19)	0.39 (0.25)	70%	2.808
0.3	0.3	0.3	0.3	0.3	2.4	0.35 (0.16)	0.64 (0.48)	0.27 (0.15)	2.05 (0.44)	68%	2.736
0.7	0	2.4	0.3	0.3	0.3	2.45 (0.2)	0.4 (0.24)	0.24 (0.16)	0.32 (0.22)	96%	3.856
0.7	0	0.3	2.4	0.3	0.3	0.3 (0.11)	2.1 (0.27)	0.28 (0.14)	0.59 (0.3)	42%	1.696
0.7	0	0.3	0.3	2.4	0.3	0.47 (0.26)	0.53 (0.4)	2.16 (0.29)	0.27 (0.12)	64%	2.568
0.7	0	0.3	0.3	0.3	2.4	0.29 (0.12)	0.79 (0.68)	0.35 (0.12)	1.85 (0.73)	65%	2.592
0	0.7	2.4	0.3	0.3	0.3	2.45 (0.14)	0.41 (0.15)	0.28 (0.12)	0.28 (0.12)	98%	3.936
0	0.7	0.3	2.4	0.3	0.3	0.26 (0.16)	1.96 (0.44)	0.34 (0.12)	0.65 (0.39)	51%	2.056
0	0.7	0.3	0.3	2.4	0.3	0.33 (0.44)	0.35 (0.17)	2.37 (0.36)	0.36 (0.25)	69%	2.776
0	0.7	0.3	0.3	0.3	2.4	0.44 (0.18)	0.78 (0.52)	0.22 (0.17)	1.85 (0.47)	66%	2.656
0.7	0.7	2.4	0.3	0.3	0.3	2.36 (0.19)	0.42 (0.26)	0.3 (0.12)	0.23 (0.16)	98%	3.904
0.7	0.7	0.3	2.4	0.3	0.3	0.26 (0.19)	1.71 (0.74)	0.41 (0.21)	0.86 (0.57)	45%	1.792
0.7	0.7	0.3	0.3	2.4	0.3	0.63 (0.26)	0.42 (0.25)	2.14 (0.22)	0.34 (0.21)	58%	2.336
0.7	0.7	0.3	0.3	0.3	2.4	0.25 (0.16)	0.77 (0.72)	0.34 (0.17)	1.72 (0.65)	65%	2.608

Note: tree sizes and the BLenD curve are used in tandem as summary statistics. The tolerance rate used is 1/4096. See Tables S1 & S2 for results for when tree sizes or the BLenD curve are used alone.

^a Fixed, known parameter values used to generate the observed data (cophylogeny) in the ABC framework. 500 replicates are generated for each set of parameter values.

^b Means (outside parentheses) and standard deviations (inside parentheses) of the rate estimates. The median BLenD density and tree sizes are taken across all 500 replicates of the observed data before being used in the ABC.

^c Percentage of the 500 replicates of observed data for which the highest rate among $\lambda_H, \lambda_S, \lambda_C, \lambda_W$ is detected correctly.

^d Actual correctness (see footnote c) divided by the expected correctness of a random guess among $\lambda_H, \lambda_S, \lambda_C, \lambda_W$ (25%).

(the third column compared to the first and second columns in Figs. 3, S1-S4). When only tree sizes are used (the first column in Figs. 3, S1-S4), the parameter estimates converge well only when λ_H is the highest among the four lambdas; otherwise the parameter estimates converge poorly, having a low precision or low accuracy. When only the BLenD curve is used (the second column in Figs. 3, S1-S4), the parameter estimates converge well when λ_S, λ_C , or λ_W is the highest among the four lambdas; however, the parameter estimates converge poorly when λ_H is the highest among the four lambdas. Thus, in terms of convergence, tree sizes alone perform well when the BLenD curve alone performs poorly; conversely, tree sizes alone perform poorly when the BLenD curve alone performs well. The performance of a combination of tree sizes and the BLenD curve (the third column in Figs. 3, S1-S4) is generally on par with the better of the two (tree sizes alone and the BLenD curve alone) and, in some cases, outperforms the better of the two (Figs. S1 & S2).

The combination of tree sizes and the BLenD curve is also shown to be generally the best at handling different types of rate heterogeneity and the least sensitive to relative extinction rates (Table 1, S1 & S2). The tree sizes, when used alone, are especially poor at handling rate heterogeneity where λ_C is the highest among the four lambdas (Table S2), but this type of rate heterogeneity is handled well by a combination of tree sizes and the BLenD curve (Table 1). Although the BLenD curve alone performs relatively well with most types of rate heterogeneity (Table S2), it is outperformed by a combination of tree sizes and the BLenD curve when λ_H is the highest among the four lambdas and the ϵ_S is as high as 0.7 (Table 1).

3.1.2 Rate Heterogeneity Detection

As for the detection of speciation rate heterogeneity, combining BLenD and tree sizes as summary statistics, again, tends to outperform the use of the BLenD curve or tree sizes alone (Table 1, S1 & S2). We are specifically interested in identifying the highest rate among the four lambdas $\lambda_H, \lambda_S, \lambda_C, \lambda_W$, and a random guess among the four lambdas is expected to have a 25% correctness. An improvement value (i.e., the actual correctness

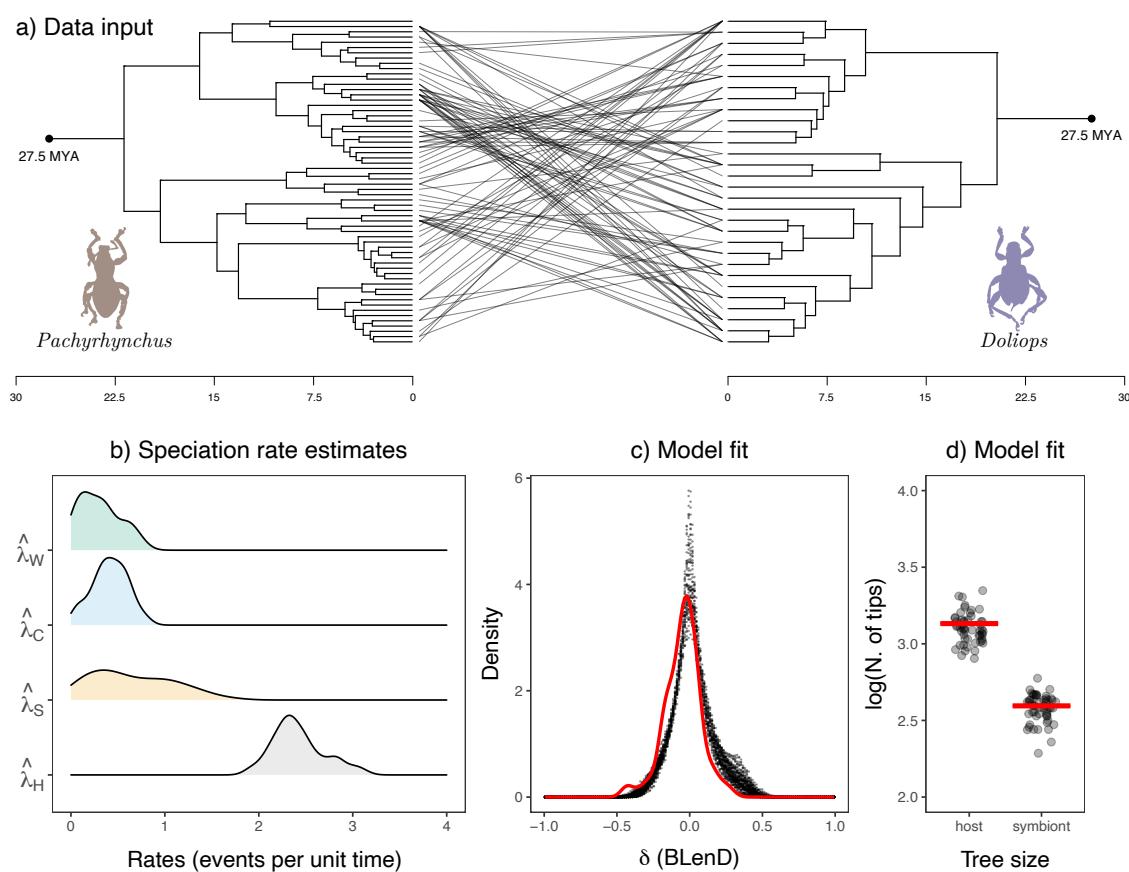


Figure 4: Speciation rate inference from the cophylogenetic dataset of beetle mimicry (Van Dam et al. 2024). Four rate parameters are considered: λ_H - host speciation; λ_S - host speciation without host switching; λ_C - cospeciation; λ_W - symbiont speciation with host switching. The BLenD curve and tree sizes are used in tandem as summary statistics. ϵ_H and ϵ_S are assumed to be 0.3. The tolerance rate used is 1/512. (a) Time-calibrated cophylogeny of *Pachyrhynchus* (models, treated as hosts) and *Doliops* (mimics, treated as symbionts). (b) Density curves of the speciation rate estimates. (c & d) Posterior predictive checks of model fit. Shown here are the BLenD curve and tree sizes of the beetle data (solid line) and accepted simulations in the ABC (dots). Results under other assumptions of ϵ_H and ϵ_S are presented in Fig. S5 and Table 2.

divided by 25%) greater than one suggests that the inference performs better than a random guess, and vice versa. Our results show that the tree sizes alone perform consistently worse than the random guess when λ_C is the highest among the four lambdas (Table S1), rendering the tree sizes unsuitable summary statistics for this purpose. The use of the BLenD curve alone performs better than a random guess in most cases (Table S2), but the performance is much improved when the BLenD curve is used in tandem with tree sizes (Table 1).

When the tree sizes and BLenD curve are used in tandem, the detection of some types of rate heterogeneity is less sensitive to extinction than others (Table 1). The detection correctness, when λ_H is the highest among the four lambdas, remains high (96–98%) regardless of ϵ_H and ϵ_S . Similarly, the detection correctness when λ_W is the highest among the four lambdas remains somewhat high (62–68%) regardless of ϵ_H and ϵ_S . However, the detection correctness for the other two types of rate heterogeneity is more sensitive to ϵ_H and ϵ_S . Notably, the detection correctness of rate heterogeneity where λ_S or λ_C is the highest among the four lambdas drops by 24–25% (from 72% to 47% and from 94% to 70%, respectively) when ϵ_H and ϵ_S increase from 0 to 0.3 (Table 1).

3.2 Rates Inferred from Beetle Mimicry

Across all the combinations of ϵ_H and ϵ_S that we consider, the ABC infers host speciation (λ_H) as the type of speciation with the highest rate (Figs. 4 & S5; Table 2). We infer the mean rate of host speciation to be 1.80–3.22 events per unit time, while the rates of symbiont speciation (with or without host switching) and cospeciation are inferred to be below 1 event per unit time. In terms of mean rate estimates, host speciation

is 4.3-5.1 times faster than the median among all four types of speciation. Regardless of assumptions of ϵ_H and ϵ_S , host speciation is consistently inferred to have the highest rate among the four types of speciation. Our performance evaluation has shown that the ABC approach generally performs well with rate heterogeneity where the host speciation rate is much higher than the other three types of speciation rates (see the Performance section in Results). In light of these performance evaluation results, the rate estimates from the beetle mimicry dataset suggest that host speciation is the fastest among all four types of speciation (i.e., host speciation, symbiont speciation without host switching, cospeciation, and symbiont speciation with host switching) in the cophylogenetic system. Our posterior predictive checks show that model fit remains reasonably good regardless of assumptions of relative extinction rates ϵ_H and ϵ_S (Figs. 4 & S5).

Table 2: Rate estimates ($\hat{\lambda}_H, \hat{\lambda}_S, \hat{\lambda}_C, \hat{\lambda}_W$) from the beetle mimicry dataset under different assumptions of ϵ_H and ϵ_S .

ϵ_H	ϵ_S	$\hat{\lambda}_H$	$\hat{\lambda}_S$	$\hat{\lambda}_C$	$\hat{\lambda}_W$
0	0	1.80 (0.21)	0.36 (0.21)	0.48 (0.12)	0.25 (0.19)
0.3	0.3	2.42 (0.27)	0.65 (0.43)	0.42 (0.18)	0.30 (0.22)
0.7	0	3.22 (0.33)	0.98 (0.43)	0.34 (0.18)	0.35 (0.28)
0	0.7	2.12 (0.26)	0.38 (0.27)	0.46 (0.16)	0.43 (0.22)
0.7	0.7	3.21 (0.42)	0.81 (0.52)	0.35 (0.26)	0.45 (0.27)

Note: for the lambdas, means are outside and standard deviations are inside parentheses. For comparability with Table 1, all lambdas are *not* converted to events/Myr.

4 Discussion

Here we present a proof of concept that an ABC approach can be useful for inferring, simultaneously, rates of different types of speciation from a cophylogenetic system. Key to our approach is the newly designed summary statistics combining both size-based (tree sizes) and size-free statistics (the BLenD curve). The new approach presented here can be used to infer the predominant mode of speciation in a cophylogenetic system, which should be an important question to biologists interested in species interactions.

We show that the new ABC approach can handle several expected technical difficulties reasonably well. Specifically, (i) the “Curse of dimensionality” (Sunnåker et al., 2013): the ABC simultaneously infers the rates of four processes (four parameters). (ii) Stochasticity: both speciation and extinction are simulated as stochastic processes in the birth-death co-diversification model, introducing stochasticity to the resulting cophylogenies (Dismukes and Heath, 2021). (iii) Partial information: a cophylogeny contains only partial information because extinct lineages and historical association between hosts and symbionts are not known from extant cophylogenies. Compared to previous approaches to estimating macroevolutionary rates of species interactions from cophylogenies (Alcalá et al., 2017; Satler et al., 2019), our new ABC approach allows speciation rate estimation under a more biologically realistic assumption that speciation-extinction dynamics occur simultaneously in the host and symbiont clades on the same timescale. In contrast to Alcalá et al. (2017), the new approach achieves satisfactory accuracy without the use of highly customized representation of cophylogenies, dimension reduction, or machine learning.

4.1 Theoretical and Practical Considerations

Recent advances have revealed that extant phylogenetic systems are consistent with a myriad of speciation and extinction configurations (Louca and Pennell, 2020). Meaningful speciation rate inference would generally require *a priori* hypotheses about the extinction rates of the system of interest (Morlon et al., 2022, 2024). This identifiability issue likely can be extrapolated to cophylogenetic systems as well - in our beetle mimicry example, we find that model fit is not obviously better or worse under any particular assumption of relative extinction rates ϵ_H and ϵ_S (Figs. 4 & S5). However, rate estimates differ considerably under different assumptions of relative extinction rates ϵ_H and ϵ_S (Table 2). Thus, biologically realistic assumptions about the relative extinction rates of hosts and symbionts are critically important for speciation rate inference. Ideally, literature should be available on what might be a reasonable assumption of relative extinction rates. Such literature is likely system-specific (e.g., on figs and fig wasps, or specialized parasitisms) and requires expert knowledge about the system of interest. If such information is unavailable, the user should perform the same speciation rate inference under different assumptions of relative extinction rates to test the robustness of their conclusions.

345 We show that, as with phylogenies of single clades (Morlon et al., 2022, 2024), cophylogenies can still provide
346 useful insights into diversification as long as realistic assumptions about extinction are made.

347 Given the numerous possibilities of rate heterogeneity between four types of speciation, we choose to
348 focus on the most basic form of rate heterogeneity, one where one type of speciation predominates in the entire
349 system. Testing the ABC approach on simulated datasets with diverse types of rate heterogeneity may extend
350 the use of this approach. Other forms of rate heterogeneity may include, for example, situations where two (or
351 three) types of speciation predominate. Those more complex forms of rate heterogeneity may not be dealt with
352 as easily as those considered in this study; they would deserve to be the focus of a separate study. However, the
353 most basic form of rate heterogeneity that we consider in this study may already be able to account for many
354 observed patterns in nature, as we show with the beetle mimicry example (Fig. 4).

355 When the user performs this inference for an empirical system, it is usually not possible to know what
356 the true parameter values are for their system of interest. Caution should be taken with the interpretation of
357 results where the four lambda estimates ($\hat{\lambda}_H, \hat{\lambda}_S, \hat{\lambda}_C, \hat{\lambda}_W$) differ only slightly. Our performance test results show
358 that, while large disparity between the four lambda estimates most likely reflects differences between their true
359 rates ($\lambda_H, \lambda_S, \lambda_C, \lambda_W$), slight differences between the four lambda estimates can arise from errors of the ABC
360 (Tables 1, S1 & S2). Thus, although strong disparity between the rate estimates of the four types of speciation
361 likely reflects a biological reality, such conclusions may not be appropriate if the rate estimate differences are
362 subtle.

363 In order to go beyond the proof of concept that we show here, technical improvements would be
364 potentially useful. Single-layered feed-forward neural networks, a machine learning technique, have been used
365 to improve the accuracy of ABC parameter inference (Csilléry et al., 2012). The improved accuracy is achieved
366 through neural-network-based regression to correct rate estimates (Blum and François, 2010). This approach
367 currently only works with a few single summary statistics. Adapting this regression-based approach for the use
368 of normalized curves (such as BLenD) as summary statistics may be an interesting future direction. Another
369 promising direction may be to enhance the ABC in this study with Sequential Monte Carlo (SMC), an algorithm
370 that improves the accuracy of inference by iteratively generating new parameter samples from the posterior
371 distribution and repeating the ABC procedure (for examples of ABC-SMC for (co)phylogenetic systems, see
372 Janzen et al. 2015; Baudet et al. 2015; Sinaimeri et al. 2023).

373 As with previous cophylogenetic studies, our ABC cophylogenetic approach is not immune to errors due
374 to phylogenetic reconstruction methods, incomplete taxon sampling, or discord between species trees and gene
375 trees (e.g., Hughes et al. 2007; Onufko et al. 2019). Additionally, apparent losses of symbionts in some host
376 lineages may be the results of incomplete sampling of host-symbiont associations (Jackson and Charleston 2004;
377 Charleston and Perkins 2006). For cophylogenetic studies in general, a statistical framework for quantifying
378 these sources of error remains undeveloped but should be a rewarding future direction.

379 A related body of research has focused on reconciling the phylogeny of symbionts with that of their hosts
380 (e.g., Merkle et al. 2010; Baudet et al. 2015; Sinaimeri et al. 2023; see Charleston and Perkins 2006; Charleston
381 and Libeskind-Hadas 2014 for an overview). These studies map the symbiont phylogeny onto the host phylogeny
382 to answer where cospeciations, duplications, host switches, and symbiont losses occur along branches of the host
383 phylogeny. As an intermediate step toward such reconciliation, recent approaches have used ABC to estimate
384 the frequencies of the aforementioned events (Baudet et al., 2015; Sinaimeri et al., 2023). It appears that one
385 could, in theory, derive the rates of these events by dividing their frequencies by time. However, such derivation
386 is impossible because of the lack of a timescale in these methods. Specifically, temporal constraints imposed by
387 tree topologies (e.g., host switches are only possible between temporally coexisting species) are enforced in these
388 methods only to the point where the *order* of events are feasible; this criterion is known as temporal feasibility
389 (Stolzer et al., 2012). This means that these methods, in contrast to our approach, do not take into account
390 branch lengths and are largely agnostic to the exact intervals between events, making it impossible to derive the
391 rates of these events from their frequencies. Therefore, to the best of our knowledge, our study represents the
392 first ABC cophylogenetic approach to offer a temporally explicit view of multiple types of macroevolutionary
393 events.

394 4.2 Broader Implications

395 The user might want to interpret the results from our approach in light of results from other methods. For
396 instance, the rate estimates from our approach offer valuable information about conservatism, a popular topic
397 in the biology of species interactions (Gómez et al., 2010). In symbiont speciation without host switching (λ_S),

398 the descendant symbiont lineages retain association with the host itself. Differently, in cospeciation (λ_C), the
399 descendant symbiont lineages retain association with the descendants of the ancestral host . In both symbiont
400 speciation without host switching (λ_S) and cospeciation (λ_C), the descendant symbiont lineages retain their
401 association with the ancestral lineage (Fig. 1), contributing to host conservatism of the symbionts. On the
402 contrary, in symbiont speciation with host switching (λ_W), one of the descendant symbiont lineages switches to
403 a new host lineage, reducing the host conservatism of the symbionts (i.e., contributing to host lability). Thus,
404 the simultaneous inference of λ_S , λ_C , and λ_W allows patterns of host conservatism to be attributed to two types
405 of speciation increasing host conservatism and one type of speciation decreasing it. The approach presented
406 here adds to the existing methods to tease apart the relative contributions of different macroevolutionary
407 processes to cophylogenetic patterns (Alcalá et al., 2017; Satler et al., 2019), but under a more biologically
408 realistic assumption that speciation and extinction occur on the timescale in the host and symbiont clades.
409 With this improved biological realism, different speciation processes' contributions may be more easily compared
410 within the *same* cophylogenetic system; additionally, a certain speciation process's contributions may also be
411 more easily compared between *multiple* cophylogenetic systems.

412 In the beetle mimicry example reanalyzed in this work, we find that the two types of speciation
413 increasing host conservatism and the one type decreasing it do not differ in rate considerably (Fig. 4). Van Dam
414 et al. (2024) reveals that, in the beetle mimicry system, a number of interactions are conserved over long periods
415 of time while others are more labile and transient. This mixed pattern of host conservatism, as our results
416 suggest, could have arisen from the fact that speciation events increasing host conservatism are balanced by
417 those decreasing it.

418 When a cophylogeny is seen as an evolving system of its own, questions can be asked about what affects
419 the rates of different types of speciation in a cophylogeny. In phylogenetic comparative research, significant
420 efforts have been devoted to studying how organismal traits affect speciation rates (Garamszegi, 2014; Revell
421 and Harmon, 2022; Morlon et al., 2024). Similar questions can be asked about cophylogenetic systems. For
422 example, key innovations have been hypothesized and debated as a driver of speciation for single clades (Miller
423 et al., 2023) - do certain types of key innovation also affect the rates of speciation in a cophylogenetic system?
424 In the beetle mimicry example, limited dispersal among islands has been shown to drive cospeciation Van Dam
425 et al. (2024). Thus, one hypothesis may be that the evolution of long-distance dispersal ability may decrease
426 the rate of cospeciation. In a different vein, the old and still influential "escape-and-radiate" hypothesis states
427 that the evolution of novel defensive traits in symbionts allows them to "escape" from the hosts and, as a
428 result, radiate (Ehrlich and Raven, 1964; Cogni et al., 2022). Then, what are the effects of these defensive traits
429 on symbiont speciation without host switching, cospeciation, and symbiont speciation with host switching?
430 Comparing the speciation rates of cophylogenetic systems with these key innovations versus those without them
431 may provide insights into these questions and hypotheses.

432 An interesting potential for the approach presented here lies in the possibility of informing real-world
433 problems about public health or conservation. For instance, the coevolutionary history of coronaviruses and
434 their mammalian hosts has been shown to be characterized by frequent host switches (Maestri et al., 2024).
435 Comparisons of host-switching speciation rates between different strains of viruses (which are essentially clades),
436 enabled by the new ABC approach, would allow one to ask whether highly virulent strains switch hosts more or
437 less frequently than less virulent strains. Alternatively, one could ask whether historical rates of host-switching
438 speciation predict pathogens' risk of spilling over to other host species in the future. In a different vein, specialism
439 (i.e., high host specificity) has been hypothesized as potential evolutionary dead ends that lead to extinction
440 (Day et al., 2016). The most extreme cases of specialism in symbionts, such as fig/fig-wasp mutualisms, tend
441 to have arisen almost exclusively through cospeciation (Machado et al., 2005). Thus, could the degree to which
442 cospeciation predominates in a cophylogenetic system serve as an indicator of extinction risk? Also interested
443 in extinction risk, Mulvey et al. (2022) presents a cophylogenetic method to estimate the extinction risk of
444 symbionts based on the number of symbiont extinctions, host-switches, and non-host-switching events. However,
445 this method does not take into account the speciation-extinction dynamics that occur simultaneously in the
446 host and symbiont clades. An interesting future direction may be to develop an extinction risk index based on
447 speciation and extinction rates estimated from a cophylogeny, such as those considered in this study.

448 By considering the cophylogenetic system as a whole, the new ABC approach shows the potential for
449 some important questions in ecology and evolution to be better understood in light of rates of speciation and
450 extinction in both the host and symbiont clades (i.e., in events per unit time). To answer these questions using
451 the new approach, more efforts to compile time-calibrated cophylogenies will be needed.

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⁴⁵⁶ **6 Supplementary Material**

⁴⁵⁷ Supplementary figures and tables are available at: <https://doi.org/10.6084/m9.figshare.29084336>.

⁴⁵⁸ **7 Data Availability**

⁴⁵⁹ Code used in this study is available at: https://github.com/yichaozeng/cophy_ABC. Datasets used in this
⁴⁶⁰ study are available at: <https://doi.org/10.6084/m9.figshare.29084336..>

461 References

- 462 Aguilée, R., Gascuel, F., Lambert, A., and Fèrriere, R. (2018). Clade diversification dynamics and the biotic
463 and abiotic controls of speciation and extinction rates. *Nature communications*, 9(1):3013.
- 464 Alcalá, N., Jenkins, T., Christe, P., and Vuilleumier, S. (2017). Host shift and cospeciation rate estimation from
465 co-phylogenies. *Ecology Letters*, 20(8):1014–1024.
- 466 Baudet, C., Donati, B., Sinaimeri, B., Crescenzi, P., Gautier, C., Matias, C., and Sagot, M.-F. (2015). Cophy-
467 logeny reconstruction via an approximate bayesian computation. *Systematic biology*, 64(3):416–431.
- 468 Blasco-Costa, I., Hayward, A., Poulin, R., and Balbuena, J. A. (2021). Next-generation cophylogeny: unravelling
469 eco-evolutionary processes. *Trends in Ecology & Evolution*, 36(10):907–918.
- 470 Blum, M. G. and François, O. (2010). Non-linear regression models for approximate bayesian computation.
471 *Statistics and computing*, 20:63–73.
- 472 Braga, M. P., Janz, N., Nylin, S., Ronquist, F., and Landis, M. J. (2021). Phylogenetic reconstruction of ances-
473 tral ecological networks through time for pierid butterflies and their host plants. *Ecology Letters*, 24(10):2134–
474 2145.
- 475 Braga, M. P., Landis, M. J., Nylin, S., Janz, N., and Ronquist, F. (2020). Bayesian inference of ancestral host–
476 parasite interactions under a phylogenetic model of host repertoire evolution. *Systematic biology*, 69(6):1149–
477 1162.
- 478 Bronstein, J. L. (2015). *Mutualism*. Oxford University Press.
- 479 Burin, G., Guimaraes Jr, P. R., and Quental, T. B. (2021). Macroevolutionary stability predicts interaction
480 patterns of species in seed dispersal networks. *Science*, 372(6543):733–737.
- 481 Chaparro-Pedraza, P. C., Roth, G., and Seehausen, O. (2022). The enrichment paradox in adaptive radiations:
482 Emergence of predators hinders diversification in resource rich environments. *Ecology letters*, 25(4):802–813.
- 483 Charleston, M. and Libeskind-Hadas, R. (2014). Event-based cophylogenetic comparative analysis. In Garam-
484 szegi, L. Z., editor, *Modern Phylogenetic Comparative Methods and Their Application in Evolutionary Biology:*
485 *Concepts and Practice*, pages 465–480. Springer.
- 486 Charleston, M. A. and Perkins, S. L. (2006). Traversing the tangle: algorithms and applications for cophyloge-
487 netic studies. *Journal of biomedical informatics*, 39(1):62–71.
- 488 Clarke, K. and Warwick, R. (2001). A further biodiversity index applicable to species lists: variation in
489 taxonomic distinctness. *Marine ecology Progress series*, 216:265–278.
- 490 Coelho, M. T. P. and Rangel, T. F. (2018). Neutral community dynamics and the evolution of species interac-
491 tions. *The American Naturalist*, 191(4):421–434.
- 492 Cogni, R., Quental, T. B., and Guimarães Jr, P. R. (2022). Ehrlich and raven escape and radiate coevolution
493 hypothesis at different levels of organization: Past and future perspectives. *Evolution*, 76(6):1108–1123.
- 494 Csilléry, K., Blum, M. G., Gaggiotti, O. E., and François, O. (2010). Approximate bayesian computation (abc)
495 in practice. *Trends in ecology & evolution*, 25(7):410–418.
- 496 Csilléry, K., François, O., and Blum, M. G. (2012). abc: an r package for approximate bayesian computation
497 (abc). *Methods in ecology and evolution*, 3(3):475–479.
- 498 Darwin, C. (1859). *On the Origin of Species by Means of Natural Selection*. John Murray, London.
- 499 Day, E. H., Hua, X., and Bromham, L. (2016). Is specialization an evolutionary dead end? testing for differences
500 in speciation, extinction and trait transition rates across diverse phylogenies of specialists and generalists.
501 *Journal of Evolutionary Biology*, 29(6):1257–1267.
- 502 de Vienne, D., Refréjier, G., López-Villavicencio, M., Tellier, A., Hood, M., and Giraud, T. (2013). Cospeciation
503 vs host-shift speciation: Methods for testing, evidence from natural associations and relation to coevolution.
504 *New Phytologist*, 198(2):347–385.
- 505 de Vienne, D. M., Giraud, T., and Martin, O. C. (2007). A congruence index for testing topological similarity
506 between trees. *Bioinformatics*, 23(23):3119–3124.

- 507 Dismukes, W., Braga, M. P., Hembry, D. H., Heath, T. A., and Landis, M. J. (2022). Cophylogenetic methods
508 to untangle the evolutionary history of ecological interactions. *Annual Review of Ecology, Evolution, and*
509 *Systematics*, 53(1):275–298.
- 510 Dismukes, W. and Heath, T. A. (2021). treeducken: An r package for simulating cophylogenetic systems.
511 *Methods in Ecology and Evolution*, 12(8):1358–1364.
- 512 Ehrlich, P. R. and Raven, P. H. (1964). Butterflies and plants: a study in coevolution. *Evolution*, 18:586–608.
- 513 Garamszegi, L. Z. (2014). *Modern Phylogenetic Comparative Methods and Their Application in Evolutionary*
514 *Biology: Concepts and Practice*. Springer.
- 515 Gómez, J. M., Verdú, M., and Perfectti, F. (2010). Ecological interactions are evolutionarily conserved across
516 the entire tree of life. *Nature*, 465(7300):918–921.
- 517 Guimaraes Jr, P. R. (2020). The structure of ecological networks across levels of organization. *Annual Review*
518 *of Ecology, Evolution, and Systematics*, 51(1):433–460.
- 519 Harmon, L. J., Andreazzi, C. S., Débarre, F., Drury, J., Goldberg, E. E., Martins, A. B., Melián, C. J.,
520 Narwani, A., Nuismer, S. L., Pennell, M. W., et al. (2019). Detecting the macroevolutionary signal of species
521 interactions. *Journal of Evolutionary Biology*, 32(8):769–782.
- 522 Harmon, L. J., Losos, J., Baum, D., Futuyma, D., Hoekstra, H., Lenski, R., Moore, A., Peichel, C., Schlüter,
523 D., and Whitlock, M. (2014). Macroevolutionary rates. *The Princeton Guide of Evolution*, pages 567–572.
- 524 Hayward, A., Poulin, R., and Nakagawa, S. (2021). A broadscale analysis of host-symbiont cophylogeny reveals
525 the drivers of phylogenetic congruence. *Ecology Letters*, 24(8):1681–1696.
- 526 Hembry, D. H. and Weber, M. G. (2020). Ecological interactions and macroevolution: a new field with old
527 roots. *Annual Review of Ecology, Evolution, and Systematics*, 51(1):215–243.
- 528 Hembry, D. H., Yoder, J. B., and Goodman, K. R. (2014). Coevolution and the diversification of life. *The*
529 *American Naturalist*, 184(4):425–438.
- 530 Hoyal Cuthill, J. and Charleston, M. (2012). Phylogenetic codivergence supports coevolution of mimetic heli-
531 conius butterflies. *PloS one*, 7(5):e36464.
- 532 Hughes, J., Kennedy, M., Johnson, K. P., Palma, R. L., and Page, R. D. (2007). Multiple cophylogenetic
533 analyses reveal frequent cospeciation between pelecaniform birds and pectinopygus lice. *Systematic biology*,
534 56(2):232–251.
- 535 Jablonski, D. (2008). Biotic interactions and macroevolution: extensions and mismatches across scales and
536 levels. *Evolution*, 62(4):715–739.
- 537 Jackson, A. P. and Charleston, M. A. (2004). A cophylogenetic perspective of rna-virus evolution. *Molecular*
538 *biology and evolution*, 21(1):45–57.
- 539 Janzen, T. and Etienne, R. S. (2024). Phylogenetic tree statistics: A systematic overview using the new r
540 package ‘treestats’. *Molecular Phylogenetics and Evolution*, page 108168.
- 541 Janzen, T., Höhna, S., and Etienne, R. S. (2015). Approximate bayesian computation of diversification rates
542 from molecular phylogenies: introducing a new efficient summary statistic, the nltt. *Methods in ecology and*
543 *evolution*, 6(5):566–575.
- 544 Kaur, K. M. and Pennell, M. (2023). Synthesizing the phylogenetic evidence for mutualism-associated diversi-
545 fication. *Evolution*, 77(8):1882–1892.
- 546 Legendre, P., Desdevises, Y., and Bazin, E. (2002). A statistical test for host-parasite coevolution. *Systematic*
547 *biology*, 51(2):217–234.
- 548 Louca, S. and Pennell, M. W. (2020). Extant timetrees are consistent with a myriad of diversification histories.
549 *Nature*, 580(7804):502–505.
- 550 Machado, C. A., Robbins, N., Gilbert, M. T. P., and Herre, E. A. (2005). Critical review of host specificity
551 and its coevolutionary implications in the fig/fig-wasp mutualism. *Proceedings of the National Academy of*
552 *Sciences*, 102(suppl_1):6558–6565.
- 553 Maestri, R., Perez-Lamarque, B., Zhukova, A., and Morlon, H. (2024). Recent evolutionary origin and localized
554 diversity hotspots of mammalian coronaviruses. *elife*, 13:RP91745.

- 555 Maliet, O., Loeuille, N., and Morlon, H. (2020). An individual-based model for the eco-evolutionary emergence
556 of bipartite interaction networks. *Ecology Letters*, 23(11):1623–1634.
- 557 Marin, J.-M., Pudlo, P., Robert, C. P., and Ryder, R. J. (2012). Approximate bayesian computational methods.
558 *Statistics and computing*, 22(6):1167–1180.
- 559 Martínez-Aquino, A. (2016). Phylogenetic framework for coevolutionary studies: a compass for exploring jungles
560 of tangled trees. *Current Zoology*, 62(4):393–403.
- 561 Merkle, D., Middendorf, M., and Wieseke, N. (2010). A parameter-adaptive dynamic programming approach
562 for inferring cophylogenies. *BMC bioinformatics*, 11:1–10.
- 563 Miller, A. H., Stroud, J. T., and Losos, J. B. (2023). The ecology and evolution of key innovations. *Trends in
564 Ecology & Evolution*, 38(2):122–131.
- 565 Morlon, H., Andréoletti, J., Barido-Sottani, J., Lambert, S., Perez-Lamarque, B., Quintero, I., Senderov, V.,
566 and Veron, P. (2024). Phylogenetic insights into diversification. *Annual Review of Ecology, Evolution, and
567 Systematics*, 55.
- 568 Morlon, H., Robin, S., and Hartig, F. (2022). Studying speciation and extinction dynamics from phylogenies:
569 addressing identifiability issues. *Trends in Ecology & Evolution*, 37(6):497–506.
- 570 Mulvey, L. P., Warnock, R. C., and De Baets, K. (2022). Where traditional extinction estimates fall flat: using
571 novel cophylogenetic methods to estimate extinction risk in platyhelminths. *Proceedings of the Royal Society
572 B*, 289(1981):20220432.
- 573 Nieberding, C. M. and Olivieri, I. (2007). Parasites: proxies for host genealogy and ecology? *Trends in Ecology
574 & Evolution*, 22(3):156–165.
- 575 Onufko, T. M., Bogusch, P., Ferrari, R. R., and Packer, L. (2019). Phylogeny and biogeography of the
576 cleptoparasitic bee genus epeolus (hymenoptera: Apidae) and cophylogenetic analysis with its host bee genus
577 colletes (hymenoptera: Colletidae). *Molecular phylogenetics and evolution*, 141:106603.
- 578 Pantel, J. H. and Becks, L. (2023). Statistical methods to identify mechanisms in studies of eco-evolutionary
579 dynamics. *Trends in Ecology & Evolution*, 38(8):760–772.
- 580 Perez-Lamarque, B. and Morlon, H. (2024). Distinguishing cophylogenetic signal from phylogenetic congruence
581 clarifies the interplay between evolutionary history and species interactions. *Systematic Biology*, 73(3):613–
582 622.
- 583 Pontarp, M., Brännström, Å., and Petchey, O. L. (2019). Inferring community assembly processes from macro-
584 scopic patterns using dynamic eco-evolutionary models and approximate bayesian computation (abc). *Methods
585 in Ecology and Evolution*, 10(4):450–460.
- 586 Pontarp, M., Lundberg, P., and Ripa, J. (2024). The succession of ecological divergence and reproductive
587 isolation in adaptive radiations. *Journal of Theoretical Biology*, 587:111819.
- 588 Pybus, O. G. and Harvey, P. H. (2000). Testing macro–evolutionary models using incomplete molecular phylo-
589 genies. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 267(1459):2267–2272.
- 590 R Core Team (2024). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical
591 Computing, Vienna, Austria.
- 592 Revell, L. J. (2020). *physketch: Drawing phylogenetic objects free-hand*. R package version 0.2.
- 593 Revell, L. J. and Harmon, L. J. (2022). *Phylogenetic Comparative Methods in R*. Princeton University Press.
- 594 Sackin, M. J. (1972). “good” and “bad” phenograms. *Systematic Biology*, 21(2):225–226.
- 595 Satler, J. D., Herre, E. A., Jandér, K. C., Eaton, D. A., Machado, C. A., Heath, T. A., and Nason, J. D.
596 (2019). Inferring processes of coevolutionary diversification in a community of panamanian strangler figs and
597 associated pollinating wasps. *Evolution*, 73(11):2295–2311.
- 598 Simmons, B. I., Sweering, M. J., Schillinger, M., Dicks, L. V., Sutherland, W. J., and Di Clemente, R. (2019).
599 bmotif: A package for motif analyses of bipartite networks. *Methods in Ecology and Evolution*, 10(5):695–701.
- 600 Sinaimeri, B., Urbini, L., Sagot, M.-F., and Matias, C. (2023). Cophylogeny reconstruction allowing for multiple
601 associations through approximate bayesian computation. *Systematic Biology*, 72(6):1370–1386.

- 602 Stadler, T. (2013). How can we improve accuracy of macroevolutionary rate estimates? *Systematic biology*,
603 62(2):321–329.
- 604 Stolzer, M., Lai, H., Xu, M., Sathaye, D., Vernot, B., and Durand, D. (2012). Inferring duplications, losses,
605 transfers and incomplete lineage sorting with nonbinary species trees. *Bioinformatics*, 28(18):i409–i415.
- 606 Sunnåker, M., Busetto, A. G., Numminen, E., Corander, J., Foll, M., and Dessimoz, C. (2013). Approximate
607 bayesian computation. *PLoS Computational Biology*, 9(1):e1002803.
- 608 Van Dam, M. H., Parisotto, A., Medina, M. N., Cabras, A. A., Gutiérrez-Trejo, N., Wilts, B. D., and Lam,
609 A. W. (2024). Biogeography confounds the signal of cospeciation in batesian mimicry. *Current Biology*,
610 34(23):5554–5563.
- 611 Yoder, J. B. and Nuismer, S. L. (2010). When does coevolution promote diversification? *The American
Naturalist*, 176(6):802–817.
- 613 Zeng, Y. and Hembry, D. H. (2024). Coevolution-induced selection for and against phenotypic novelty shapes
614 species richness in clade co-diversification. *Journal of Evolutionary Biology*, 37(12):1510–1522.
- 615 Zeng, Y. and Wiens, J. J. (2021). Species interactions have predictable impacts on diversification. *Ecology
Letters*, 24(2):239–248.