# REPORT - PROJECT 01 - GROUP 5

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### INTRODUCTION

Drug repurposing describes the strategy of reusing drugs that have already been approved for other medical applications. These drugs can obtain a new purpose by using them in a different therapeutic indication. This approach offers several advantages compared to the development and research of novel drugs. Drug repurposing allows researchers to use already approved pharmaceuticals, which helps them face less obstacles regarding potential side effects and the general approval of these drugs. This is due to the fact that the safety of the drugs has already been tested and validated in prior researches. Therefore, costs and legal requirements can be reduced which results in a quicker development of new treatment options (Pushpakom et al. 2019).

According to the WHO<sup>1</sup>, cancer is the second leading cause of death in the United States and a major global health problem. In addition, the progress of diagnosing and treating this disease was set back due to the coronavirus disease 2019 (COVID-19) pandemic which resulted in reduced health care capacities (Siegel et al. 2021). Hence, finding new treatment options by repurposing drugs in order to safe resources and time seems to be essential for our society.

For our project, we used data obtained from a large screening of 1396 oncological and non-oncological drugs that were tested on 481 different cancer cell lines (Corsello et al. 2020). While also working with the whole data consisting of several different cancer cell lines, in our analysis we particularly focused on the included 30 ovarian cancer cell lines. The American Cancer Society states that ovarian cancer causes more deaths than any other form of cancer being related to the female reproductive system.<sup>2</sup>

In our project, we aimed to answer two main questions:

- 1. How do different drugs influence ovary cancer cell lines and which are particularly noticeable? How do these drugs affect the other cell lines?
- 2. Is the sensitivity of different drugs on ovary cancer cell lines connected to specific cancerrelated genes or gene knock-outs?

We subdivided our project into smaller steps in order to find answers to these questions.

First, we analyzed the drugs and their influence on the proliferation of cancer cell lines. We were able to identify the most promising drugs and categorized them based on their mechanism of action (MOA). In addition, we analyzed to what extent the used drug dosage had an effect on the proliferation values of the cancer cell lines and found out that in most cases the higher the dosage the greater the effect on proliferation. We also investigated the drug response of the ovarian cancer cell lines based on certain gene expression patterns and knockdown scores which showed less correlations than we initially anticipated. Finally, we used the data to perform a linear regression model in order to predict drug efficiency from certain gene mutations or expressions.

<sup>&</sup>lt;sup>1</sup>https://www.who.int/health-topics/cancer#tab=tab 1

<sup>&</sup>lt;sup>2</sup>https://www.cancer.org/cancer/ovarian-cancer/about/key-statistics.html

# MOST EFFECTIVE DRUGS AMONG OVARIAN CELL LINES

#### GENERAL OVERVIEW

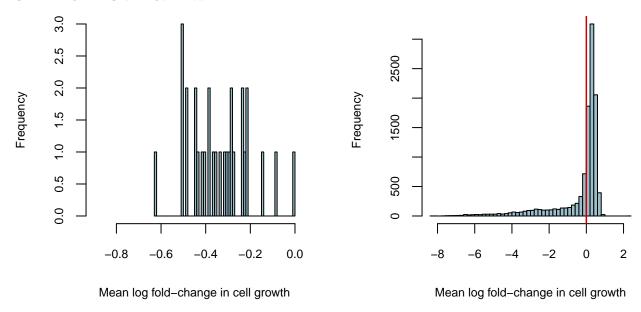


Figure 1: **1.a.** Ovarian Cancer Cell Lines Mean Sensitivity towards Drugs **1.b.** Mean Drug Efficiency regarding Proliferation in Ovarian Cell Lines

Using the provided prism data frame, we extracted the ovarian DepMap\_IDs with their proliferation values after drug treatment. In order to get a general overview of our data, we computed two histograms showing the distribution of the mean sensitivity across all ovarian cell lines and all drugs (see Fig. 1). Fig. 1.a. shows that all ovarian cancer cell lines had a mean sensitivity towards the drugs of less than or equal to zero, which showed us that in general, the applied drugs seemed to have a promising effect on their cell growth. The right plot (Fig. 1.b.) shows all values for the mean drug efficiency in ovarian cell lines. Values below zero indicate negative proliferation. In total, 8652 drugs lie below zero. This means that these drugs generally—with regard to their mean values—caused a reduction in cell growth when they were administered to the ovarian cell lines.

#### SETTING A THRESHOLD

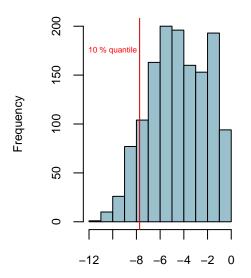
In our next step, we further analyzed the prism data set. Our aim was to determine the strongest effect each drug had on the ovarian cancer cell lines in terms of proliferation values - independent of the dosage that was used. For each drug we chose its minimum value which reflects the strongest negative effect on proliferation of the ovarian cell lines. In order to be able to work with a manageable amount of drugs, we decided to set a threshold at the 10 % quantile (see Fig. 2.a.). By doing so, we could focus on the 140 most effective drugs which caused the lowest negative proliferation values in the ovarian cancer cell lines.

#### MECHANISM OF ACTION

While further analyzing the drugs that lie below the 10 % quantile, we focused on categorizing them after their mechanism of action which we found in the data set prism.treat. According to the original study, clustering by the MOAs seems to be a good choice in order to determine whether the drugs are normally used for treating cancer or not (Corsello et al. 2020).

Our results are shown in Fig. 2.b. These are the 45 most common MOAs regarding the drugs that had the most promising effects on the ovarian cancer cell lines.

We further investigated these MOAs and came to the following results.



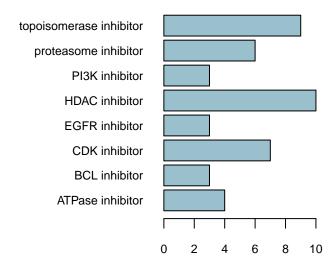


Figure 2: **2.a.** Most Effective Drug-Proliferation Values in Ovarian Cell Lines **2.b.** Most Frequent MOAs Among the Most Effective Drugs

10 out of the 140 drugs below our threshold are Histone deacetylase (HDAC) inhibitors. Their targets include among other things HDAC3, HDAC4, HDAC8. These are epigenetic targets for noncancer drug repurposing candiates (Moreira-Silva et al. 2020) and were also found to be potential anti-cancer agents for inhibiting cancer cell migration and invasion in ovarian cancer (Ahn et al. 2012).

With 9 occurrences, the topoisomerase inhibitors are the second most frequent MOAs we found. However, this is not surprising since these drugs are commonly used in cancer therapy, thus they showed strong effects on the cancer cell lines (Parchment and Pessina 1998). Interestingly, one of the drugs, mitoxantrone, is also used in the disease area neurology/psychiatry and multiple sclerosis<sup>3</sup>.

In addition, cyclin-dependent kinase (CDK) inhibitors appeared seven times in our findings regarding the most effective and common MOAs for treating ovarian cancer. Especially the targets CDK4 and CDK6 showed promising results in a different study (Iyengar et al. 2018) as a new therpeutic approach to treat ovarian cancer.

We found 6 proteasome inhibitors to be most effective against ovarian cancer cell lines. They are an important class of drugs against multiple myeloma and cell lymphoma. However, they are currently in clinical trials for other types of cancer (Fricker 2020) which is why it makes sense that we also found them to have promising effects in our analysis.

In a different study, researchers found that epidermal growth factor receptor (EGFR) is not a suitable target for ovarian cancer therapy (Mehner et al. 2017) and seems to be more effective for lung cancer and pancreas cancer therapy. However, we found three EGFR inhibitors with effective impacts on ovarian cancer cell lines.

The remaining three MOAs: Phosphoinositide 3-kinase (PI3K)- (Vanhaesebroeck et al. 2021), B-cell lymphoma (BCL)- (Montero and Letai 2018) and ATPase-inhibitors (Mijatovic, Dufrasne, and Kiss 2012), are all cancer related MOAs. Therefore, all of them are commonly used in cancer therapy which is why it is not surprising that they appeard in our findings.

# DIMENSION REDUCTION

Our next step included performing k-means and finding the optimal number of clusters which we visualized by computing a principal component analysis (PCA).

<sup>&</sup>lt;sup>3</sup>National Center for Biotechnology Information (2021). PubChem Compound Summary for CID 4212, Mitoxantrone. Retrieved July 11, 2021 from https://pubchem.ncbi.nlm.nih.gov/compound/Mitoxantrone.

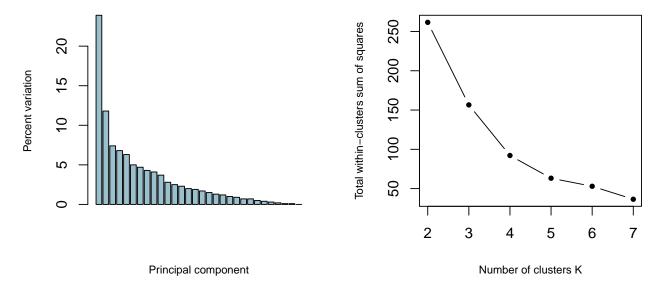


Figure 3: **3.a.** Variance Explained by Mechanism of Action PCs **3.b.** Elbow Method for Finding Optimal Number of Clusters

Fig. 3.a. shows that the majority of the variance is explained by using the first two principal components. In order to find the most optimal number of clusters for our PCA of PC1 and PC2, we used the elbow method. As a result, the optimal number of clusters seems to be 4 (see Fig. 3.b.).

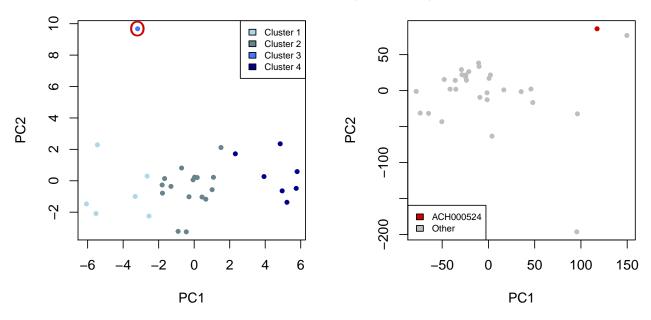


Figure 4: **4.a.** PC1 vs. PC2 for Ovary Cell Line Clusters **4.b.** PC1 vs. PC2 for CN Values in Ovary Cell Lines

### **OUTLIER**

Fig. 4.a. shows the clustering for the ovarian cell lines according to their reactions towards the 45 selected drugs (see Fig. 2.b.). They do not cluster in completely distinct clusters but one cell line (ACH-000524) forms its own cluster. We found that interesting, which is why we looked further into that one. According to the data set prism.cl, the outlier cell line belongs to the most prominent lineage subtype: ovary adenocarcinoma. This subtype makes up 28 of the 30 lineage subtypes in our data with the remaining two being brenner tumor

and ovary carcinoma. This is why we figured that the lineage subtype could not have been the reason for the cell line forming its own cluster.

We performed several PCAs with the other provided data sets (prism.achilles, prism.ecp and prism.cnv) and inspected the behavior of the outlier cell line. When we analyzed the gene transcripts per million (TPM) values and the gene knockdown scores, the cell line ACH-000524 clustered in the biggest clusters together with the other cell lines (not shown). However, the cell line did show a different behavior and clustered separately from the majority according to its gene copy number (CN) values (see Fig. 4.b.). This led us to the hypothesis that its differing behavior (see Fig. 4.a.) could be a result of distinct CN values.

#### GENERALLY EFFECTIVE DRUGS

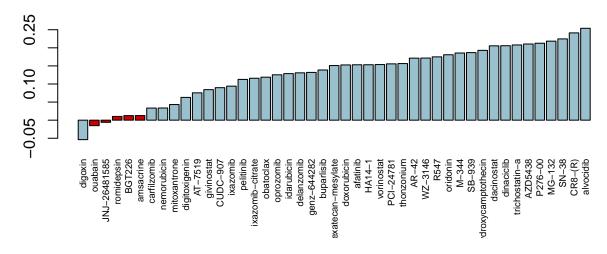


Figure 5: Contribution to PC1

In Fig. 5, the contribution to PC1 for ovary cell line clusters (see Fig. 4.a.) is shown. Drugs with values close to zero show a similar effect on the proliferation among all cell lines - thus, these are generally effective when administered to ovary cancer cell lines. This is the case for the following drugs (see highlighted red in Fig. 5).

The ATPase inhibitor ouabain is normally used to treat hypertension and cardiac arrhythmia - however, a study found that it can have promising effects as a cancer treatment for ER+ breast cancer (Chen et al. 2006). In another study, researchers found that one in three ovarian cancers is ER+ (Harding et al. 1990), which leads to the conclusion that this is why we found that ouabain generally showed positive effects among the ovarian cancer cell lines.

The drugs JNJ-26481585 (Arts et al. 2009) and romidepsin (Bertino and Otterson 2011) are HDAC inhibitors that, as mentioned before, in general show promising effects on ovarian cancer cell lines. Therefore, it makes sense that we found them among the five generally most effective drugs.

BGT226 is a PI3K inhibitor that is actually being considered as a potential therapeutic treatment option in different solid tumors, including ovarian cancer (Simioni et al. 2015).

Amsacrine, a topoisomerase inhibitor, is a chemotherapy drug that is commonly used for acute leukemia as well as Hodgkin's and non-Hodgkin's lymphomas (Ketron et al. 2012). Hence, a general effect on the proliferation of other cancer cell lines such as ovarian cancer seems to be logical.

### REFERENCES

- Ahn, Mee Young, Dong O. Kang, Yong Jin Na, Sungpil Yoon, Whan Soo Choi, Keun Wook Kang, Hae Young Chung, Jee H. Jung, Do Sik Min, and Hyung Sik Kim. 2012. "Histone Deacetylase Inhibitor, Apicidin, Inhibits Human Ovarian Cancer Cell Migration via Class II Histone Deacetylase 4 Silencing." Cancer Letters 325 (2): 189–99. https://doi.org/https://doi.org/10.1016/j.canlet.2012.06.017.
- Arts, Janine, Peter King, Ann Mariën, Wim Floren, Ann Beliën, Lut Janssen, Isabelle Pilatte, et al. 2009. "JNJ-26481585, a Novel "Second-Generation" Oral Histone Deacetylase Inhibitor, Shows Broad-Spectrum Preclinical Antitumoral Activity." *Clinical Cancer Research* 15 (22): 6841–51. https://doi.org/10.1158/1078-0432.CCR-09-0547.
- Bertino, Erin M, and Gregory A Otterson. 2011. "Romidepsin: A Novel Histone Deacetylase Inhibitor for Cancer." Expert Opinion on Investigational Drugs 20 (8): 1151–58. https://doi.org/10.1517/13543784.201 1.594437.
- Chen, Jin-Qiang, Ruben G. Contreras, Richard Wang, Sandra V. Fernandez, Liora Shoshani, Irma H. Russo, Marcelino Cereijido, and Jose Russo. 2006. "Sodium/Potasium ATPase (Na+, k+-ATPase) and Ouabain/Related Cardiacglycosides: A New Paradigm for Development of Anti- Breast Cancer Drugs?" Breast Cancer Research and Treatment 96 (1): 1–15. https://doi.org/10.1007/s10549-005-9053-3.
- Corsello, Steven M., Rohith T. Nagari, Ryan D. Spangler, Jordan Rossen, Mustafa Kocak, Jordan G. Bryan, Ranad Humeidi, et al. 2020. "Discovering the Anticancer Potential of Non-Oncology Drugs by Systematic Viability Profiling." *Nature Cancer* 1 (2): 235–48. https://doi.org/10.1038/s43018-019-0018-6.
- Fricker, Lloyd D. 2020. "Proteasome Inhibitor Drugs." Annual Review of Pharmacology and Toxicology 60 (1): 457–76. https://doi.org/10.1146/annurev-pharmtox-010919-023603.
- Harding, M., S. Cowan, D. Hole, L. Cassidy, H. Kitchener, J. Davis, and R. Leake. 1990. "Estrogen and Progesterone Receptors in Ovarian Cancer." Cancer 65 (3): 486–91. https://doi.org/10.1002/1097-0142(19900201)65:3%3C486::AID-CNCR2820650319%3E3.0.CO;2-C.
- Iyengar, Mangala, Patrick O'Hayer, Alex Cole, Tara Sebastian, Kun Yang, Lan Coffman, and Ronald J. Buckanovich. 2018. "Cdk4/6 Inhibition as Maintenance and Combination Therapy for High Grade Serous Ovarian Cancer." Oncotarget 9 (21): 15658–72. https://pubmed.ncbi.nlm.nih.gov/29644000.
- Ketron, Adam C., William A. Denny, David E. Graves, and Neil Osheroff. 2012. "Amsacrine as a Topoisomerase II Poison: Importance of Drug-DNA Interactions." *Biochemistry* 51 (8): 1730–39. https://doi.org/10.1021/bi201159b.
- Mehner, Christine, Ann L. Oberg, Krista M. Goergen, Kimberly R. Kalli, Matthew J. Maurer, Aziza Nassar, Ellen L. Goode, et al. 2017. "EGFR as a Prognostic Biomarker and Therapeutic Target in Ovarian Cancer: Evaluation of Patient Cohort and Literature Review." Genes & Cancer 8 (5-6): 589–99. https://pubmed.ncbi.nlm.nih.gov/28740577.
- Mijatovic, Tatjana, François Dufrasne, and Robert Kiss. 2012. "Na+/k+-ATPase and Cancer." *Pharmaceutical Patent Analyst* 1 (1): 91–106. https://doi.org/10.4155/ppa.12.3.
- Montero, Joan, and Antony Letai. 2018. "Why Do BCL-2 Inhibitors Work and Where Should We Use Them in the Clinic?" Cell Death and Differentiation 25 (1): 56–64. https://doi.org/10.1038/cdd.2017.183.
- Moreira-Silva, Filipa, Vânia Camilo, Vítor Gaspar, João F. Mano, Rui Henrique, and Carmen Jerónimo. 2020. "Repurposing Old Drugs into New Epigenetic Inhibitors: Promising Candidates for Cancer Treatment?" *Pharmaceutics* 12 (5): 410. https://doi.org/10.3390/pharmaceutics12050410.
- Parchment, R. E., and A. Pessina. 1998. "Topoisomerase i Inhibitors and Drug Resistance." Cytotechnology 27 (1-3): 149–64. https://doi.org/10.1023/A:1008008719699.
- Pushpakom, Sudeep, Francesco Iorio, Patrick A. Eyers, K. Jane Escott, Shirley Hopper, Andrew Wells, Andrew Doig, et al. 2019. "Drug Repurposing: Progress, Challenges and Recommendations." *Nature Reviews Drug Discovery* 18 (1): 41–58. https://doi.org/10.1038/nrd.2018.168.

- Siegel, Rebecca L., Kimberly D. Miller, Hannah E. Fuchs, and Ahmedin Jemal. 2021. "Cancer Statistics, 2021." CA: A Cancer Journal for Clinicians 71 (1): 7–33. https://doi.org/https://doi.org/10.3322/caac.21654.
- Simioni, Carolina, Alice Cani, Alberto M. Martelli, Giorgio Zauli, Ayman A. M. Alameen, Simona Ultimo, Giovanna Tabellini, James A. McCubrey, Silvano Capitani, and Luca M. Neri. 2015. "The Novel Dual Pi3k/mTOR Inhibitor NVP-Bgt226 Displays Cytotoxic Activity in Both Normoxic and Hypoxic Hepatocarcinoma Cells." Oncotarget 6 (19): 17147–60. https://pubmed.ncbi.nlm.nih.gov/26003166.
- Vanhaesebroeck, Bart, Matthew W. D. Perry, Jennifer R. Brown, Fabrice André, and Klaus Okkenhaug. 2021. "Pi3k Inhibitors Are Finally Coming of Age." Nature Reviews Drug Discovery, June. https://doi.org/10.1038/s41573-021-00209-1.