NZ SRW Calving interval report

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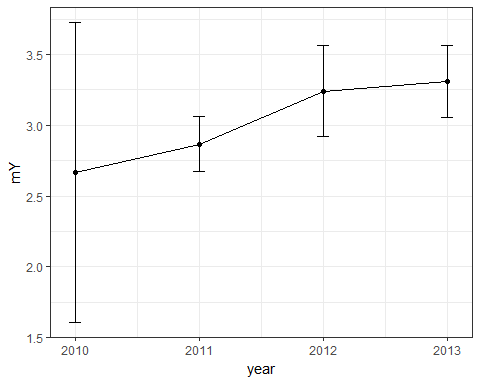
# Raw data

The data is then converted into intervals for each of the years 2010, 2011, 2012 and 2013.

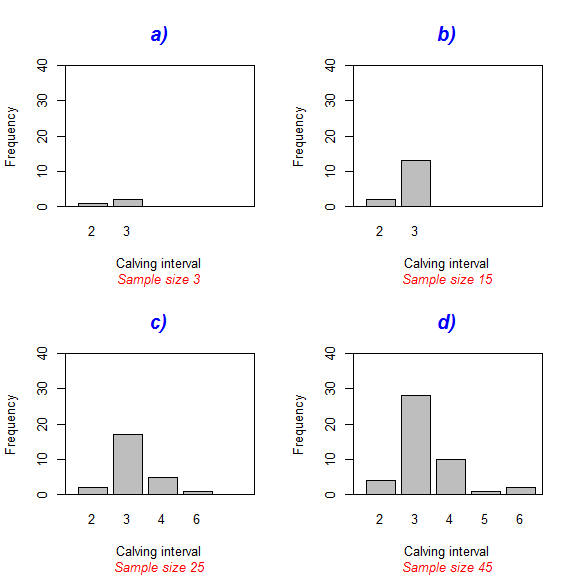
A table of the raw results as the year increase

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| year | n | mY | low.qt | high.qt | sd |
| 2010 | 3 | 2.666667 | 1.605851 | 3.727482 | 0.5773503 |
| 2011 | 15 | 2.866667 | 2.673022 | 3.060312 | 0.3518658 |
| 2012 | 25 | 3.240000 | 2.919170 | 3.560830 | 0.7788881 |
| 2013 | 45 | 3.311111 | 3.056488 | 3.565734 | 0.8480518 |

The raw intervals can be presented as different means and confidence intervals as the length of the study increases:



We can also plot these as bar charts that show this more obviously:

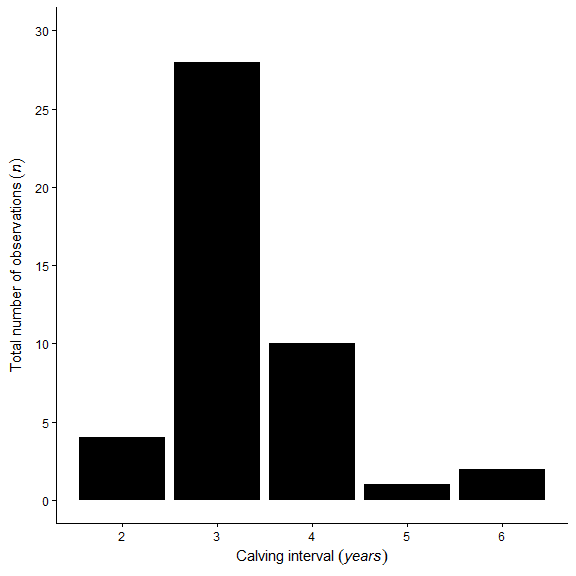


What is interesting is that in 2011 there were more intervals that subsequently reduced the standard error of the estimate and the precision increased.

**My idea of why this has happened is as follows:**

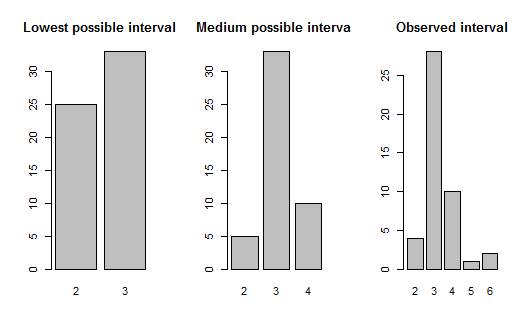
At the time of collection in 2011 the number of possible intervals greater than 5 was very unlikely (as there where only 5 years of research). This meant that the overall error of the estimate was reduced as there was very few "chances" of obtaining a calving interval of 5 or 6 even though there may have been quite a few. The following estimates then picked these up and the error in the estimate increased again.

### Publication plot (Figure 2)

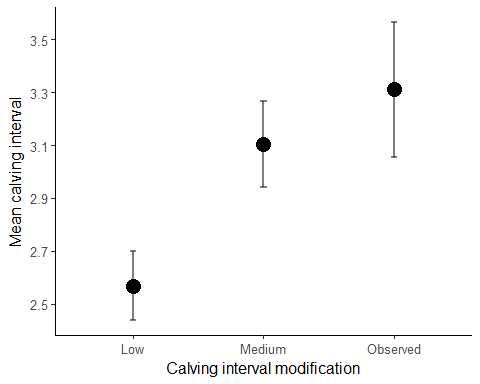


# Missing intervals (Bradford et al. 2008)

This is a way to see what the calving interval might be if we had in fact missed calving events that happened before or after the study period.



### Publication plot (Not included in final publication)



The error gets smaller as the variance in the estimate decreases. We are artificially doing this here by modifying data to match what maybe more biologically possible.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Interval | Sample size | Mean | Lower limit | Higher limit | SD |
| Low | 58 | 2.568966 | 2.437666 | 2.700265 | 0.4995461 |
| Medium | 48 | 3.104167 | 2.943089 | 3.265244 | 0.5550382 |
| Observed | 45 | 3.311111 | 3.056488 | 3.565734 | 0.8480518 |

# Bootstrapping

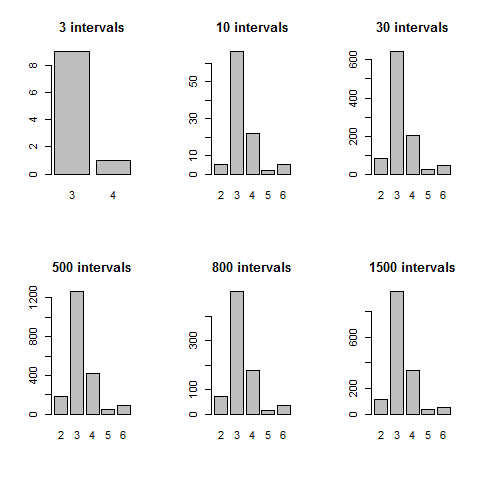
Below is the estimates from other populations of SRWs around the world.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Sample size | Mean | Lower limit | Higher limit | SE | Author | Location |
| NA | 3.12 | 3.07 | 3.17 | NA | Best et al. 2001 | South Africa |
| 1504 | 3.15 | 3.11 | 3.18 | NA | Best et al. 2005 | South Africa (1971-2003 Updated) |
| NA | 3.16 | 3.13 | 3.19 | NA | Brandao et al 2010 | South Africa ( 1971-2006 Updated) |
| NA | 3.35 | NA | NA | 0.05 | Cooke et al. 2001 | Argentina |
| 749 | 3.42 | NA | NA | 0.11 | Cooke et al. 2003 | Argentina |
| NA | 3.63 | NA | NA | 0.13 | Burnell 2001 | Australia |

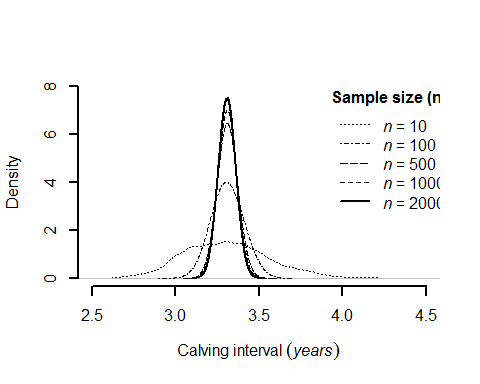
**Table of parameters:** This is taken directly from my Master's thesis. There may be updated estimates that need to be checked here.

## Single bootstrap with increased sample size

Bootstrap mean calving interval 1000 times and save the mean for each bootstrap sample. Here I investigate the effect of a sample size of 10,100,1000,2000 from the observed NZ calving interval.



### Publication plot (Figure 3)



# Referees additional analysis

## Suggestion one

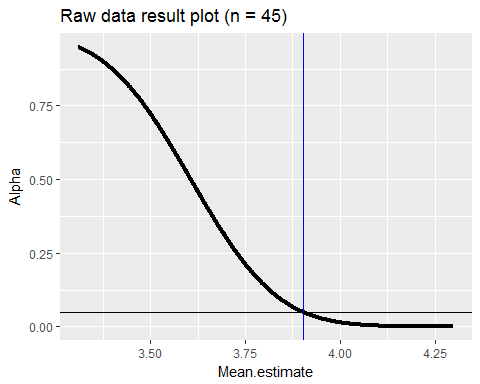
**"If another 45 samples were collected over the next 8 years, but the mean moved from 3.31 to e.g., 3.7 years, would it be detectable with alpha = 0.05, 0.10 … ? Given that other authors have hypothesised that poor food availability would translate to increased calving intervals/calf mortalities (e.g., Cooke et al 2003, Rowntree et al 2013), being able to detect such a change would be important."**

To do this it is simply a old school power analysis with the focus on the type one error (alpha) as below:

#observed sample  
rev.one <- bdata$RealCI[1:45]  
  
#sample 45 times  
sample.true <- year2013$interval  
  
#power analysis  
pwr.test.results <- power.t.test(n = 45,# sample size  
 delta = seq(0,0.99,0.001), #difference between means  
 sd = sd(sample.true), #observed variation  
 alternative = "one.sided", #observed test type  
 sig.level = 0.05) #significance level  
  
#additional packages are avaliable for more complex analysis  
#but have not done this as don't think it is needed

Here I have looked at the effect of the difference in the mean to range from the same 3.31 (delta = 0) to 4.31 (delta = 1) with the same variation as observed in the observed data.

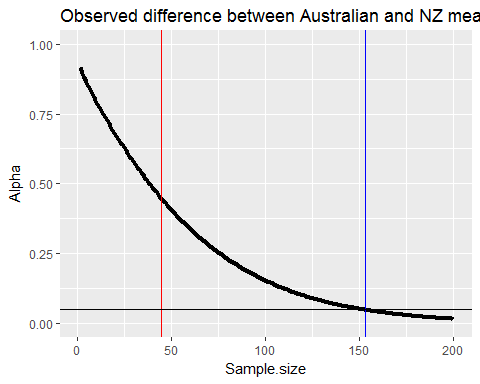
## Power Mean.difference Alpha Mean.estimate  
## 1 0.9501505 0.593 0.04984946 3.903



It is interesting to note that we the minimum mean difference with a sample size of 45 that a difference between populations was observed at the 95% level would be a mean calving interval of 3.9.

Below is an additional analysis to look at the number of samples needed to find a difference the same or greater than the mean estimate for the Australian population recorded as 3.63.

## Power Sample.size Alpha  
## 1 0.9503366 153 0.0496634



Red line represents the current sample size of NZ population estimate and blue line represents the size needed to have alpha=0.05

**Important note**

I do not this is the best approach because...

if the sampling continues over the next 8 years, the number of observations will not increase in a linear function as the number of individuals available for recapture is growing (more individuals in database) at a different rate to the capture probability (that is hopefully constant). I will do the simulation recommended because both the editor and reviewer wanted it.

I previously approached this problem by using bootstrapping and looking at the effect of the sampling size (n) increasing.

## Suggestion three

**"Is there any information for the females with the potentially missed calving events in the intervening years; e.g., were they seen without calves?"**

### Individual status

I only have data for these overall data points up to 2012 so have done the analysis with this and can just add an additional year to check this.

To do this the following steps are taken:

1. Raw data I have currently looks like so:

## Observations: 180  
## Variables: 10  
## $ ID <fctr> AI06006, AI06007, AI06015, AI06022, AI06038, AI10...  
## $ X2006 <int> 1, 2, 2, 1, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,...  
## $ X2007 <int> 0, 1, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,...  
## $ X2008 <int> 1, 0, 1, 0, 2, 0, 1, 0, 0, 0, 0, 0, 0, 0, 1, 0, 0,...  
## $ X2009 <int> 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,...  
## $ X2010 <int> 0, 0, 2, 0, 0, 6, 6, 5, 5, 3, 4, 2, 5, 4, 5, 3, 2,...  
## $ X2011 <int> 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,...  
## $ X2012 <int> 0, 1, 2, 4, 0, 0, 0, 0, 0, 0, 5, 0, 0, 12, 0, 0, 0...  
## $ total <int> 2, 4, 8, 5, 3, 6, 7, 5, 5, 3, 9, 2, 5, 16, 6, 3, 2...  
## $ X..yrs.seen <int> 2, 3, 5, 2, 2, 1, 2, 1, 1, 1, 2, 1, 1, 2, 2, 1, 1,...

## Observations: 41  
## Variables: 8  
## $ ID <fctr> AI10124, AI10070, AI10086, AI08340, AI08341, AI...  
## $ Yr.first.seen <int> 2007, 2008, 2007, 2008, 2008, 2008, 2008, 2008, ...  
## $ Calves <int> 2007, 2008, 2007, 2008, 2008, 2008, 2008, 2008, ...  
## $ Calves.1 <int> 2010, 2010, 2010, 2011, 2011, 2011, 2011, 2011, ...  
## $ Calves.2 <int> 2013, 2013, 2013, NA, NA, NA, NA, NA, NA, NA, NA...  
## $ Interval.1 <int> 3, 2, 3, 3, 3, 3, 3, 3, 3, 3, 3, 3, 3, 3, 2, 6, ...  
## $ Interval.2 <int> 3, 3, 3, NA, NA, NA, NA, NA, NA, NA, NA, NA, NA,...  
## $ X <lgl> NA, NA, NA, NA, NA, NA, NA, NA, NA, NA, NA, NA, ...

1. The two data sets can be combined to reduce the data down to just individuals that have been seen with a calf at least once.

This is important because we do not know the reproductive state of individuals until they are first seen with a calf so therefore any individuals that are in the database and have not been identified with a calf maybe juvenile and therefore not reproducing and we would be overestimating the number of possible reproductive events.

## Observations: 237  
## Variables: 7  
## $ ID <chr> "AI06006", "AI06007", "AI06015", "AI06022", "AI0...  
## $ year <chr> "X2006", "X2006", "X2006", "X2006", "X2006", "X2...  
## $ count <int> 1, 2, 2, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, ...  
## $ Yr.first.seen <int> NA, NA, NA, 2006, NA, NA, NA, 2007, 2007, NA, NA...  
## $ Calves <int> NA, NA, NA, 2006, NA, NA, NA, 2007, 2007, NA, NA...  
## $ Calves.1 <int> NA, NA, NA, 2012, NA, NA, NA, 2013, 2010, NA, NA...  
## $ Calves.2 <int> NA, NA, NA, NA, NA, NA, NA, NA, 2013, NA, NA, 20...

## # A tibble: 1 x 4  
## ID year Calves Calves.1  
## <chr> <chr> <int> <int>  
## 1 AI09216 2007 2009 2011

1. And overall between 2006 and 2012 only a single individual (AI09216) was identified as being a mother and was seen in 2007 without a calf.

This individual had also not been seen with a calf until 2009 so even this individual may have been juvenile at this stage.

## Suggestions on variations of detection error and removing/increasing observed estimates

These comments are about additional data manipulation and the following estimates are plotted in the same figure as the Bradford et al 2008 modifications below.

**"Another possibility not explored is that the reported two year calving intervals could have been an error; what would the mean calving interval be in this case?"**

greater.than.2 <- sample.true[sample.true>2]  
  
#greater.than.2  
mean.2<-sum(greater.than.2)/length(greater.than.2)  
s.2<-sd(greater.than.2)  
SE.2<-s2013/(sqrt(length(greater.than.2)))  
n.2<-length(greater.than.2)  
low.qt.2<- mean.2-(qt(0.975,length(greater.than.2))\*SE.2)  
high.qt.2 <- mean.2+(qt(0.975,length(greater.than.2))\*SE.2)  
  
#add it to the table from bradford data  
Sumtable[4,] <- c("miss2year",n.2,mean.2,low.qt.2,  
 high.qt.2,sd(greater.than.2))

**"The existing simulation could be improved by assuming imperfect detection of calving events. For example, how does the variance around the estimate of mean calving interval change when 5% of calving events are not detected, increasing the number of 5 or 6 year observed calving intervals?"**

########################### 2.2%  
#parameters  
boots <- 1000  
n <- c(1:1000)  
  
###round all percentages upwards  
detect1 <- 44 # (45\*1.02) - 45 = 0.9  
detect2 <- 42 # (45\*1.05) - 45 = 2.25  
detect3 <- 40 # (45\*1.10) - 45 = 4.5  
  
sample2 <-rep(NA, 1000)  
sample5 <-rep(NA, 1000)  
sample10 <-rep(NA, 1000)  
  
for (i in 1:boots) {  
 sample2[i]<-mean(sample(year2013$interval,detect1,replace=T))  
 sample5[i]<-mean(sample(year2013$interval,detect2,replace=T))  
 sample10[i]<-mean(sample(year2013$interval,detect3,replace=T))  
 } #i  
  
######################estimates##############  
sample2 <- sort(sample2)  
#low = 25/1000  
sample2.2.5 <- sample2[25]  
#median  
sample2.50 <- sample2[500]  
#high = 975/1000  
sample2.975 <- sample2[975]  
  
sample5 <- sort(sample5)  
#low = 25/1000  
sample5.2.5 <- sample5[25]  
#median  
sample5.50 <- sample5[500]  
#high = 975/1000  
sample5.975 <- sample5[975]  
  
sample10 <- sort(sample10)  
#low = 25/1000  
sample10.2.5 <- sample10[25]  
#median  
sample10.50 <- sample10[500]  
#high = 975/1000  
sample10.975 <- sample10[975]  
  
  
#add it to the table from bradford data  
Sumtable[5,] <- c("detect1",detect1,sample2.50,sample2.2.5,sample2.975,NA)  
Sumtable[6,] <- c("detect2",detect2,sample5.50,sample5.2.5,sample5.975,NA)  
Sumtable[7,] <- c("detect5",detect3,sample10.50,sample10.2.5,sample10.975,NA)

**"or increasing the number of 5 or 6 year observed calving intervals?"**

Here I have added twice an many 5 and 6 year intervals (5yr = 2, 6yr = 4)

longer5.6 <- c(sample.true,5,6,6)  
  
#greater.than.2  
mean.56<-sum(longer5.6)/length(longer5.6)  
s.56<-sd(longer5.6)  
SE.56<-s.56/(sqrt(length(longer5.6)))  
n.56<-(length(longer5.6))  
low.qt.56<- mean.56-(qt(0.975,length(longer5.6))\*SE.56)  
high.qt.56 <- mean.56+(qt(0.975,length(longer5.6))\*SE.56)  
  
#add it to the table from bradford data  
Sumtable[8,] <- c("longer.56",n.56,mean.56,low.qt.56,high.qt.56,sd(longer5.6))  
  
###sort out numbering in dataframe  
Sumtable <- as.data.frame(Sumtable)  
  
 Sumtable$n <- as.numeric(as.character(Sumtable$n))  
 Sumtable$mY <- as.numeric(as.character(Sumtable$mY))  
 Sumtable$low.qt <- as.numeric(as.character(Sumtable$low.qt))  
 Sumtable$high.qt <- as.numeric(as.character(Sumtable$high.qt))  
 Sumtable$sd <- as.numeric(as.character(Sumtable$sd))  
 Sumtable$interval <- as.character(Sumtable$interval)

### Table of results

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Interval | Sample size | Mean | Lower limit | Higher limit | SD |
| Low | 58 | 2.568966 | 2.437666 | 2.700265 | 0.4995461 |
| Medium | 48 | 3.104167 | 2.943089 | 3.265244 | 0.5550382 |
| Observed | 45 | 3.311111 | 3.056488 | 3.565734 | 0.8480518 |
| miss2year | 41 | 3.439024 | 3.171549 | 3.706499 | 0.7761695 |
| detect1 | 44 | 3.318182 | 3.068182 | 3.568182 | NA |
| detect2 | 42 | 3.309524 | 3.095238 | 3.571429 | NA |
| detect5 | 40 | 3.300000 | 3.050000 | 3.575000 | NA |
| longer.56 | 48 | 3.458333 | 3.165307 | 3.751360 | 1.0097047 |

### Publication plot (Not included in final publication)

