**Setting Up the Chip Priming Station**

1. Assure the base plate in in position C
2. Assure the Syringe clip is in the lowest position

**Prepare the Gel-Dye Mix**

1. Allow all reagents to equilibrate to room temperature for 30 minutes
2. Add 15 ul of High Sensitivity DNA dye concentrate (blue) to a High Sensitivity DNA gel matrix vial (red)
3. Vortex the solution well and spin down. Transfer to spin filter
4. Centrifuge at 2250 \* g for 15 min. Protect solution from light. Store at 4C. Use prepared gel dye mix within 6 weeks of preparation

**Loading the Gel-Dye Mix**

1. Allow all reagents to equilibrate to room temperature for 30 minutes
2. Put a new High Sensitivity DNA chip on the chip priming station
3. Pipette 9 ul of gel Dye mix into the well marked with a “G” with a dark circle
4. Make sure the plunger is positioned at 1 mL and then close the chip priming station
5. Press the plunger until it is held by the clip. Wait 60 seconds and release the clip. Wait for 5 seconds and slowly pull the plunger to the 1 mL position
6. Open the chip priming station and pipette 9 ul of gel dye mix in the wells marked “G”

**Loading the Marker, Ladder, and samples**

1. Pipette 5 ul of marker (green) in all sample and ladder wells. Do not leave any wells empty
2. Pipette 1 ul of High Sensitivity DNA ladder (yellow) in the well marker with a ladder
3. In each of the 11 sample wells pipette 1 ul of sample
4. Put the chip in the adapter and vortex for 1 min at 2400 rpm
5. Run the chip in the Agilent 2100 Bioanlayzer instrument within 5 min