

DisperseR: Calculating Seed Dispersal In R

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Contents

1	Introduction	1
2	Generating Plot Map	2
2.1	generatePlotMap	2
3	Sampling The Plot Map	3
3.1	findAdultTrees	4
4	Calculating Parameters for the Ribbens Equation	5

1 Introduction

This is a small package intended to help users calculating seed dispersal in R. Although the R base machinery is capable of doing so, this package streamlines the process and enables you to focus more on the important aspects of data analysis instead of data generation or clean-up.

This code operates as follows. Ideally, you'll need a dataframe that contains the following data: (x,y) coordinates of each tree and seedling in a plot; and dbh measurements of any tree large enough. A tree is any individual that can be measured for diameter at breast height, and all trees are assumed to be reproductively active; a seedling is any individual that is new in the calendar year.

Spatial seed dispersal is characterized by a single equation,

$$R_i = STR * \sum_{k=1}^T \left(\frac{DBH_k}{30} \right)^\beta e^{-Dm_{ik}^3} * \left(\frac{1}{n} \right) \quad (1)$$

where n is a normalizer function that standardizes the equation to values between 0 and 1,

$$n = \int_0^\infty e^{-Dm_{ik}^3} \quad (2)$$

and where STR is the standardized number of tree recruits, DBH is the diameter at breast height, β is a modifier of DBH and STR, D is a species-specific parameter estimated by this equation, and m is the distance between the measured point i and adult tree k , summed over each adult tree ($k=1$ to T adult trees). These equations were originally established by Ribbens et al. (1994), in an experiment where seedling per m^2 along a belt transect were correlated to the number and size of any adults within a $20m$ radius.

The first piece of the equation, containing STR, establishes the number of recruits produced for a tree of a standard DBH (30cm), and the second piece of the equation establishes the mean density of recruits found in a $1m^2$ quadrat centered at m distance away from the parent tree. Finally, $\frac{1}{n}$ serves as a normalizer to standardize the equation across species.

The parameters STR and D are both needed by SORTIE-ND, an individual tree neighborhood dynamics forest gap model (say that five times fast!), to calculate seed dispersal for target species in its simulations. SORTIE-ND, unfortunately, does not come packaged with a magic bullet that offers species-specific parameters, and therefore, we must parameterize the model ourselves. This package is intended to help create estimates of both STR and D quickly, so that other parameters may be addressed.

What follows is a list of functions alongside example usage. To start, you must import or generate a plot map of all trees in a given area. This plot map must include a species identifier, an x coordinate, a y coordinate, and DBH (or NA) for each individual.

2 Generating Plot Map

2.1 generatePlotMap

We can generate a sample plot easily with `generatePlotMap()`. As you can see below, this function generates a plot map with NA's for seedlings and actual values of DBH for adult trees. See `?generatePlotMap()` for information on how to customize your random plot map.

```
> library(disperseR)
> myplot <- generatePlotMap()
> head(myplot)
```

	treeid	species	x	y	dbh	stage
1	1	1	379.6264	316.2253	NA	seedling
2	2	1	705.8937	686.4814	NA	seedling
3	3	1	743.1112	300.8632	NA	seedling
4	4	1	485.6738	631.5433	NA	seedling
5	5	1	757.9238	324.6329	NA	seedling
6	6	1	523.2300	298.1684	NA	seedling

```
> tail(myplot)
```

	treeid	species	x	y	dbh	stage
745	745	5	700.99618	839.6000	75.34190	tree
746	746	5	782.15567	919.0318	81.64657	tree
747	747	5	81.38552	974.0578	53.24793	tree
748	748	5	314.30493	517.1162	33.62520	tree
749	749	5	992.02656	275.7232	10.41214	tree
750	750	5	501.88144	606.8550	83.27242	tree

If you do have your own data, just make sure that it matches the column names of the plot map generated above, and also the data types. You can check the structure of a dataframe using `str()` and then `as.numeric()` or `as.character()` to adjust as needed. In our case, you need five columns: `treeid`, `species`, `x`, `y`, and `dbh`. `x`, `y`, and `dbh` should all be numeric. “species” can be a character vector or a numeric vector, as long as the species names are unique. “treeid” is a unique identifier for each tree. If you generate randomly, it will match the rownumbers of your resulting data.frame. You’ll also need a “stage” column, which is just a shortcut for figuring out if a record is for a seedling or a tree.

```
> ## exploring the structure of myplot
> str(myplot)

'data.frame':      750 obs. of  6 variables:
 $ treeid : int  1 2 3 4 5 6 7 8 9 10 ...
 $ species: int  1 1 1 1 1 1 1 1 1 1 ...
 $ x      : num  380 706 743 486 758 ...
 $ y      : num  316 686 301 632 325 ...
 $ dbh    : num  NA NA NA NA NA NA NA NA NA NA ...
 $ stage  : chr  "seedling" "seedling" "seedling" "seedling" ...

> ## if we needed to convert a column
> myplot$species <- as.numeric(myplot$species)
```

3 Sampling The Plot Map

Now that we have a plot map ready, we need to be able to sample the plot. Ribbens et al. (1994) sampled using a belt transect, stopping every so often to count all of the seedlings in a $1m^2$ plot, and all adult trees within 20m of the seedling plot. We have an advantage, in that we have the exhaustive map and can just sample for every individual or clump of seedlings there is.

We can do this with a “find and eliminate” approach. For each possible seedling, subsetted from the plot map, we can search around it at “m” distance to see if there are other seedlings. We can count the total number of seedlings in that box, store the (x,y) and number (n) in a data.frame, prevent the counted seedlings from getting re-counted, and move to the next row. This functionality is wrapped up in the `findSeedPlots()` function in `disperseR`, demonstrated below. Remember, your input data.frame will need all of the columns generated by `generatePlotMap()`.

```

> ## make a sample seed data.frame by subsetting the included
> ## expandedTrees data.frame..
> seeds <- myplot[myplot$stage=="seedling",]
> ## we'll need this one later...
> adults <- myplot[myplot$stage=="tree",]
> ## show the start of results
> myseedplots <- findSeedPlots(seeds, 1)
> head(myseedplots)

```

```

      x      y n
1 379.6264 316.2253 1
2 705.8937 686.4814 1
3 743.1112 300.8632 1
4 485.6738 631.5433 1
5 757.9238 324.6329 1
6 523.2300 298.1684 1

```

```

> ## What are the possible densities of our seedplots?
> unique(myseedplots$n)

```

```
[1] 1
```

Now that we have seedling density in our subplots, we need to figure out how many possible parent trees there are for each of the positive hits. We can do that using the `findAdultTrees()` function.

3.1 findAdultTrees

The `findAdultTrees()` function works by searching a full plot for trees that are “m” distance away from the points provided (seedling plot points).

```

> parentTrees <- findAdultTrees(myseedplots, adults, 20)
> head(parentTrees)

```

```

  treeid ri      m      dbh
1    136  1 14.995687 88.381839
2    416  1 13.445802 70.020780
3    435  1 19.449352 30.993858
4    590  1  8.931349  6.342278
5    136  1  5.846505 88.381839
6    717  1 10.383254 77.669515

```

```

> nrow(parentTrees)

```

```
[1] 65
```

Now, you can see that the random plot map that we’ve generated doesn’t work too well on its default values at getting “good” values for this equation.

“ri”, in particular, should have more than one unique value if it’s going to be used as a predictor value. Now that we’ve shown proof of concept for these functions, however, we’re going to get around the limitation of random data by using some data that comes in the package, known as `expandedTrees`. This data is obfuscated real data, as you’ll see below.

4 Calculating Parameters for the Ribbens Equation

Unfortunately, because the data above are randomly generated, they will not allow a NLS model to converge on a meaningful parameter set. To get around this and demonstrate the model, we’ve included a large plot-year dataset called “`expandedTrees`”. This dataset represents unique data when subsetting at the “`plot`” and “`measyear`” columns. You can use `?expandedTrees` to find out more about the dataset and how it functions. `expandedTrees` is generated from `ssdAllTrees` and `ssdPlotDesc`, so check those data.frames out too if you need more information.

Since `expandedTrees` is organized by plot and year, we can take one of those plot-year combinations and run the model. We will also need to generate a seedling map and all of the other steps from above.

Let’s look at `expandedTrees`, and take the first plot-year combination available.

```
> head(expandedTrees)
```

	plot	treeid	species	ingrowth	firstrec	deathyear	x	y	measyear	dbh
2	bellow	7093	PIMO	2003	2003	NA	100733.1	51405.46	2003	1.2
3	bellow	7093	PIMO	2003	2003	NA	100733.1	51405.46	2008	2.1
4	bellow	7093	PIMO	2003	2003	NA	100733.1	51405.46	2013	2.7
5	bellow	7094	PIMO	2004	2004	NA	100732.7	51405.07	2008	2.1
6	bellow	7094	PIMO	2004	2004	NA	100732.7	51405.07	2013	3.8
7	bellow	7095	PIMO	2004	2004	NA	100733.2	51405.02	2008	2.4

```

      stage
2 seedling
3      tree
4      tree
5 seedling
6      tree
7 seedling

> str(expandedTrees)
```

```
'data.frame':      51493 obs. of  11 variables:
 $ plot      : chr  "bellow" "bellow" "bellow" "bellow" ...
 $ treeid    : num  7093 7093 7093 7094 7094 ...
 $ species   : chr  "PIMO" "PIMO" "PIMO" "PIMO" ...
```

```

$ ingrowth : num 2003 2003 2003 2004 2004 ...
$ firstrec : num 2003 2003 2003 2004 2004 ...
$ deathyear: num NA NA NA NA NA NA NA NA NA ...
$ x         : num 100733 100733 100733 100733 100733 ...
$ y         : num 51405 51405 51405 51405 51405 ...
$ measyear  : num 2003 2008 2013 2008 2013 ...
$ dbh       : num 1.2 2.1 2.7 2.1 3.8 2.4 4.1 8.1 8.1 8.2 ...
$ stage     : chr "seedling" "tree" "tree" "seedling" ...

> ## get unique plot/year combos
> plotlist <- unique(expandedTrees[,c("plot", "measyear")])
> rownames(plotlist) <- 1:nrow(plotlist)
> ## count the number of adult trees in a plot/year combination
> plotlist$tree <- NA
> for(i in 1:nrow(plotlist)){
+   plotlist[i, "tree"] <- nrow(expandedTrees[expandedTrees$plot==plotlist[i, "plot"] & expa
+ }
> ## get number of seedlings in a plot/year combo
> plotlist$seedlings <- NA
> for(i in 1:nrow(plotlist)){
+   plotlist[i, "seedlings"] <- nrow(expandedTrees[expandedTrees$plot==plotlist[i, "plot"] &
+ }
> ## eliminate any plots that have only trees or seedlings
> plotlist <- plotlist[plotlist$tree!=0 & plotlist$seedlings!=0,]

For now, let's take a large plot, like trinity, in one of its middle years. The
data.frame knows exactly when seedlings established in middle years, because
plots were checked years for establishment. Let's do trinity in 2001:

> trinity01 <- expandedTrees[expandedTrees$plot=="trinity" & expandedTrees$measyear==2001,]
> nrow(trinity01[trinity01$stage=="seedling",])

[1] 431

> str(trinity01)

'data.frame':      2402 obs. of  11 variables:
 $ plot      : chr "trinity" "trinity" "trinity" "trinity" ...
 $ treeid    : num 1532 1534 1535 1536 1537 ...
 $ species   : chr "ABCO" "ABCO" "ABCO" "ABCO" ...
 $ ingrowth  : num NA NA NA NA NA NA NA NA NA ...
 $ firstrec  : num 1991 1991 1991 1991 1991 ...
 $ deathyear : num NA NA 2012 NA 2003 ...
 $ x         : num 88904 88903 88900 88900 88902 ...
 $ y         : num 48744 48741 48738 48737 48736 ...
 $ measyear  : num 2001 2001 2001 2001 2001 ...
 $ dbh       : num 15.3 13.9 7.5 15.5 2 18.3 32.5 10.6 22.5 17.2 ...
 $ stage     : chr "tree" "tree" "tree" "tree" ...

```

```

> ## set up for plot
> ## by stage
> trinity01$colors <- ifelse(trinity01$stage=="seedling", "red", "blue")
> ## by species
> specieslist <- unique(trinity01$species)
> for(i in 1:length(specieslist)){
+   trinity01[trinity01$species==specieslist[i],"pch"] <- as.numeric(i)
+ }

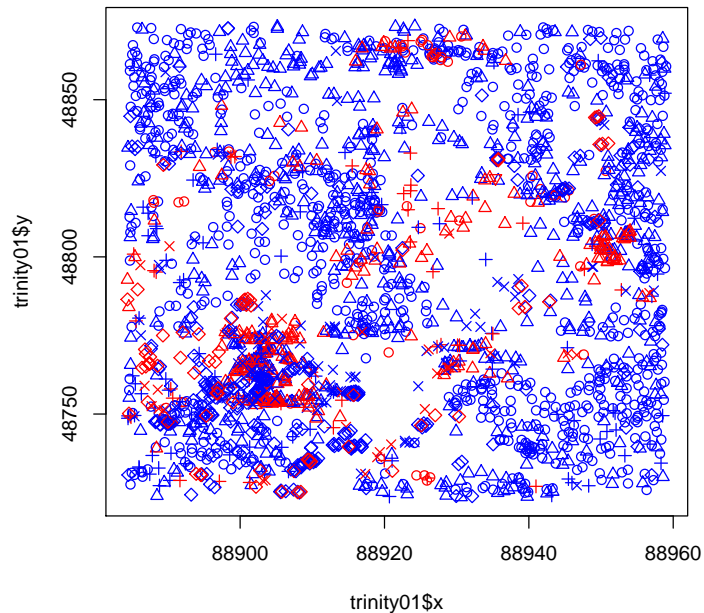
```

Now, let's take a look at the distribution of seedlings and adults by species. This is just a simple graph of trinity01, not scaled to dbh at all. See the code above for the color/species designations

```

> plot(trinity01$x, trinity01$y, pch=trinity01$pch, col=trinity01$colors)

```



And now let's separate out the seedlings from trees, and run our scripts to get parentTrees. The scripts also only deal with a single species, so we'll pick one.

```

> ## get seeds and adults ready
> trinSeeds <- trinity01[trinity01$stage=="seedling" &
+   trinity01$species=="ABC0", ]
> trinAdults <- trinity01[trinity01$stage=="tree" &

```

```

+                               trinity01$species=="ABCO", ]
> seedlingPlots <- findSeedPlots(trinSeeds, 1)
> parentTrees <- findAdultTrees(seedlingPlots, trinAdults, 20)
> ## check that there are multiple seedling densities,
> ## and parentTrees looks right.
> str(parentTrees)

'data.frame':      3352 obs. of  4 variables:
 $ treeid: chr  "1532" "1534" "1535" "1536" ...
 $ ri     : num  1 1 1 1 1 1 1 1 1 1 ...
 $ m      : num  15 15.8 14.5 15.9 18 ...
 $ dbh    : num  15.3 13.9 7.5 15.5 2 18.3 32.5 10.6 22.5 3 ...

> unique(parentTrees$ri)

[1] 1 2 4

```

Yay, we have a functioning parentTrees dataframe that we can use to estimate parameters! Onto the parameters. If you remember from above, we are trying to find parameters for this equation:

$$R_i = STR * \sum_{k=1}^T \left(\frac{DBH_k}{30} \right)^\beta e^{-Dm_{ik}^3} * \left(\frac{1}{n} \right) \quad (3)$$

To estimate STR, β , and D, we'll need to run a non-linear least squares model. But, to be honest, those are super-breakable and suck in R. If you notice, however, this equation only non-linear because of the second half (e^{\dots}). As all ecologists love to do, we can take the natural log of both sides of this equation and turn it into a linear model that looks something like this:

$$\ln(R_i) = \ln(STR) + \beta * \ln\left(\frac{DBH}{30}\right) + D * m^3 \quad (4)$$

This transformation means that the parameters we are estimating, β , STR, and D, are all related linearly to the other parameters. In layman's terms, that means that we can now predict the parameters using the much more robust and unbreakable function `glm()` in R. The formula we'll be using looks like this:

```
> formula <- "log(ri)~log(dbh/30) + m^3"
```

As a reminder, the `log()` function in R gives the natural log (\ln , or \log_e), not the \log_{10} value.

We will do the normalizer afterwards, because it should not affect the outcome of the model. Now that we have the dataframe and the model, it's a simple matter of running it, then taking a look at the transformed parameters and converting them back to the form we need them in.

```

> myModel <- glm(formula, data=parentTrees)
> summary(myModel)

```



```

Call:
glm(formula = formula, data = parentTrees)

Deviance Residuals:
    Min       1Q   Median       3Q      Max
-0.2281  -0.1608  -0.1381  -0.1120   1.2928

Coefficients:
              Estimate Std. Error t value Pr(>|t|)
(Intercept)  0.175139   0.018673   9.379  < 2e-16 ***
log(dbh/30) -0.013730   0.005548  -2.475 0.013385 *
m            -0.003941   0.001120  -3.519 0.000438 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for gaussian family taken to be 0.1053276)

Null deviance: 353.31  on 3333  degrees of freedom
Residual deviance: 350.85  on 3331  degrees of freedom
(18 observations deleted due to missingness)
AIC: 1962.7

```

Number of Fisher Scoring iterations: 2

As you can see, that's about as simple as you can get! But what do these parameters mean? The intercept in this is actually the $\ln(\text{STR})$, so we need to do e^i to find the value of STR. The parameter generated for dbh is representative of β without transformation. And the parameter generated for m is equivalent to -D, so we need to get the negative value for it to be equal to D.

```

> STR <- exp(myModel$coefficients[1])
> beta <- myModel$coefficients[2]
> D <- -myModel$coefficients[3]
> STR
(Intercept)
  1.191412
> beta
log(dbh/30)
-0.01372956
> D
m
0.003941383
>

```

And that is the simple way to get parameters for these equations!

References

- Ribbens, E., J. A. Silander, and S. W. Pacala, 1994. Seedling recruitment in forests : Calibrating models to predict patterns of tree seedling dispersion. *Ecology* **75**:1794–1806.