

CMEE MiniProject:

# **Comparison of Models used to explain the Response of Metabolic Rate to Temperature**

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**Word Count:** 2533

## **1 Abstract**

2 Metabolism is at the heart of biology; this rate at which organisms convert resources  
3 to energy dictates process across organisation levels up to ecosystem functions. Ther-  
4 mal Performance Curves (TPCs) are used to describe how these rates of metabolism in  
5 organisms responds to temperature. With rising global temperatures an understanding  
6 of these processes at a mechanistic level will enable more informed predictions about  
7 species responses to be made. This study fitted and compared 5 models to hundred's  
8 of TPCs, a phenomenological cubic model, three variations of Schoolfield model and the  
9 enzyme-assisted Arrhenius-model. The cubic model provided the best fit to the data,  
10 while out of the mechanistic models the EAAR performed poorly in comparison to the  
11 Schoolfield.

## **12 1 Introduction**

13 Metabolic rate is a fundamental measure in biology influencing levels from biochemistry  
14 through to ecosystem functioning (Brown et al. 2004, Gillooly et al. 2001). This rate is  
15 temperature dependant due to nature of chemical reactions, where increases in energy  
16 lead to a higher rate of molecule collisions of and so higher rate of reaction. However  
17 in biology higher temperatures eventually lead to a reduction in metabolic rate due to  
18 decreased protein stability (Schulte 2015). There is therefore an optimum temperature  
19  $T_{opt}$  at which organisms most efficiently convert resources to energy - these **optimums**  
20 vary among taxa (Dell et al. 2011). Changes in metabolic rate have impacts across  
21 all levels of biology leading to affects on ecological processes such species interaction,  
22 population growth and ecosystem functioning (Savage et al. 2004). Metabolism has even  
23 been linked to evolution with higher rates associated with greater genetic divergence  
24 and higher rates of speciation (Allen et al. 2006). Given the continuing rise in global  
25 temperatures it is becoming increasingly important for us to understand how changes in  
26 temperature effects metabolism allowing us to better predict species' responses (Schulte  
27 2015).

28 Thermal performance curves (TPCs) are used to describe how metabolic rate are in-  
29 fluenced by temperature (Kontopoulos et al. 2018). TPCs have a general right skewed  
30 unimodal shape, rates rise to, and level off at an optimum temperature for the given trait  
31 ( $T_{opt}$ ) before a sharp fall with further temperature increases (Dell et al. 2011). There  
32 has been much debate recently around the mechanisms which underlie the shape of this  
33 curve with a variety of models proposed (DeLong et al. 2017, Kontopoulos et al. 2018,  
34 Schoolfield et al. 1981). Models can be phenomenological, describing the data but not  
35 the underlying mechanisms, or mechanistic and attempt to describe the data by predict-  
36 ing the mechanisms which underpin it. This enable us to understand what influences  
37 observations by describing in a general way the biological processes which govern them.  
38 This study will use model fitting and selection (Johnson & Omland 2004) to identify which  
39 of five models (one phenomenological and four mechanistic), best fit and describe data  
40 of hundreds of TPCs from a wide range of taxa. This will increase our understanding of  
41 which mechanisms are involved in the response of metabolism to temperature.

## 42 1.1 The Models

43 In this analysis five models were compared:  
44 A general cubic polynomial model (eq. 1) which is able to describe unimodal data such  
45 as TPCs but has parameters with no biological meaning. This equation gives the trait  
46 value  $B$  at the temperature  $T$  measured in °C.

$$B = B_0 + B_1T + B_2T^2 + B_3T^3 \quad (1)$$

47 The Full Schoolfield model (eq. 2) (Schoolfield et al. 1981), which attempts to explain the  
48 rise and fall of TPCs using understanding of how protein stability is affected by changes  
49 in temperature, impacting the efficiency of a rate-limiting enzyme. The equation gives the  
50 trait value  $B$  at the given temperature  $T$  measured in Kelvin.

$$B = \frac{B_0 e^{\frac{-E}{k}(\frac{1}{T} - \frac{1}{283.15})}}{1 + e^{\frac{E_l}{k}(\frac{1}{T_l} - \frac{1}{T})} + e^{\frac{E_h}{k}(\frac{1}{T_h} - \frac{1}{T})}} \quad (2)$$

51 In this equation  $B_0$  represents the trait value at  $T_{ref}$  in this study a reference temperature  
 52 of 283.15 K was used.  $E$  is the activation energy of the reaction. While  $E_l$  and  $E_h$  are  
 53 the low and high deactivation energies which dictate the change in rate of reaction at  
 54 low and high temperatures.  $T_l$  is the temperature at which enzyme activity is reduced to  
 55 50% due to low temperatures and  $T_h$  where enzyme activity drops to 50% due to high  
 56 temperatures.  $k$  is Boltzmann's constant ( $8.61 \times 10^{-5} \text{eV K}^{-1}$ ). In many cases high or low  
 57 temperature deactivation is not detectable within the data (with too few measurements  
 58 taken at low and high temperatures being a common problem) and so two simplified  
 59 version of the model were used which exclude these parameters respectively (eq. 3 and  
 60 4) (Kontopoulos et al. 2018).

$$61 B = \frac{B_0 e^{\frac{-E}{k}(\frac{1}{T} - \frac{1}{283.15})}}{1 + e^{\frac{E_h}{k}(\frac{1}{T_h} - \frac{1}{T})}} \quad (3) \qquad B = \frac{B_0 e^{\frac{-E}{k}(\frac{1}{T} - \frac{1}{283.15})}}{1 + e^{\frac{E_l}{k}(\frac{1}{T_l} - \frac{1}{T})}} \quad (4)$$

62 The enzyme-assisted Arrhenius Model (EAAR) proposed by DeLong et al. (2017) at-  
 63 tempts to explain the rise, peak and fall of TPCs in a slightly different way. Unlike the  
 64 Schoolfield model which assumes a maximum metabolic rate which is reduced at low  
 65 and high temperatures due to lowered enzyme performance, the EAAR assumes a base-  
 66 line reaction rate with a high activation energy. This is lowered with the assistance of  
 67 an enzyme and so the metabolic rate changes with temperature in line with changes in  
 68 enzyme activity. The equation gives the trait value  $V$  at a given temperature  $T$  again  
 69 measured in Kelvin.

$$V = A_0 e^{\frac{-(E_b - (E_{\Delta H}(1 - \frac{T}{T_m}) + E_{\Delta Cp}(T - T_m - T \ln \frac{T}{T_m})))}{kT}} \quad (5)$$

70 In this equation  $A_0$  is a constant unique to each reaction.  $E_b$  is the baseline activation  
 71 energy of the reaction.  $E_{\Delta Cp}$  and  $E_{\Delta H}$  are both changes in the activation energy of the re-  
 72 action associated with enthalpy change and heat capacity of the enzyme respectively.  $T_m$   
 73 is the melting point, equivalent to  $T_h$  in the Schoolfield model and  $k$  is again Boltzmann's  
 74 constant.

75 **2 Methods**

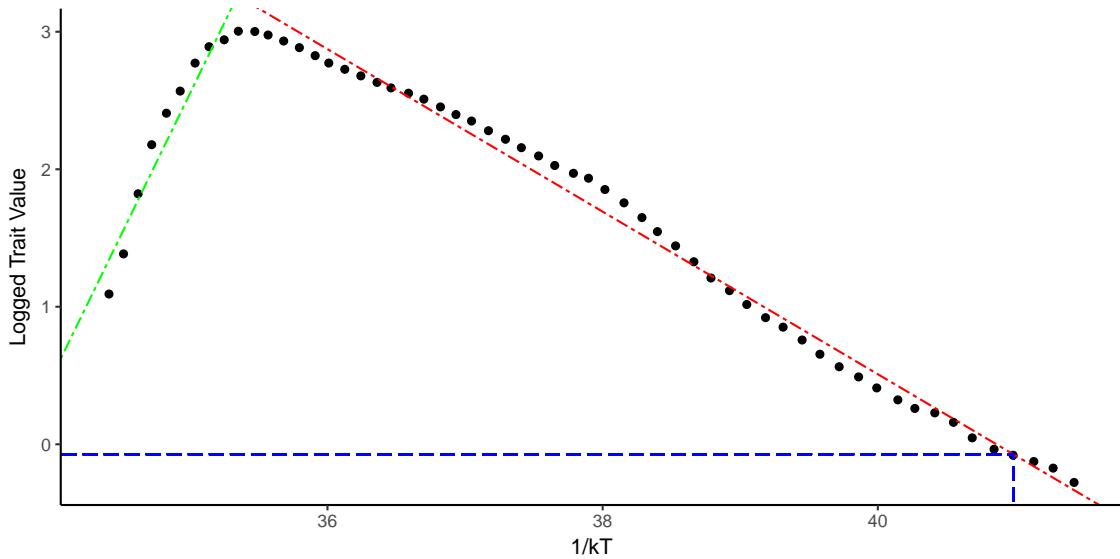
76 **2.1 Data**

77 Data for the analysis was taken from the Biotraits database (Dell et al. 2013) a large  
78 resource containing measures of ecological rates and traits across a variety of tempera-  
79 ture points, for species ranging from bacteria to terrestrial plants attempting to cover the  
80 extensive variation in metabolic responses to temperature found in life on Earth. This  
81 database contained over 25826 rows of data from 2165 experiments compiled from hun-  
82 dreds of published sources.

83 In order to ensure that all models could be fitted (or an attempt at fitting made), data was  
84 filtered to only include groups which contained a minimum of six points with trait values  
85 greater than zero. This is because the full Schoolfield model has six parameters and so a  
86 minimum of six points are needed to perform the non-linear least-squared fitting. Groups  
87 where all temperature values or all trait values were identical were also removed as these  
88 would be impossible to fit.

89 **2.2 Calculating Starting Values and Fitting Models**

90 To increase the likelihood of the Schoolfield models converging, appropriate starting val-  
91 ues were calculated from the data for the remaining groups.  $E$  was taken as the slope of  
92 the linear regression of points to the right hand side of the peak of  $\log(\text{traitValue})$  by  $\frac{1}{kT}$   
93 curve and  $E_h$  the left hand side see figure 1. In cases where the peak of the curve was  
94 at the left or right  $E$  and  $E_h$  starting values were set as the same; the slope through all  
95 points.  $E_l$  was taken as  $0.5 \times E$ .  $B_0$  which represents the trait value at the reference tem-  
96 perature was calculated from the line used for  $E$  where the the temperature is 283.15 K  
97 ( $\frac{1}{kT} = 40.98$ ).  $T_h$  was estimated as the temperature with the highest trait value while  $T_l$   
98 was the lowest temperature for which a trait value existed.



**Figure 1:** log transformed trait values against  $\frac{1}{kT}$ . Starting values for  $E$  were taken as the slope of the right hand side (red line), and  $E_h$  the left side (green line).  $B_0$  was predicted where the line used to calculate  $E$  was at  $\frac{1}{kT_{\text{ref}}}$  (Blue line). The data for this example is taken from O'Sullivan et al. (2013) showing the Leaf Respiration Rate of *Eucalyptus pauciflora*.

99 Starting values for the EAAR could not be predicted from the data and so random val-  
 100 ues were chosen: between 0:10 for  $A_0$  and  $E_b$ , between -10:10 for  $E_{\Delta C_p}$  and  $E_{\Delta H}$  and  
 101 between 280 K and 350 K for  $T_m$ .

102 All models were fitted for each group with the Non-Linear Least-Squares method using  
 103 the python package LMFIT (Newville et al. 2014), attempting to minimize the residuals  
 104 of the given model with the Levenberg-Marquardt algorithm. The cubic model was run  
 105 through the fitting using the original (not log transformed data) trait values and temper-  
 106 ature in °C. As the model is phenomenological and has no biological meaning starting  
 107 values could not be predicted from the data and as such were set at 0.

108 The three versions of the Schoolfield model were fitted on the log transformed trait data  
 109 and temperature in Kelvin. Initially **staring** values as describe above were used, further  
 110 fits were then attempted on each group a minimum of two more times with starting values  
 111 randomised between zero and twice the calculated starting values, the output with lowest  
 112 AIC was then chosen. For groups which had not converged after these 3 attempts up to  
 113 a further 22 (total of 25) attempts were made with these randomised starting values in an  
 114 attempt to get as many curves as possible to converge. Fitting of the EAAR followed the

115 same method, taking the best fit of the first three tries and continuing up to 25 attempts if  
116 fitting still failed for particular groups.

### 117 2.3 Comparison of Models

118 As different models were fitted on the logged and non-logged data, Rsquared and AIC  
119 values needed to be converted to be comparable. Models which had been fitted on the log  
120 transformed data had their residuals un-transformed fitted parameters were put through  
121 the model and residuals calculated from the un-logged data. Residual Sum of Squares  
122 (*RSS*) and Total Sum of Squares (*TSS*) were recalculated using these un-logged residu-  
123 als and from these an Rsquared value and AIC were calculated using the below equation  
124 where *N* is the number of observations and *P* is the number of parameters in the model.  
125 AIC provides a measure of fit which penalises for over-fitting and so allows for comparison  
126 of models with varying numbers of parameters.

$$AIC = N \ln \frac{RSS}{N} + 2P \quad (6)$$

127 Models were compared within each group; a  $\Delta AIC$  was calculated as the difference be-  
128 tween the lowest AIC and the AIC for that model. Where  $\Delta AIC$  was less than or equal to  
129 two the models were considered to be the best or comparable the best while when  $\Delta AIC$   
130 rose above ten then no support for the model could be inferred (Burnham & Anderson  
131 2004). An Akaike weight  $W_i(AIC)$  was also calculated for each model using the equation  
132 below, this gives a probability that the model best describes the data (Johnson & Omland  
133 2004).

$$W_i(AIC) = \frac{\exp\{-\frac{1}{2}\Delta_i(AIC)\}}{\sum_{k=1}^K \exp\{-\frac{1}{2}\Delta_k(AIC)\}} \quad (7)$$

134 Comparison of the model (calculation of  $\Delta AIC$  and  $W_i(AIC)$ ) was performed using all  
135 models as well as excluding the cubic model. This enable the best mechanistic model  
136 to be identified and so the most likely mechanisms explaining the relationship between  
137 temperature and metabolism.

138 **2.4 Computing Languages Used**

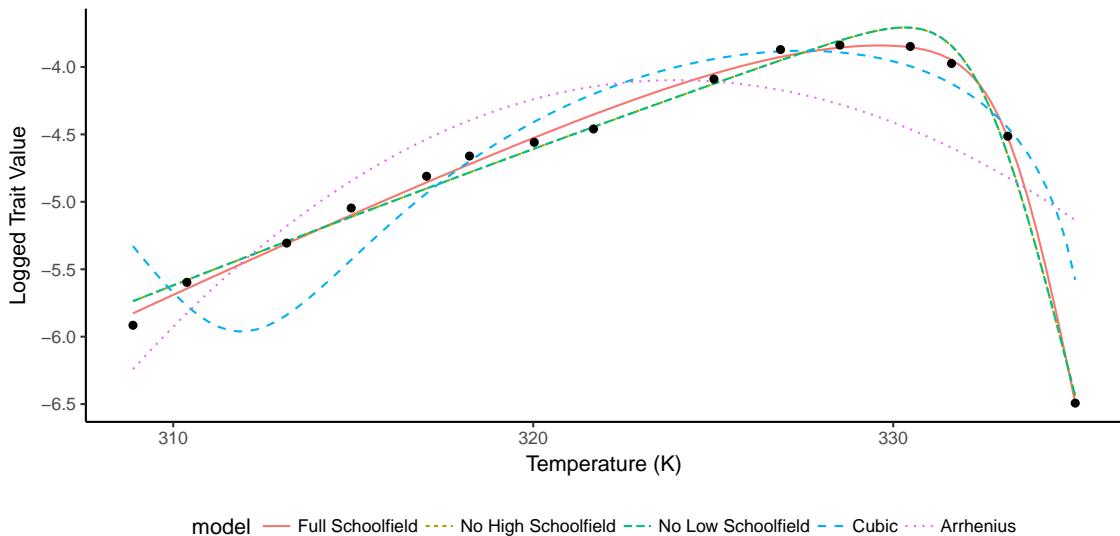
139 Three computing languages were used throughout the analysis:

- 140 • Python 3.6 was used in the data wrangling stage using the library pandas for efficient data manipulation. Estimating starting values used the libraries SciPy and NumPy for scientific and numerical functions and model fitting with NLLS was performed with the library lmfit.
- 144 • R 3.4.2 (R Core Team 2017) was used for general analysis of the results with the wide variety of inbuilt functions, and to produce easily readable plots with the package ggplot2.
- 147 • The command line language Bash (4.4.12) was used to tie the project together into one repeatable workflow and compile this writeup from L<sup>A</sup>T<sub>E</sub>X into pdf.

149 **3 Results**

150 Once the data had been sorted to remove curves with too few points 1582 groups remained with an average of 13.83 observations each. During the fitting process curves 151 converged on all groups for both the Cubic and EAAR models, while all but 11 of the Full 152 Schoolfield model converged, and all but 8 of the No High and 3 of the No Low Schoolfield 153 models converged within the 25 attempts permitted.

155 Figure 2 shows an example curve where model fitting was successful for all models, with 156 the Full Schoolfield model having the lowest AIC and highest RSquared values and so 157 the best fit. The mean Rsquared values 1 show that the cubic model tended to provide 158 the better fit. The incredibly low mean Rsquared value for the EAAR model is explained 159 by a few curves where the Rsquared values are anomalously low. Not including these 160 values the mean Rsquare of the EAAR is 0.707.



**Figure 2:** The five models fitted on a TPC. The data from this example shows the doubling rate of *Peptoclostridium paradoxum* taken from Li et al. (1993). RSquared and AIC values are shown in table.1.

**Table 1:** Rsquared and AIC for each fitted model shown in figure 2 and mean values across all groups.

Model	RSquared (fig.2)	AIC (fig.2)	Mean Rsquared	Mean AIC
Cubic	0.915	-178.78	0.887	-57.17
Full Schoolfield	0.993	-212.96	0.845	-62.17
No High Schoolfield	0.964	-191.57	0.777	-51.67
No Low Schoolfield	0.964	-191.57	0.764	-49.37
EAAR	0.507	-150.43	-28.76	-44.01

161 AIC is a measure of fit which penalises for over-fitting, allowing models with different  
 162 numbers of parameters to be compared (Johnson & Omland 2004). Where the  $\Delta\text{AIC} \leq 2$   
 163 there is significant support for the model, with less at support at  $4 < \Delta\text{AIC} \leq 7$  and no  
 164 support at  $\Delta\text{AIC} > 10$  (Burnham & Anderson 2004). It is clear from table 2 that the Cubic  
 165 model was consistently able to better fit the data in comparison to the other models, with  
 166 more than twice as many groups having the cubic model as the best or comparable to  
 167 best model.

**Table 2:** The number of  $\Delta$ AIC's for each model which fall into the respective categories when all models are compared.

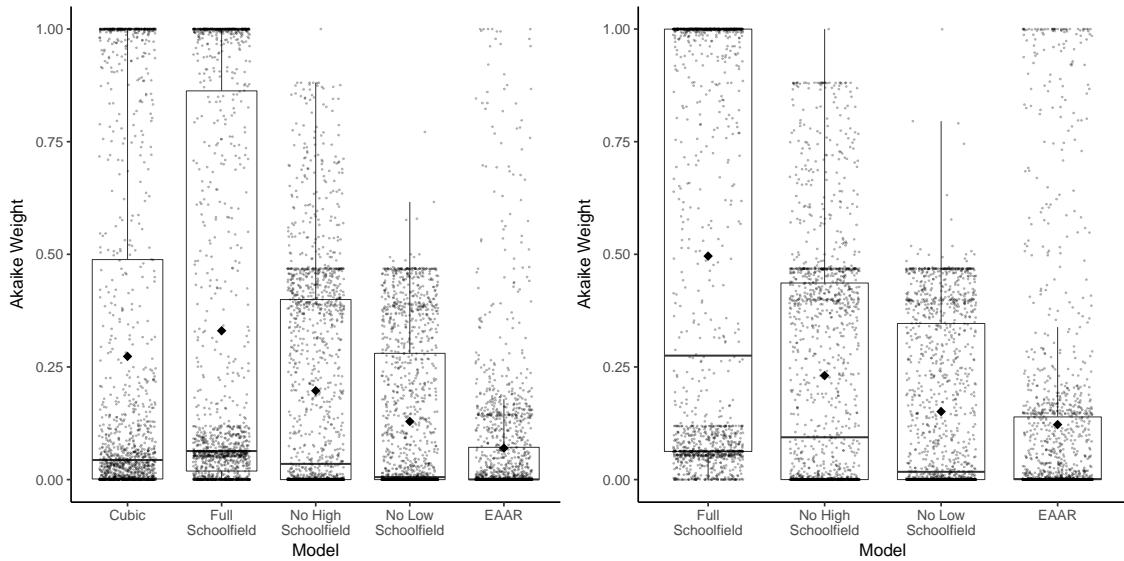
	$\Delta \leq 2$	$2 < \Delta \leq 4$	$4 < \Delta \leq 7$	$7 < \Delta \leq 10$	$\Delta > 10$
Cubic	990	90	105	73	324
Full Schoolfield	454	134	418	87	478
No High Schoolfield	471	119	106	91	787
No Low Schoolfield	370	125	115	93	876
EAAR	19	281	138	119	1025

<sup>168</sup> The cubic model is phenomenological and so does not provide any insight into the mech-  
<sup>169</sup> anisms involved in TPCs and so the same comparisons were made between the four  
<sup>170</sup> mechanistic models (table 3). Here the Schoolfield model without the high temperature  
<sup>171</sup> deactivation component performed better, being the best or comparable to best model  
<sup>172</sup> most frequently.

**Table 3:** The number of  $\Delta$ AIC's for each model which fall into the respective categories when only mechanistic models are compared.

	$\Delta \leq 2$	$2 < \Delta \leq 4$	$4 < \Delta \leq 7$	$7 < \Delta \leq 10$	$\Delta > 10$
Full Schoolfield	597	95	253	83	115
No High Schoolfield	701	27	42	27	652
No Low Schoolfield	542	56	59	40	743
EAAR	301	111	71	58	883

<sup>173</sup> The Akaike weights  $W_i(\text{AIC})$  tell a similar story (figure 3) with the cubic model having  
<sup>174</sup> a greater proportion with higher weights and so higher probabilities of being the model  
<sup>175</sup> which best describes the data. For a significant proportion of groups the Akaike weights  
<sup>176</sup> were bimodal with the majority having one model with a weight approaching 1 with the  
<sup>177</sup> other models almost at zero. When only the mechanistic models are compared the Full  
<sup>178</sup> Schoolfield has the higher average  $W_i(\text{AIC})$ , given that this was not the case when look-  
<sup>179</sup> ing at  $\Delta$ AIC's it suggests that though the Full Schoolfield is not the best fit as often, when  
<sup>180</sup> it is it provides a significantly better fit.



**Figure 3:** Distribution of Akaike weights for each model. Left: when all models are compared. Right: When only mechanistic models are compared.

181 Banding can be seen in the distributions of  $W_i(\text{AIC})$  in the various models (figure 3). This  
 182 occurs where models have essentially identical fits but differing numbers of parameters,  
 183  $\Delta\text{AIC}$  is identical for each group where this occurs and as it is dictated only by the param-  
 184 eter penalisation. This translates directly to  $W_i(\text{AIC})$  and is the cause of the observed  
 185 bands. In these circumstances it is the model with fewer parameters which is preferable.

## 186 4 Discussion

187 When looking at the results from all the models it is clear that the Cubic model was better  
 188 able to fit the data, with a far greater number of groups where the  $\Delta\text{AIC}$ 's suggested  
 189 it to be the best or comparable to best model, and a larger mean  $W_i(\text{AIC})$ . This may  
 190 seem unexpected as the mechanistic models it was compared against were designed to  
 191 explain the thermal performance data the models were fitted to (Schoolfield et al. 1981,  
 192 DeLong et al. 2017). **However as the parameters of the cubic model are unbounded they**  
 193 **are better able to adjust and fit the data.** To understand the mechanisms which underlie  
 194 TPCs it was therefore necessary to recalculate the  $\Delta\text{AIC}$ 's and  $W_i(\text{AIC})$ 's with the cubic  
 195 model excluded from the analysis.

196 When this was done the Schoolfield model without high temperature deactivation was  
197 able to fit the data best with 701 of the 1582 TPCs having it as the best or equivalent  
198 to best fit according to  $\Delta$ AIC. However looking at the mean Akaike weights implies that  
199 the full Schoolfield model provided the best fit. This suggests that on many occasions  
200 the full schoolfield provided a identical or near identical fit to the simplified version  
201 however when  $\Delta$ AIC was calculated it was discounted as with two parameters more it  
202 would have a  $\Delta$ AIC of at least four. The higher average  $W_i(\text{AIC})$  for the full Schoolfield  
203 may indicate that when it did provide a better fit than the simplified versions it was a  
204 substantially better fit. This makes it difficult to categorically state which model is best,  
205 with the quality of available data and nature of the thermal response having a significant  
206 impact.

207 It is interesting that the enzyme-assisted Arrhenius model (DeLong et al. 2017) performed  
208 so poorly with its  $\Delta$ AIC being too high to infer any support for the model in the vast  
209 majority of groups. This is likely confounded by the fact that the starting values for the non-  
210 linear least-squared model fitting for the EAAR were randomly chosen and not calculated  
211 from the data - this initially put the model at a disadvantage. Only the best of three  
212 fitting attempts were taken to save on computer time, it may be the case that with greater  
213 resources better fits may have been found.

214 This study looked at TPCs across a very wide range of taxa adapted for various climates  
215 and a large number of different trait measurements as proxies for metabolism. Further  
216 analysis sub-setting by these various categories may provide better insight into how the  
217 mechanisms of metabolism work across biology.



## 218 References

- 219 Allen, A. P., Gillooly, J. F., Savage, V. M. & Brown, J. H. (2006), 'Kinetic effects of tem-  
220 perature on rates of genetic divergence and speciation', *Proceedings of the National  
221 Academy of Sciences* **103**(24), 9130–9135.  
222 URL: <http://www.pnas.org/cgi/doi/10.1073/pnas.0603587103>

- 223 Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M. & West, G. B. (2004), Toward a  
224 metabolic theory of ecology, *in* ‘Ecology’, Vol. 85, pp. 1771–1789.
- 225 Burnham, K. P. & Anderson, D. R. (2004), ‘Multimodel inference: Understanding AIC and  
226 BIC in model selection’, *Sociological Methods and Research* **33**(2), 261–304.
- 227 Dell, A. I., Pawar, S. & Savage, V. M. (2011), ‘Systematic variation in the temperature de-  
228 pendence of physiological and ecological traits’, *Proceedings of the National Academy  
229 of Sciences* **108**(26), 10591–10596.
- 230 **URL:** <http://www.pnas.org/cgi/doi/10.1073/pnas.1015178108>
- 231 Dell, A., Pawar, S. & Savage, V. M. (2013), ‘The thermal dependence of biological traits’,  
232 *Ecology* **94**(5), 1205.
- 233 DeLong, J. P., Gilbert, J. P., Luhring, T. M., Bachman, G., Reed, B., Neyer, A. & Montooth,  
234 K. L. (2017), ‘The combined effects of reactant kinetics and enzyme stability explain the  
235 temperature dependence of metabolic rates’, *Ecology and Evolution* **7**(11), 3940–3950.
- 236 Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M. & Charnov, E. L. (2001), ‘Effects  
237 of size and temperature on metabolic rate’, *Science* **293**(September), 2248–2251.  
238 **URL:** [Gillooly\\_etal\\_science\\_01.pdf%5Cn10.1126/science.1061967%5Cn11567137](https://science.org/doi/10.1126/science.1061967)
- 239 Johnson, J. B. & Omland, K. S. (2004), ‘Model selection in ecology and evolution’, *Trends  
240 in Ecology & Evolution* **19**(2), 101–108.
- 241 Kontopoulos, D. G., García-Carreras, B., Sal, S., Smith, T. P. & Pawar, S. (2018), ‘Use  
242 and misuse of temperature normalisation in meta-analyses of thermal responses of  
243 biological traits’, *PeerJ* **6**, e4363.  
244 **URL:** <https://peerj.com/articles/4363>
- 245 Li, Y., Mandelco, L. & Wiegel, J. (1993), ‘Isolation and Characterization of a Moderately  
246 Thermophilic Anaerobic Alkaliphile,’, *Cultures* **3**(July), 450–460.
- 247 Newville, M., Stensitzki, T., Allen, D. B. & Ingargiola, A. (2014), ‘LMFIT: Non-Linear Least-  
248 Square Minimization and Curve-Fitting for Python’.  
249 **URL:** <https://doi.org/10.5281/zenodo.11813#.WqES7r0w8iQ.mendeley>
- 250 O’Sullivan, O. S., Weerasinghe, K. W. K., Evans, J. R., Egerton, J. J., Tjoelker, M. G.

- 251 & Atkin, O. K. (2013), 'High-resolution temperature responses of leaf respiration in  
252 snow gum (*Eucalyptus pauciflora*) reveal high-temperature limits to respiratory func-  
253 tion', *Plant, Cell and Environment* **36**(7), 1268–1284.
- 254 R Core Team (2017), *R: A Language and Environment for Statistical Computing*, R Foun-  
255 dation for Statistical Computing, Vienna, Austria.  
256 **URL:** <https://www.r-project.org/>
- 257 Savage, V. M., Gilloly, J. F., Brown, J. H. & Charnov, E. L. (2004), 'Effects of body size  
258 and temperature on population growth.', *The American Naturalist* **163**(3), 429–441.
- 259 Schoolfield, R. M., Sharpe, P. J. H. & Magnuson, C. E. (1981), 'Non-linear Regression  
260 of Biological Temperature-dependent Rate Models Based on Absolute Reaction-rate  
261 Theory', *Journal of Theoretical Biology* **88**, 719–731.
- 262 Schulte, P. M. (2015), 'The effects of temperature on aerobic metabolism: towards a  
263 mechanistic understanding of the responses of ectotherms to a changing environment',  
264 *Journal of Experimental Biology* **218**(12), 1856–1866.  
265 **URL:** <http://jeb.biologists.org/cgi/doi/10.1242/jeb.118851>