Differences between flower and leaf phenological responses to environmental variation drive shifts in spring phenological sequences of temperate woody plants

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# Abstract

The relative timing of growth and reproduction is an important driver of plant fitness. For deciduous woody species in temperate regions leaves and flowers both appear in the early spring, but the order and duration of these phenological events vary among species, populations, and individuals. Researchers have long hypothesized that this variation in flower-leaf sequences (FLSs) may be important—affecting the reproduction, recruitment and survival of individuals. Further, FLSs appear to be shifting with climate change; thus anticipating the extent of these shifts may influence projections of how climate change may impact species' performance and reshape forest communities. Predicting FLS shifts requires an improved understanding of how environmental variation dictates FLS patterns. To address this, we compared the phenological responses of flowers and developing leaves for 10 temperate woody 11 species to varying levels of temperature and photoperiod in a lab experiment. Our experi-12 mental design allowed us to test competing hypotheses for how environmental cues determine 13 FLS variation—specifically whether forcing (warm temperatures) alone drives variation or differential sensitivity to chilling (cool temperatures generally in the fall and winter) and/or photoperiod matter. Within species, we found that flower and leaf phenology responded 16 with differential sensitivity to environmental cues, with differences in their response to chilling being the dominant driver of FLS variation. These differences between flowering and leaf responses were consistent across species, but because species differ the order of phenologi-19 cal events in their FLSs (flowering-first vs. leafing-first), differences between flower and leaf 20 phenology will have contrasting impacts on FLS variation across species. Because climate 21 change will amplify variability in chilling across time and space, our findings suggest that FLS shifts may be large, but are likely to vary substantially among populations and species. Simple projections of FLS shifts with climate change, based on our results, showed large shifts in wind-pollinated species that flower before leafing, with flower-leaf interphases substantially shortened. This shorter interphase would reduce the time period for efficient pollen

- transfer, and thus raises the possibility that wind-pollinated taxa especially may experience reproductive declines due to FLS shifts in the decades to come.
- Keywords: chilling, climate change, deciduous forests, flower-leaf sequences, forcing, hyster anthy, phenology, wind-pollination

# 2 Introduction

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Among the most widely documented biological effects of anthropogenic climate change are shifts in plant phenology, the timing of life cycle events (Parmesan & Yohe, 2003; Menzel et al., 2006; Cleland et al., 2007). While phenology is generally advancing with climate change, the strength of these phenological shifts can vary substantially among specific phenological phases (Augspurger & Zaya, 2020). These differences alter the timing of phases relative to each other, changing the duration between events that make up phenological sequences (Ettinger et al., 2018). Phenological sequences are a major driver of plant fitness that impact plant life history, resource allocation, demography and ecosystem processes (Post et al., 2008). Thus, shifting sequences with climate change will likely impact many of these processes. The effects of these shifts, however, depend both on their direction—whether distinct phases are shifting closer together or farther apart—and magnitude—how much they are shifting relative to each other.

For deciduous woody plants, the relative timing of flower and leaf phenology, or flower-leaf sequences (FLSs), may be particularly consequential to fitness in temperate regions where flowering prior to leaf development is common (Rathcke & Lacey, 1985). There are several hypotheses regarding the function of FLS variation (see Gougherty & Gougherty, 2018), and it is likely that the adaptive significance of FLSs vary among species, and may co-vary with

other plant traits.

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The flowering-first FLS is strongly correlated with wind-pollination (Buonaiuto et al., 2021; Friedman & Barrett, 2009) and models of pollen movement show that for wind-pollinated species, flowering-first increases pollen dispersal distances and significantly reduces the amount of pollen intercepted by non-reproductive structures (Di-Giovanni et al., 1989; Tauber, 1967; Whitehead, 1969). Flowering-first is also prevalent in some biotically-pollinated taxa, but its function is less clear. Some authors suggest that flowering-first impacts floral visibility to pollinators (Janzen, 1967; Bukovac et al., 2017; Forrest & Thomson, 2009) or modifies hydraulic demand (Gougherty & Gougherty, 2018; Franklin, 2016), while, others suggest that in biotically-pollinated taxa there is no unique function to the sequence and flowering-first a by-product of selection for early flowering in general (Primack, 1987).

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Phenological observations over the last several decades indicate that, like other phenological sequences, FLSs are shifting with climate change (Ma et al., 2020). For several species, the time between flowering and leafing appears to be increasing, but the strength of this trend varies among species and the direction of FLS shifts are not consistent across populations (Buonaiuto et al., 2021; Ma et al., 2020). These changes could affect the important functions of FLSs, potentially putting some species at greater risk for fitness declines, while benefiting others.

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The impact on FLS shifts with climate change on the fitness of woody plants depends on 1)
the function of FLS for that species and 2) the direction and magnitude of the shift. For
example, in wind-pollinated species that rely on a substantial flower-leaf interphase for effective pollen transport, decreasing FLS interphases with climate change may drive a reduction
in pollination success as more pollen is intercepted by vegetation. Conversely, pollination

efficiency could improve for species with lengthening FLS interphases. However, a proportionate FLS shift in biotically-pollinated taxa may have different fitness implications because the contrasting function of FLS variation in these species.

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While several recent analyses have examined the function of FLS variation (e.g. Buonaiuto et al., 2021; Gougherty & Gougherty, 2018), the factors that influence the magnitude and direction of FLS shifts are less well studied (but see Ma et al., 2020). Predicting FLS shifts requires identifying the proximate mechanisms that drive and constrain FLS variation, and how these mechanisms differ among species.

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Decades of research suggest that cool winter temperatures (chilling), warm spring temperatures (forcing), and day-length (photoperiod) are the primary drivers of both reproductive and vegetative phenology for woody plants in temperate regions (Körner & Basler, 2010; Flynn & Wolkovich, 2018). However, observed FLS shifts indicate that there must be differences in how these cues influence the phenology of flowers and leaves (Buonaiuto *et al.*, 2021).

It is also likely that FLS variation is mediated by other internal mechanisms like developmental construction (Diggle, 1995), or other physical constraints like inflorescence architecture or bud type (Pope et al., 2013). For example, FLS variation in species with separate buds (buds containing either embryonic leaves or flowers) may be less constrained than species with mixed buds (buds containing both embryonic leaves and flowers together). Other factors like growth form (tree vs. shrub) or colonization-competition tradeoffs that have been show to influence the phenological sensitivity of specific phenophases (Basler & Körner, 2012; Donnelly & Yu, 2021) may also influence the sensitivity of phenological sequences to climate.

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While FLS variation in woody plants is no doubt the product of interactions between species-

specific biology and complex environmental inputs, identifying the differences in how flower and leaf phenology responds to environmental change is a necessary step for predicting the direction, magnitude and—ultimately—fitness impacts of FLS shifts with climate change. Studies that have attempted to identify the differences between reproductive and vegetative phenology in woody plants (mostly focused on crop species) have yielded two common explanations which we present below.

#### $_{09}$ Hypotheses for FLS variation

One hypothesis suggests that reproductive and vegetative buds utilize the same underlying environmental cues, but have different threshold responses to forcing, with whichever bud type bursts later—leaves or flowers—having a higher thermal requirement (that is, they need a greater sum of warm temperature to trigger the phenological event, Guo et al., 2014; Cosmulescu & Calusaru, 2020; Cosmulescu & Ionescu, 2018). Under this hypothesis, which we call the forcing hierarchy hypothesis (FHH), leaf and flower buds share the same suite of cues and develop similarly to non-forcing cues (i.e., chilling and photoperiod), but differ in the thermal requirement for budburst.

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In contrast, an alternative hypothesis suggests that flower and leaf buds differ in the strength of their phenological responses to multiple environmental cues (Citadin *et al.*, 2001; Gariglio *et al.*, 2006; Aslamarz *et al.*, 2009; Mehlenbacher, 1991). Under this hypothesis, which we call the differential sensitivity hypothesis (DSH), each bud type relies more or less on certain cues, generating FLS variability.

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While observational studies of phenology in the field are commonly used to evaluate the relationship between environmental conditions and phenology (Cleland *et al.*, 2007), under

current and recent climatic conditions, the FHH and DSH may produce similar phenological 127 patterns, making it difficult to evaluate these hypotheses with such methods. However, ex-128 periments designed to isolate all three environmental cues have the potential to disentangle 129 the two hypotheses. Studies aiming to differentiate these hypotheses can look for two dif-130 ferent signatures. The key signature of the FHH is that the sensitivity to forcing ( $\Delta$  day of 131 phenological event/ $\Delta$ °C) of the second phase in the phenological sequence is always greater 132 than that of the first phase with sensitivity differences being inversely proportional to the 133 difference in thermal requirement among bud types (2x in our simulations; Fig. 1 a.). By 134 contrast, the signature of the DSH is that sensitivity estimates of flower and leaf buds to chilling and/or photoperiod will diverge (Fig. 1 b.), but there is no characteristic pattern 136 as to the strength or direction of this divergence (e.g., leaves may require longer or shorter 137 photoperiods than flowers, or more or less chilling). 138

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The clarity of these signatures from each hypothesis, however, may be obscured in certain 140 situations. First, a species' buds could be both differentially sensitive to photoperiod or chill-141 ing (supporting the DSH) and also respond to forcing in a forcing hierarchy (that is, both 142 hypotheses can operate at once, see Fig. 1 c.). Second, the differential sensitivity framework 143 can generate the signature of the FHH when other cues are at high levels due to interactions 144 between the chilling or photoperiod response and the thermal requirement, which make forc-145 ing the dominant phenological cue once other cue requirements have been met (see Fig. 1 b., chill x force interaction). When this happens the FHH would effectively be a special case 147 of the DSH. 148

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While these two hypothesized mechanisms may produce similar phenology patterns under current conditions, differentiating them is important, as they have substantially different implications regarding the potential for FLS shifts with climate change. The FHH suggests that FLS variation is largely a product of climate variation during the interphase. If spring temperatures increase with climate change, the second phenophase of the FLS will be accelerated
relative to the first and FLS interphases will decrease, but given the relative auto-correlation
of spring temperatures (Di Cecco & Gouhier, 2018), these shifts should be muted. If FLSs are
structured by a forcing hierarchy, the direction of FLS shifts are relatively straight forward
to predict, and it is less likely that shifts will be large enough in magnitude to significantly
affect woody plant fitness.

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Climate change would also drive FLS shifts under the DSH, but the trajectory of these shifts 161 are more complicated to predict. The DSH suggests that there could be strongly localized 162 or regional effects of climate change on FLSs. Shifts in FLS variation will depend on the 163 direction and rate of change in cues at given locations and the species-specific differential sensitivity of reproductive and vegetative phenology to cue combinations. This hypothesis 165 allows for larger magnitude shift in FLSs, on a scale that could impact pollen transport or the 166 physiological functioning of woody plants. Further, the DSH also suggests that the magnitude 167 of shifts may be highly divergent both among species in a community, and among popula-168 tions of the same species which could impact gene flow, population structure and demography. 169

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In this study we tested the FHH and DSH hypotheses in 10 temperate shrub and tree species.

Using a full-factorial growth-chamber experiment we manipulated chilling, forcing and photoperiod cues for flower and leaf buds. We then leveraged these data to examine how FLSs
may shift with climate change.

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## <sub>76</sub> Materials and Methods

#### $_{\scriptscriptstyle 177}$ Growth chamber study

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We sampled plant material from Harvard Forest in Petersham, MA, USA (42.5314°N, 72.1900°W) 178 on 25 October 2017, just as most individuals dropped their leaves, to capture the period 179 immediately after most plants in the area entered endo-dormancy but before they could ac-180 cumulate significant chilling in the field. We collected branch cuttings from 7-13 individuals 181 of 12 woody plant species (4-12 cuttings per individual for a total of 48-56 per species), con-182 sisting of a mix of deciduous shrubs, understory and canopy trees commonly found in mesic 183 hardwood forests of the eastern United States. The use of branch cuttings to study woody 184 plant phenology in artificial environments is common (Ettinger et al., 2020), and these meth-185 ods have been shown to match whole-plant phenological patterns (Vitasse & Basler, 2014; 186 Primack et al., 2015). We selected species displaying variation of the three major FLS pat-187 terns; species for which flowers appear before leaves (flowering-first; Acer rubrum, Betula alleghaniensis, Comptonia peregrina, and Corylus cornuta), species for which flowers appear 189 with their leaves (concurrent; Acer pensylvanicum, Acer saccharum, Ilex mucronata, Prunus 190 pensylvanica, Vaccinium corymbosum), and species for which flowers appear after leaf devel-191 opment (leafing-first; *Ilex verticillata*, *Prunus virginiana*, *Viburnum acerifolium*). Because 192 we expected that other characteristics such as pollination syndrome, bud type, and growth 193 habit were likely to impact FLS sensitivity, we made sure that the species we chose exhibited 194 variation in these traits as well (see Tab. S1 for details and full species list). We included sev-195 eral congenerics in our study, explicitly choosing species with different FLS patterns (which therefore could not have been inherited from a common ancestor) to mitigate phylogenetic 197 non-independence in our trait of interest (FLS) (Revell, 2010). 198

• We transported all cuttings to the Arnold Arboretum in Boston, MA (USA) where they were

re-cut in water to prevent callousing and minimize embolism and placed in 500 ml Erlenmeyer flasks with distilled water.

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We randomly assigned cuttings to a fully crossed set of eight experimental treatments; two 204 levels of chilling (4 vs 8 weeks at 4°C), two levels of temperature (24°C:18°C (day/night) 205 warm vs 18°C:12°C (day/night) cool) and two levels of photoperiod (12 vs 8 hours). We 206 alternated day/night temperature periodicity on a 12 hour schedule to reduce co-variation 207 with photo-periodicity. We re-cut all twigs and changed the water every 7-10 days and ro-208 tated all treatments between growth chambers every two weeks to minimize chamber effects. 209 We made phenological observations every 2-3 days using a modified BBCH scale for woody 210 plants (Finn et al., 2007) for three months following release from chilling conditions. In 211 this period we assessed three phenological phases: leaf budburst (BBCH phase 07), leafout (BBCH phase 15) and first flower open (BBCH 60). While leaf budburst is the first stage 213 of vegetative expansion and therefore most directly comparable to first flowering, we also 214 included leafout in our observations because several of the functional hypotheses for FLS 215 variation are predicated on developing leaves be sufficiently large enough to impact canopy 216 structure, which would only happen at later stages of leaf development. 217

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At the conclusion of this period we assessed all individuals that did not undergo budburst and excluded 56 dead twigs from our analyses.

# Data analysis

To assess the phenological sensitivity of each phase, we fit mixed-effects hierarchical models with chilling, forcing, photoperiod and all two-way interactions as the fixed effects and species as a grouping factor on both the slopes and the intercepts. We chose a Bayesian hierarchical approach in order to identify systematic trends across species' responses while accounting
for sample size, variance, and the unique effect of each species. We modeled the effects of
environmental parameters on flower opening, leaf budburst and leafout separately. We also
fit a model with FLS interphase (day of budburst - day of flowering) as a response variable
to compare these estimates with field observations. Two species, *Betula allegheniensis* and *Acer saccharum*, produced no flowers in our trial, likely because flowering branches were out
of reach during our field sampling, so we excluded them from our analysis. In total, our
analyses included 464 twigs from 10 species.

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The models we fit appear below:

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$$y_{[i]} \sim N(\alpha_{sp_{[i]}} + \beta_{forcing_{sp_{[i]}}} + \beta_{chilling_{sp_{[i]}}} + \beta_{photoperiod_{sp_{[i]}}} + \beta_{forcingxchilling_{sp_{[i]}}} + \beta_{forcingxphotoperiod_{sp_{[i]}}} + \beta_{sp_{ij}} +$$

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Where  $y_{[i]}$  is either the day of the experiment of leaf budburst, day of first flower opening, day of leafout or FLS interphase length, and  $\sigma_y^2$  the error. We modeled the intercept  $(\alpha)$  and each slope  $(\beta)$  parameter at the species level using the formula:

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$$\alpha_{x_{sp}}$$
 or  $\beta_{x_{sp}} \sim N(\mu_x, \sigma_x^2)$ 

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To test the hypothesis that the FHH is a special case of the DSH that occurs when other cue (i.e., chilling and photoperiod) requirements are met, we re-ran our leaf budburst and flowering models on a subset of our data which included both levels of the forcing treatment but only the high photoperiod and chilling treatment levels. This model included forcing as the only main effect but, like our main models written above, included species as a grouping

factor on the slopes and intercept.

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We fit all models using the R package "brms" (Bürkner, 2018). We ran each model on four 252 chains with 4000 iterations and a 3000 iteration warm up for a total of 4000 posterior draws 253 for each parameter. In all models we used weakly informative priors and increasing the priors 254 5-fold did not affect the model results. We assessed model performance through ensuring Rs255 were between 1 and 1.01 and bulk and tail effective sample sizes were high (1800-2800 for 256 most parameters, but as low as 800-900 for some). We present 50% credible intervals in 257 figures because they are the most computationally stable (Gelman et al., 2013), but provide 258 other intervals in the Supporting Information (Tab. S3, Tab. S4, Tab. S5). All of our 259 estimates of phenological sensitivity ( $\Delta$  day of phenological event/ $\Delta$  environmental cue) are 260 scaled by treatment level (chilling:  $\Delta$  4 weeks, forcing:  $\Delta$  6°C, photoperiod:  $\Delta$  4 hours).

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#### 3 Climate change scenarios

To apply our model results to general climate change scenarios we chose environmental treatments in the experiment to broadly reflect historic and future conditions at our sampling site. 265 Our low forcing treatment approximated average spring temperature (March/April) at the 266 site while our high temperature treatment reflects a 6 °C increase. Average field chilling 267 (calculated from 15 October - 15 April, measured in Chill Hours, Weinberger, 1950) at Har-268 vard Forest is 1170.7, approximately 63% of the difference between our low and high chilling 269 treatment (Tab. S2). Thus, our low chilling treatment represents a feasible estimate for 270 a decrease in chilling with climate change and our high chilling treatment approximates a 271 reasonable increase (Luedeling, 2012). Our low photoperiod treatment (8 hours of daylight) is well below the photoperiod experienced at Harvard Forest, but given that the photoperiod 273 effects are expected to be small (Laube et al., 2014), we chose more extreme values in order 274

to robustly estimate an effect (i.e., increasing statistical power). For this reason, our climate change scenarios for FLS variation are based on our high photoperiod treatment alone.

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<sup>278</sup> We used our flower, leafout and leaf budburst models to project for each species in our study:

- 1. FLSs under average environmental conditions (low forcing, 6.5 weeks of chilling treatment)
- 281 2. FLS shifts with spring warming only (high forcing, 6.5 weeks of chilling treatment)
- 3. FLS shifts with warming and increased chilling (high forcing, 8 weeks of chilling treatment)
- 4. FLS shifts with warming and decreased chilling (high forcing, 4 weeks of chilling treatment)
- To validate our predictions, we compared our FLS interphase model estimates of "average" condition FLS interphases to long term phenological records from Harvard Forest (O'Keefe, 2015) for five species common to both datasets (Fig. S1), and found them comparable.

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Given the variable dynamics of shifts in environmental forcing and chilling with climate change over time and space (Luedeling *et al.*, 2011), these scenarios should not be treated as absolute predictions of the magnitude of FLS shifts with climate change. Instead, we provide these scenarios to identify general trends in how FLSs could shift with warming and demonstrate that the range of possibilities vary based on characteristics of plant species and the specific climate dynamics.

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Finally, we characterized FLS shifts for three broad FLS groups (flowering-first, concurrent, leafing-first) by extracting all sampling posterior estimates from our main models using the

R package tidybayes (Kay, 2020), and grouping them by FLS pattern. We also include the species-specific estimates on which these grouped estimates are based in Fig. S3.

Results

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#### Growth chamber study

Flowering, leaf budburst and leafout advanced with higher forcing and longer chilling dura-304 tion (flowering: chilling effect: -21.2 days, forcing effect: -19 days, leaf budburst: chilling 305 effect: -30.4 days, forcing effect: -17.8 days, leafout: chilling effect: -39.7 days, forcing ef-306 fect: -32.6 days; Fig. 2, Fig. S2, Tab. S3, Tab. S4—all sensitivity estimates are scaled by treatments; see "Data analysis" section above), but increases in both of these cues together 308 offset these advances (flowering: force x chill effect: 7.0 days, leaf budburst: force x chill 309 effect: 12.4 days, leafout: force x chill effect: 21.4 days; Fig. 2, Fig. S2, Tab. S3, Tab. S4). 310 Leaf and flower phenologies diverged in their responses to photoperiod, with flower phenol-311 ogy advancing at longer photoperiod and leaf phenology remaining unaffected (leafout) with 312 photoperiod changes, or delaying (budburst) at longer photoperiods when the other two cues 313 were at low levels (Fig. 2, Fig. S2, Tab. S3, Tab. S4). As seen in the interactions between 314 photoperiod and chilling and photoperiod and forcing, increasing chilling or forcing with 315 longer photoperiod advanced the phenology of all three phases. For flowering, leaf budburst 316 and leafout, chilling and forcing were the dominant cues, while photoperiod produced a more 317 muted phenological response (Fig. 2, Fig. S2, Tab. S3, Tab. S4). 318

While leaf and flower phenological responses to environmental cues were qualitatively similar, the strength of their responses to each cue differed substantially. Leaf budburst and

leafout responded more strongly to chilling than flowering (budburst: 1.4x, leafout: 1.9x),
and had stronger responses to all cue interactions (leaf budburst: forcing x chilling: 1.8x,
photoperiod x chilling: 5.8x, photoperiod x forcing: 2.2x, leafout: forcing x chilling: 3.1x,
photoperiod x chilling: 2.4x, photoperiod x forcing: 1x; Fig. 2, Fig. S2, Tab. S3, Tab. S4).
Across all species, both leaf budburst and flowering displayed a similar magnitude advance
with increased forcing, while leafout responded comparatively much stronger (Fig. 2, Fig.
S2, Tab. S3, Tab. S4).

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Our results did not show the characteristic sensitivity pattern of the FHH across most species in our experiment (Fig. 2, Fig. S2, see Fig. 1,a.,b.), though there was significant variation among species in the relative strength of their response to forcing between flowering and leaf budburst. However, when we re-ran our flowering and leaf budburst models on the subset of data which included phenological observations at only high levels of chilling and photoperiod, we found the the sensitivity to forcing for 9/10 species followed the predicted pattern of the FHH, with the second phases of the FLS showing stronger sensitivity to forcing than the first phases (Fig. 3, Tab. S5).

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# 39 Climate change scenarios

Our models project that flowering, budburst and leafout will advance in our generalized future scenarios. FLS shifts depended strongly on how forcing levels change relative to chilling exposure (Fig. 4). Given the significant differences in sensitivity to chilling between flowering and leafing phenology we found in our experiment, our scenarios showed that FLS interphases will be more strongly influenced by changes in chilling duration than increased forcing alone.

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The direction and magnitude of shifts in FLS interphases varied among species, varying espe-346 cially by whether or not they displayed a flowering-first FLS. Under some warming scenarios, 347 flower-leafout interphases were strongly reduced in a number of taxa, and flower-budburst 348 interphases for some species effectively disappeared or the order of phenophases in the FLS 349 switched (Fig. 4, Fig. S3). Several species, Acer rubrum, Ilex verticillata, Prunus pensyl-350 vanica, Prunus virginiana, and Viburnum acerifolium, had FLSs that were relatively robust 351 to changing environments. For other species, Acer pensylvanicum, Vaccinium corymbosum 352 and *Ilex mucronata*, which typically begin to produce leaves shortly before flowers open, the 353 magnitudes of projected FLS shifts were moderate. The two species with the most significant FLS shifts in both direction and magnitude across treatment combinations and climate 355 change scenarios were the flowering-first, wind-pollinated shrubs Comptonia peregrina and 356 Corylus cornuta (Fig. S3). In all of our climate change scenarios, the FLS interphase was 357 dramatically reduced in these taxa. 358

# Discussion

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Our experiment supports the hypothesis that flower-leaf sequences are structured by differ-361 ential sensitivity (DSH) to the environment between flower and leaf phenology. Specifically, 362 differences in the chilling response among flowering and leaf phenology was the strongest 363 driver of FLS variation. We also found that under high chilling and long photoperiods, the 364 FLSs of most species followed the predicted sensitivity pattern of the FHH, supporting the 365 hypothesis that the FHH is likely a special case of the DSH that occurs when other phenological cues are met for both vegetative and flowering phenology. Together, these results 367 explain why the two FLS hypotheses have been difficult to distinguish, and suggest that cli-368 mate change has the potential to significantly disrupt FLSs as global warming alters historic 369 chilling patterns across the temperate zone (Morin et al., 2009).

# Reconciling the differential sensitivity and the forcing hierarchy hypotheses

The relationship we found between the FHH and the DSH in our experiment offers a path forward for reconciling previous studies that have favored one hypothesis over the other.

Support for the FHH is most often associated with observational studies (e.g. Cosmulescu & Calusaru, 2020; Guo et al., 2014). By contrast, experimental studies which manipulate chilling levels beyond historically observed minima in the field tend to support the DSH (e.g. Aslamarz et al., 2009; Gariglio et al., 2006). This pattern can be explained by the FHH being a special case of the DSH, if historic climate regimes make detection of the DSH in the field difficult.

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Under historic climate regimes, seasonal chilling requirements were usually met (Gauzere 382 et al., 2019; Chuine et al., 2016), making it difficult to detect any effect of chilling in ob-383 servational studies, let alone differences in flower versus leaf responses. It would take larger 384 reductions in chilling to reveal the signature of the DSH, which at present, rarely occurs 385 in the field, but can be induced in artificial environments, which explains the discrepancy 386 between these two methods. These dynamics are consistent with findings in other phenolog-387 ical studies that suggest simple growing degree models (which underlie the FHH) accurately 388 predict phenology under current climate, but under-perform under climate change scenarios when shifts in chilling accumulation become more pronounced (Linkosalo et al., 2008; Chuine 390 et al., 2016). Our findings suggest that as climate continues to change, differential sensitivity 391 to the environment between flower and leaf phenology should become more apparent in field 392 observations, and that the magnitude of interannual FLS variation is likely to extend beyond historically observed reaction norms. 394

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#### Population-level implications of the DSH with climate change

Predicted shifts in chilling are highly variable across both time and space—because chilling 397 only accumulates at intermediately low temperatures, warming may increase chilling at some 398 locations while decreasing it in others (Man et al., 2017; Zhang et al., 2007). Layered onto 399 these variable shifts in chilling, spring warming from climate change also differs by location 400 (Karmalkar & Bradley, 2017; Loarie et al., 2009), suggesting that forcing may increase more 401 rapidly in some places than others. These shifts in both chilling and forcing suggest that the direction and magnitude of FLS shifts is likely to vary substantially among populations based 403 on the specific cue combinations at a given locality (Chmielewski et al., 2012). Long-term 404 phenology records show there was already substantial intra-specific variation in FLSs at the 405 population level (Buonaiuto et al., 2021) and our findings suggest that these population-level 406 differences may be further amplified by climate change. In this way, all the three generic FLS 407 climate change scenarios depicted in Fig. 4 should not be considered alternatives to each 408 other, but could occur contemporaneously across a species' range. 409

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This increased population-level heterogeneity in FLSs that our models project has the po-411 tential to influence patterns of pollen dispersal across the landscape (Borycka et al., 2017; 412 Pace et al., 2018). For example, in wind-pollinated species advancing canopy closure relative 413 to flowering can impede long-distance pollen transport (Milleron et al., 2012), which may 414 in turn alter patterns of gene flow across a landscape. In biotically-pollinated species FLS 415 shifts may interact with other drivers of global change such as phenological mismatches with 416 pollinators (Burkle et al., 2013) or flowering asynchrony among populations (Cresti et al., 417 2013; Zohner et al., 2018) to impact gene flow, but more research is needed to determine if and how FLS variation affects the foraging behavior of pollinators. Our findings regarding 419 FLS shifts should be integrated with projections of pollinator movement or prevailing wind 420 directions (Kling & Ackerly, 2020) to better understand how FLS variation may contribute 421

to population structure in the long term.

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While changes in chilling and forcing from climate change may increase FLS variation, the 424 climate change implications of strong differential sensitivity to photoperiod among flower 425 and leaf phenology are more difficult to predict. Climate change does not directly impact 426 photoperiod, but may alter the photoperiods plants experience as they approach their tem-427 perature optima for phenological activity. However, in most temperate regions, temperature 428 would have to drive phenological shifts by a minimum of several weeks before the experienced 429 photoperiod of plants would change substantially. Because these dynamics are more extreme 430 than most studies suggest (Thackeray et al., 2016) and the photoperiod effect we observed on 431 our study was relatively weak, we modeled climate change scenarios with a constant photope-432 riod in our FLS scenarios. Such an approach may be useful at temperate and some boreal 433 latitudes, but may not work at high latitudes, where photoperiod changes more rapidly over 434 the season and thus temperature-driven phenological shifts may significantly alter the expe-435 rienced photoperiod of plants. Our results suggest that such shifts could mute or amplify the 436 FLS shifts, which may be particularly important as species shift their distribution poleward 437 with climate change and begin to encounter novel photoperiod regimes (Way & Montgomery, 438 2015). 439

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# Species-level implications of the DSH with climate change

The impact of FLS shifts with climate change rely on both the function of FLS variation and the magnitude of FLS shifts. Just as previous analyses suggest that the function of FLS variation may differ among species (Buonaiuto et al., 2021; Gougherty & Gougherty, 2018), our study highlights that the direction and magnitude of FLS shifts with climate change are also species-specific. We found that FLSs of some species were very sensitive to changing

climate conditions, while other remain fairly resilient (Fig. 4, Fig. S3).

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The different FLS response patterns we observed across species may correlate with suite of interrelated anatomical, physiological and phenological traits. Just as pollination syndrome appears to be a major driver of differences in the function of FLSs in woody plants (Buonaiuto et al., 2021), we also found significant differences in the magnitude and direction of FLS shifts between biotically and wind-pollinated species.

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The species that maintained FLS structure across our climate change scenarios were bioticallypollinated. They also generally shared a strongly leafing-first FLS, with a fairly long FLS
interphase (Fig 4). As expected, these species tended to have mixed buds so there may be
strong physical constraints on their FLSs. Our models suggest that the FLSs of a subset
of biotically-pollinated taxa, those classified with "concurrent", are expected to shift under
some scenarios (see Fig 4), but it is unclear if these FLS shifts would be large enough in
magnitude to impact plant performance.

462

By contrast, the species that were most sensitive to FLS shifts were monoecious, flowering-463 first, wind-pollinated shrubs (Fig 4, Fig S3). This result supports other evidence that the 464 reproductive phenology of wind-pollinated species may be more sensitive to climate change 465 compared to biotically pollinated taxa (Ziello et al., 2012). In all scenarios, our projections 466 suggest that the flower-leaf interphase will decrease for these species. Given this period is 467 considered critical for successful pollination in these taxa (Whitehead, 1969), this pattern 468 suggests that these species, and flowering-first, wind-pollinated taxa in general, may face 469 particular risk for reproductive performance reductions with climate change. 470

471

However, there are several caveats to this prediction. The function of FLS variation in wind-

pollinated species suggests that leaves must create a substantial barrier to pollen movement. 473 In our study, we observed both leaf budburst, a stage that is unlikely to contribute much 474 to canopy structure, and leafout, a stage more likely to impact airflow though the canopy. 475 It is unclear at what point in leaf development wind-pollination becomes impaired, so the 476 effective duration of the FLS interphase that impacts wind pollination may be somewhere in between the two interphases we measured (flower-leaf budburst and flower-leafout) or be-478 yond the interphases used in our projections. Secondly, in growth chamber experiments such 479 as ours, it is difficult to evaluate the full duration of phenological episode (Primack et al., 480 2015). Shifts in the duration of flowering or period of leaf expansion could compensate for, 481 or exacerbate the shifts in FLSs we observed. 482

483

Finally, many wind-pollinated species are monoecious or dioecious (Ackerman, 2000) and in 484 several families (e.g. Betulaceae, Juglandaceace), male flowers occur in separate buds, while 485 female flowers occur in mixed buds (Pope et al., 2013). This suggest that the phenology of 486 female flowers may be more tightly constrained by leaf phenology and if this is the case, even 487 increases in the FLS interphases for flowering-first wind-pollinated taxa such as those ob-488 served in the field (e.g. Ma et al., 2020) could adversely affect wind-pollination by disrupting 489 floral synchrony of male and female flowers (Alizoti et al., 2010; Mutke et al., 2005; Elkassaby 490 & Davidson, 1991). 491

492

While much of the public interest around phenology and pollination in the context of global change has centered around trophic mismatches between pollinator and floral phenology (Memmott *et al.*, 2007), our study identified the possibility that the effects of FLS shifts with climate change may be particularly important for wind-pollinated woody plants. The direction and magnitude of FLS shifts we observed in these taxa, coupled with the hypothesized function of a flowering-first FLS in wind-pollinated species, suggests that FLS variation

in this functional group should be explored in greater detail in the future. More research is needed to identify species' traits that may correlate with the potential for FLS shifts, but flowering-first, wind-pollinated species may be particular sensitive to FLS shifts. Our results suggest that species in this functional group should be considered a research priority for the study of spring phenological sequences in deciduous woody plants.

504

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508

## **Author Contributions:**

DMB and EMW conceived of the experiment; DMB collected the data and performed the statistical analyses, DMB and EMW wrote the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

513

# Data Availability Statement:

Data and code will be available on KNB (https://knb.ecoinformatics.org/) upon publication.

# 516 References

- Ackerman, J.D. (2000) Abiotic pollen and pollination: ecological, functional, and evolutionary perspectives. *Plant Systematics and Evolution* **222**, 167–185.
- Alizoti, P.G., Kilimis, K. & Gallios, P. (2010) Temporal and spatial variation of flowering
- among Pinus nigra Arn. clones under changing climatic conditions. Forest Ecology and
- Management 259, 786–797, Conference on Adaptation of Forests and Forest Management
- to Changing Climate with Emphasis on Forest Health, Umea, Sweden, AUG 25-28, 2008.
- Aslamarz, A.A., Vahdati, K., Rahemi, M. & Hassani, D. (2009) Estimation of chilling and
- heat requirements of some persian walnut cultivars and genotypes. HortScience 44, 697–
- <sub>525</sub> 701.
- <sup>526</sup> Augspurger, C.K. & Zaya, D.N. (2020) Concordance of long-term shifts with climate warming
- varies among phenological events and herbaceous species. Ecological Monographs n/a,
- e01421.
- Basler, D. & Körner, C. (2012) Photoperiod sensitivity of bud burst in 14 temperate forest
- tree species. Agricultural and Forest Meteorology 165, 73–81.
- Borycka, K., Ortyl, B. & Kasprzyk, I. (2017) Temporal variations and spatial differentiation in
- the black alder and silver birch pollination pattern-the impact of local climate or something
- more? Agricultural and Forest Meteorology **247**, 65–78.
- Bukovac, Z., Shrestha, M., Garcia, J.E., Burd, M., Dorin, A. & Dyer, A.G. (2017) Why
- background colour matters to bees and flowers. Journal of Comparative Physiology A 203,
- <sub>536</sub> 369–380.
- Buonaiuto, D.M., Morales-Castilla, I. & Wolkovich, E.M. (2021) Reconciling competing hy-
- potheses regarding flower-leaf sequences in temperate forests for fundamental and global
- change biology. New Phytologist 229, 1206–1214.

- Burkle, L.A., Marlin, J.C. & Knight, T.M. (2013) Plant-pollinator interactions over 120 years: loss of species, co-occurrence, and function. *Science* **339**, 1611–1615.
- Bürkner, P.C. (2018) Advanced bayesian multilevel modeling with the r package brms. RJournal 10, 395–411.
- Chmielewski, F.M., Blümel, K. & Pálesová, I. (2012) Climate change and shifts in dormancy
   release for deciduous fruit crops in germany. Climate Research 54, 209–219.
- Chuine, I., Bonhomme, M., Legave, J.M., García de Cortázar-Atauri, I., Charrier, G., Lacointe, A. & Améglio, T. (2016) Can phenological models predict tree phenology accurately
  in the future? the unrevealed hurdle of endodormancy break. *Global Change Biology* 22,
  3444–3460.
- Citadin, I., Raseira, M.C.B., Herter, F.G. & da Silva, J.B. (2001) Heat requirement for blooming and leafing in peach. *HortScience HortSci* **36**, 305–307.
- Cleland, E.E., Chuine, I., Menzel, A., Mooney, H.A. & Schwartz, M.D. (2007) Shifting plant phenology in response to global change. *Trends in Ecology Evolution* **22**, 357 365.
- Cosmulescu, S. & Calusaru, F.G. (2020) Influence of temperature on blackthorn (prunus spinosa l.) phenophases in spring season. *Journal of Agricultural Meteorology* **76**, –.
- Cosmulescu, S. & Ionescu, M. (2018) Phenological calendar in some walnut genotypes grown
   in romania and its correlations with air temperature. *International Journal of Biometeo-* rology 62.
- Cresti, M., Aroca, R., Khanduri, V.P., Sharma, C.M., Kumar, K.S. & Ghildiyal, S.K. (2013)
   Annual variation in flowering phenology, pollination, mating system, and pollen yield in
   two natural populations of schima wallichii (dc.) korth. The Scientific World Journal 2013,
   350157.

- Di Cecco, G.J. & Gouhier, T.C. (2018) Increased spatial and temporal autocorrelation of temperature under climate change. *Scientific Reports* 8, 14850.
- Di-Giovanni, F., Beckett, P.M. & Flenley, J.R. (1989) Modelling of dispersion and deposition of tree pollen within a forest canopy. *Grana* 28, 129–139.
- Diggle, P.K. (1995) Architectural effects and the interpretation of patterns of fruit and seed development **26**, 531–552.
- Donnelly, A. & Yu, R. (2021) Temperate deciduous shrub phenology: the overlooked forest layer. *International journal of biometeorology* **65**, 343–355.
- Elkassaby, Y. & Davidson, R. (1991) Impact of pollination environment manipulation on the apparent outcrossing rate in a Douglas Fir seed orchard. *Heredity* **66**, 55–59.
- Ettinger, A., Gee, S. & Wolkovich, E. (2018) Phenological sequences: how early season events
  define those that follow. *American Journal of Botany* **105**.
- Ettinger, A.K., Chamberlain, C.J., Morales-Castilla, I., Buonaiuto, D.M., Flynn, D.F.B.,
   Savas, T., Samaha, J.A. & Wolkovich, E.M. (2020) Winter temperatures predominate in
   spring phenological responses to warming. *Nature Climate Change* 10, 1137–1142.
- Finn, G.A., Straszewski, A.E. & Peterson, V. (2007) A general growth stage key for describing trees and woody plants. *Annals of Applied Biology* **151**, 127–131.
- Flynn, D.F.B. & Wolkovich, E.M. (2018) Temperature and photoperiod drive spring phenology across all species in a temperate forest community. *New Phytologist* **219**, 1353–1362.
- Forrest, J. & Thomson, J.D. (2009) Background complexity affects colour preference in bumblebees. *Naturwissenschaften* **96**, 921–925.
- Franklin, D.C. (2016) Flowering while leafess in the seasonal tropics need not be cued by

- leaf drop: evidence from the woody genus brachychiton (malvaceae). Plant Ecology and
  Evolution 149, 272–279.
- Friedman, J. & Barrett, S.C.H. (2009) Wind of change: new insights on the ecology and evolution of pollination and mating in wind-pollinated plants. *Annals of Botany* **103**, 1515–1527.
- Gariglio, N., González Rossia, D.E., Mendow, M., Reig, C. & Agusti, M. (2006) Effect of artificial chilling on the depth of endodormancy and vegetative and flower budbreak of peach and nectarine cultivars using excised shoots. *Scientia Horticulturae* **108**, 371–377.
- Gauzere, J., Lucas, C., Ronce, O., Davi, H. & Chuine, I. (2019) Sensitivity analysis of tree phenology models reveals increasing sensitivity of their predictions to winter chilling temperature and photoperiod with warming climate. *Ecological Modelling* **411**, 108805.
- Gellman, A. & Hill, J. (2007) Data Analysis Using Regression and Multilevel/Hierarchical
   Models. Cambridge University Press.
- Gelman, A., Carlin, J., Stern, H., Dunson, D., Vehtari, A. & Rubin, D. (2013) Bayesian Data
   Analysis. Chapman and Hall/CRC.
- Gougherty, A.V. & Gougherty, S.W. (2018) Sequence of flower and leaf emergence in deciduous trees is linked to ecological traits, phylogenetics, and climate. New Phytologist 220, 121–131.
- Guo, L., Luedeling, E., Dai, J. & Xu, J. (2014) Differences in heat requirements of flower
   and leaf buds make hysteranthous trees bloom before leaf unfolding. *Plant Diversity and Resources* 36, 245–253.
- Janzen, D.H. (1967) Synchronization of sexual reproduction of trees within the dry season in central america. *Evolution* **21**, 620–637.

- $_{608}$  Karmalkar, A.V. & Bradley, R.S. (2017) Consequences of global warming of 1.5  $\,^{\circ}\mathrm{c}$  and 2  $\,^{\circ}\mathrm{c}$
- for regional temperature and precipitation changes in the contiguous united states. PLOS
- ONE **12**, e0168697–.
- Kay, M. (2020) tidybayes: Tidy Data and Geoms for Bayesian Models. R package version
- 612 2.1.1.
- Kling, M.M. & Ackerly, D.D. (2020) Global wind patterns and the vulnerability of wind-
- dispersed species to climate change. Nature Climate Change 10, 868–875.
- Körner, C. & Basler, D. (2010) Phenology under global warming. Science 327, 1461–1462.
- Laube, J., Sparks, T.H., Estrella, N., Höfler, J., Ankerst, D.P. & Menzel, A. (2014) Chilling
- outweighs photoperiod in preventing precocious spring development. Global Change Biology
- **20**, 170–182.
- Linkosalo, T., Lappalainen, H.K. & Hari, P. (2008) A comparison of phenological models of
- leaf bud burst and flowering of boreal trees using independent observations. Tree Physiology
- **28**, 1873–1882.
- Loarie, S.R., Duffy, P.B., Hamilton, H., Asner, G.P., Field, C.B. & Ackerly, D.D. (2009) The
- velocity of climate change. Nature 462, 1052–1055.
- 624 Luedeling, E. (2012) Climate change impacts on winter chill for temperate fruit and nut
- production: A review. Scientia Horticulturae 144, 218–229.
- Luedeling, E., Girvetz, E.H., Semenov, M.A. & Brown, P.H. (2011) Climate change affects
- winter chill for temperate fruit and nut trees. PLOS ONE 6, e20155-.
- Ma, Q., Huang, J.G., Hänninen, H., Li, X. & Berninger, F. (2020) Climate warming prolongs
- the time interval between leaf-out and flowering in temperate trees: Effects of chilling,
- forcing and photoperiod. Journal of Ecology n/a.

- Man, R., Lu, P. & Dang, Q.L. (2017) Insufficient chilling effects vary among boreal tree species and chilling duration. Frontiers in Plant Science 8, 1354.
- Mehlenbacher, S.A. (1991) Chilling requirements of hazelnut cultivars. Scientia Horticulturae
   47, 271–282.
- Memmott, J., Craze, P.G., Waser, N.M. & Price, M.V. (2007) Global warming and the disruption of plant–pollinator interactions. *Ecology Letters* **10**, 710–717.
- 637 Menzel, A., Sparks, T.H., Estrella, N., Koch, E., Aasa, A., Ahas, R., Alm-Kuebler, K.,
- Bissolli, P., Braslavska, O., Briede, A., Chmielewski, F.M., Crepinsek, Z., Curnel, Y.,
- Dahl, A., Defila, C., Donnelly, A., Filella, Y., Jatcza, K., Mage, F., Mestre, A., Nordli, O.,
- Penuelas, J., Pirinen, P., Remisova, V., Scheifinger, H., Striz, M., Susnik, A., Van Vliet,
- A.J.H., Wielgolaski, F.E., Zach, S. & Zust, A. (2006) European phenological response to
- climate change matches the warming pattern. Global Change Biology 12, 1969–1976.
- 643 Milleron, M., Lopez de Heredia, U., Lorenzo, Z., Perea, R., Dounavi, A., Alonso, J., Gil,
- 644 L. & Nanos, N. (2012) Effect of canopy closure on pollen dispersal in a wind-pollinated
- species (Fagus sylvatica L.). Plant Ecology 213, 1715–1728.
- Morin, X., Lechowicz, M.J., Augspurger, C.K., O'Keefe, J., Viner, D. & Chuine, I. (2009)
- Leaf phenology in 22 north american tree species during the 21st century. Global Change
- Biology **15**, 961–975.
- Mutke, S., Gordo, J. & Gil, L. (2005) Variability of mediterranean stone pine cone production:
- Yield loss as response to climate change. Agricultural and Forest Meteorology 132, 263–272.
- <sup>651</sup> O'Keefe, J. (2015) Phenology of Woody Species at Harvard Forest since 1990. Harvard Forest
- Data Archive: HF003., Petersham, MA, USA.
- Pace, L., Boccacci, L., Casilli, M., Di Carlo, P. & Fattorini, S. (2018) Correlations between

- weather conditions and airborne pollen concentration and diversity in a mediterranean high-altitude site disclose unexpected temporal patterns. *Aerobiologia* **34**, 75–87.
- Parmesan, C. & Yohe, G. (2003) A globally coherent fingerprint of climate change impacts across natural systems. *Nature* **421**, 37 EP –.
- Pope, K.S., Dose, V., Da Silva, D., Brown, P.H., Leslie, C.A. & DeJong, T.M. (2013) Detect-
- ing nonlinear response of spring phenology to climate change by bayesian analysis. Global
- 660 Change Biology 19, 1518–1525.
- Post, E.S., Pedersen, C., Wilmers, C.C. & Forchhammer, M.C. (2008) Phenological sequences reveal aggregate life history response to climatic warming. *Ecology* **89**, 363–370.
- Primack, R.B. (1987) Relationships among flowers, fruits, and seeds. Annual Review of Ecology and Systematics 18, 409–430.
- Primack, R.B., Laube, J., Gallinat, A.S. & Menzel, A. (2015) From observations to experiments in phenology research: investigating climate change impacts on trees and shrubs using dormant twigs. *Annals of Botany* **116**, 889–897.
- Rathcke, B. & Lacey, E.P. (1985) Phenological patterns of terrestrial plants. *Annual Review*of Ecology and Systematics **16**, 179–214.
- Revell, L.J. (2010) Phylogenetic signal and linear regression on species data. *Methods in Ecology and Evolution* 1, 319–329.
- Tauber, H. (1967) Investigations of the mode of pollen transfer in forested areas. Review of

  Palaeobotany and Palynology 3, 277–288.
- Thackeray, S.J., Henrys, P.A., Hemming, D., Bell, J.R., Botham, M.S., Burthe, S., Helaouet,
- P., Johns, D.G., Jones, I.D., Leech, D.I., Mackay, E.B., Massimino, D., Atkinson, S., Bacon,
- P.J., Brereton, T.M., Carvalho, L., Clutton-Brock, T.H., Duck, C., Edwards, M., Elliott,

- J.M., Hall, S.J.G., Harrington, R., Pearce-Higgins, J.W., Høye, T.T., Kruuk, L.E.B.,
- Pemberton, J.M., Sparks, T.H., Thompson, P.M., White, I., Winfield, I.J. & Wanless,
- S. (2016) Phenological sensitivity to climate across taxa and trophic levels. *Nature* 535,
- 680 241-245.
- Vitasse, Y. & Basler, D. (2014) Is the use of cuttings a good proxy to explore phenological
- responses of temperate forests in warming and photoperiod experiments? Tree Physiology
- 683
- Way, D.A. & Montgomery, R.A. (2015) Photoperiod constraints on tree phenology, perfor-
- mance and migration in a warming world. Plant, Cell & Environment 38, 1725–1736.
- Weinberger, J.H. (1950) Chilling requirements of peach varieties. id 19511602967. Proceed-
- ings. American Society for Horticultural Science **56**, 122–28.
- Whitehead, D.R. (1969) Wind pollination in the angiosperms: Evolutionary and environ-
- mental considerations. Evolution 23, 28–35.
- <sup>690</sup> Zhang, X., Tarpley, D. & Sullivan, J.T. (2007) Diverse responses of vegetation phenology to
- a warming climate. Geophysical Research Letters 34.
- <sup>692</sup> Ziello, C., Böck, A., Estrella, N., Ankerst, D. & Menzel, A. (2012) First flowering of wind-
- pollinated species with the greatest phenological advances in europe. Ecography 35, 1017–
- 694 1023.
- Zohner, C.M., Mo, L. & Renner, S.S. (2018) Global warming reduces leaf-out and flowering
- synchrony among individuals. *eLife* 7, e40214.
- Zohner, C.M., Mo, L., Renner, S.S., Svenning, J.C., Vitasse, Y., Benito, B.M., Ordonez, A.,
- Baumgarten, F., Bastin, J.F., Sebald, V., Reich, P.B., Liang, J., Nabuurs, G.J., de Miguel,
- S., Alberti, G., Antón-Fernández, C., Balazy, R., Brändli, U.B., Chen, H.Y.H., Chisholm,
- C., Cienciala, E., Dayanandan, S., Fayle, T.M., Frizzera, L., Gianelle, D., Jagodzinski,

A.M., Jaroszewicz, B., Jucker, T., Kepfer-Rojas, S., Khan, M.L., Kim, H.S., Korjus, H.,
Johannsen, V.K., Laarmann, D., Lang, M., Zawila-Niedzwiecki, T., Niklaus, P.A., Paquette, A., Pretzsch, H., Saikia, P., Schall, P., Šebeň, V., Svoboda, M., Tikhonova, E., Viana,
H., Zhang, C., Zhao, X. & Crowther, T.W. (2020) Late-spring frost risk between 1959
and 2017 decreased in north america but increased in europe and asia. *Proceedings of the*National Academy of Sciences 117, 12192–12200.

# Figures 707

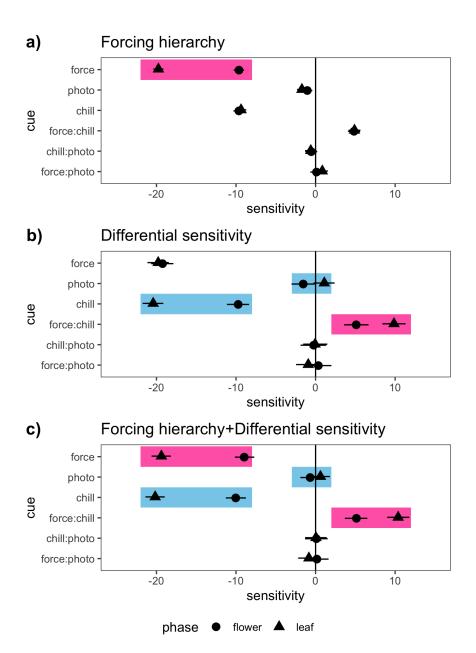


Figure 1: Characteristic sensitivity ( $\Delta$  day of phenological event/ $\Delta$  environmental cue) patterns of the phenological response to changing cue levels for the two major flower-leaf sequence hypotheses. a) displays a signature pattern of the forcing hierarchy hypothesis (FHH, pink boxes)—with the second phenophase in the sequence (in this case leafing) having a higher sensitivity to forcing than the first. b) Highlights a typical sensitivity pattern produced by the differential sensitivity hypothesis (DSH). c) Depicts a scenario where both the FHH and the DSH contribute to flower-leaf sequence variation. Here the characteristic forcing sensitivity of the FHH is still apparent but the differential sensitivity to chilling and photoperiod is seen as well (blue boxes). All plots above are based on simulations (see Supporting Information: Methods). Shapes indicate mean estimates and lines depict 95% credible intervals from Bayesian hierarchical models with advances in phenology shown as negative numbers, and delays in phenology as positive numbers.

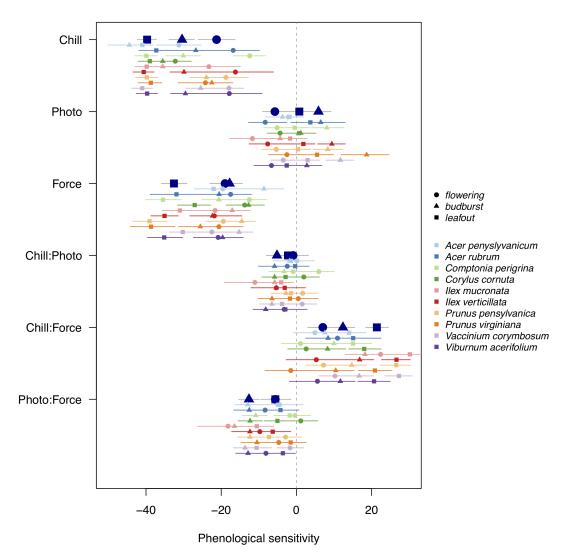


Figure 2: Effects of forcing temperature, chilling duration, and photoperiod on the leaf budburst (triangles), leafout (squares) and flowering (circles) phenology of 10 temperate woody plant species collected from Harvard Forest (Petersham, MA, USA). Shapes indicate mean estimates and lines depict 50% credible intervals (See Tab. S3, Tab. S4 for other intervals) from Bayesian hierarchical models with advances in phenology shown as negative numbers, and delays in phenology as positive numbers. Flower and leaf phenology differs in sensitivity ( $\Delta$  day of phenological event/ $\Delta$  environmental cue; 4 weeks chilling/6 °C forcing/4 hours photoperiod) to these environmental cues. See Fig. S2 for an alternative presentation of these results that depicts the difference between the mean estimates of each phase (shapes).

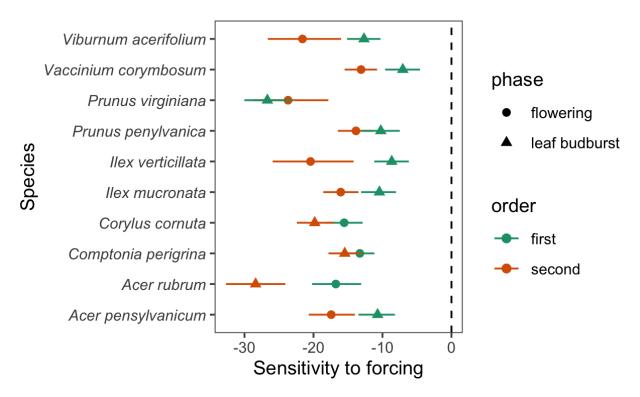


Figure 3: Phenological sensitivity ( $\Delta$  days of phenological event/  $\Delta$  6°C) to forcing temperatures of leaf budburst (triangles) and flowering (circles) phenology from 10 temperate deciduous woody plants at long (12 hour) photoperiod and long chilling duration treatments (8 weeks at 4°C). Shapes indicate mean estimates and lines depict 50% credible intervals (See Tab. S5 for other intervals) from Bayesian hierarchical models with advances in phenology shown as negative numbers. When photoperiod and chilling are high, most species follows the predicted pattern of the forcing hierarchy hypothesis (FHH), with the second phenophase of the sequence consistently more sensitive to forcing than the first. This result suggests that the FHH should be considered a special case of the differential sensitivity hypothesis (DSH) that occurs when the chilling and photoperiod requirements are met for both tissue types.

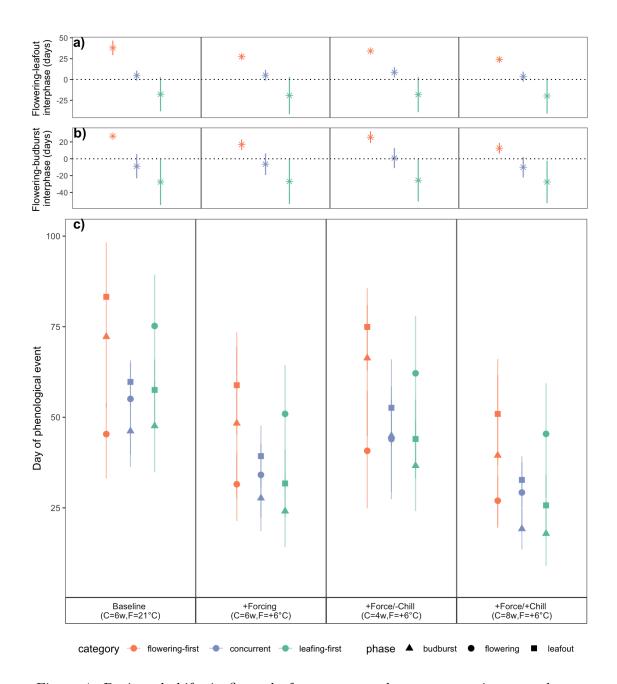


Figure 4: Projected shifts in flower-leaf sequences under current environmental conditions (Baseline) and three climate change scenarios (increase forcing, increase forcing/decrease chilling, increase forcing/increase chilling) predict that FLS shifts differ among the three major FLS types, and will be strongest is flowering-first species. Panels a) and b) show the mean time between flowering and vegetative phenological events (shapes) with 50% credible intervals (lines). Panel c) shows the predicted event day for each phase. Predictions are based on species-level posterior estimates grouped by FLS category (flowering-first, concurrent, leafing-first) from Bayesian hierarchical models comparing flowering (circles), leaf budburst (triangles) and leafout (squares) phenological responses to variable chilling duration and forcing temperatures. Shapes represent the mean estimates and lines represent the 50% credible intervals.