- Differences in flower and leaf bud environmental responses drive
- shifts in spring phenological sequences of temperate woody plants
  - D.M. Buonaiuto <sup>1,2,a</sup>, E.M. Wolkovich<sup>3</sup>
- 4 Author affiliations:
- <sup>5</sup> Arnold Arboretum of Harvard University, Boston, Massachusetts, USA. ORCID: 0000-0003-4022-
- 6 2591

3

- <sup>7</sup> Department of Organismic and Evolutionary Biology, Harvard University, Cambridge, Massachusetts,
- 8 USA

12

16

- <sup>9</sup> Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, Vancouver,
- 10 British Columbia, Canada
- <sup>a</sup>Corresponding author: 617.823.0687; dbuonaiuto@g.harvard.edu
- 13 Acknowledgements: We thank C. de Keyzer for generously helping to collect and prepare plant
- 14 materials for the experiment. We also thank R. Lee for help with monitoring and maintaining the
- 15 experiment.
- 17 Author Contributions: DMB and EMW conceived of the experiment; DMB collected the data
- and performed the statistical analyses, DMB and EMW wrote the manuscript. All authors con-
- 19 tributed critically to the drafts and gave final approval for publication.
- Data Availability Statement: Data will be available on KNB upon publication. All modeling
- code will be made available upon request.

#### 23 Abstract

The relative timing of growth and reproduction is an important driver of plant fitness. For deciduous woody species in temperate regions leaves and flowers both appear in the early spring, but the order and duration of these phenological events vary among species, populations, and individuals. This observed variation in flower-leaf sequences (FLSs) suggests the relative timing of these events 27 may be important—affecting the reproduction, recruitment and survival of individuals. Further, 28 FLSs appear to be shifting with climate change, and anticipating the extent of these shifts is key to 29 projecting how climate change may impact species' performance and reshape forest communities. Predicting FLS shifts, however, requires an improved understanding of how the environment dic-31 tates FLS patterns. To address this, we compared the phenological responses of flower and leaf buds for 10 temperate woody species to varying levels of temperature and light in a lab experiment. Our 33 experimental design allowed us to test competing hypotheses for how environmental cues determine FLS variation—specifically whether forcing (warm temperatures) alone drivers variation or differential sensitivity to chilling (cool temperatures generally in the fall and winter) and/or photoperiod matter. We found that flower and leaf buds respond with differential sensitivity to environmental 37 cues, with differences in their response to chilling being the dominant driver of FLS variation. Be-38 cause climate change will amplify variability in chilling across time and space, these findings suggest 39 that FLS shifts may be large, but are likely to vary substantially among populations and species. In our study, FLS shifts were largest in wind-pollinated species that flower before leafing, raising the possibility that, more generally, wind-pollinated taxa may experience reproductive declines due to FLS shifts in the decades to come.

44

45 Keywords: chilling, climate change, deciduous forests, differential sensitivity, flower-leaf sequences,

46 forcing, hysteranthy, phenology

# 18 Introduction

Among the most widely documented biological effects of anthropogenic climate change are shifts in plant phenology, the timing of life cycle events (Parmesan & Yohe, 2003; Menzel et al., 2006; 50 Cleland et al., 2007). While phenology is generally advancing with climate change, the strength 51 of these phenological shifts can vary substantially among specific phenological phases (Augspurger 52 & Zaya, 2020). These differences alter the timing of phases relative to each other, changing the 53 duration between events that make up phenological sequences (Ettinger et al., 2018). Phenological 54 sequences are a major driver of plant fitness that impacts plant life history, resource allocation, demography and ecosystem processes (Post et al., 2008). Thus, shifting sequences with climate 56 change will likely impact many of these processes. The effects of these shifts, however, depend 57 both on their direction—whether distinct phases are shifting closer together or farther apart—and 58 magnitude—how much they are shifting relative to each other. 59

60

Among deciduous woody plants, the relative timing of flower and leaf phenology, or flower-leaf sequences (FLSs), may be particularly consequential to fitness in temperate regions where flowering prior to leaf development is common (Rathcke & Lacey, 1985; Gougherty & Gougherty, 2018). Flowering before leafing may be a critical adaptation for pollination efficiency in wind-pollinated taxa, with the leafless canopy minimizing obstacles for air flow and pollen transfer (Whitehead, 1969). In insect-pollinated taxa, flowering-first may increase the visibility of flowers to pollinators (Janzen, 1967; Savage, 2019) or alleviate hydraulic demand in dry conditions (Gougherty & Gougherty, 2018; Franklin, 2016).

69

Phenological observations over the last several decades indicate that, like other phenological sequences, FLSs are shifting due to anthropogenic climate change (Buonaiuto *et al.*, 2020). For several species, the time between flowering and leafing appears to be increasing, but the strength of this trend varies among species and the direction of FLS shifts are not consistent across populations (Buonaiuto *et al.*, 2020). These changes could affect the important functions of FLSs, potentially

<sub>75</sub> putting some species at greater risk for fitness declines, while benefiting others.

76

For example, species with decreasing FLS interphases with climate change may see a reduction in pollination success as more pollen is intercepted by vegetation or flowers are obscured by developing leaves. Conversely, pollination efficiency could improve for species with lengthening FLS interphases. A change in the FLS interphase of just a few days would likely have little impact on these processes, but if shifts were on the order of weeks, the impact on the pollination biology of a species could be significant. For example, because canopy structure influences the speed and direction of air flow (Niklas, 1985; Jackson & Lyford, 1999), decreasing FLS interphases could reduce pollen dispersal distances for wind-pollinated taxa (Milleron et al., 2012), which would ultimately impact population structure and gene flow across a landscape.

86

Predicting FLS shifts requires identifying the proximate mechanisms that drive FLS variation.

Decades of research suggest that cool winter temperatures (chilling), warm spring temperatures (forcing), and day-length (photoperiod) are the primary drivers of both reproductive and vegetative phenology for woody plants in temperate regions (Forrest & Miller-Rushing, 2010; Flynn & Wolkovich, 2018). However, observed FLS shifts indicate that there must be differences in how these cues influence the phenology of flowers and leaves (Buonaiuto et al., 2020). Identifying these differences is a necessary step for predicting the direction, magnitude and—ultimately—fitness impacts of FLS shifts with climate change.

### 95 Hypotheses for FLS variation

Studies that have attempted to identify the differences between reproductive and vegetative phenology in woody plants (mostly focused on crop species) have yielded two common explanations:

One hypothesis suggests that reproductive and vegetative buds utilize the same underlying environmental cues, but have different threshold responses to forcing, with whichever bud type bursts later—leaves or flowers—having a higher thermal requirement (that is, they need a great sum of warm temperature to trigger the phenological event, Guo et al., 2014; Cosmulescu & Calusaru, 2020;
Cosmulescu & Ionescu, 2018). Under this hypothesis, which we call the forcing hierarchy hypothesis
(FHH), leaf and flower buds share the same suite of cues and develop similarly to non-forcing cues
(i.e., chilling and photoperiod), but differ in the thermal requirement for budburst.

105

In contrast, an alternative hypothesis suggests that flower and leaf buds differ in the strength of their phenological responses to multiple environmental cues (Citadin *et al.*, 2001; Gariglio *et al.*, 2006; Aslamarz *et al.*, 2009; Mehlenbacher, 1991). Under this hypothesis, which we call the differential sensitivity hypothesis (DSH), each bud type relies more or less on certain cues, generating FLS variability.

111

While observational studies of phenology in the field are commonly used to evaluate the relationship 112 between environmental conditions and phenology (Cleland et al., 2007), under current and recent 113 climatic conditions, the FHH and DSH may produce similar phenological patterns, making it dif-114 ficult to evaluate these hypotheses with such methods. However, experiments designed to isolate all three environmental cues have the potential to disentangle the two hypotheses. Studies aiming 116 to differentiate these hypotheses can look for two different signatures. The key signature of the 117 FHH is that the sensitivity to forcing ( $\Delta$  day of phenological event/ $\Delta$ °C) of the second phase in 118 the phenological sequence is always greater than that of the first phase with sensitivity differences 119 being inversely proportional to the difference in thermal requirement among bud types (2x in our 120 simulations; Fig. 1 a.). By contrast, the signature of the DSH is that sensitivity estimates of flower 121 and leaf buds to chilling and/or photoperiod will diverge (Fig. 1 c.), but there is no characteristic 122 pattern as to the strength or direction of this divergence (e.g., the leaves may require longer or 123 shorter photoperiods than the flowers, or more or less chilling). 124

125

The clarity of these signatures from each hypothesis, however, may be obscured in certain situations.

First, a species' buds could be both differentially sensitive to photoperiod or chilling (supporting the DSH) and also respond to forcing in a forcing hierarchy (that is, both hypotheses can operate

at once, see Fig. 1 b.). Second, the differential sensitivity framework can generate this signature of
the FHH when other cues are at high levels due to interactions between the chilling or photoperiod
response and thermal requirement, which makes forcing the dominant phenological cue once other
cue requirements have been met (see Fig. 1 c., chill x force interaction). When this happens the
FHH would effectively be a special case of the DSH. That is, the presence of FHH does not exclude
the DSH (Gariglio et al., 2006).

While these two hypothesized mechanisms may produce similar phenology patterns under current conditions, differentiating them is important, as they have substantially different implications re-garding the potential for FLS shifts with climate change. The FHH suggests that FLS variation is largely a product of climate variation during the interphase. If spring temperatures increase with climate change, the second phenophase of the FLS with be accelerated relative to the first and FLS interphases will decrease, but given the relative auto-correlation of spring temperatures (Di Cecco & Gouhier, 2018), these shifts should be muted. If FLSs are structured by a forcing hierarchy, the direction of FLS shifts are relatively straight forward to predict, and it is less likely that shifts will be large enough in magnitude to significantly affect woody plant fitness. 

Climate change would also drive FLS shifts under the DSH, but the trajectory of these shifts are more complicated to predict. The DSH suggests that there could be strongly localized effects of climate change on FLSs. Shifts in FLS variation will depend on the direction and rate of change in cues at given locations and the species-specific differential sensitivity of reproductive and vegetative phenology to cue combinations. This hypothesis allows for larger magnitude shift in FLSs, on a scale that could impact pollen transport or the physiological functioning of woody plants. Further, the DSH also suggests that the magnitude of shifts may be highly divergent both among species in a community, and among populations of the same species which could impact gene flow, population structure and demography.

In this study we tested the FHH and DSH hypotheses in 10 temperate shrub and tree species. Using

a full-factorial growth-chamber experiment we manipulated chilling, forcing and photoperiod cues 157 for flower and leaf buds. We then leveraged these data to examine how FLSs may shift with climate 158 change. 159

160

#### Materials and Methods

#### Growth chamber study

We sampled plant material from Harvard Forest in Petersham, MA, USA (42.5314°N, 72.1900°W) 163 on 25 October 2017, immediately after most plants in the area entered dormancy but before they 164 could accumulate significant chilling in the field. We collected branch cuttings from 7-13 individuals 165 of 12 woody plant species (4-12 cuttings per individual for a total of 48-56 per species), consisting 166 of a mix of deciduous shrubs, understory and canopy trees commonly found in mesic hardwood 167 forests of the eastern United States. We selected species displaying variation of the three major 168 FLS patterns; species for which flowers appear before leaves (flowering-first; Acer rubrum, Betula 169 alleghaniensis, Comptonia peregrina, and Corulus cornuta), species for which flowers appear with 170 their leaves (concurrent; Acer pensylvanicum, Acer saccharum, Ilex mucronata, Prunus pensylvanica, 171 Vaccinium corymbosum), and species for which flowers appear after leaf development (leafingfirst; Ilex verticillata, Prunus virginiana, Viburnum acerifolium), These species also vary in other important functional traits including pollination syndrome, bud type, and growth habit (see tab. 174 S1 for details and full species list). 175

We transported all cuttings to the Arnold Arboretum in Boston, MA (USA) where they were re-cut 176 in water to prevent callousing and cavitation and placed in 500 ml Erlenmeyer flasks with distilled 177 water. 178

179

We randomly assigned cuttings to a fully crossed set of eight experimental treatments; two lev-180 els of chilling (4 vs 8 weeks at 4°C), two levels of temperature (24°C:18°C (day/night) warm vs 18°:12°C (day/night) cool) and two levels of photoperiod (12 vs 8 hours). We alternated day/night
temperature periodicity on a 12 hour schedule to reduce co-variation with photo-periodicity. We
re-cut all twigs and changed the water every 7-10 days and rotated all treatments between growth
chambers every two weeks to minimize chamber effects. We made phenological observations every
2-3 days using a modified BBCH scale for woody plants (Finn et al., 2007) for three months following release from chilling conditions. In this period we assessed two phenological phases: budburst
(BBCH phase 07) and first flower open (BBCH 60). At the conclusion of this period we assessed
all individuals that did not undergo budburst and excluded 56 dead twigs from our analyses.

#### Data analysis

To assess the phenological sensitivity of each phase, we fit mixed-effects hierarchical models with 191 chilling, forcing, photoperiod and all two-way interactions as the fixed effects and species as a group-192 ing factor on both the slopes and the intercepts. We chose a Bayesian, hierarchical approach in 193 order to identify systematic trends across species' responses while accounting for sample size, vari-194 ance, and the unique effect of each species. We also used z-scoring to standardized the units of each 195 environmental predictor to allow for appropriate comparisons between them (Gellman & Hill, 2007). 196 We modeled the effects of environmental parameters on flower opening and leaf budburst separately. 197 We also fit a model with FLS interphase (day of budburst - day of flowering) as a response variable 198 to compare these estimates with field observations. Two species, Betula allegheniensis and Acer 199 saccharum, produced no flowers in our trial, so we excluded them from our analysis. In total, our 200 analyses included 464 twigs from 10 species. 201

202

203 The models we fit appear below:

204

205 
$$y_{[i]} \sim N(\alpha_{sp_{[i]}} + \beta_{forcing_{sp_{[i]}}} + \beta_{chilling_{sp_{[i]}}} + \beta_{photoperiod_{sp_{[i]}}} + \beta_{forcingxchilling_{sp_{[i]}}} + \beta_{forcingxphotoperiod_{sp_{[i]}}} + \beta_{sp_{in}} +$$

Where  $y_{[i]}$  is either the day of the experiment of leaf budburst, day of first flower opening or FLS interphase length, and  $\sigma_y^2$  the error. We modeled the intercept  $(\alpha)$  and each slope  $(\beta)$  parameter at the species level using the formula:

212  $\alpha_{x_{sp}}$  or  $\beta_{x_{sp}} \sim N(\mu_x, \sigma_x^2)$ 

211

213

219

220

221

222

225

226

227

228

To test the hypothesis that the FHH is a special case of the DSH that occurs when other cue (i.e., chilling and photoperiod) requirements are met, we re-ran our models on a subset of our data which included both levels of the forcing treatment but only the high photoperiod and chilling treatment levels. This model included forcing as the only main effect but, like our main models written above, included species as a grouping factor on the slopes and intercept.

We fit all models using the R package "brms" (Bürkner, 2018). We ran each model on four chains with 4000 iterations and a 3000 iteration warm up for a total of 4000 posterior draws for each parameter. In all models we used weakly informative priors and increasing the priors 5-fold did not affect the model results. We assessed model performance through ensuring  $\hat{R}$ s were between 1 and 1.01 and bulk and tail effective sample sizes were high (1800-2800 for most parameters, but as low as 800-900 for some.) we present 50% intervals in figures generally because they are the most computationally stable (Gelman *et al.*, 2013) but provide other intervals in the Supporting Information (Tab. S3,Tab. S4, Tab. ??.

#### Climate change scenarios

To apply our model results to general climate change scenarios we chose environmental treatments in the experiment to broadly reflect historic and future conditions at our sampling site. Our low forcing treatment approximated average spring temperature (March/April) at the site while our high temperature treatment reflects a 5 °C increase. Average field chilling (calculated from 15 October - 15 April, measured in Chill Hours, Weinberger, 1950) at Harvard Forest is 1170.7, approximately 63% of the difference between our low and high chilling treatment (Tab. S2). Thus, our low chilling treatment represents a feasible estimate for a decrease in chilling with climate change and our high chilling treatment approximates a reasonable increase (Luedeling, 2012). Our low photoperiod treatment (8 hours of daylight) is well below the photoperiod experienced at Harvard Forest, but given that the photoperiod effects are expected to be small, we chose more extreme values in order to robustly estimate an effect (i.e., increasing statistical power). For this reason, our climate change scenarios for FLS variation are based on our high photoperiod treatment alone.

242

We used our flower and budburst models to project for each species in our study:

- 1. FLSs under average environmental conditions (low forcing, 6.5 weeks of chilling treatment)
- 2. FLS shifts with spring warming only (high forcing, 6.5 weeks of chilling treatment)
- 3. FLS shifts with warming and increased chilling (high forcing, 8 weeks of chilling treatment)
- 4. FLS shifts with warming and decreased chilling (high forcing, 4 weeks of chilling treatment)

To validate our predictions, we compared our FLS interphase model estimates of "average" condition FLS interphases to long term phenological records from Harvard Forest (O'Keefe, 2015) for five species common to both datasets (Fig. S1), and found them comparable.

251

Given the variable dynamics of shifts in environmental forcing and chilling with climate change over time and space (Luedeling *et al.*, 2011), these scenarios should not be treated as absolute predictions of the magnitude of FLS shifts with climate change. Instead, we provide these scenarios to identify general trends in how FLSs could shift with warming and demonstrate that the range of possibilities vary based on characteristics of plant species and the specific climate dynamics.

257

Finally, we characterized FLS shifts for three broad FLS groups (flowering-first, concurrent, leafingfirst) by we extracting all sampling posterior estimates from our main models using the R package tidybayes (Kay, 2020), and grouping them by FLS pattern. We also include the species-specific estimates on which these grouped estimates are based in Fig. S2.

262

#### ${f Results}$

## 264 Growth chamber study

Both flower and leaf phenology advanced with higher forcing and longer chilling duration (flowers: 265 chilling effect: -21.2, forcing effect: -19.0, leaves: chilling effect: -30.4, forcing effect: -17.8; Fig. 266 2, Tab. S3, Tab. S4, all standardized units; see "Data analysis" section above), but increases in 267 both of these cues together offset these advances (flowers: force x chill effect: 7.0, leaves: force x 268 chill effect: 12.4; Fig. 2, Tab. S3, Tab. S4). Leaf and flower buds diverged in their responses 269 to increasing photoperiod, with flower phenology advancing and leaf phenology delaying when the 270 other two cues were at low levels (Fig. 2, Tab. S3, Tab. S4). As seen in the interactions between 271 photoperiod and chilling and photoperiod and forcing, increasing chilling or forcing with longer 272 photoperiod advanced the phenology of both bud types. For both bud types, chilling and forcing 273 were the dominant cues, while increasing photoperiod produced a more muted phenological response 274 (Fig. 2, Tab. S3, Tab. S4). 275

276

While leaf and flower bud phenological responses to environmental cues were qualitatively similar,
the strength of their responses to each cue differed substantially. Leaf buds responded more strongly
to chilling than flower buds (1.4x), and had a stronger response to all cue interactions (forcing x
chilling: 1.8x, photoperiod x chilling: 5.8x, photoperiod x forcing: 2.2x) (Fig. 2, Tab. S3, Tab.
S4). Across all species both bud types displayed a similar magnitude advance with increased forcing
(Fig. 2, Tab. S3, Tab. S4).

283

Our results did not showed the characteristic sensitivity pattern of the FHH across most species in

our experiment (Fig. 2, see Fig. 1,a.,b.), though there was significant variation among species in
the strength of their response to forcing between bud types. However, when we re-ran our models
on the subset of data which included phenological observations at only high levels of chilling and
photoperiod, we found the sensitivity to forcing for 9/10 species followed the predicted pattern
of the FHH, with the second phases of the FLS showing stronger sensitivity to forcing than the first
phases (Fig. 3, Tab. S5).

291

292

# Climate change scenarios

Our models project that both flower and leaf phenology will advance in our generalized future scenarios. FLS shifts depended strongly on how forcing levels change relative to chilling exposure (Fig. 4). Given the significant differences in sensitivity to chilling between flowering and leafing phenology we found in our experiment, our scenarios showed that FLS interphases will be more strongly influenced by changes in chilling duration than increased forcing alone.

298

The direction and magnitude of shifts in FLS interphases varied among species, varying especially by 299 whether or not they displayed a flowering-first FLS. Under some warming scenarios, our model pre-300 dicted that FLS interphases for some species may effectively disappear or the order of phenophases 301 in the FLS may switch (Fig. 4, Fig. S2). Several species, Acer rubrum, Ilex verticillata, Prunus 302 pensylvanica, Prunus virginiana, and Viburnum acerifolium, had FLSs that were relatively robust 303 to changing environments. For other species, Acer pensulvanicum, Vaccinium corymbosum and Ilex 304 mucronata, which typically begin to produce leaves shortly before flowers open, the magnitudes 305 of projected FLS shifts were moderate. The two species with the most significant FLS shifts in 306 both direction and magnitude across treatment combinations and climate change scenarios were 307 Comptonia peregrina and Corylus cornuta (Fig. S2). In all of our climate change scenarios, the 308 FLS interphase was dramatically reduced in these taxa. 309

# Discussion

Our experiment supports the hypothesis that flower-leaf sequences are structured by differential 312 sensitivity (DSH) to the environment between flower and leaf buds. Specifically, differences in 313 the chilling response among bud types was the strongest driver of FLS variation. We also found that under high chilling and long photoperiods, the FLSs of most species followed the predicted 315 sensitivity pattern of the FHH, supporting that the FHH is likely a special case of the DSH that 316 occurs when other phenological cues are met for both bud types. Together, these results explain 317 why the two FLS hypotheses have been difficult to distinguish, and suggest that climate change has 318 the potential to significantly disrupt FLSs as global warming alters historic chilling patterns across 319 the temperate zone (Morin et al., 2009). 320

#### Reconciling the differential sensitivity and the forcing hierarchy hypotheses

The relationship we found between the FHH and the DSH in our experiment offers a path forward for reconciling previous studies that have favored one hypothesis over the other. Support for the FHH is most often associated with observational studies (e.g. Cosmulescu & Calusaru, 2020; Guo et al., 2014). By contrast, experimental studies which manipulate chilling levels beyond historically observed minima in the field tend to support the DSH (e.g. Aslamarz et al., 2009; Gariglio et al., 2006). This pattern can be explained by the FHH being a special case of the DSH, if historic climate regimes make detection of the DSH in the field difficult.

329

330

331

332

333

335

336

Under historic climate regimes, seasonal chilling requirements were usually met (Gauzere et al., 2019; Chuine et al., 2016), making it difficult to detect any effect of chilling in observational studies, let alone differences in flower versus leaf buds. It would take larger reductions in chilling to reveal the signature of the DSH, which at present, rarely occurs in the field, but can be induced in artificial environments which explains the discrepancy between these two methods. These dynamics are consistent with findings in other phenological studies that suggest simple growing degree models (which underlie the FHH) accurately predict phenology under current climate, but under-perform under

climate change scenarios when shifts in chilling accumulation become more pronounced (Linkosalo et al., 2008; Chuine et al., 2016). Our findings suggest that as climate continues to change, differential sensitivity to the environment between flower and leaf phenology should become more apparent in field observations, and that the magnitude of interannual FLS variation is likely to extend beyond historically observed reaction norms.

342

# Population-level implications of the DSH with climate change

Predicted shifts in chilling are highly variable across both time and space—because chilling only 344 accumulates at intermediately low temperatures warming may increase chilling at some locations 345 while decreasing it in others (Man et al., 2017; Zhang et al., 2007). Layered onto these variable 346 shifts in chilling, spring warming from climate change also differs by location (Karmalkar & Bradley, 347 2017; Loarie et al., 2009), suggesting that forcing may increase more rapidly in some places than others. These shifts in both chilling and forcing suggest that the direction and magnitude of FLS 349 shifts is likely to vary substantially among populations based on the specific cue combinations at 350 a given locality (Chmielewski et al., 2012). Long-term phenology records show there was already 351 substantial intra-specific variation in FLSs at the population level (Buonaiuto et al., 2020) and our 352 findings suggest that these population-level differences may be further amplified by climate change. 353 In this way, all the three generic FLS climate change scenarios depicted in Fig. 4 should not be 354 considered alternatives to each other, but could occur contemporaneously across a species' range. 355

356

This increased population-level heterogeneity in FLSs that our models project has the potential to influence patterns of pollen dispersal across the landscape (Borycka *et al.*, 2017; Pace *et al.*, 2018). For example, advancing canopy closure relative to flowering can impede long-distance pollen transport (Milleron *et al.*, 2012), which may in turn alter patterns of gene flow across a landscape.

Our findings regarding FLS shifts should be integrated with projections of pollinator movement or prevailing wind directions (Kling & Ackerly, 2020) to better understand how FLS variation may contribute to population structure in the long term.

364

While changes in chilling and forcing from climate change may increase FLS variation, the climate 365 change implications of strong differential sensitivity to photoperiod among bud types are more 366 difficult to predict. Climate change does not directly impact photoperiod, but may alter the photoperiods plants experience as they approach their temperature optima for phenological activity. 368 However, in most temperate regions, temperature would have to drive phenological shifts by a min-369 imum of several weeks before the experienced photoperiod of plants would change substantially. 370 Because these dynamics are more extreme than most studies suggest (Thackeray et al., 2016), we 371 modeled climate change scenarios with a constant photoperiod in our FLS scenarios. Such an approach may be useful at temperate and some boreal latitudes, but may not work at high latitudes, 373 where photoperiod changes more rapidly over the season and thus temperature-driven phenological 374 shifts may significantly alter the experienced photoperiod of plants. Our results suggest that such 375 shifts could mute or amplify the FLS shifts, which may be particularly important as species shift 376 their distribution poleward with climate change and begin to encounter novel photoperiod regimes (Way & Montgomery, 2015). 378

379

## Species-level implications of the DSH with climate change

Our study highlights that the direction and magnitude of FLS shifts with climate change are speciesspecific. Not only is it likely that the function of FLS variation differs among species (Buonaiuto
et al., 2020), but we found that FLSs of some species were very sensitive to changing climate conditions, while other remain fairly resilient (Fig. 4, Fig. S2).

385

386

387

388

389

These differences suggest that FLS shifts will impact some species more than others and successful forest conservation and management programs may require identifying the species that are most vulnerable to extreme FLS shifts. To aid this effort, there is a strong need for generalizing principles to identify species with potential for consequential FLS shifts with climate change. While our one

study does not begin to represent the taxonomic diversity of a temperate forest, several patterns in the FLS responses of our multi-species experiment may serve as starting point for further inquiry.

The different FLS response patterns we observed across species may correlate with suite of interrelated anatomical, physiological and phenological traits. The species that maintained FLS structure across our climate change scenarios generally shared a strongly leafing-first FLS, with a fairly long FLS interphase (Fig 4). These species tended to have mixed buds (leaf and flower primordia together in one bud) so there may be strong physical constraints on their FLSs. By contrast, the species that were most sensitive to FLS shifts were monoecious, flowering-first, wind-pollinated shrubs (Fig 4, Fig S2). This result supports other evidence that the reproductive phenology of wind-pollinated species may be more sensitive to climate change compared to biotically pollinated taxa (Ziello et al., 2012). Given the hypothesized function of FLS in wind-pollinated species, the significant reductions in FLS interphases we observed suggest that these species, and flowering-first, wind-pollinated taxa in general, may face particular risk for reproductive performance reductions.

While much of the public interest around phenology and pollination in the context of global change has centered around trophic mismatches between pollinator and floral phenology (Memmott et al., 2007), which is of little relevance to abiotically pollinated taxa, our study identified the possibility that the effects of FLS shifts with climate change may be particularly important for wind-pollinated woody plants. The direction and magnitude of FLS shifts we observed in these taxa, coupled with the hypothesized function of a flowering-first FLS in wind-pollinated species, suggests that FLS variation in this functional group should be explored in greater detail in the future. More research is needed to identify species' traits that may correlate with the potential for FLS shifts, but flowering-first, wind-pollinated species may be particular sensitive to FLS shifts. Our results suggest that species in this functional group should be considered a research priority for the study of spring phenological sequences in deciduous woody plants.

# 117 References

- Aslamarz, A.A., Vahdati, K., Rahemi, M. & Hassani, D. (2009) Estimation of chilling and heat requirements of some persian walnut cultivars and genotypes. *HortScience* **44**, 697–701.
- Augspurger, C.K. & Zaya, D.N. (2020) Concordance of long-term shifts with climate warming varies
  among phenological events and herbaceous species. *Ecological Monographs* n/a, e01421.
- Borycka, K., Ortyl, B. & Kasprzyk, I. (2017) Temporal variations and spatial differentiation in the
- black alder and silver birch pollination pattern-the impact of local climate or something more?
- Agricultural and Forest Meteorology 247, 65–78.
- Buonaiuto, D.M., Morales-Castilla, I. & Wolkovich, E.M. (2020) Reconciling competing hypotheses
- regarding flower-leaf sequences in temperate forests for fundamental and global change biology.
- New Phytologist n/a.
- Bürkner, P.C. (2018) Advanced bayesian multilevel modeling with the r package brms. *R Journal*10, 395–411.
- Chmielewski, F.M., Blümel, K. & Pálesová, I. (2012) Climate change and shifts in dormancy release
   for deciduous fruit crops in germany. Climate Research 54, 209–219.
- 432 Chuine, I., Bonhomme, M., Legave, J.M., García de Cortázar-Atauri, I., Charrier, G., Lacointe, A.
- & Améglio, T. (2016) Can phenological models predict tree phenology accurately in the future?
- the unrevealed hurdle of endodormancy break. Global Change Biology 22, 3444–3460.
- Citadin, I., Raseira, M.C.B., Herter, F.G. & da Silva, J.B. (2001) Heat requirement for blooming
  and leafing in peach. *HortScience HortSci* **36**, 305–307.
- Cleland, E.E., Chuine, I., Menzel, A., Mooney, H.A. & Schwartz, M.D. (2007) Shifting plant phenology in response to global change. *Trends in Ecology Evolution* **22**, 357 365.
- Cosmulescu, S. & Calusaru, F.G. (2020) Influence of temperature on blackthorn (¡i¿prunus spinosa¡/i¿ l.) phenophases in spring season. advpub, –.

- Cosmulescu, S. & Ionescu, M. (2018) Phenological calendar in some walnut genotypes grown in 441 romania and its correlations with air temperature. International Journal of Biometeorology 62. 442
- Di Cecco, G.J. & Gouhier, T.C. (2018) Increased spatial and temporal autocorrelation of tempera-443 ture under climate change. Scientific Reports 8, 14850.

- Ettinger, A., Gee, S. & Wolkovich, E. (2018) Phenological sequences: how early season events define 445 those that follow. American Journal of Botany 105. 446
- Finn, G.A., Straszewski, A.E. & Peterson, V. (2007) A general growth stage key for describing trees and woody plants. Annals of Applied Biology 151, 127–131. 448
- Flynn, D.F.B. & Wolkovich, E.M. (2018) Temperature and photoperiod drive spring phenology 449 across all species in a temperate forest community. New Phytologist 219, 1353–1362. 450
- Forrest, J. & Miller-Rushing, A.J. (2010) Toward a synthetic understanding of the role of phenology 451 in ecology and evolution. Philosophical Transactions of the Royal Society B: Biological Sciences 452 **365**, 3101–3112. 453
- Franklin, D.C. (2016) Flowering while leafess in the seasonal tropics need not be cued by leaf drop: 454 evidence from the woody genus brachychiton (malvaceae). Plant Ecology and Evolution 149, 455 272 - 279.456
- Gariglio, N., González Rossia, D.E., Mendow, M., Reig, C. & Agusti, M. (2006) Effect of artificial 457 chilling on the depth of endodormancy and vegetative and flower budbreak of peach and nectarine 458 cultivars using excised shoots. Scientia Horticulturae 108, 371–377. 459
- Gauzere, J., Lucas, C., Ronce, O., Davi, H. & Chuine, I. (2019) Sensitivity analysis of tree phenol-460 ogy models reveals increasing sensitivity of their predictions to winter chilling temperature and 461 photoperiod with warming climate. Ecological Modelling 411, 108805. 462
- Gellman, A. & Hill, J. (2007) Data Analysis Using Regression and Multilevel/Hierarchical Models. 463 Cambridge University Press. 464

- Gelman, A., Carlin, J., Stern, H., Dunson, D., Vehtari, A. & Rubin, D. (2013) Bayesian Data
   Analysis. Chapman and Hall/CRC.
- Gougherty, A.V. & Gougherty, S.W. (2018) Sequence of flower and leaf emergence in deciduous trees is linked to ecological traits, phylogenetics, and climate. *New Phytologist* **220**, 121–131.
- Guo, L., Luedeling, E., Dai, J. & Xu, J. (2014) Differences in heat requirements of flower and leaf
- buds make hysteranthous trees bloom before leaf unfolding. Plant Diversity and Resources 36,
- 471 245-253.
- Jackson, S.T. & Lyford, M.E. (1999) Pollen dispersal models in quaternary plant ecology: Assumptions, parameters, and prescriptions **65**, 39–75.
- Janzen, D.H. (1967) Synchronization of sexual reproduction of trees within the dry season in central america. Evolution 21, 620–637.
- Karmalkar, A.V. & Bradley, R.S. (2017) Consequences of global warming of 1.5 °c and 2 °c for regional temperature and precipitation changes in the contiguous united states. *PLOS ONE* **12**, e0168697—.
- 479 Kay, M. (2020) tidybayes: Tidy Data and Geoms for Bayesian Models. R package version 2.1.1.
- Kling, M.M. & Ackerly, D.D. (2020) Global wind patterns and the vulnerability of wind-dispersed species to climate change. *Nature Climate Change* **10**, 868–875.
- Linkosalo, T., Lappalainen, H.K. & Hari, P. (2008) A comparison of phenological models of leaf
- bud burst and flowering of boreal trees using independent observations. Tree Physiology 28,
- 484 1873–1882.
- Loarie, S.R., Duffy, P.B., Hamilton, H., Asner, G.P., Field, C.B. & Ackerly, D.D. (2009) The velocity of climate change. *Nature* **462**, 1052–1055.
- Luedeling, E. (2012) Climate change impacts on winter chill for temperate fruit and nut production:
- A review. Scientia Horticulturae 144, 218–229.

- Luedeling, E., Girvetz, E.H., Semenov, M.A. & Brown, P.H. (2011) Climate change affects winter chill for temperate fruit and nut trees. *PLOS ONE* **6**, e20155—.
- Man, R., Lu, P. & Dang, Q.L. (2017) Insufficient chilling effects vary among boreal tree species and chilling duration. Frontiers in Plant Science 8, 1354.
- Mehlenbacher, S.A. (1991) Chilling requirements of hazelnut cultivars. *Scientia Horticulturae* 47, 271–282.
- Memmott, J., Craze, P.G., Waser, N.M. & Price, M.V. (2007) Global warming and the disruption of plant–pollinator interactions. *Ecology Letters* **10**, 710–717.
- Menzel, A., Sparks, T.H., Estrella, N., Koch, E., Aasa, A., Ahas, R., Alm-Kuebler, K., Bissolli, P.,
- Braslavska, O., Briede, A., Chmielewski, F.M., Crepinsek, Z., Curnel, Y., Dahl, A., Defila, C.,
- Donnelly, A., Filella, Y., Jatcza, K., Mage, F., Mestre, A., Nordli, O., Penuelas, J., Pirinen, P.,
- Remisova, V., Scheifinger, H., Striz, M., Susnik, A., Van Vliet, A.J.H., Wielgolaski, F.E., Zach,
- S. & Zust, A. (2006) European phenological response to climate change matches the warming
- pattern. Global Change Biology 12, 1969–1976.
- Milleron, M., Lopez de Heredia, U., Lorenzo, Z., Perea, R., Dounavi, A., Alonso, J., Gil, L. &
- Nanos, N. (2012) Effect of canopy closure on pollen dispersal in a wind-pollinated species (Fagus
- sylvatica L.). *Plant Ecology* **213**, 1715–1728.
- Morin, X., Lechowicz, M.J., Augspurger, C.K., O'Keefe, J., Viner, D. & Chuine, I. (2009) Leaf
- phenology in 22 north american tree species during the 21st century. Global Change Biology 15,
- <sub>508</sub> 961–975.
- Niklas, K.J. (1985) The aerodynamics of wind pollination. The Botanical Review 51, 328–386.
- O'Keefe, J. (2015) Phenology of Woody Species at Harvard Forest since 1990. Harvard Forest Data
- Archive: HF003., Petersham, MA, USA.
- Pace, L., Boccacci, L., Casilli, M., Di Carlo, P. & Fattorini, S. (2018) Correlations between weather
- conditions and airborne pollen concentration and diversity in a mediterranean high-altitude site
- disclose unexpected temporal patterns. Aerobiologia **34**, 75–87.

- Parmesan, C. & Yohe, G. (2003) A globally coherent fingerprint of climate change impacts across natural systems. *Nature* **421**, 37 EP –.
- Post, E.S., Pedersen, C., Wilmers, C.C. & Forchhammer, M.C. (2008) Phenological sequences reveal aggregate life history response to climatic warming 89, 363–370.
- Rathcke, B. & Lacey, E.P. (1985) Phenological patterns of terrestrial plants. *Annual Review of Ecology and Systematics* **16**, 179–214.
- Savage, J.A. (2019) A temporal shift in resource allocation facilitates flowering before leaf out and spring vessel maturation in precocious species. *American Journal of Botany* **106**, 113–122.
- Thackeray, S.J., Henrys, P.A., Hemming, D., Bell, J.R., Botham, M.S., Burthe, S., Helaouet, P.,
- Johns, D.G., Jones, I.D., Leech, D.I., Mackay, E.B., Massimino, D., Atkinson, S., Bacon, P.J.,
- Brereton, T.M., Carvalho, L., Clutton-Brock, T.H., Duck, C., Edwards, M., Elliott, J.M., Hall,
- S.J.G., Harrington, R., Pearce-Higgins, J.W., Høye, T.T., Kruuk, L.E.B., Pemberton, J.M.,
- Sparks, T.H., Thompson, P.M., White, I., Winfield, I.J. & Wanless, S. (2016) Phenological sen-
- sitivity to climate across taxa and trophic levels. Nature 535, 241–245.
- Way, D.A. & Montgomery, R.A. (2015) Photoperiod constraints on tree phenology, performance and migration in a warming world. *Plant, Cell & Environment* 38, 1725–1736.
- Weinberger, J.H. (1950) Chilling requirements of peach varieties. id 19511602967. Proceedings.

  American Society for Horticultural Science 56, 122–28.
- Whitehead, D.R. (1969) Wind pollination in the angiosperms: Evolutionary and environmental considerations. *Evolution* **23**, 28–35.
- Zhang, X., Tarpley, D. & Sullivan, J.T. (2007) Diverse responses of vegetation phenology to a warming climate. *Geophysical Research Letters* **34**.
- Ziello, C., Böck, A., Estrella, N., Ankerst, D. & Menzel, A. (2012) First flowering of wind-pollinated
   species with the greatest phenological advances in europe. *Ecography* 35, 1017–1023.

# Figures 539

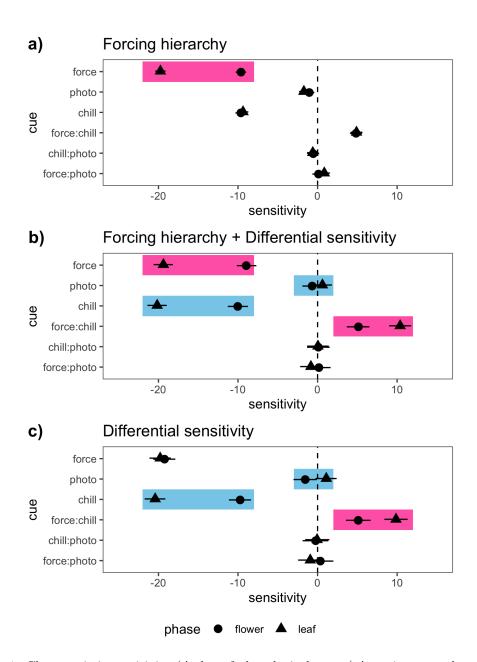


Figure 1: Characteristic sensitivity ( $\Delta$  day of phenological event/  $\Delta$  environmental cue) patterns of the phenological response to changing cue levels for the two major flower-leaf sequence hypotheses. a) displays a signature pattern of the forcing hierarchy hypothesis (FHH, pink boxes)—with the second phenophase in the sequence (in this case leafing) having a higher sensitivity to forcing than the first. b) depicts a scenario where both the FHH and the differential sensitivity hypothesis (DSH) contribute to flower-leaf sequence variation. Here the characteristic forcing sensitivity of the FHH is still apparent but the differential sensitivity to chilling and photoperiod is seen as well (blue boxes). c) highlights a typical sensitivity pattern produced by the DSH without the FHH. All plots above are based on simulations (see Supporting Information: Methods). Shapes indicate mean estimates and lines depict 95% credible intervals from Bayesian hierarchical models with advances in phenology shown as negative numbers, and delays in phenology as positive numbers.

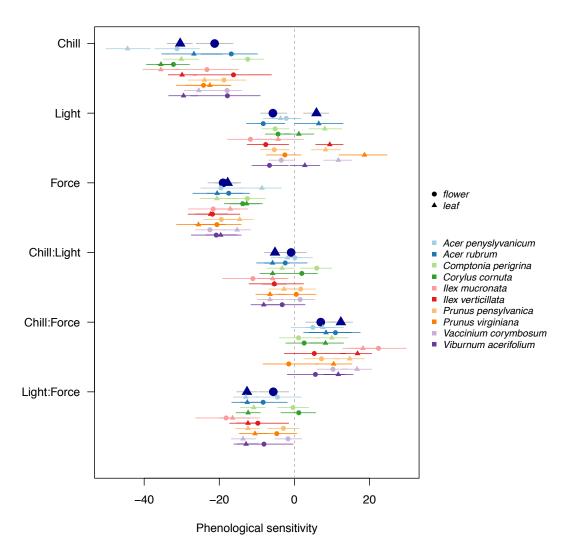


Figure 2: Effects of forcing temperature, chilling duration, and photoperiod on the leaf (circles) and flower (triangles) phenology of 10 temperate woody plant species collected from Harvard Forest (Petersham, MA, USA). Shapes indicate mean estimates and lines depict 50% credible intervals (See Tab. S3, Tab. S4 for other intervals) from Bayesian hierarchical models with advances in phenology shown as negative numbers, and delays in phenology as positive numbers. Flower and leaf phenology differs in sensitivity ( $\Delta$  day of phenological event/ $\Delta$  environmental cue) to these environmental cues.

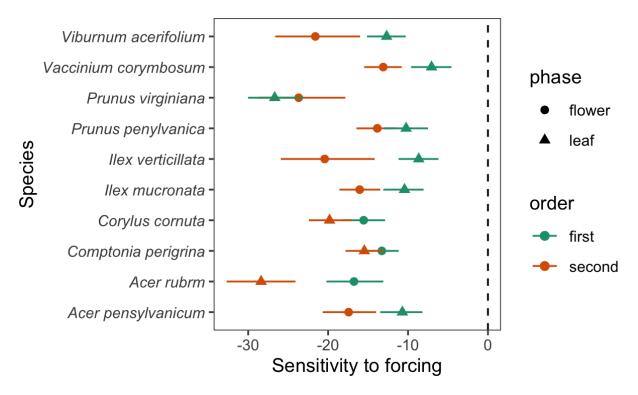


Figure 3: Phenological sensitivity ( $\Delta$  phenological event/  $\Delta$  C°) to forcing temperatures of leaf (circles) and flower (triangles) phenology from 10 temperate deciduous woody plants at long (12 hour) photoperiod and long chilling duration treatments (8 weeks at 4°C). Shapes indicate mean estimates and lines depict 50% credible intervals (See Tab. S5 for other intervals) from Bayesian hierarchical models with advances in phenology shown as negative numbers. When photoperiod and chilling are high, most species follows the predicted pattern of the forcing hierarchy hypothesis (FHH), with the second phenophase of the sequence consistently more sensitive to forcing than the first. This result suggests that the FHH should be considered a special case of the differential sensitivity hypothesis (DSH) that occurs when the chilling and photoperiod requirements are met for both tissue types.

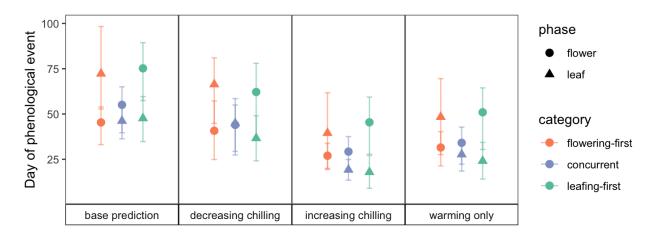


Figure 4: Projected shifts in flower-leaf sequences under current environmental conditions (base prediction) and three climate change scenarios predict that FLS differ among the three major FLS types and will be strongest is flowering-first species. Predictions are based on species-level posterior estimates grouped by FLS category (flowering-first, concurrent, leafing-first) from Bayesian hierarchical models comparing flower (circles) and leaf (triangles) phenological responses to variable chilling duration and forcing temperatures. Shapes represent the mean estimates and lines represent the 50% credible intervals.