

# Experimental designs for testing the interactive effects of temperature and light in ecology and the problem of periodicity

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## 1 Introduction

Across the tree of life, temperature and light availability shape a number of important biological processes including growth and metabolic rates (MacLean & Gilchrist, 2019) sex determination (Brown *et al.*, 2014), acclimatization to seasonal environments (Hamilton *et al.*, 2016) and the timing of life cycle transitions (phenology) (Forrest & Miller-Rushing, 2010). These biological responses in turn dictate broad scale ecological processes and patterns ranging from biogeochemical cycling (Piao *et al.*, 2007) to species range limits (Chaine & Beaubien, 2001). Characterizing the specific dynamics of how these environmental factors synergistically affect biological processes across a wide range of taxa has become even more important as anthropogenic global change continues to expose organisms to novel environmental conditions (Pörtner & Farrell, 2008).

Because temperature and light availability often co-vary in the field (for example, in most temperate ecosystems, daylength and temperature both increase as the season progresses (Rosenberg, 1974)) it can be difficult to disentangle their relative contributions to biological processes. Experimental manipulations of climate variables in artificial environments are ideal for mechanistically characterizing biological responses to environmental fluctuations (Ettinger *et al.*, 2020; Primack *et al.*, 2015). Growth chambers of all shapes and sizes have been used to this end (Downs, 1980) and these efforts have greatly advanced researchers understanding of the fundamental biology of a wide variety of organisms and their ability to predict the responses to current and future climate change. (Stewart *et al.*, 2013).

However, controlled environment experiments have their own challenges. Experimentalists must balance biological realism with statistical inference, experimental effort with statistical power, and account for the effects of unmanipulated or unmeasured variables (Scheiner & Gurevitch, 2001). Because biological responses to the environment are generally the product of complex interactions between multiple environmental signals (Casal, 2002), seemingly small choices about experimental design can generate significant differences in outcomes. Experimental treatments are rarely standardized among researchers even within disciplines (Wolkovich *et al.*, in Review.) and these

complexities may in part contribute to many discrepancies between experimental studies and observation data (Poorter *et al.*, 2016).

Even with these limitations, growth chamber studies remain a powerful tool for mechanistically assessing organismic responses to the environment provided that the implications of treatment designs are well understood and well matched with the scope of the research question. Below we highlight a particular challenge that can arise in experiments seeking to estimate the interactive effects of temperature and light on biological outcomes when the periodicity of both variables is included. Our example deals with the phenology (timing of recurring life cycle events, e.g. leaf budburst, flowering) in temperate woody plants. However, the issues and solutions we present below are broadly applicable to studies of any other organisms and biological processes that utilize temperature and daylength signals.

We begin by briefly detailing how temperature and light treatments are generally applied in experimental phenology studies and reviewing the minimum experimental elements required to robustly test interactions between two or more environmental variables. We then detail the problem of inference that can arise manipulating the periodicity of both temperature and light in experiments and demonstrate the extent to which this is an issue with a mathematical and experimental example. Finally, we conclude by outlining several possible solutions for overcoming these issues.

## Estimating phenological cues from experiments

Decades of experimental work in growth chambers have demonstrated that temperature (both cool temperatures in fall/winter and warming temperatures in spring) and photoperiod are the primary cues of phenology for plants in the temperate/boreal zones (Ettinger *et al.*, 2020). While exposure to cool winter temperatures (chilling) strongly impacts phenology (Laube *et al.*, 2014), our discussion will focus primarily warm temperature treatments, and light treatments, as chilling levels are usually applied in a pre-treatment either by sequentially removing plant materials from the field throughout the winter dormancy period (Wolkovich *et al.*, in Review.) or applying experimental chilling in a cold chamber before warm temperature and light treatments are applied in tandem (Weinberger, 1950; Flynn & Wolkovich, 2018). It choices that about how to apply these two treatments in particular that can compromise inference on their effects, so we will focus of these two cues.

While a large variety of experimental designs have been used to study plant phenology, generally phenology experiments tend to manipulated two major axes of light and warm temperature variation:

1. Intensity: The amount or quality of a variable. Here we define temperature intensity as the amount of heat present in the system (measured in degrees). In the phenology literature this measurement is generally referred to as forcing. We define light intensity as the luminosity or irradiance present in the system (measured in lumens or watts).

2. Periodicity: The interval at which the intensity of the variable is applied. Hereafter, we refer to the periodicity of light as photoperiod (often used synonymously with “daylength”) and the periodicity of temperature as thermoperiod.

For phenology, it is generally accepted that photoperiodicity is the primary light cue to which plants respond ((Way & Montgomery, 2015) though see (Brelsford & Robson, 2018; Cober *et al.*, 1996) regarding light intensity and phenology). For temperature, conventionally both intensity and periodicity drive phenological activity and several metrics, (e.g. growing degree hours, thermal sums, growing degree days) that combine these two axes have been developed (Gu, 2016). This assumption is well supported; under natural conditions diurnal temperature fluctuations in temperate regions can be quite large in the spring, and studies have found that diurnal temperature variation strongly influences plant phenology (Burghardt *et al.*, 2016). In fact, even if thermoperiodicity is not an explicit treatment variable (i.e. manipulated systematically), incorporating it in experiments is essential for translating experimental results into real world predictions (Chiang *et al.*, 2020).

Like many other biological processes, recent advances have demonstrated that plant phenological responses are nonlinear, due largely to interactions between cues (Wolkovich *et al.*, in Review.; Fu *et al.*, 2015), highlighting the need for experiments to be designed to evaluate the strength of these interactions. In order to have the statistical power to partition the individual and interactive effects of two or more variables in an experiment, one must:

1. Have at minimum of two treatment levels of at least two variables.
2. Treatment levels must be full factorial (Fig. 1a.). Full factorial designs are both balanced (Fig. 1b.) and orthogonal (Fig. 1c.); which is to say that all possible treatment combinations are applied and each treatment is independent of all others (Cheng, 2016).

These two critical elements may seem obvious but can be conspicuously absent from many published studies. In the case of woody plant phenology, using a recently published database (OSPREE: Observed spring phenological responses in experimental environments (Wolkovich *et al.*, 2019)) we found that out of 152 controlled environment experiments (across 93 studies) only 64 of them manipulated both light and forcing cues in the same experiment and only 15 of those did so with a design that was both balanced and orthogonal (see Supplement for details). This notable dearth of robust tests of light and temperature interactions may stem from the common limitations of time, space, and resources that experimentalists often face, but it may equally relate to a fundamental issues that arises from the fact that these variables themselves are comprised of multiple axes of variability.

## The problem of periodicity

A common approach in phenology experiments that seems to balance prior knowledge about the underlying physiology of phenology, biological realism and experimental inferences is to vary pho-

toperiodicity, and thermal intensity and periodicity

Yet the orthogonality of this design is based on the assumption to a 12 hour thermoperiod. If, rather the thermoperiod is coupled with the photoperiod, this is not the case because the daily mean temperature of the long/high treatment will be higher than that of the short/high treatment, and the long/low treatment slightly warmer than the short low (Fig 2.a). This is because the warmer day time temperatures are applied for different duration across the high temperature treatments. While this covariation among the photoperiod and temperature treatments is biologically realistic, it make it statistically impossible to differentiate their independent and interactive effects on any given biological process.

**Thing to think on: covariation or periodicity is a problem in studies that don't manipulate forcing e.g. Zohner paper. If you vary thermoperiod in step with photoperiod, your long photoperiod treatment gets more GDDs. I need to make this point somewhere, ideally, without restructuring the whole paper. But as I am saying this, I am realizing this problem might be less about interactions and more just about evaluating photoperiod effects in general.**

Of the studies in the OSPREE database that manipulated both photoperiod and temperature interactively, we found that around 45% of them may have this issue, suggesting that the true interactive effects of these cues on spring phenology is still quite poorly characterized. This may be in part why the relative contribution of temperature and photoperiod cues to spring phenology remains a contentious debate in the phenology literature (Koerner & Basler, 2010).

## Periodicity and inference

Forcing is the more dominant cue than photoperiod for spring phenology (CHUINE *et al.*, 2010), and therefore we would expect the covariation between photo- and thermo-period may result in an over-estimation of the photoperiod effect. This prediction is supported mathematically by transposing the treatment levels and phenological responses into 3-dimensional geometric planes and solving for the differences. (Fig. 3, Supporting info)(*Maybe Megan or Lizzie could help me here say this better elaborate on this?*)

While we are aware of no experiments that explicitly compare the effects of co-varying vs. independent photo- and thermo- periods, we identified two phenology experiments in the literature which utilized many overlapping treatment levels and species from the same sampling sites however in one study, Flynn & Wolkovich (2018), photo- and thermo- period co-vary, while in the other Buonaiuto & Wolkovich (2021) photo- and thermo- period were varied independently. Comparing the cue estimates from these two studies offers an opportunity to test the theoretical and mathematical predictions, and further understanding the uncertainty in cue estimate due to periodicity.

We subset each dataset (publicly available at the Harvard Forest Data Archive and KNB) to in-

clude only the species shared among the two studies, and re-analyzed the data using Bayesian hierarchical models to compare difference in the photoperiod and forcing estimates (see Supplement for Methods). We found that the estimated differences in the mean response to photoperiod and forcing among study designs were on the same order our mathematical predictions, and that the un-coupled design estimated a weaker (less negative) photoperiod effect, and stronger forcing effects than the coupled experimental design (4,??).

It is worth noting that there are almost certainly other factors driving the differences between these experiments. Both were conducted in different years, sampled different individual from the population, and used different methods for applying an chilling pre-treatments (Flynn & Wolkovich, 2018; Buonaiuto & Wolkovich, 2021). However, because this comparison is well matched to our mathematical predictions and prior knowledge about how temperature and photoperiod are expected to interacting in phenology, we argue that the influence of periodicity covariation on statistical inference is apparent enough to take seriously .

## 2 Paths Forward

In the sections above we have systematically demonstrated that experiments which co-vary thermoperiod and photoperiod cannot robustly differentiate the individual effect of temperature and photoperiod on a spring phenology (or any other biological process) or quantify their interactive influence. Given the paucity of interactive studies in the literature, it is clear that more well designed studies will be needed to better characterize the effects of these cues. Below we offer several generalized experiment designs that improve statistical orthogonality of controlled environment experiments which could be further developed and adjusted to fit the needs of experimentalists across many sub-fields of biology.

1. **Covarying photo- thermo- period with quantified uncertainty.** It may be that the experimental design that best balances environmental realism, statistical inferences and translatability to observational studies are designs that co-vary periodicity to mimic natural systems (2a.). Moving forward, researchers using this design need to be aware of the non-orthogonality of this design, and be sure to present the uncertainty surrounding their cue effect estimate, which could be done using a similar mathematical approach to the one we present in this paper (see Supplement).
2. **Manipulate photoperiod and temperature intensity with no thermoperiodicity.** This approach allows for the maintenance of statistical orthogonality across treatment combinations (2b.). The main drawback is that this design sacrifices the biological realism of diurnal temperature variation, which may make it more difficult to translate estimates from experiments to real world applications.
3. **Compensatory diurnal temperature fluctuations.** There are almost unlimited pairs of in-

ters that can reduce to the same mean (e.g.  $(24 + 26)/2 = (30 + 20)/2 = 25$ ) and the non-orthogonality of the mean daily temperature that arises in a coupled photo-thermoperiod design could be corrected for by proportionately increasing the diurnal temperature fluctuation of the short photoperiod treatment relative to the long treatments (2c.). However, if the differences between day and night temperature has a meaningful biological effect, this introduces another confounding, non-orthogonal factor for interpreting temperature and photoperiod effects. For example, the influence of day time warming of phenology can be as much as three times stronger than proportionate night time warming (Rossi & Isabel, 2017; Meng *et al.*, 2020).

4. **Uncouple thermoperiod and photoperiod.** By varying thermoperiod and photoperiod independently (2d.), statistical orthogonality can be maintained across treatment. However, this approach may also introduce new artifacts that occur from the biological rather than statistical interactions between light and temperature. For example, there is evidence that increasing temperatures in the first two hours of daylight can be almost as effective for stimulating shoot elongation as similar temperature increases for the whole photoperiod (Erwin, 1998). With this design, treatments must inherently differ in the amount of time the warmer daytime temperature extends into the dark nighttime light regime, introducing a new axes of non-orthogonality.

In correcting one problem, each one of these designs introduces another, which may in fact be an intrinsic property of any experimental manipulation. It would certainly be useful for researchers to explicitly test how cue estimates vary among these experimental designs, and which design is most useful for predicting phenology in field under current and future climate conditions. In the meantime, we hope the treatment reminds experimentalists that we must continue to be thoughtful about matching design to the goals of a study, and being transparent about uncertainty around our experimental inferences.

## References

- Brelsford, C.C. & Robson, T.M. (2018) Blue light advances bud burst in branches of three deciduous tree species under short-day conditions. *TREES-STRUCTURE AND FUNCTION* **32**, 1157–1164.
- Brown, E.E., Baumann, H. & Conover, D.O. (2014) Temperature and photoperiod effects on sex determination in a fish. *Journal of Experimental Marine Biology and Ecology* **461**, 39–43.
- Buonaiuto, D.M. & Wolkovich, E.M. (2021) Differences between flower and leaf phenological responses to environmental variation drive shifts in spring phenological sequences of temperate woody plants. *Journal of Ecology* **109**, 2922–2933.
- Burghardt, L.T., Runcie, D.E., Wilczek, A.M., Cooper, M.D., Roe, J.L., Welch, S.M. & Schmitt, J. (2016) Fluctuating, warm temperatures decrease the effect of a key floral repressor on flowering time in *Arabidopsis thaliana*. *New Phytologist* **210**, 564–576.

- 216 Casal, J.J. (2002) Environmental cues affecting development. *Current Opinion in Plant Biology* **5**,  
217 37–42.
- 218 Cheng, C.S. (2016) *Theory of Factorial Design*. Chapman and Hall/CRC.
- 219 Chiang, C., Bånkestad, D. & Hoch, G. (2020) Reaching natural growth: The significance of light  
220 and temperature fluctuations in plant performance in indoor growth facilities. *Plants* **9**.
- 221 Chuine, I. & Beaubien, E. (2001) Phenology is a major determinant of tree species range. *Ecology*  
222 *Letters* **4**, 500–510.
- 223 CHUINE, I., MORIN, X., BUGMANN, H., KÖRNER, C. & BASLER, D. (2010) *Strong* warming,  
224 photoperiods, and tree phenology *Strong* [with response]. *Science* **329**, 277–278.
- 225 Cober, E., Tanner, J. & Voldeng, H. (1996) Soybean photoperiod-sensitivity loci respond differen-  
226 tially to light quality. *CROP SCIENCE* **36**, 606–610.
- 227 Downs, R.J. (1980) Phytotrons. *Botanical Review* **46**, 447–489.
- 228 Erwin, J. (1998) Temperature and light effects on stem elongation. *JOURNAL OF THE JAPANESE*  
229 *SOCIETY FOR HORTICULTURAL SCIENCE* **67**, 1113–1120, Session of Plant Growth Regu-  
230 lation by Physical and Mechanical Stimuli at the Commemorative Symposium of the 75th An-  
231 niversary of Japanese-Society-for-Horticultural-Science, JAPAN, APR 03-04, 1998.
- 232 Ettinger, A.K., Chamberlain, C.J., Morales-Castilla, I., Buonaiuto, D.M., Flynn, D.F.B., Savas, T.,  
233 Samaha, J.A. & Wolkovich, E.M. (2020) Winter temperatures predominate in spring phenological  
234 responses to warming. *Nature Climate Change* **10**, 1137–1142.
- 235 Flynn, D.F.B. & Wolkovich, E.M. (2018) Temperature and photoperiod drive spring phenology  
236 across all species in a temperate forest community. *New Phytologist* **219**, 1353–1362.
- 237 Forrest, J. & Miller-Rushing, A.J. (2010) Toward a synthetic understanding of the role of phenology  
238 in ecology and evolution. *Philosophical Transactions of the Royal Society of London B: Biological*  
239 *Sciences* **365**, 3101–3112.
- 240 Fu, Y.S.H., Zhao, H.F., Piao, S.L., Peaucelle, M., Peng, S.S., Zhou, G.Y., Ciais, P., Huang, M.T.,  
241 Menzel, A., Uelas, J.P., Song, Y., Vitasse, Y., Zeng, Z.Z. & Janssens, I.A. (2015) Declining global  
242 warming effects on the phenology of spring leaf unfolding. *Nature* **526**, 104–107.
- 243 Gu, S. (2016) Growing degree hours - a simple, accurate, and precise protocol to approximate  
244 growing heat summation for grapevines. *International Journal of Biometeorology* **60**, 1123–1134.
- 245 Hamilton, J.A., El Kayal, W., Hart, A.T., Runcie, D.E., Arango-Velez, A. & Cooke, J.E. (2016)  
246 The joint influence of photoperiod and temperature during growth cessation and development of  
247 dormancy in white spruce (*Picea glauca*). *Tree Physiology* **36**, 1432–1448.
- 248 Koerner, C. & Basler, D. (2010) Phenology under global warming. *Science* **327**, 1461–1462.

- Laube, J., Sparks, T.H., Estrella, N., Höfler, J., Ankerst, D.P. & Menzel, A. (2014) Chilling out-weighs photoperiod in preventing precocious spring development. *Global Change Biology* **20**, 170–182.
- MacLean, H.J. & Gilchrist, G.W. (2019) Temperature, photoperiod and life history traits in *Drosophila subobscura*. *bioRxiv* p. 717967.
- Meng, L., Zhou, Y., Li, X., Asrar, G.R., Mao, J., Wanamaker, A.D. & Wang, Y. (2020) Divergent responses of spring phenology to daytime and nighttime warming. *Agricultural and Forest Meteorology* **281**, 107832.
- Piao, S., Friedlingstein, P., Ciais, P., Viovy, N. & Demarty, J. (2007) Growing season extension and its impact on terrestrial carbon cycle in the northern hemisphere over the past 2 decades. *Global Biogeochemical Cycles* **21**.
- Poorter, H., Fiorani, F., Pieruschka, R., Wojciechowski, T., van der Putten, W.H., Kleyer, M., Schurr, U. & Postma, J. (2016) Pampered inside, pestered outside? differences and similarities between plants growing in controlled conditions and in the field. *New Phytologist* **212**, 838–855.
- Pörtner, H.O. & Farrell, A.P. (2008) Physiology and climate change. *Science* **322**, 690–692.
- Primack, R.B., Laube, J., Gallinat, A.S. & Menzel, A. (2015) From observations to experiments in phenology research: investigating climate change impacts on trees and shrubs using dormant twigs. *ANNALS OF BOTANY* **116**, 889–897.
- Rosenberg, N.J. (1974) *Microclimate: the biological environment*. Wiley, New York.
- Rossi, S. & Isabel, N. (2017) Bud break responds more strongly to daytime than night-time temperature under asymmetric experimental warming. *Global Change Biology* **23**, 446–454.
- Scheiner, S.M. & Gurevitch, J. (2001) *Design and analysis of ecological experiments*. Oxford University Press.
- Stewart, R.I.A., Dossena, M., Bohan, D.A., Jeppesen, E., Kordas, R.L., Ledger, M.E., Meerhoff, M., Moss, B., Mulder, C., Shurin, J.B., Suttle, B., Thompson, R., Trimmer, M. & Woodward, G. (2013) *Chapter Two - Mesocosm Experiments as a Tool for Ecological Climate-Change Research*, vol. 48, pp. 71–181. Academic Press.
- Way, D.A. & Montgomery, R.A. (2015) Photoperiod constraints on tree phenology, performance and migration in a warming world. *Plant, Cell & Environment* **38**, 1725–1736.
- Weinberger, J.H. (1950) Chilling requirements of peach varieties. id - 19511602967. *Proceedings. American Society for Horticultural Science* **56**, 122–28.
- Wolkovich, E., Chamberlin, C., Buonaiuto, D., Ettinger, A. & Morales-Castilla, I. (in Review.) Integrating experiments to predict interactive cue effects on spring phenology with warming .



282 Wolkovich, E.M., Ettinger, A.K., Flynn, D., Savas, T., Chamberlain, C., Buonaiuto, D. &  
283 Samaha, J. (2019) Observed spring phenology responses in experimental environments (OS-  
284 PREE). doi:10.5063/F1CZ35KB.

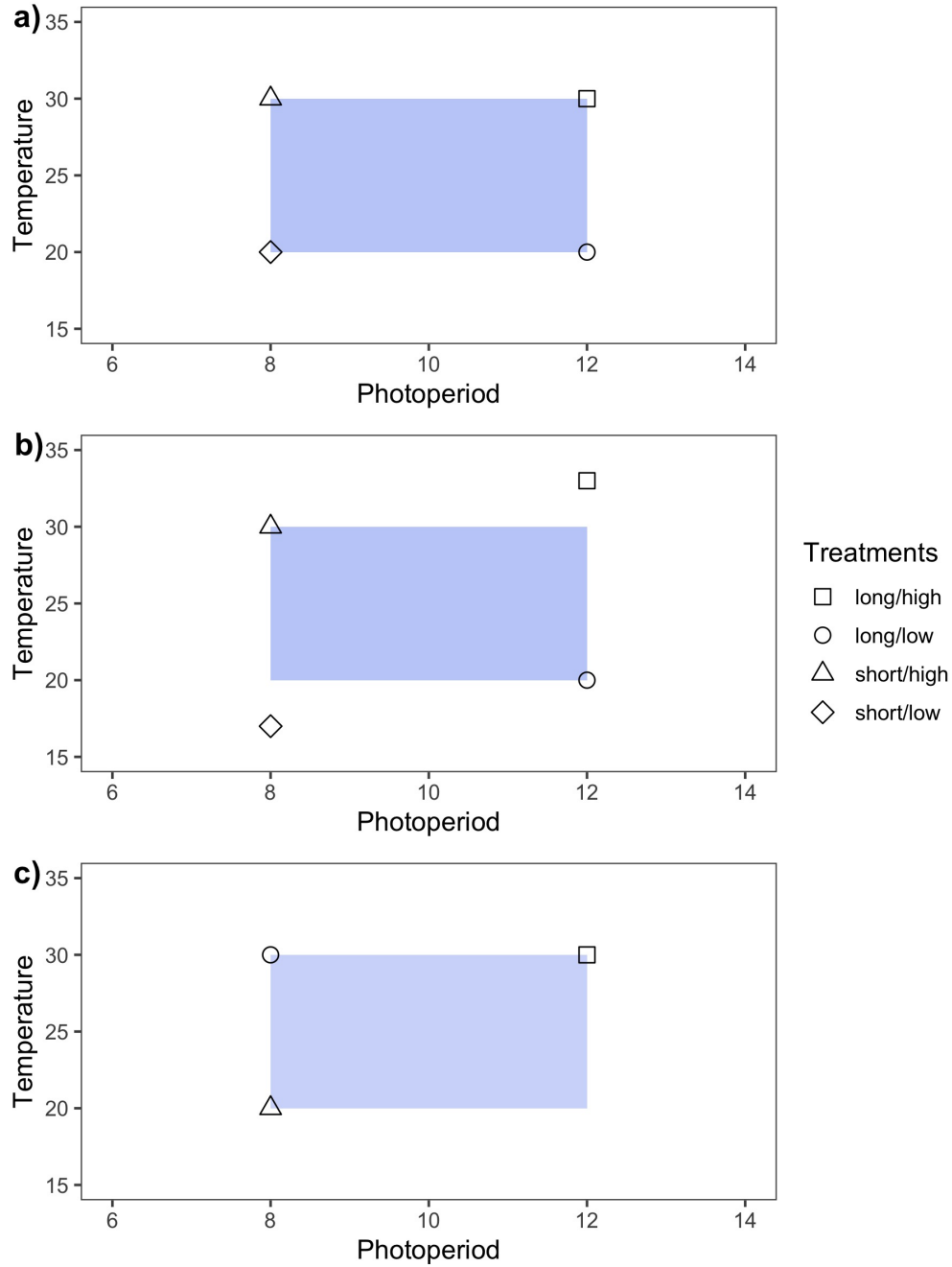


Figure 1: Idealized experimental designs demonstrate three approaches for varying temperature and light treatment level in controlled environment experiments. Design **a)** is fully factorial in that treatments levels are balanced and orthogonal. This design is appropriate for testing interactions between two or more variables. In **b)** the design is balanced both not orthogonal. Non-orthogonality in experiments often arises in experiments when there is covariation among the test variables is unaccounted for. In **c)**, the experimental design is orthogonal but unbalanced. Lack of balance in experiments often arises due to time, space or resource limitations.

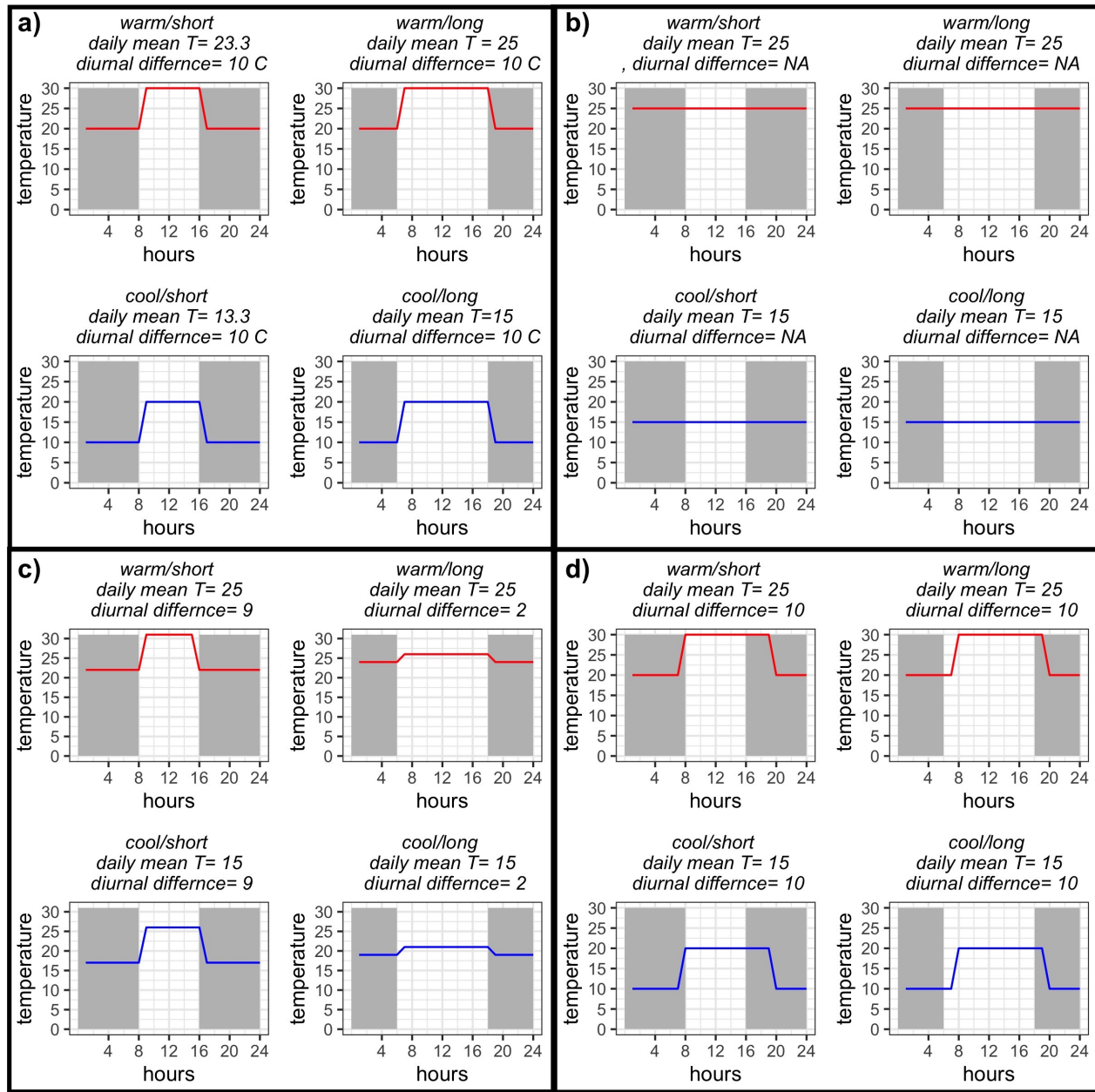


Figure 2: Conceptualized experimental designs to test temperature and daylength interactions on a biological response. In **a)** the design incorporates a standardize diurnal temperature fluctuation across all treatment. Because this thermoperiod is coupled with the photoperiod, while the same day and night temperatures are applied for the high and low temperature treatments respectively, the mean daily temperatures differ across each photoperiod treatment generating non-orthogonality. Designs **b)**, **c)** and **d)** are all designs that can correct this non-orthogonality. Design **b)** manipulated temperature intensity only (no thermoperiodicity). In **c)** photo- and thermo- periods are still are coupled but the orthogonality of mean daily temperature is maintained by proportionately varying the diurnal temperature fluctuations across treatments. In design **d)** standard diurnal temperature fluctuations are maintained but, thermoperiod and photoperiod are decoupled and varied independently, maintaining orthogonality daily mean temperatures.

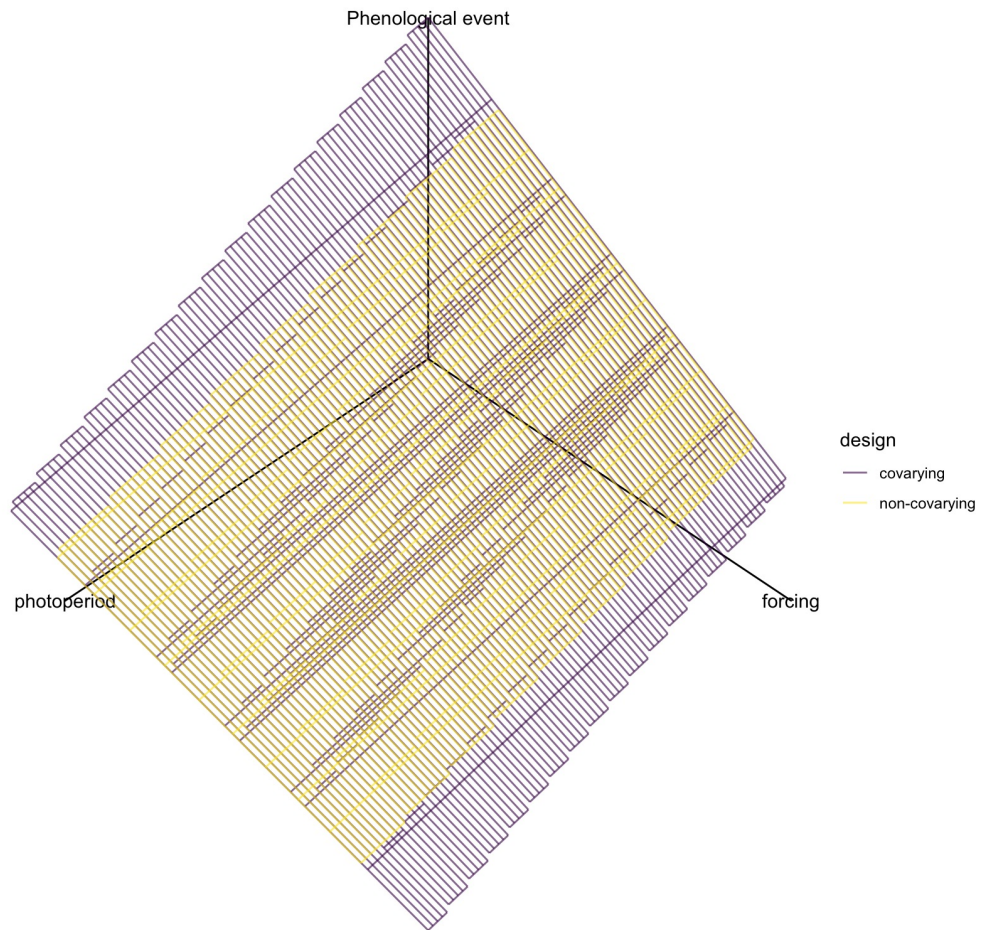


Figure 3: Can we make a good 3d version of this figure?

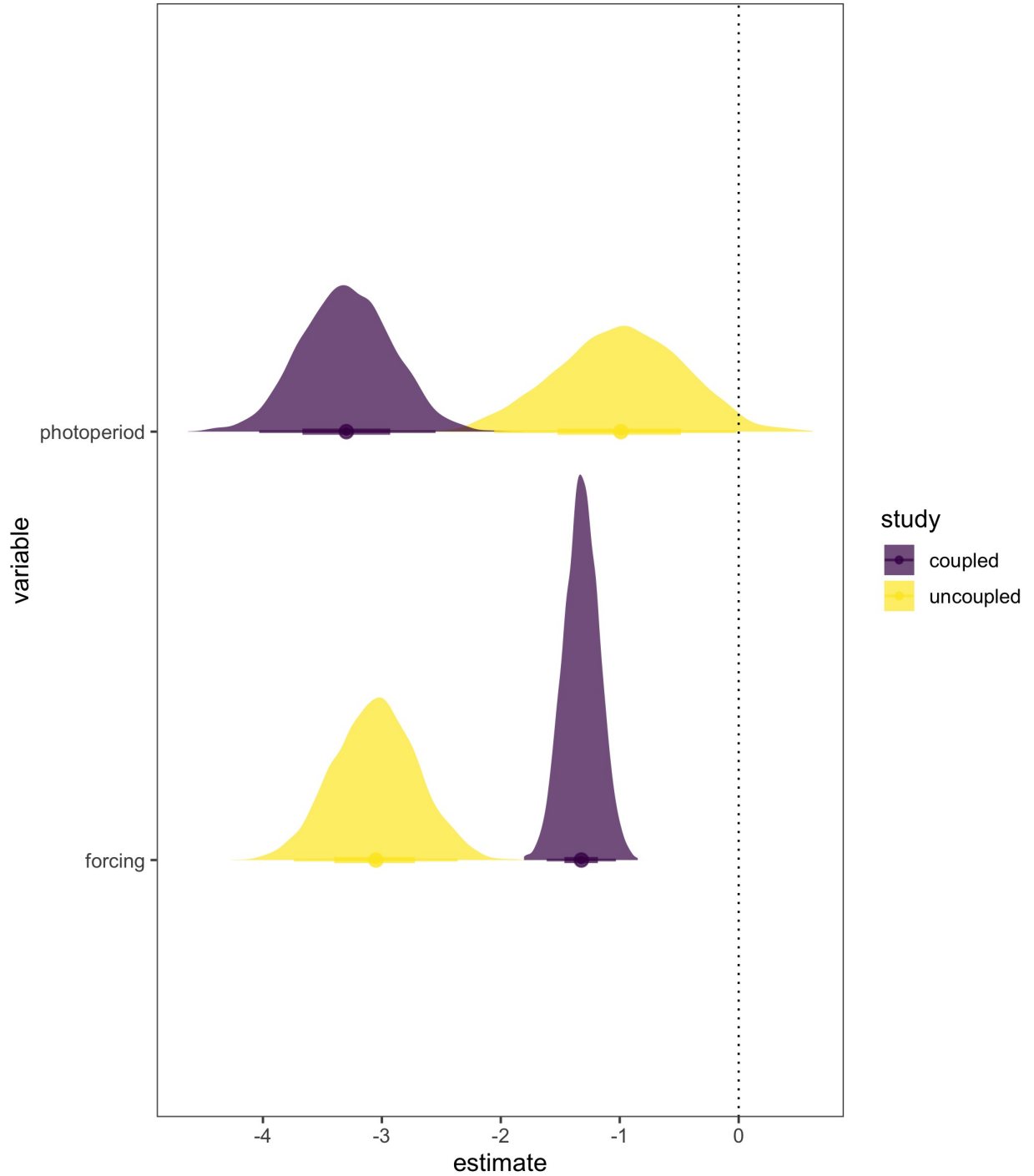


Figure 4: Estimated effects of one hour increase in photoperiod and one degree C increase in forcing on the day of leaf expansion using an alternative methods of varying thermo-period relative to photoperiod. Dots indicate the estimated mean effect, thick and thin bars the 50% and 97.5% credible intervals respectively. The full posterior distributions are also depicted for each parameter estimate. The colors represent the study design effect, with purple representing study design in which photo- and thermo-period co-vary, and yellow representing a design where there periodicity of each variable is independent of the other.