# Differences in flower and leaf bud environmental responses drive shifts in spring phenological sequences of temperate woody plants

D.M. Buonaiuto  $^{1,2,a},$  E.M. Wolkovich  $^3$ 

#### Author affiliations:

<sup>1</sup>Arnold Arboretum of Harvard University, Boston, Massachusetts, USA. ORCID: 0000-0003-4022-2591

<sup>2</sup>Department of Organismic and Evolutionary Biology, Harvard University, Cambridge, Massachusetts, USA

<sup>3</sup>Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, Vancouver, British Columbia, Canada

<sup>a</sup>Corresponding author: 617.823.0687; dbuonaiuto@g.harvard.edu

#### Abstract

The relative timing of growth and reproduction is an important driver of plant fitness. For deciduous woody species in temperate regions leaves and flowers both appear in the early spring, but the order and duration of these phenological events vary among species, populations, and individuals. This observed variation in flower-leaf sequences (FLSs) suggests that the relative timing of these events may be important—affecting the reproduction, recruitment and survival of individuals. Further, FLSs appear to be shifting with climate change, and anticipating the extent of these shifts is key to projecting how climate change may impact species' performance and reshape forest communities. Predicting FLS shifts, however, requires an improved understanding of how the environment dictates FLS patterns. To address this, we compared the phenological responses of flower and leaf buds for 10 temperate 11 woody species to varying levels of temperature and light in a lab experiment. Our experi-12 mental design allowed us to test competing hypotheses for how environmental cues determine 13 FLS variation—specifically whether forcing (warm temperatures) alone drives variation or differential sensitivity to chilling (cool temperatures generally in the fall and winter) and/or 15 photoperiod matter. We found that flower and leaf buds respond with differential sensitivity 16 to environmental cues, with differences in their response to chilling being the dominant driver 17 of FLS variation. Because climate change will amplify variability in chilling across time and space, these findings suggest that FLS shifts may be large, but are likely to vary substan-19 tially among populations and species. In our study, FLS shifts were largest in wind-pollinated 20 species that flower before leafing, raising the possibility that, more generally, wind-pollinated 21 taxa may experience reproductive declines due to FLS shifts in the decades to come.

23

Keywords: chilling, climate change, deciduous forests, differential sensitivity, flower-leaf sequences, forcing, hysteranthy, phenology

26

#### 27 Introduction

Among the most widely documented biological effects of anthropogenic climate change are shifts in plant phenology, the timing of life cycle events (Parmesan & Yohe, 2003; Menzel et al., 2006; Cleland et al., 2007). While phenology is generally advancing with climate change, the strength of these phenological shifts can vary substantially among specific phenological phases (Augspurger & Zaya, 2020). These differences alter the timing of phases relative to each other, changing the duration between events that make up phenological sequences (Ettinger et al., 2018). Phenological sequences are a major driver of plant fitness that impacts plant life history, resource allocation, demography and ecosystem processes (Post et al., 2008). Thus, shifting sequences with climate change will likely impact many of these processes. The effects of these shifts, however, depend both on their direction—whether distinct phases are shifting closer together or farther apart—and magnitude—how much they are shifting relative to each other.

40

Among deciduous woody plants, the relative timing of flower and leaf phenology, or flowerleaf sequences (FLSs), may be particularly consequential to fitness in temperate regions
where flowering prior to leaf development is common (Rathcke & Lacey, 1985; Gougherty &
Gougherty, 2018). Flowering before leafing may be a critical adaptation for pollination efficiency in wind-pollinated taxa, with the leafless canopy minimizing obstacles for air flow and
pollen transfer (Whitehead, 1969). In insect-pollinated taxa, flowering-first may increase the
visibility of flowers to pollinators (Janzen, 1967; Savage, 2019) or alleviate hydraulic demand
in dry conditions (Gougherty & Gougherty, 2018; Franklin, 2016).

49

Phenological observations over the last several decades indicate that, like other phenological sequences, FLSs are shifting due to anthropogenic climate change (Buonaiuto *et al.*, 2020).

For several species, the time between flowering and leafing appears to be increasing, but the strength of this trend varies among species and the direction of FLS shifts are not consistent across populations (Buonaiuto *et al.*, 2020). These changes could affect the important functions of FLSs, potentially putting some species at greater risk for fitness declines, while benefiting others.

57

For example, species with decreasing FLS interphases with climate change may see a reduction in pollination success as more pollen is intercepted by vegetation, or flowers are obscured by developing leaves. Conversely, pollination efficiency could improve for species with lengthening FLS interphases. A change in the FLS interphase of just a few days would likely have little impact on these processes, but if shifts were on the order of weeks, the impact on the pollination biology of a species could be significant. For example, because canopy structure influences the speed and direction of air flow (Niklas, 1985; Jackson & Lyford, 1999), decreasing FLS interphases could reduce pollen dispersal distances for wind-pollinated taxa (Milleron et al., 2012), which would ultimately impact population structure and gene flow across a landscape.

68

Predicting FLS shifts requires identifying the proximate mechanisms that drive FLS variation. Decades of research suggest that cool winter temperatures (chilling), warm spring temperatures (forcing), and day-length (photoperiod) are the primary drivers of both reproductive and vegetative phenology for woody plants in temperate regions (Körner & Basler, 2010; Flynn & Wolkovich, 2018). However, observed FLS shifts indicate that there must be differences in how these cues influence the phenology of flowers and leaves (Buonaiuto et al., 2020). Identifying these differences is a necessary step for predicting the direction, magnitude and—ultimately—fitness impacts of FLS shifts with climate change.

## 77 Hypotheses for FLS variation

Studies that have attempted to identify the differences between reproductive and vegetative phenology in woody plants (mostly focused on crop species) have yielded two common explanations:

One hypothesis suggests that reproductive and vegetative buds utilize the same underlying environmental cues, but have different threshold responses to forcing, with whichever bud type bursts later—leaves or flowers—having a higher thermal requirement (that is, they need a greater sum of warm temperature to trigger the phenological event, Guo et al., 2014; Cosmulescu & Calusaru, 2020; Cosmulescu & Ionescu, 2018). Under this hypothesis, which we call the forcing hierarchy hypothesis (FHH), leaf and flower buds share the same suite of cues and develop similarly to non-forcing cues (i.e., chilling and photoperiod), but differ in the thermal requirement for budburst.

89

In contrast, an alternative hypothesis suggests that flower and leaf buds differ in the strength of their phenological responses to multiple environmental cues (Citadin *et al.*, 2001; Gariglio *et al.*, 2006; Aslamarz *et al.*, 2009; Mehlenbacher, 1991). Under this hypothesis, which we call the differential sensitivity hypothesis (DSH), each bud type relies more or less on certain cues, generating FLS variability.

95

While observational studies of phenology in the field are commonly used to evaluate the relationship between environmental conditions and phenology (Cleland *et al.*, 2007), under current and recent climatic conditions, the FHH and DSH may produce similar phenological patterns, making it difficult to evaluate these hypotheses with such methods. However, experiments designed to isolate all three environmental cues have the potential to disentangle the two hypotheses. Studies aiming to differentiate these hypotheses can look for two dif-

ferent signatures. The key signature of the FHH is that the sensitivity to forcing ( $\Delta$  day of phenological event/ $\Delta$ °C) of the second phase in the phenological sequence is always greater than that of the first phase with sensitivity differences being inversely proportional to the difference in thermal requirement among bud types (2x in our simulations; Fig. 1 a.). By contrast, the signature of the DSH is that sensitivity estimates of flower and leaf buds to chilling and/or photoperiod will diverge (Fig. 1 c.), but there is no characteristic pattern as to the strength or direction of this divergence (e.g., leaves may require longer or shorter photoperiods than flowers, or more or less chilling).

110

The clarity of these signatures from each hypothesis, however, may be obscured in certain 111 situations. First, a species' buds could be both differentially sensitive to photoperiod or chill-112 ing (supporting the DSH) and also respond to forcing in a forcing hierarchy (that is, both hypotheses can operate at once, see Fig. 1 b.). Second, the differential sensitivity framework 114 can generate the signature of the FHH when other cues are at high levels due to interactions 115 between the chilling or photoperiod response and the thermal requirement, which make forc-116 ing the dominant phenological cue once other cue requirements have been met (see Fig. 1 c., 117 chill x force interaction). When this happens the FHH would effectively be a special case of 118 the DSH. 119

120

While these two hypothesized mechanisms may produce similar phenology patterns under current conditions, differentiating them is important, as they have substantially different implications regarding the potential for FLS shifts with climate change. The FHH suggests that FLS variation is largely a product of climate variation during the interphase. If spring temperatures increase with climate change, the second phenophase of the FLS with be accelerated relative to the first and FLS interphases will decrease, but given the relative auto-correlation of spring temperatures (Di Cecco & Gouhier, 2018), these shifts should be

muted. If FLSs are structured by a forcing hierarchy, the direction of FLS shifts are relatively straight forward to predict, and it is less likely that shifts will be large enough in magnitude to significantly affect woody plant fitness.

131

Climate change would also drive FLS shifts under the DSH, but the trajectory of these shifts 132 are more complicated to predict. The DSH suggests that there could be strongly localized 133 effects of climate change on FLSs. Shifts in FLS variation will depend on the direction and 134 rate of change in cues at given locations and the species-specific differential sensitivity of re-135 productive and vegetative phenology to cue combinations. This hypothesis allows for larger 136 magnitude shift in FLSs, on a scale that could impact pollen transport or the physiological 137 functioning of woody plants. Further, the DSH also suggests that the magnitude of shifts 138 may be highly divergent both among species in a community, and among populations of the same species which could impact gene flow, population structure and demography. 140

141

In this study we tested the FHH and DSH hypotheses in 10 temperate shrub and tree species.

Using a full-factorial growth-chamber experiment we manipulated chilling, forcing and photoperiod cues for flower and leaf buds. We then leveraged these data to examine how FLSs
may shift with climate change.

146

# Materials and Methods

## 148 Growth chamber study

We sampled plant material from Harvard Forest in Petersham, MA, USA (42.5314°N, 72.1900°W) on 25 October 2017, immediately after most plants in the area entered dormancy but before

they could accumulate significant chilling in the field. We collected branch cuttings from 151 7-13 individuals of 12 woody plant species (4-12 cuttings per individual for a total of 48-56 152 per species), consisting of a mix of deciduous shrubs, understory and canopy trees commonly 153 found in mesic hardwood forests of the eastern United States. The use of branch cuttings 154 to study woody plant phenology in artificial environments is common (Ettinger et al., 2020), 155 and these methods have been shown to match whole-plant phenological patterns (Vitasse & 156 Basler, 2014; Primack et al., 2015). We selected species displaying variation of the three ma-157 jor FLS patterns; species for which flowers appear before leaves (flowering-first; Acer rubrum, 158 Betula alleghaniensis, Comptonia peregrina, and Corylus cornuta), species for which flowers 159 appear with their leaves (concurrent; Acer pensylvanicum, Acer saccharum, Ilex mucronata, 160 Prunus pensylvanica, Vaccinium corymbosum), and species for which flowers appear after 161 leaf development (leafing-first; *Ilex verticillata, Prunus virginiana, Viburnum acerifolium*), 162 These species also vary in other important functional traits including pollination syndrome, 163 bud type, and growth habit (see tab. S1 for details and full species list). 164

We transported all cuttings to the Arnold Arboretum in Boston, MA (USA) where they were re-cut in water to prevent callousing and cavitation and placed in 500 ml Erlenmeyer flasks with distilled water.

168

We randomly assigned cuttings to a fully crossed set of eight experimental treatments; two levels of chilling (4 vs 8 weeks at 4°C), two levels of temperature (24°C:18°C (day/night) warm vs 18°C:12°C (day/night) cool) and two levels of photoperiod (12 vs 8 hours). We alternated day/night temperature periodicity on a 12 hour schedule to reduce co-variation with photo-periodicity. We re-cut all twigs and changed the water every 7-10 days and rotated all treatments between growth chambers every two weeks to minimize chamber effects. We made phenological observations every 2-3 days using a modified BBCH scale for woody plants (Finn et al., 2007) for three months following release from chilling conditions. In this

period we assessed two phenological phases: budburst (BBCH phase 07) and first flower open (BBCH 60). At the conclusion of this period we assessed all individuals that did not undergo budburst and excluded 56 dead twigs from our analyses.

#### Data analysis

To assess the phenological sensitivity of each phase, we fit mixed-effects hierarchical models 181 with chilling, forcing, photoperiod and all two-way interactions as the fixed effects and species 182 as a grouping factor on both the slopes and the intercepts. We chose a Bayesian hierarchical 183 approach in order to identify systematic trends across species' responses while accounting 184 for sample size, variance, and the unique effect of each species. We also used z-scoring to 185 standardized the units of each environmental predictor to allow for appropriate comparisons 186 between them (Gellman & Hill, 2007). We modeled the effects of environmental parameters 187 on flower opening and leaf budburst separately. We also fit a model with FLS interphase (day 188 of budburst - day of flowering) as a response variable to compare these estimates with field 189 observations. Two species, Betula allegheniensis and Acer saccharum, produced no flowers 190 in our trial, likely because flowering branches were out of reach during our field sampling, so 191 we excluded them from our analysis. In total, our analyses included 464 twigs from 10 species. 192

194 The models we fit appear below:

193

195

198

196 
$$y_{[i]} \sim N(\alpha_{sp_{[i]}} + \beta_{forcing_{sp[i]}} + \beta_{chilling_{sp[i]}} + \beta_{photoperiod_{sp[i]}} + \beta_{forcingxchilling_{sp[i]}} + \beta_{forcingxphotoperiod_{sp[i]}} + \beta_{forc$$

Where  $y_{[i]}$  is either the day of the experiment of leaf budburst, day of first flower opening or FLS interphase length, and  $\sigma_y^2$  the error. We modeled the intercept  $(\alpha)$  and each slope  $(\beta)$  parameter at the species level using the formula:

202

$$\alpha_{x_{sn}}$$
 or  $\beta_{x_{sn}} \sim N(\mu_x, \sigma_x^2)$ 

204

To test the hypothesis that the FHH is a special case of the DSH that occurs when other cue (i.e., chilling and photoperiod) requirements are met, we re-ran our models on a subset of our data which included both levels of the forcing treatment but only the high photoperiod and chilling treatment levels. This model included forcing as the only main effect but, like our main models written above, included species as a grouping factor on the slopes and intercept.

210

We fit all models using the R package "brms" (Bürkner, 2018). We ran each model on four chains with 4000 iterations and a 3000 iteration warm up for a total of 4000 posterior draws for each parameter. In all models we used weakly informative priors and increasing the priors 5-fold did not affect the model results. We assessed model performance through ensuring  $\hat{R}$ s were between 1 and 1.01 and bulk and tail effective sample sizes were high (1800-2800 for most parameters, but as low as 800-900 for some). We present 50% credible intervals in figures generally because they are the most computationally stable (Gelman *et al.*, 2013) but provide other intervals in the Supporting Information (Tab. S3,Tab. S4, Tab. S5.

219

## Climate change scenarios

To apply our model results to general climate change scenarios we chose environmental treatments in the experiment to broadly reflect historic and future conditions at our sampling site. Our low forcing treatment approximated average spring temperature (March/April) at the site while our high temperature treatment reflects a 5 °C increase. Average field chilling

(calculated from 15 October - 15 April, measured in Chill Hours, Weinberger, 1950) at Har-225 vard Forest is 1170.7, approximately 63\% of the difference between our low and high chilling 226 treatment (Tab. S2). Thus, our low chilling treatment represents a feasible estimate for 227 a decrease in chilling with climate change and our high chilling treatment approximates a 228 reasonable increase (Luedeling, 2012). Our low photoperiod treatment (8 hours of daylight) is well below the photoperiod experienced at Harvard Forest, but given that the photoperiod 230 effects are expected to be small (Laube et al., 2014), we chose more extreme values in order 231 to robustly estimate an effect (i.e., increasing statistical power). For this reason, our climate 232 change scenarios for FLS variation are based on our high photoperiod treatment alone. 233

234

We used our flower and budburst models to project for each species in our study:

- 1. FLSs under average environmental conditions (low forcing, 6.5 weeks of chilling treatment)
- 238 2. FLS shifts with spring warming only (high forcing, 6.5 weeks of chilling treatment)
- 3. FLS shifts with warming and increased chilling (high forcing, 8 weeks of chilling treatment)
- 4. FLS shifts with warming and decreased chilling (high forcing, 4 weeks of chilling treatment)
- To validate our predictions, we compared our FLS interphase model estimates of "average" condition FLS interphases to long term phenological records from Harvard Forest (O'Keefe, 2015) for five species common to both datasets (Fig. S1), and found them comparable.

246

Given the variable dynamics of shifts in environmental forcing and chilling with climate change over time and space (Luedeling *et al.*, 2011), these scenarios should not be treated

as absolute predictions of the magnitude of FLS shifts with climate change. Instead, we provide these scenarios to identify general trends in how FLSs could shift with warming and demonstrate that the range of possibilities vary based on characteristics of plant species and the specific climate dynamics.

253

Finally, we characterized FLS shifts for three broad FLS groups (flowering-first, concurrent, leafing-first) by extracting all sampling posterior estimates from our main models using the R package tidybayes (Kay, 2020), and grouping them by FLS pattern. We also include the species-specific estimates on which these grouped estimates are based in Fig. S2.

258

#### 59 Results

## 260 Growth chamber study

Both flower and leaf phenology advanced with higher forcing and longer chilling duration 261 (flowers: chilling effect: -21.2, forcing effect: -19.0, leaves: chilling effect: -30.4, forcing ef-262 fect: -17.8; Fig. 2, Tab. S3, Tab. S4—all standardized units; see "Data analysis" section 263 above), but increases in both of these cues together offset these advances (flowers: force x 264 chill effect: 7.0, leaves: force x chill effect: 12.4; Fig. 2, Tab. S3, Tab. S4). Leaf and flower 265 buds diverged in their responses to increasing photoperiod, with flower phenology advancing 266 and leaf phenology delaying when the other two cues were at low levels (Fig. 2, Tab. S3, 267 Tab. S4). As seen in the interactions between photoperiod and chilling and photoperiod 268 and forcing, increasing chilling or forcing with longer photoperiod advanced the phenology 269 of both bud types. For both bud types, chilling and forcing were the dominant cues, while 270 increasing photoperiod produced a more muted phenological response (Fig. 2, Tab. S3, Tab.

272 S4).

273

While leaf and flower bud phenological responses to environmental cues were qualitatively similar, the strength of their responses to each cue differed substantially. Leaf buds responded more strongly to chilling than flower buds (1.4x), and had a stronger response to all cue interactions (forcing x chilling: 1.8x, photoperiod x chilling: 5.8x, photoperiod x forcing: 2.2x; Fig. 2, Tab. S3, Tab. S4). Across all species both bud types displayed a similar magnitude advance with increased forcing (Fig. 2, Tab. S3, Tab. S4).

280

Our results did not showed the characteristic sensitivity pattern of the FHH across most species in our experiment (Fig. 2, see Fig. 1,a.,b.), though there was significant variation among species in the strength of their response to forcing between bud types. However, when we re-ran our models on the subset of data which included phenological observations at only high levels of chilling and photoperiod, we found the sensitivity to forcing for 9/10 species followed the predicted pattern of the FHH, with the second phases of the FLS showing stronger sensitivity to forcing than the first phases (Fig. 3, Tab. S5).

288

# Climate change scenarios

Our models project that both flower and leaf phenology will advance in our generalized future scenarios. FLS shifts depended strongly on how forcing levels change relative to chilling exposure (Fig. 4). Given the significant differences in sensitivity to chilling between flowering and leafing phenology we found in our experiment, our scenarios showed that FLS interphases will be more strongly influenced by changes in chilling duration than increased forcing alone.

295

The direction and magnitude of shifts in FLS interphases varied among species, varying espe-296 cially by whether or not they displayed a flowering-first FLS. Under some warming scenarios, 297 our model predicted that FLS interphases for some species may effectively disappear or the 298 order of phenophases in the FLS may switch (Fig. 4, Fig. S2). Several species, Acer rubrum, 299 Ilex verticillata, Prunus pensylvanica, Prunus virginiana, and Viburnum acerifolium, had FLSs that were relatively robust to changing environments. For other species, Acer pen-301 sylvanicum, Vaccinium corymbosum and Ilex mucronata, which typically begin to produce 302 leaves shortly before flowers open, the magnitudes of projected FLS shifts were moderate. 303 The two species with the most significant FLS shifts in both direction and magnitude across 304 treatment combinations and climate change scenarios were Comptonia peregrina and Corylus 305 cornuta (Fig. S2). In all of our climate change scenarios, the FLS interphase was dramati-306 cally reduced in these taxa. 307

# Discussion

308

Our experiment supports the hypothesis that flower-leaf sequences are structured by dif-310 ferential sensitivity (DSH) to the environment between flower and leaf buds. Specifically, 311 differences in the chilling response among bud types was the strongest driver of FLS varia-312 tion. We also found that under high chilling and long photoperiods, the FLSs of most species followed the predicted sensitivity pattern of the FHH, supporting the hypothesis that the 314 FHH is likely a special case of the DSH that occurs when other phenological cues are met 315 for both bud types. Together, these results explain why the two FLS hypotheses have been 316 difficult to distinguish, and suggest that climate change has the potential to significantly disrupt FLSs as global warming alters historic chilling patterns across the temperate zone 318 (Morin et al., 2009). 319

# Reconciling the differential sensitivity and the forcing hierarchy hypotheses

The relationship we found between the FHH and the DSH in our experiment offers a path forward for reconciling previous studies that have favored one hypothesis over the other.

Support for the FHH is most often associated with observational studies (e.g. Cosmulescu & Calusaru, 2020; Guo et al., 2014). By contrast, experimental studies which manipulate chilling levels beyond historically observed minima in the field tend to support the DSH (e.g. Aslamarz et al., 2009; Gariglio et al., 2006). This pattern can be explained by the FHH being a special case of the DSH, if historic climate regimes make detection of the DSH in the field difficult.

330

Under historic climate regimes, seasonal chilling requirements were usually met (Gauzere 331 et al., 2019; Chuine et al., 2016), making it difficult to detect any effect of chilling in observa-332 tional studies, let alone differences in flower versus leaf buds. It would take larger reductions 333 in chilling to reveal the signature of the DSH, which at present, rarely occurs in the field, but 334 can be induced in artificial environments which explains the discrepancy between these two 335 methods. These dynamics are consistent with findings in other phenological studies that sug-336 gest simple growing degree models (which underlie the FHH) accurately predict phenology 337 under current climate, but under-perform under climate change scenarios when shifts in chilling accumulation become more pronounced (Linkosalo et al., 2008; Chuine et al., 2016). Our 339 findings suggest that as climate continues to change, differential sensitivity to the environ-340 ment between flower and leaf phenology should become more apparent in field observations, 341 and that the magnitude of interannual FLS variation is likely to extend beyond historically observed reaction norms. 343

344

#### Population-level implications of the DSH with climate change

Predicted shifts in chilling are highly variable across both time and space—because chilling 346 only accumulates at intermediately low temperatures warming may increase chilling at some 347 locations while decreasing it in others (Man et al., 2017; Zhang et al., 2007). Layered onto 348 these variable shifts in chilling, spring warming from climate change also differs by location 349 (Karmalkar & Bradley, 2017; Loarie et al., 2009), suggesting that forcing may increase more 350 rapidly in some places than others. These shifts in both chilling and forcing suggest that the direction and magnitude of FLS shifts is likely to vary substantially among populations based 352 on the specific cue combinations at a given locality (Chmielewski et al., 2012). Long-term 353 phenology records show there was already substantial intra-specific variation in FLSs at the 354 population level (Buonaiuto et al., 2020) and our findings suggest that these population-level 355 differences may be further amplified by climate change. In this way, all the three generic FLS 356 climate change scenarios depicted in Fig. 4 should not be considered alternatives to each 357 other, but could occur contemporaneously across a species' range. 358

359

This increased population-level heterogeneity in FLSs that our models project has the potential to influence patterns of pollen dispersal across the landscape (Borycka et al., 2017;
Pace et al., 2018). For example, advancing canopy closure relative to flowering can impede long-distance pollen transport (Milleron et al., 2012), which may in turn alter patterns of gene flow across a landscape. Our findings regarding FLS shifts should be integrated with projections of pollinator movement or prevailing wind directions (Kling & Ackerly, 2020) to better understand how FLS variation may contribute to population structure in the long term.

367

While changes in chilling and forcing from climate change may increase FLS variation, the climate change implications of strong differential sensitivity to photoperiod among bud types are more difficult to predict. Climate change does not directly impact photoperiod, but

may alter the photoperiods plants experience as they approach their temperature optima 371 for phenological activity. However, in most temperate regions, temperature would have to 372 drive phenological shifts by a minimum of several weeks before the experienced photoperiod 373 of plants would change substantially. Because these dynamics are more extreme than most 374 studies suggest (Thackeray et al., 2016), we modeled climate change scenarios with a constant photoperiod in our FLS scenarios. Such an approach may be useful at temperate and 376 some boreal latitudes, but may not work at high latitudes, where photoperiod changes more 377 rapidly over the season and thus temperature-driven phenological shifts may significantly 378 alter the experienced photoperiod of plants. Our results suggest that such shifts could mute 379 or amplify the FLS shifts, which may be particularly important as species shift their dis-380 tribution poleward with climate change and begin to encounter novel photoperiod regimes 381 (Way & Montgomery, 2015). 382

383

# Species-level implications of the DSH with climate change

Our study highlights that the direction and magnitude of FLS shifts with climate change are species-specific. Not only is it likely that the function of FLS variation differs among species (Buonaiuto *et al.*, 2020), but we found that FLSs of some species were very sensitive to changing climate conditions, while other remain fairly resilient (Fig. 4, Fig. S2).

389

These differences suggest that FLS shifts will impact some species more than others and successful forest conservation and management programs may require identifying the species that are most vulnerable to extreme FLS shifts. To aid this effort, there is a strong need for generalizing principles to identify species with potential for consequential FLS shifts with climate change. While our one study does not begin to represent the taxonomic diversity of a temperate forest, several patterns in the FLS responses of our multi-species experiment

may serve as starting point for further inquiry.

397

The different FLS response patterns we observed across species may correlate with suite of 398 interrelated anatomical, physiological and phenological traits. The species that maintained 399 FLS structure across our climate change scenarios generally shared a strongly leafing-first 400 FLS, with a fairly long FLS interphase (Fig 4). These species tended to have mixed buds 401 (leaf and flower primordia together in one bud) so there may be strong physical constraints on 402 their FLSs. By contrast, the species that were most sensitive to FLS shifts were monoecious, 403 flowering-first, wind-pollinated shrubs (Fig 4, Fig S2). This result supports other evidence 404 that the reproductive phenology of wind-pollinated species may be more sensitive to climate 405 change compared to biotically pollinated taxa (Ziello et al., 2012). Given the hypothesized 406 function of FLSs in wind-pollinated species, the significant reductions in FLS interphases we 407 observed suggest that these species, and flowering-first, wind-pollinated taxa in general, may 408 face particular risk for reproductive performance reductions. 409

410

While much of the public interest around phenology and pollination in the context of global 411 change has centered around trophic mismatches between pollinator and floral phenology 412 (Memmott et al., 2007), which is of little relevance to abiotically pollinated taxa, our study 413 identified the possibility that the effects of FLS shifts with climate change may be particu-414 larly important for wind-pollinated woody plants. The direction and magnitude of FLS shifts 415 we observed in these taxa, coupled with the hypothesized function of a flowering-first FLS in 416 wind-pollinated species, suggests that FLS variation in this functional group should be ex-417 plored in greater detail in the future. More research is needed to identify species' traits that 418 may correlate with the potential for FLS shifts, but flowering-first, wind-pollinated species 419 may be particular sensitive to FLS shifts. Our results suggest that species in this functional 420 group should be considered a research priority for the study of spring phenological sequences in deciduous woody plants.

423

# 424 Acknowledgements:

We thank C. de Keyzer for generously helping to collect and prepare plant materials for the

experiment. We also thank R. Lee for help with monitoring and maintaining the experiment.

427

# Author Contributions:

DMB and EMW conceived of the experiment; DMB collected the data and performed the

430 statistical analyses, DMB and EMW wrote the manuscript. All authors contributed critically

431 to the drafts and gave final approval for publication.

432

# Data Availability Statement:

Data will be available on KNB (https://knb.ecoinformatics.org/) upon publication. All mod-

eling code will be made available upon request.

# References

Aslamarz, A.A., Vahdati, K., Rahemi, M. & Hassani, D. (2009) Estimation of chilling and

heat requirements of some persian walnut cultivars and genotypes. HortScience 44, 697–

439 701.

- Augspurger, C.K. & Zaya, D.N. (2020) Concordance of long-term shifts with climate warming
  varies among phenological events and herbaceous species. *Ecological Monographs* n/a,
  e01421.
- Borycka, K., Ortyl, B. & Kasprzyk, I. (2017) Temporal variations and spatial differentiation in the black alder and silver birch pollination pattern-the impact of local climate or something more? Agricultural and Forest Meteorology 247, 65–78.
- Buonaiuto, D.M., Morales-Castilla, I. & Wolkovich, E.M. (2020) Reconciling competing hypotheses regarding flower-leaf sequences in temperate forests for fundamental and global change biology. New Phytologist n/a.
- Bürkner, P.C. (2018) Advanced bayesian multilevel modeling with the r package brms. *R*Journal 10, 395–411.
- Chmielewski, F.M., Blümel, K. & Pálesová, I. (2012) Climate change and shifts in dormancy
   release for deciduous fruit crops in germany. Climate Research 54, 209–219.
- Chuine, I., Bonhomme, M., Legave, J.M., García de Cortázar-Atauri, I., Charrier, G., Lacointe, A. & Améglio, T. (2016) Can phenological models predict tree phenology accurately
  in the future? the unrevealed hurdle of endodormancy break. *Global Change Biology* 22,
  3444–3460.
- Citadin, I., Raseira, M.C.B., Herter, F.G. & da Silva, J.B. (2001) Heat requirement for
   blooming and leafing in peach. HortScience HortSci 36, 305–307.
- Cleland, E.E., Chuine, I., Menzel, A., Mooney, H.A. & Schwartz, M.D. (2007) Shifting plant
   phenology in response to global change. Trends in Ecology Evolution 22, 357 365.
- Cosmulescu, S. & Calusaru, F.G. (2020) Influence of temperature on blackthorn (prunus spinosa l.) phenophases in spring season. *Journal of Agricultural Meteorology* **76**, –.

- Cosmulescu, S. & Ionescu, M. (2018) Phenological calendar in some walnut genotypes grown in romania and its correlations with air temperature. *International Journal of Biometeo-*
- rology **62**.
- Di Cecco, G.J. & Gouhier, T.C. (2018) Increased spatial and temporal autocorrelation of temperature under climate change. *Scientific Reports* 8, 14850.
- Ettinger, A., Gee, S. & Wolkovich, E. (2018) Phenological sequences: how early season events define those that follow. *American Journal of Botany* **105**.
- Ettinger, A.K., Chamberlain, C.J., Morales-Castilla, I., Buonaiuto, D.M., Flynn, D.F.B.,
- Savas, T., Samaha, J.A. & Wolkovich, E.M. (2020) Winter temperatures predominate in
- spring phenological responses to warming. Nature Climate Change 10, 1137–1142.
- Finn, G.A., Straszewski, A.E. & Peterson, V. (2007) A general growth stage key for describing trees and woody plants. *Annals of Applied Biology* **151**, 127–131.
- Flynn, D.F.B. & Wolkovich, E.M. (2018) Temperature and photoperiod drive spring phenology across all species in a temperate forest community. *New Phytologist* **219**, 1353–1362.
- Franklin, D.C. (2016) Flowering while leafess in the seasonal tropics need not be cued by leaf drop: evidence from the woody genus brachychiton (malvaceae). *Plant Ecology and Evolution* **149**, 272–279.
- Gariglio, N., González Rossia, D.E., Mendow, M., Reig, C. & Agusti, M. (2006) Effect of artificial chilling on the depth of endodormancy and vegetative and flower budbreak of peach and nectarine cultivars using excised shoots. *Scientia Horticulturae* **108**, 371–377.
- Gauzere, J., Lucas, C., Ronce, O., Davi, H. & Chuine, I. (2019) Sensitivity analysis of tree phenology models reveals increasing sensitivity of their predictions to winter chilling temperature and photoperiod with warming climate. *Ecological Modelling* **411**, 108805.

- Gellman, A. & Hill, J. (2007) Data Analysis Using Regression and Multilevel/Hierarchical

  Models. Cambridge University Press.
- Gelman, A., Carlin, J., Stern, H., Dunson, D., Vehtari, A. & Rubin, D. (2013) Bayesian Data

  Analysis. Chapman and Hall/CRC.
- Gougherty, A.V. & Gougherty, S.W. (2018) Sequence of flower and leaf emergence in deciduous trees is linked to ecological traits, phylogenetics, and climate. New Phytologist 220, 121–131.
- Guo, L., Luedeling, E., Dai, J. & Xu, J. (2014) Differences in heat requirements of flower and leaf buds make hysteranthous trees bloom before leaf unfolding. *Plant Diversity and Resources* **36**, 245–253.
- Jackson, S.T. & Lyford, M.E. (1999) Pollen dispersal models in quaternary plant ecology:

  Assumptions, parameters, and prescriptions. *The Botanical Review* **65**, 39–75.
- Janzen, D.H. (1967) Synchronization of sexual reproduction of trees within the dry season in central america. *Evolution* **21**, 620–637.
- Karmalkar, A.V. & Bradley, R.S. (2017) Consequences of global warming of 1.5 °c and 2 °c for regional temperature and precipitation changes in the contiguous united states. *PLOS*ONE 12, e0168697—.
- Kay, M. (2020) tidybayes: Tidy Data and Geoms for Bayesian Models. R package version 2.1.1.
- Kling, M.M. & Ackerly, D.D. (2020) Global wind patterns and the vulnerability of winddispersed species to climate change. *Nature Climate Change* **10**, 868–875.
- Körner, C. & Basler, D. (2010) Phenology under global warming. Science 327, 1461–1462.

- Laube, J., Sparks, T.H., Estrella, N., Höfler, J., Ankerst, D.P. & Menzel, A. (2014) Chilling outweighs photoperiod in preventing precocious spring development. *Global Change Biology*20, 170–182.
- Linkosalo, T., Lappalainen, H.K. & Hari, P. (2008) A comparison of phenological models of leaf bud burst and flowering of boreal trees using independent observations. *Tree Physiology*28, 1873–1882.
- Loarie, S.R., Duffy, P.B., Hamilton, H., Asner, G.P., Field, C.B. & Ackerly, D.D. (2009) The velocity of climate change. *Nature* **462**, 1052–1055.
- Luedeling, E. (2012) Climate change impacts on winter chill for temperate fruit and nut production: A review. *Scientia Horticulturae* **144**, 218–229.
- Luedeling, E., Girvetz, E.H., Semenov, M.A. & Brown, P.H. (2011) Climate change affects winter chill for temperate fruit and nut trees. *PLOS ONE* **6**, e20155—.
- Man, R., Lu, P. & Dang, Q.L. (2017) Insufficient chilling effects vary among boreal tree species and chilling duration. *Frontiers in Plant Science* 8, 1354.
- Mehlenbacher, S.A. (1991) Chilling requirements of hazelnut cultivars. *Scientia Horticulturae*47, 271–282.
- Memmott, J., Craze, P.G., Waser, N.M. & Price, M.V. (2007) Global warming and the disruption of plant–pollinator interactions. *Ecology Letters* **10**, 710–717.
- Menzel, A., Sparks, T.H., Estrella, N., Koch, E., Aasa, A., Ahas, R., Alm-Kuebler, K.,
- Bissolli, P., Braslavska, O., Briede, A., Chmielewski, F.M., Crepinsek, Z., Curnel, Y.,
- Dahl, A., Defila, C., Donnelly, A., Filella, Y., Jatcza, K., Mage, F., Mestre, A., Nordli, O.,
- Penuelas, J., Pirinen, P., Remisova, V., Scheifinger, H., Striz, M., Susnik, A., Van Vliet,
- A.J.H., Wielgolaski, F.E., Zach, S. & Zust, A. (2006) European phenological response to
- climate change matches the warming pattern. Global Change Biology 12, 1969–1976.

- Milleron, M., Lopez de Heredia, U., Lorenzo, Z., Perea, R., Dounavi, A., Alonso, J., Gil,
- L. & Nanos, N. (2012) Effect of canopy closure on pollen dispersal in a wind-pollinated
- species (Fagus sylvatica L.). Plant Ecology 213, 1715–1728.
- Morin, X., Lechowicz, M.J., Augspurger, C.K., O'Keefe, J., Viner, D. & Chuine, I. (2009)
- Leaf phenology in 22 north american tree species during the 21st century. Global Change
- Biology **15**, 961–975.
- Niklas, K.J. (1985) The aerodynamics of wind pollination. The Botanical Review 51, 328–
- <sub>539</sub> 386.
- O'Keefe, J. (2015) Phenology of Woody Species at Harvard Forest since 1990. Harvard Forest
- Data Archive: HF003., Petersham, MA, USA.
- Pace, L., Boccacci, L., Casilli, M., Di Carlo, P. & Fattorini, S. (2018) Correlations between
- weather conditions and airborne pollen concentration and diversity in a mediterranean
- high-altitude site disclose unexpected temporal patterns. Aerobiologia 34, 75–87.
- Parmesan, C. & Yohe, G. (2003) A globally coherent fingerprint of climate change impacts
- across natural systems. Nature **421**, 37 EP -.
- Post, E.S., Pedersen, C., Wilmers, C.C. & Forchhammer, M.C. (2008) Phenological sequences
- reveal aggregate life history response to climatic warming. Ecology 89, 363–370.
- Primack, R.B., Laube, J., Gallinat, A.S. & Menzel, A. (2015) From observations to exper-
- iments in phenology research: investigating climate change impacts on trees and shrubs
- using dormant twigs. Annals of Botany 116, 889–897.
- Rathcke, B. & Lacey, E.P. (1985) Phenological patterns of terrestrial plants. Annual Review
- of Ecology and Systematics 16, 179–214.

- Savage, J.A. (2019) A temporal shift in resource allocation facilitates flowering before leaf out and spring vessel maturation in precocious species. *American Journal of Botany* **106**, 113–122.
- Thackeray, S.J., Henrys, P.A., Hemming, D., Bell, J.R., Botham, M.S., Burthe, S., Helaouet,
- P., Johns, D.G., Jones, I.D., Leech, D.I., Mackay, E.B., Massimino, D., Atkinson, S., Bacon,
- P.J., Brereton, T.M., Carvalho, L., Clutton-Brock, T.H., Duck, C., Edwards, M., Elliott,
- J.M., Hall, S.J.G., Harrington, R., Pearce-Higgins, J.W., Høye, T.T., Kruuk, L.E.B.,
- Pemberton, J.M., Sparks, T.H., Thompson, P.M., White, I., Winfield, I.J. & Wanless,
- S. (2016) Phenological sensitivity to climate across taxa and trophic levels. *Nature* 535,
- <sub>563</sub> 241–245.
- Vitasse, Y. & Basler, D. (2014) Is the use of cuttings a good proxy to explore phenological responses of temperate forests in warming and photoperiod experiments? *Tree Physiology*
- 566
- Way, D.A. & Montgomery, R.A. (2015) Photoperiod constraints on tree phenology, performance and migration in a warming world. *Plant, Cell & Environment* **38**, 1725–1736.
- Weinberger, J.H. (1950) Chilling requirements of peach varieties. id 19511602967. Proceed ings. American Society for Horticultural Science 56, 122–28.
- Whitehead, D.R. (1969) Wind pollination in the angiosperms: Evolutionary and environmental considerations. *Evolution* **23**, 28–35.
- Zhang, X., Tarpley, D. & Sullivan, J.T. (2007) Diverse responses of vegetation phenology to
   a warming climate. Geophysical Research Letters 34.
- Ziello, C., Böck, A., Estrella, N., Ankerst, D. & Menzel, A. (2012) First flowering of wind pollinated species with the greatest phenological advances in europe. *Ecography* 35, 1017–
   1023.

# Figures

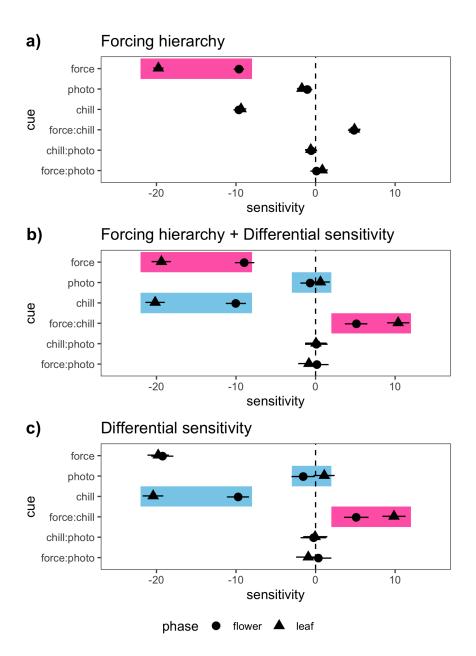


Figure 1: Characteristic sensitivity ( $\Delta$  day of phenological event/  $\Delta$  environmental cue) patterns of the phenological response to changing cue levels for the two major flower-leaf sequence hypotheses. a) displays a signature pattern of the forcing hierarchy hypothesis (FHH, pink boxes)—with the second phenophase in the sequence (in this case leafing) having a higher sensitivity to forcing than the first. b) depicts a scenario where both the FHH and the differential sensitivity hypothesis (DSH) contribute to flower-leaf sequence variation. Here the characteristic forcing sensitivity of the FHH is still apparent but the differential sensitivity to chilling and photoperiod is seen as well (blue boxes). c) highlights a typical sensitivity pattern produced by the DSH without the FHH. All plots above are based on simulations (see Supporting Information: Methods). Shapes indicate mean estimates and lines depict 95% credible intervals from Bayesian hierarchical models with advances in phenology shown as negative numbers, and delays in phenology as positive numbers.

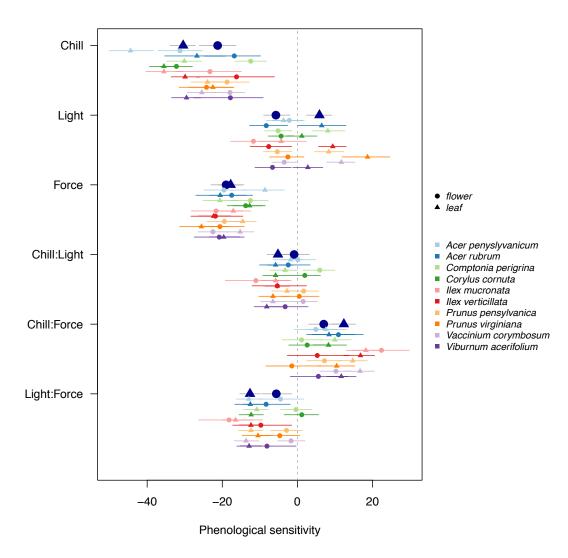


Figure 2: Effects of forcing temperature, chilling duration, and photoperiod on the leaf (circles) and flower (triangles) phenology of 10 temperate woody plant species collected from Harvard Forest (Petersham, MA, USA). Shapes indicate mean estimates and lines depict 50% credible intervals (See Tab. S3, Tab. S4 for other intervals) from Bayesian hierarchical models with advances in phenology shown as negative numbers, and delays in phenology as positive numbers. Flower and leaf phenology differs in sensitivity ( $\Delta$  day of phenological event/ $\Delta$  environmentals cue) to these environmental cues.

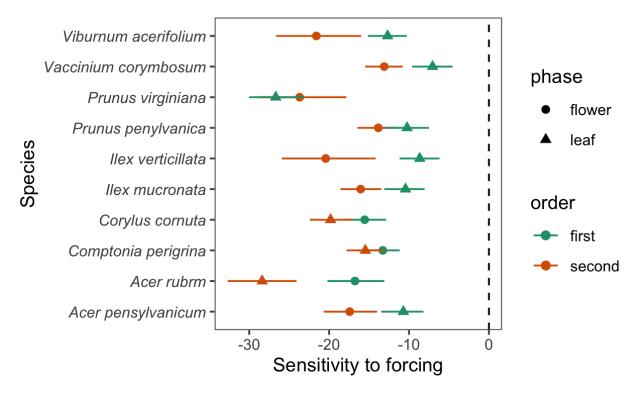


Figure 3: Phenological sensitivity ( $\Delta$  phenological event/  $\Delta$  C°) to forcing temperatures of leaf (circles) and flower (triangles) phenology from 10 temperate deciduous woody plants at long (12 hour) photoperiod and long chilling duration treatments (8 weeks at 4°C). Shapes indicate mean estimates and lines depict 50% credible intervals (See Tab. S5 for other intervals) from Bayesian hierarchical models with advances in phenology shown as negative numbers. When photoperiod and chilling are high, most species follows the predicted pattern of the forcing hierarchy hypothesis (FHH), with the second phenophase of the sequence consistently more sensitive to forcing than the first. This result suggests that the FHH should be considered a special case of the differential sensitivity hypothesis (DSH) that occurs when the chilling and photoperiod requirements are met for both tissue types.

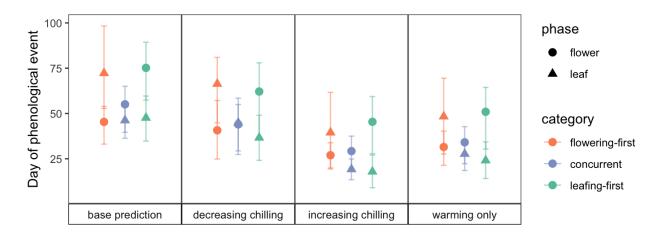


Figure 4: Projected shifts in flower-leaf sequences under current environmental conditions (base prediction) and three climate change scenarios predict that FLS differ among the three major FLS types and will be strongest is flowering-first species. Predictions are based on species-level posterior estimates grouped by FLS category (flowering-first, concurrent, leafing-first) from Bayesian hierarchical models comparing flower (circles) and leaf (triangles) phenological responses to variable chilling duration and forcing temperatures. Shapes represent the mean estimates and lines represent the 50% credible intervals.