- Ecological drivers of flower-leaf sequences: aridity and floral traits select for flowering-first in the American Plums
- D.M. Buonaiuto ^{1,2,3,a}, T.J. Davies ^{4,5}, S. Collins ⁴ & E.M. Wolkovich ^{2,3,4}
- 4 Author affiliations:
- ⁵ Department of Environmental Conservation, University of Massachusetts, Amherst, Massachusetts, USA.
- 6 ORCID: 0000-0003-4022-2591
- ²Arnold Arboretum of Harvard University, Boston, Massachusetts, USA.
- ³Department of Organismic and Evolutionary Biology, Harvard University, Cambridge, Massachusetts, USA
- ⁹ Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, Vancouver, British
- 10 Columbia, Canada
- Department of Botany, University of British Columbia, Vancouver, British Columbia, Canada
- ¹² Corresponding author: 617.823.0687; dbuonaiuto@umass.edu
- Word Count: Introduction 944:, Materials and Methods: 1722, Results: 501, Discussion:1394, Total: 4561
- Figures: 4

16

17 Summary

- Across temperate forests many tree species produce flowers before their leaves emerge. This flower-leaf phenological sequence, known as hysteranthy, is generally described as an adaptation for wind-pollination. However, this explanation does not address why hysteranthy is also common in biotically-pollinated taxa.
- We quantified flower-leaf sequence variation in the American plums (*Prunus*, subspp. *Prunus* sect.

 Prunocerasus), a clade of insect-pollinated trees, using herbaria specimens and Bayesian hierarchical modeling. We tested two common, but rarely interrogated hypotheses—that hysteranthy confers aridity tolerance and/or pollinator visibility—by modeling the associations between hysteranthy and related traits. To understand how these phenology-trait associations were sensitive to taxonomic scale and flower-leaf sequence classification, we then extended these analyses to all *Prunus* species in North America.
- Our findings across two taxonomic levels support the hypotheses that hysteranthy may help temporally partition hydraulic demand to reduce water stress, and increase pollinator visibility and thereby reduce selective pressure on inflorescence size.
- Our results provide foundational insights into the evolution of flower-leaf sequences in the genus *Prunus*, with implications for understanding these patterns in biotically-pollinated plants in general. Our approach suggests a path to advance these hypotheses to other clades, but teasing out drivers fully will require new experiments.
- Keywords: Deciduous forests, Flower-leaf sequences, Hysteranthy, Phenology, Plant hydraulics, Pollination,
 Phylogeny

38 Introduction

- ³⁹ Woody perennials are among a subset of plant types with the unique ability to seasonally begin reproduction
- 40 prior to vegetative growth. This flowering-first phenological sequence, known as hysteranthy, proteranthy or
- precocious flowering, is apparent in temperate deciduous forests around the globe (Rathcke & Lacey, 1985).
- A number of studies suggest that this flower-leaf sequence is under selection, and that hysteranthy can confer
- performance advantages (Guo et al., 2014; Gougherty & Gougherty, 2018; Buonaiuto et al., 2021), but the
- 44 importance of variation in flower-leaf sequences for maintaining fitness may vary across functional types, taxa
- and biomes.
- 46 The most common, and well-tested explanation for the evolution of hysteranthy in temperate forests is that it
- 47 is adaptive for wind-pollination, as leafless canopies increase wind speeds for pollen transport and reduce the
- 48 likelihood of pollen interception by vegetation (Whitehead, 1969; Niklas, 1985). However, this explanation
- does not address the widespread prevalence of hysteranthy in biotically-pollinated taxa found in temperate
- 50 regions. This number is not trivial; a recent analysis found that approximately 20% of the hysteranthy species
- in Eastern Temperate Forests of North America are biotically-pollinated (Buonaiuto et al., 2021).
- 52 Alternative hypotheses have been put forward to explain the advantage of hysteranthy in biotically-pollinated
- 53 species, but they have not been widely evaluated in the literature. Below, we briefly review these hypotheses
- and their predictions, then test their predictions using the American plums (*Prunus* subspp. *Prunus* sect.
- 55 Prunocerasus)—a widespread clade with high variability in flower-leaf sequences, as a case-study. Our study
- both clarifies the hypothesized function of flower-leaf sequence variation in the genus *Prunus* and lays the
- 57 groundwork for understanding the origins of flower-leaf sequence variation in biotically-pollinated taxa more
- 58 generally.

59 Hypotheses of hysteranthous flowering in biotically-pollinated taxa

- 60 Water limitation hypothesis: In the dry-deciduous tropics of South and Central America, hysteranthy is
- 61 common (Rathcke & Lacey, 1985; Franklin, 2016), and is regarded as an important adaptation to alleviate
- ₆₂ water stress by partitioning the hydraulic demand of flowers and leaves across the season (Borchert, 1983;
- Reich & Borchert, 1984; Franklin, 2016; Gougherty & Gougherty, 2018). Under this hypothesis, the function
- of hysteranthous flowering in temperate regions parallels that in the dry tropics. While temperate forests are
- rarely water-limited in the early season during which flowering and leafing occur (Polgar & Primack, 2011),
- there is still considerable variation in water availability in space and time within temperate regions of the
- 67 globe. With this hypothesis, we would expect to find hysteranthous taxa in locations that are, on average,
- drier than their non-hysteranthous relatives.
- 69 Insect visibility hypothesis: Hysteranthous flowers are visually conspicuous in the landscape. Thus, as in
- wind-pollinated taxa, hysteranthy in biotically-pollinated taxa may be an adaptation for pollination efficiency
- as flowering-first species are easier for insect pollinators to locate (Janzen, 1967). A challenge to evaluating this

hypothesis is that correlated selection between flower-leaf sequences and pollinator visibility could have either
a positive or negative relationship depending on the pollination environment. In one scenario, hysteranthy
may be associated with smaller floral displays: because flowers are not obscured by leaves, they are easier
to see and there is weaker selection for increasing floral display size. In an alternative scenario, hysteranthy
could be associated with larger floral displays, especially in environments where plants are more often pollenlimited and selection may favor both hysteranthy and increased floral display size to augment floral attraction
to visual pollinators.

In contrast to these functional hypotheses, hysteranthous flowering could simply be a by-product of selection for early flowering. Species that flower before their leaves inherently flower early in the season. For example, fruit development or dispersal constraints may drive early flowering (Primack, 1987), and because spring flower phenology is less constrained by prior phenological events than leaf phenology (Ettinger *et al.*, 2018; Savage, 2019), this selection for early flowering could incidentally produce the hysteranthous phenological sequence. Here, there is no specific adaptive advantage to hysteranthy; selection is not operating on the relative timing of flower and leaf emergence, but rather the absolute flowering time alone. Rejection of the above functional hypotheses might provide support to this null explanation.

A significant challenge for robust testing of hysteranthy hypotheses is that most characterizations of flowerleaf phenological sequences are based on expert-opinion verbal descriptions (e.g. "flowers before leaves" or
"flower before/with leaves"), which make comparisons across taxa, time and space difficult and sensitive
to observer bias (see Buonaiuto et al., 2021). This problem can be overcome by adopting standardized
quantitative measures of plant phenology for observational studies and applying them to historical data
records. Herbarium records are an excellent source of data that can be leveraged for quantitative phenological
measurements (Willis et al., 2017), but have not been widely used to investigate variability of flower-leaf
sequences.

The American plums are useful model clade to investigate drivers of hysteranthous flowering in bioticallypollinated species. The species that make up this group are distributed across the temperate zone of North
America and, like the genus *Prunus* more generally show pronounced inter-specific variation in flower-leaf
sequences. Usefully, species in this clade are well represented in herbaria records (Fig. 1a), making them a
tractable group to measure and assess variation in flower-leaf sequences.

To interrogate the functional hypotheses for hysteranthous flowering described above, we used herbaria records to quantify variation in flower-leaf sequences of the American plums. Then we combined environmental attributes, biological traits and phylogenetic data in statistical models designed to evaluate whether the observed associations between flower-leaf sequences and morphological and environmental traits match the predicted associations of the hysteranthy hypotheses. Finally, we compared our findings in this clade to patterns observed in larger genus *Prunus* to test whether these phenology-trait associations were sensitive to taxonomic scale and flower-leaf sequence classification.

of Materials and Methods

Quantifying flower-leaf sequence variation

We obtained digital herbarium specimens of the American plums from the Consortium of Midwest Herbaria 109 (CMH) Database (of Midwest Herbaria, 2023). Specimen collection dates ranged from 1844-2020, with the 110 majority collected between 1950-2000. To quantify flower-leaf sequence variation in this group we randomly 111 sampled 200 specimens for each species and scored the phenological development of flowers and leaves; we used a modified BBCH scale for woody plants designed to evaluate vegetative and reproductive phenological 113 progress through a standardized quantitative index (Finn et al., 2007). For species with less than 200 114 specimens in the collection, we included all available specimens. In total, we evaluated the phenology of 2521 115 specimens, but only specimens with visible flowers were included in this analysis. We also removed specimens 116 with flowering dates that were major outliers from the observed flowering period of each species. We removed outliers visually, and by excluding observations that were beyond three standard deviations of the median 118 flowering time for each species (n=9). Our final analyses included 1000 specimens (see Tab. S1 for number 119 of observations/species). 120

We reconstructed the phylogenetic relationships among species in this group based on the tree topology in Shaw & Small (2004). We inferred branch lengths following the method of Grafen & Hamilton (1989) in which node heights are estimated in proportion to number of subtending taxa using the R package "ape" (E. Paradis and K. Schliep, 2019).

To quantify flower-leaf sequence variation, we fit an ordinal, hierarchical, Bayesian phylogenetic mixed model 125 (de Villemeruil P. & Nakagawa, 2014) designed to assess the likelihood an individual would be at any given 126 vegetative BBCH phase while flowering. Our model predicted leaf stage $(y_i, \text{ ordinal}, \text{ with six categories})$ 127 representing stage from 1 for "buds closed" and 6 for "leaf expansion complete") as a function of species and additional phylogenetic effects. Because hysteranthy co-varies with flowering time (i.e., flowering first species will generally flower earlier than other species, on average), and collection dates were not evenly distributed 130 across the flowering season (see Fig. S1), we included day of year of observation as an additional predictor. 131 Additionally, because climate change could affect the interval between flowering and leafout over the course 132 of our time series, we included the year of collection of each specimen as a covariate. Following previous 133 conventions for modeling the possible effects of climate change on spring phenology, we parameterized year as a hinge variable, using 1980 as a break point (Stocker et al., 2013; Buonaiuto et al., 2021). 135

136 The model is written below:

137

$$y_i = \begin{cases} 1 & if \quad z_i < 0 \\ 2 & if \quad z_i \in (0, c_2) \\ 3 & if \quad z_i \in (c_2, c_3) \\ 4 & if \quad z_i \in (c_3, c_4) \\ 5 & if \quad z_i \in (c_4, c_5) \\ 6 & if \quad z_i > c_5 \end{cases}$$

141

148

150

138
$$z_i = \alpha + \alpha_{phylo} + \alpha_{sp} + \beta_{\rm day\ of\ year[sp]} * X_{\rm day\ of\ year} + \beta_{year} * X_{year} + \epsilon_i$$
139
$$\epsilon_i \sim logistic(0,1)$$

where y_i is the ordinal outcome (leaf stage; as 1,2,...6 categories). $c_{2...5}$ are the estimated cutpoints between leaf stages on the logit scale and year is: (the year the specimen was collected - 1980). z_i is the linear component of the underlying latent variable model.

 α describes an intercept for each category [1,2,...6] and slopes ($\beta_{\rm day\ of\ year}$ and β_{year}) are constant across cutpoints. $\beta_{\rm day\ of\ year}$ also varies among *species* while β_{year} is a pooled estimate across species.

The influence of the phylogeny (α_{phylo}) was modeled as:

$$\alpha_{phylo} \sim \text{normal}(0, COR[\sigma_{phylo}^2])$$

The α for species effects independent of the phylogeny was modeled as:

$$\alpha_{sp} \sim \text{normal}(0, \sigma_{species}^2)$$

We used our model to predict the probability that each species would be observed at a given vegetative 151 BBCH stage during flowering for each day of the flowering period of each species by extracting 1000 ran-152 dom draws from the posterior distribution. Next, for each day of the flowering season, we summed the 153 predicted likelihood that species would be at BBCH 0 ("bud closed"), BBCH 07/09 ("bud break") or BBCH 11 ("start of leaf unfolding) vs. BBCH 15 ("leaf unfolding"), BBCH 17 ("most leaves unfolded"), BBCH 19 155 ("leaf expansion complete")—this allowed us to quantify the likelihood that a species would be hysteranthous 156 or non-hysteranthy respectively for each day of the season. We chose the BBCH 11/BBCH 15 boundary to 157 define hysteranthous flowering because this is the earliest point in development when most leaves are unfurled enough to visually obscure flowers and transpire. Finally, we used these estimates to develop a flower-leaf sequence index: for this, we summed the likelihood of hysteranthy vs. non-hysteranthy across the full flower-160 ing period of each species, with 0 being never hysteranthous and 1 being always hysteranthous. To evaluate 161 the sensitivity of our model to choice of cutoff, we also calculated a hysteranthy index using an alternative 162

cutoff at the BBCH 09/BBCH 11, which did not alter the species' ranks on the index (see Tab. S2).

To better understand how within-season dynamics affected our inference, we also refit our model excluding day of year as a predictor. This version of the model did not substantially alter the species' ranks on the index or our inference about the relationships between flower-leaf sequence variation and the trait representing the main hysteranthy hypotheses (Tab. S2, Tab. S3).

Evaluating hysteranthy hypotheses

To test the hypotheses of hysteranthy, we first recorded petal length measurements directly from herbarium specimens. For these morphological measurements, we sampled 321 specimens and measured the petal length of up to 10 randomly selected petals per specimen (n=2757) using ImageJ image processing software (see Tab. S2, for n per species).

To assess aridity tolerance, we computed the average Palmer Modified Drought Index score from June-August (hereafter: PDSI), obtained from Gille *et al.* (2017) for every *Prunocerasus* specimen in the database (n=2305, see Tab. S2, for n per species). PDSI is a unitless, standardized drought index that integrates temperature and precipitation data to estimate relative dryness in time and space (Heim, 2002). Negative PDSI values indicate more arid conditions and positive values wetter conditions. For any specimens that lacked accurate geo-location information, we extracted PDSI values at the county centroid of the herbaria specimen.

Because all of our measurements were made on different individuals—with different sample sizes—we used two different modeling approaches to test the relationship between flower-leaf sequence index scores, aridity tolerance and floral displays.

First we computed species-level means of PDSI and petal length and used a beta regression to evaluate the relationship between flower-leaf sequences, PDSI, petal length and their interaction. We standardized the units of all predictors through z-scoring (Gellman & Hill, 2007) to make their effect size estimates directly comparable within the following model structure:

187
$$y_i = (\mu, \mu(1-\mu)/(1+\phi)$$

186

188

192

where μ and ϕ are the two shape parameters of the beta regression. Due to the limited sample size of this analysis (13 species), we only modeled the effect of our predictors on the mean parameter μ and fit a grand intercept for the precision parameter ϕ . We modeled the μ parameter as:

$$\mu = \alpha + \beta_{PDSI} * \overline{X_{PDSI}} + \beta_{\text{petal length}} * \overline{X_{\text{petal length}}} + \beta_{PDSI_x \text{petal length}} * (\overline{X_{PDSI}}) (\overline{X_{\text{petal length}}})$$

We chose this model structure because it allowed us to assess the additive and interactive effects of PDSI and

petal size on flower-leaf sequences. However, by using mean trait values as predictors, we could not incorporate within-species variation in these trait/environmental predictors or account for their phylogenetic structure.

To understand how these factors affected our inferences about the relationship between flower-leaf sequences and traits, we fit two additional models to estimate relationship between flower-leaf sequences index values and PDSI, and between flower-leaf sequences index values and petal size separately which included the intraspecific variation and phylogenetic structure of each of these traits (see Supporting Information: Extended Methods for details). This alternative modeling approach produced similar results about the phenology-trait relationships investigated in our main model.

Hysteranthy in the larger genus Prunus

To better understand how the patterns we identified in the American Plums clade scaled across coarser taxonomic resolution, we also evaluated the relationship between hysteranthous flowering and hypothesisrelated traits for additional *Prunus* species native to, or established in, North America (n=32). For this
analysis, we obtained categorical descriptions of flower-leaf sequences and mean estimates of the number of
flowers per inflorescence as a proxy for floral investment from the <u>Flora of North America</u> (Rohrer, 1993+).
We extracted PDSI values for all herbaria observations of those species in the Consortium of Midwest Herbaria
database (n=23,272) as described above.

To account for the influence of evolutionary relationships among species, we reconstructed the phylogenetic relationships in the genus based on the tree topology in Chin *et al.* (2014). As above, we computed branch lengths with the R package "ape" (E. Paradis and K. Schliep, 2019).

As above, we standardized the units of all predictors through z-scoring (Gellman & Hill, 2007). The model structure is:

$$y_i = \begin{cases} 1 & if \quad z_i < 0 \\ 2 & if \quad z_i \in (0, c_2) \\ 3 & if \quad z_i \in (c_2, c_3) \\ 4 & if \quad z_i > c_3 \end{cases}$$

 $z_i = \alpha + \alpha_{phylo} + \beta_{PDSI} * X_{PDSI} + \beta_{\text{floral investment}} * X_{\text{flowers/inflorescence}} + \beta_{PDSI_x \text{floral investment}} (X_{PDSI}) (X_{\text{flowers/inflorescence}}) + \epsilon_i$

 $\epsilon_i \sim logistic(0,1)$

219

where y_i is the ordinal outcome of flower-leaf sequence category ("flowers after leaves"=1, "flowers with leaves"=2, "flowers before/with leaves"=3 and "flowers before leaves"=4) and $c_{2...3}$ are the estimated cutpoints between categories on the logit scale. z_i is the linear component of the underlying latent variable model. α

describes a grand intercept, and we modeled the influence of phylogeny (α_{phylo}) as above. Note that this model includes four ordinal categories while our model of the American Plums clade included six, due to the different underlying structures of the two datasets.

$_{26}$ Model runs

We fit all models in the R package "brms" (Bürkner, 2018) using weakly informative priors, and four chains. For the models aimed at "Quantifying flower-leaf sequence variation" and "Evaluating hysteranthy hypotheses" in the American plums, we ran the models with a warm-up of 3000 iterations, and 4000, and 5000 sampling iterations respectively, for a total of 4000 and 8000 sampling iterations across all chains. For the "Hysteranthy in the larger genus Prunus" model, we used a warm up of 6,000 iterations and 8,000 sampling iterations for a total of 8000 sampling iterations to maximize the effective sampling size. Model fits were assessed with \hat{R} <1.01, high effective sample sizes, and no divergent transitions. We provide mean estimates with uncertainty intervals in-text, and 50% and 89% intervals for all figures and tables.

35 Results

Quantifying flower leaf sequences in the American plums

We found substantial inter-specific differences in flower-leaf sequences within the American plums (Fig. 1b, Fig. 2a). Several species (*P. mexicana*, *P. umbellata*, *P. angustifolia*, *P. maritima* and *P. gracilis*) were most likely to be hysteranthous for all—or most—of their flower period, while for others, (*P. americana*, *P. munsoniana*, *P. alleghaniensis*, *P. nigra*, *P. hortulana*, *P. texana* and *P. rivularis*), hysteranthous flowering was only likely in the early part of their flowering session. One species, *P. subcordata*, was unlikely to be hysteranthous at any point in its flowering period (Fig. 2a). These relative ranking of species' hysteranthy likelihoods were consistent with our alternative method for constructing the hysteranthy index (Tab. S2, Fig. S2).

Across all species of American Plums, day of year increased the likelihood of flowering during a later vegetative phenological stage (Fig. 2b). Year of observation did not substantially impact the likelihood of hysteranthy for this taxonomic group (Fig. 2b).

Associations between hysteranthy and environmental and morphological traits

In the American plums, predominately hysteranthous species had marginally smaller flowers and occurred in historically drier localities than species with more overlap between flowers and leaves (i.e., increased likelihood of hysteranthy was negatively associated with PDSI and petal length without a substantial interaction between them, Fig. 3a; parameter estimates from this model were β_{PDSI} : $-0.47, UI_{89}[-0.96, 0.01], \beta_{petal length}$: $-0.14, UI_{89}[-0.54, 0.24]$ $\beta_{PDSI_x petal length}: -0.14, UI_{89}[-0.91, 0.65])$. These estimates were comparable to estimates from models where we treated each predictor separately and accounted for phylogeny (Fig. S3), and where we used the hysteranthy index derived from models that did not include day of year as a predictor (Tab. S3). The direction and magnitude of the estimated effects support the predictors of the water-limitation hypothesis and marginally support the predictions of the insect-visibility hypothesis.

In the larger genus Prunus, hysteranthous species had smaller inflorescences and were found in drier locations (Fig. 4a, b; i.e., there was a negative association between hysteranthy and PDSI and number of flowers per inflorescence, as well as a substantial negative interaction between them, parameter estimates from this model were β_{PDSI} : $-8.0, UI_{89}[-16.6, -2.44], \beta_{flowers/inflorescence}$: $-15.5, UI_{89}[-31.46, -5.56]$ and $\beta_{PDSIxflowers/inflorescence}$: $-13.06, UI_{89}[-28.53, -2.93]$). The direction and magnitude of the estimated effects support the predictors of both the water-limitation hypothesis and the insect-visibility hypothesis.

The estimated effects of floral traits and their interactions with PDSI were stronger in the larger genus *Prunus*than in the American plums clade. This is not surprising given that all species in the American plums clade
have solitary flowers, making the variation in floral display size highly constrained. By contrast, *Prunus*species included in our secondary analysis include those with solitary flowers and species with as many as 100
flowers per inflorescence—substantially more variation in both floral investment and in hydraulic demand.
This suggests that the correlated selection between flower-leaf sequences and these floral traits may be more
pronounced at coarser taxonomic resolutions, where underlying trait variation is greater.

Discussion Discussion

Using North American *Prunus* species as a case study, our analyses indicate that flower-leaf sequences are likely under selection. We show that variation in flower-leaf sequences across species may reflect adaptive tradeoffs between a) the timing of investment in reproduction relative to the timing of resumption of carbon acquisition through leafout, and b) other aspects of plant performance, such as environmental tolerance and pollinator attraction strategies that we investigated in this study. We show that hysteranthous flowering is associated with historic aridity (PDSI) and smaller flower displays in both the American plums, and more broadly across *Prunus* species native to, or established in North America. The relationships between hysteranthy and aridity, and hysteranthy and floral display size support the predictions of the water limitation hypothesis and the insect visibility hypothesis.

Our models estimated a strong relationship between aridity (PDSI) and flower-leaf sequences at both taxonomic scales we studied, but the relationship between floral display size and flower-leaf sequences predicted by the insect visibility was better supported at the coarser taxonomic scale of the full genus *Prunus* than in the American Plums clade. While the flowers of American Plums clade are solitary, they are still clustered on branches, so it is unlikely the unit of attraction that pollinators are responding to is the individual flower, which may explain why the relationships between hysteranthy and petal size we observed were weak (Fig.

3b). The inflorescences of the larger genus *Prunus*, are probably a better proxy for differences in units of attraction, which may explain their stronger association with flower-leaf sequences variation among species (Fig 4). This contrast may suggest that associated selection between flower-leaf sequences and pollinator traits has more strongly influenced inflorescence architecture than the morphology of individual flowers, our estimates at both scales agreed in directionality (i.e., hysteranthy associated with smaller floral displays).

Under the insect visibility hypothesis, floral display size could either be positively or negatively associated with hysteranthy depending on the pollination environment. The association between hysteranthy and smaller flower displays we found supports the prediction that increased visibility of hysteranthous flowers reduces selection pressure on flower display size. These results fit with other comparative anatomy studies in plants that report hysteranthous species typically have smaller inflorescences than non-hysteranthous relatives (Gunatilleke & Gunatilleke, 1984), and studies on pollinator foraging behavior that suggest the presence of leaves substantially alters the visual perception of pollinators (Rivest et al., 2017; Forrest & Thomson, 2009).

Our support for both the water limitation hypothesis and insect visibility hypothesis (especially in the larger 299 genus Prunus), and the strong positive interactions between PDSI and floral investment that we observed in the larger genus *Prunus* highlight that these hypotheses are not mutually exclusive, and could be related. 301 Selection on floral size represents a classic evolutionary tradeoff where larger floral displays may generally be more effective for attracting pollinators but demand more resources, including water, to maintain turgor 303 and reproductive function than smaller ones (Galen et al., 1999; Lambrecht & Dawson, 2007). With this trade-off, reproductive displays are often small in harsher environments (Lambrecht, 2013; Teixido et al., 2016), and hysteranthy could represent a compensatory mechanism that both reduces hydraulic demand 306 while increasing pollination efficiency in these environments. Studies that have compared the transpiration 307 rates among flowers and leaves provide insights to the potential importance of hysteranthy for maintaining 308 water status. Measurements of water movement (transpiration rates, sap flow, hydraulic conductivity) to flowers range from 20%-60% of that of leaves under comparable conditions (Whiley et al., 1988; Roddy & Dawson, 2012; Liu et al., 2017; McMann et al., 2022). This level of additional hydraulic demand can drive 311 loss of stomatal conductance and decrease photosynthetic rates (Galen et al., 1999). 312

The 2000+ year of PDSI records we incorporated in this study offer additional insights into why the water limitation hypothesis may still be relevant to temperate forest regions that are typically well saturated in the spring. These records reveal that the southeastern United States (the region in which much of our data originates) was once much drier than it has been in contemporary times (Cook *et al.*, 2010, Fig S4), suggesting that the spatial patterns of hysteranthy we see in the American plums may represent a lagged-correlation with historical climate conditions, rather than a pattern driven by current precipitation patterns.

Despite this evidence that hysteranthy can reduce hydraulic demand in dry environments, hysteranthous species in the American plum clade are not found—even historically—in extremely arid locations (PDSI values typically range from -4 to 4, although the values that we observed in our analyses were more restricted, ranging from -0.5 to 0.2). This contrasts with hysteranthous species in the dry tropics where this phenological pattern appears to allow them to tolerate more extreme aridity (Franklin, 2016). But the flower-leaf sequences
of the hysteranthous species in our study were markedly different from patterns of hysteranthy in these drytropics where the water limitation hypothesis was initially proposed. While flowering can precede leafout
by as much several weeks for species in the American plums, the process of fruit development, which is also
water intensive, occurs when leaves are present. By contrast, in the dry tropics hysteranthous flowering is
initiated at the time of leaf drop (Borchert, 1983; Franklin, 2016); thus, the full reproductive cycle occurs in
the leafless period. The comparatively small window of leafless reproductive development in our temperate
clade suggests that hysteranthy may allow temperate species to occupy marginally drier environments than
non-hysteranthous species, but may not facilitate species' persistence under extreme aridity.

Inter-and intra-specific variation in flower-leaf sequences

We developed a novel approach to assessing flower-leaf sequences that scales from quantitative, individuallevel observations to species-level characterizations. With this approach, we were able to—for the first
time—quantitatively assess intermediate cases of hysteranthy (such as those that are typically described as
"flowers before/with leaves"). Previous studies of hysteranthous flowering have either excluded these cases
from their analyses (e.g.; Gougherty & Gougherty, 2018) or binned them with the well defined cases (e.g.;
Buonaiuto et al., 2021). We found that many American plum species expressed this intermediate flower-leaf
sequence. Further, while our classifications broadly matched previous species-level analyses in this group by
Shaw & Small (2004), our approach identified substantial differences in flower-leaf sequences among these
intermediate cases (Fig. 1b), which allowed us to assess the trait associations with this phenotype.

Our quantitative analysis of the American plums clade revealed that flower-leaf sequences—often described 342 as a species-level trait—are highly variable within species (Fig. 2a). For almost all members of the clade, hysteranthy was strongly predicted by the day of the observation ("day of year" in our model, Fig. 2b). In many cases there was a high likelihood that individuals of a species may be observed at different vegetative 345 stages during flowering (Fig. 2a, Fig. S2). The variation we observed here could either suggest high levels of 346 local adaptation in flower-leaf sequences or, alternatively, high levels of plasticity through which flower-leaf 347 sequences respond to interannual variation in environmental conditions. Because our study was based on herbaria records collected on different individuals across space and time without repeat sampling, we could not robustly estimate how much flower-leaf sequences vary within vs. among species. However, this would be 350 an important next step for understanding how the environment and species interactions have shaped these 351 phenological patterns. 352

Interestingly, while there is substantial evidence that both flowering and leaf phenology have advanced over the last several decades in response to anthropogenic climate change (Menzel et al., 2006; Cleland et al., 2007; Augspurger & Zaya, 2020), we did not observe changes in flower-leaf sequences over that time scale in our dataset (Fig. 2b). This supports a recent finding that despite changes in both flowering and leafout, the time interval between them has remained relatively stable (Guo et al., 2023), but does not preclude that possibility that these the sequences will eventually be disrupted as climate change continues to become more extreme in the future (Buonaiuto & Wolkovich, 2021).

Future directions

We focused on a well-studied, and economically important clade of morphologically similar species. Our case-study provides a road map for evaluating the role of hysteranthy more generally in temperate biotically-pollinated plant taxa (other groups with high interspecific flower-leaf sequence variation include *Magnolia*, *Rhododendron*, *Acer* and *Cornus*), and more broadly across taxa and biomes.

Combining the observational approach with novel experiments could further advance our collective under-365 standing of the adaptive significance of flower-leaf sequences. To test the water-limitation hypothesis, researchers could plant sister-taxa with contrasting flower-leaf sequences in common environments across a 367 gradient of aridity, and evaluate their performance. To test the insect visibility hypothesis, researchers should consider hysteranthy—and phenology in general—in the broader framework of tradeoffs in pollination biology. The tradeoff between phenology and pollination investment could not only consider flower size, but also the number of flowers, nectar and pollen reward investment, volatiles between related hysteranthous and 371 non-hysteranthous taxa. Findings that hysteranthous species invest fewer resources into these other pollinator 372 attraction traits than non-hysteranthous relatives would support the insect visibility hypothesis. For a simple experiment to test the pollinator visibility hypothesis, researchers could force hysteranthy/non-hysteranthy phenotypes for the same genotype using environmental cues, and systematically release pollinators to ob-375 serve their preferences, search times and foraging behavior. If pollinators are more readily drawn to the hysteranthous individuals, it would support hysteranthy as an adaptive trait for pollinator attraction. 377

With a better mechanistic understanding of the relationship between flower-leaf sequences and ecological performance, researchers could then use experiments to assess how differences in floral and leaf physiological responses to temperature variation shape flower-leaf sequences. The measurement and modeling approaches we developed in our observational study can be readily implemented to analyze data from such experimental settings, presenting an important opportunity to unite observations of broad ecological patterns with targeted experimental manipulations to better understand both the evolutionary past and ecological future of flower-leaf sequences.

Competing Interests:

The authors declare no conflict of interest.

387 Author contributions

- DMB, and EMW conceived of the manuscript; DMB and SC collected the data; DMB led the statistical
- analyses with TJD and EMW; DMB led the writing of the manuscript. All authors contributed to writing
- 390 and gave approval for the submission.

391 Data Availability

- The phenology and trait data collected for this study will be made available and archived at KNB: The
- Knowledge Network for Biocomplexity (https://knb.ecoinformatics.org/) at the time of publication.

394 References

- Augspurger CK, Zaya DN. 2020. Concordance of long-term shifts with climate warming varies among phenological events and herbaceous species. *Ecological Monographs*, n/a: e01421.
- Borchert R. 1983. Phenology and control of flowering in tropical trees. Biotropica, 15: 81–89.
- Buonaiuto DM, Morales-Castilla I , Wolkovich EM. 2021. Reconciling competing hypotheses regarding
- flower-leaf sequences in temperate forests for fundamental and global change biology. New Phytologist,
- **229**: 1206–1214.
- Buonaiuto DM, Wolkovich EM. 2021. Differences between flower and leaf phenological responses to envi-
- ronmental variation drive shifts in spring phenological sequences of temperate woody plants. Journal of
- Ecology, **109**: 2922–2933.
- Bürkner PC. 2018. Advanced bayesian multilevel modeling with the r package brms. R Journal, 10: 395–411.
- ⁴⁰⁵ Chin SW, Shaw J, Haberle R, Wen J, Potter D. 2014. Diversification of almonds, peaches, plums and
- cherries –molecular systematics and biogeographic history of prunus (rosaceae). Molecular Phylogenetics
- and Evolution, **76**: 34–48.
- Cleland EE, Chuine I, Menzel A, Mooney HA, Schwartz MD. 2007. Shifting plant phenology in response
- to global change. Trends in Ecology Evolution, 22: 357 365.
- 410 Cook ER, Seager R, Heim Jr RR, Vose RS, Herweijer C, Woodhouse C. 2010. Megadroughts in north
- america: placing ipcc projections of hydroclimatic change in a long-term palaeoclimate context. Journal
- of Quaternary Science, 25: 48–61.
- E. Paradis and K. Schliep. 2019. ape 5.0: an environment for modern phylogenetics and evolutionary analyses
- in R. Bioinformatics, **35**: 526–528.

- Ettinger A, Gee S, Wolkovich E. 2018. Phenological sequences: how early season events define those that follow. American Journal of Botany, 105.
- Finn GA, Straszewski AE, Peterson V. 2007. A general growth stage key for describing trees and woody plants. Annals of Applied Biology, 151: 127–131.
- Forrest J, Thomson JD. 2009. Background complexity affects colour preference in bumblebees. *Naturwis-*senschaften, 96: 921–925.
- Franklin DC. 2016. Flowering while leafess in the seasonal tropics need not be cued by leaf drop: evidence from the woody genus brachychiton (malvaceae). Plant Ecology and Evolution, 149: 272–279.
- Galen C, Sherry R, Carroll A. 1999. Are flowers physiological sinks or faucets? costs and correlates of water
 use by flowers of polemonium viscosum. *OECOLOGIA*, 118: 461–470.
- Gellman A , Hill J. 2007. Data Analysis Using Regression and Multilevel/Hierarchical Models. Cambridge
 University Press.
- Gille E, Wahl E, Vose R, Cook E. 2017. Noaa/wds paleoclimatology living blended drought atlas (lbda)
 version 2 recalibrated reconstruction of united states summer pmdi over the last 2000 years.
- Gougherty AV, Gougherty SW. 2018. Sequence of flower and leaf emergence in deciduous trees is linked to ecological traits, phylogenetics, and climate. New Phytologist, 220: 121–131.
- Grafen A , Hamilton WD. 1989. The phylogenetic regression. Philosophical Transactions of the Royal Society
 of London. B, Biological Sciences, 326: 119–157.
- Gunatilleke CVS, Gunatilleke IAUN. 1984. Some observations on the reproductive biology of three species of cornus (cornaceae). Journal of the Arnold Arboretum, 65: 419–427.
- Guo L, Liu X, Alatalo JM, Wang C, Xu J, Yu H, Chen J, Yu Q, Peng C, Dai J et al. 2023. Climatic
 drivers and ecological implications of variation in the time interval between leaf-out and flowering. Current
 Biology, 33: 3338-3349.e3.
- Guo L, Luedeling E, Dai J, Xu J. 2014. Differences in heat requirements of flower and leaf buds make
 hysteranthous trees bloom before leaf unfolding. *Plant Diversity and Resources*, **36**: 245–253.
- Heim RR. 2002. A review of twentieth-century drought indices used in the united states. Bulletin of the
 American Meteorological Society, 83: 1149–1166.
- Janzen DH. 1967. Synchronization of sexual reproduction of trees within the dry season in central america.

 Evolution. 21: 620-637.
- Lambrecht SC. 2013. Floral water costs and size variation in the highly selfing leptosiphon bicolor (polemoniaceae). *International Journal of Plant Sciences*, 174: 74–84.

- Lambrecht SC, Dawson TE. 2007. Correlated variation of floral and leaf traits along a moisture availability gradient. *Oecologia*, **151**: 574–583.
- Liu H, Xu QY, Lundgren MR, Ye Q. 2017. Different water relations between flowering and leaf periods: a case
 study in flower-before-leaf-emergence <i>magnolia</i> species. Functional Plant Biology, 44: 1098-1110.
- McMann N, Peichel A, Savage JA. 2022. Early spring flowers rely on xylem hydration but are not limited by stem xylem conductivity. New Phytologist, 233: 838–850.
- ⁴⁵² Menzel A, Sparks TH, Estrella N, Koch E, Aasa A, Ahas R, Alm-Kuebler K, Bissolli P, Braslavska O, Briede
- A et al. 2006. European phenological response to climate change matches the warming pattern. Global
- 454 Change Biology, **12**: 1969–1976.
- of Midwest Herbaria C. 2023. Consortium of midwest herbaria.
- 456 Niklas KJ. 1985. The aerodynamics of wind pollination. The Botanical Review, 51: 328–386.
- Polgar C , Primack R. 2011. Leaf-out phenology of temperate woody plants: From trees to ecosystems. New
 Phytologist, 191: 926-41.
- Primack RB. 1987. Relationships among flowers, fruits, and seeds. Annual Review of Ecology and Systematics,
 18: 409-430.
- Rathcke B, Lacey EP. 1985. Phenological patterns of terrestrial plants. Annual Review of Ecology and

 Systematics, 16: 179–214.
- Reich P, Borchert R. 1984. Water-stress and tree phenology in a tropical dry forest in the lowlands of costa-rica. Journal of Ecology, 72: 61–74.
- Rivest SA, Austen EJ, Forrest JRK. 2017. Foliage affects colour preference in bumblebees (bombus impatiens): a test in a three-dimensional artificial environment. Evolutionary Ecology, 31: 435–446.
- Roddy AB, Dawson TE. 2012. Determining the water dynamics of flowering using miniature sap flow sensors.
 International Society for Horticultural Science (ISHS), Leuven, Belgium, 951, pp. 47–53.
- Rohrer J. 1993+. Prunus. In: Flora of North America North of Mexico (ed. of North America Editorial Committee F.). New York and Oxford.
- Savage JA. 2019. A temporal shift in resource allocation facilitates flowering before leaf out and spring vessel maturation in precocious species. American Journal of Botany, 106: 113–122.
- Shaw J, Small RL. 2004. Addressing the "hardest puzzle in american pomology:" phylogeny of prunus sect.

 prunocerasus (rosaceae) based on seven noncoding chloroplast dna regions. Am J Bot, 91: 985–996.
- Stocker T, Qin D, Plattner GK, Tignor M, Allen S, Boschung J, Nauels A, Xia Y, Bex V, Midgley P.

 2013. Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth

- Assessment Report of the Intergovernmental Panel on Climate Change. IPCC, Cambridge, United Kingdom and New York, NY.
- Teixido AL, Barrio M, Valladares F. 2016. Size matters: Understanding the conflict faced by large flowers in mediterranean environments. *The Botanical Review*, 82: 204–228.
- de Villemeruil P., Nakagawa S. 2014. Modern Phylogenetic Comparative Methods and Their Application in
- Evolutionary Biology, Springer, New York, chap. General quantitative genetic methods for comparative
- biology, pp. pp. 287–303.
- Whiley A, Chapman K, Saranah J. 1988. Water loss by floral structures of avocado (<i>persea americana</i>cana</i>cy. fuerte) during flowering. Australian Journal of Agricultural Research, 39: 457–467.
- Whitehead DR. 1969. Wind pollination in the angiosperms: Evolutionary and environmental considerations.
- Evolution, 23: 28–35.
- Willis CG, Ellwood ER, Primack RB, Davis CC, Pearson KD, Gallinat AS, Yost JM, Nelson G, Mazer SJ,
- Rossington NL et al. 2017. Old plants, new tricks: Phenological research using herbarium specimens.
- 490 Trends in Ecology & Evolution, **32**: 531–546.

Figures Figures

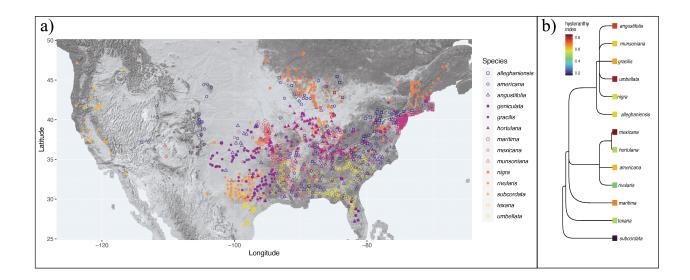


Figure 1: Geographic distribution and taxonomic relationships among the American plums. a) Maps the localities of all the herbaria records used in this study. b) Depicts phylogenetic relationships among the American plums and the likelihood they each species is hysteranthous across its full flowering period, represented by a hysteranthy index where 0 is never hysteranthous and 1 is always hysteranthous. These designations are based on ordinal phylogenetic mixed models. Tree topology is from Shaw & Small (2004)

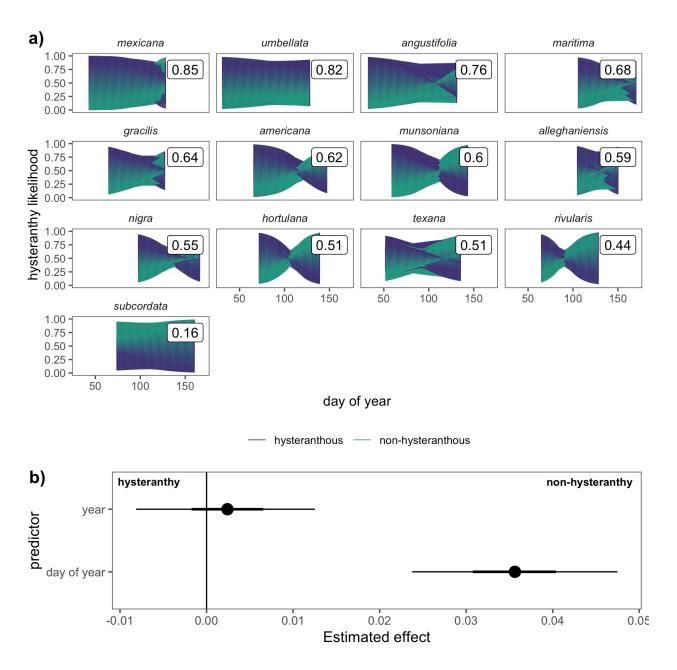


Figure 2: Predicted likelihood of hysteranthy across the flowering period of 13 American plum species and the temporal predictors that drive these patterns. Panel a) depicts the predicted likelihood that each species would express hysteranthy on each day of their flowering season based on 1000 draws from the posterior distribution of Bayesian hierarchical models. The colored shapes represent how the likelihood changes over time and the boxed numerical values represent the average likelihood a species would express hysteranthy, summed across the full flowering period. Panel b) depicts the influence of among season (year of sample) and within season (day of year of sample) trends on the likelihood species would express hysteranthy. Points are the mean effect size estimates, while thick and thin bars represent the 50% and 89% uncertainty intervals respectively.

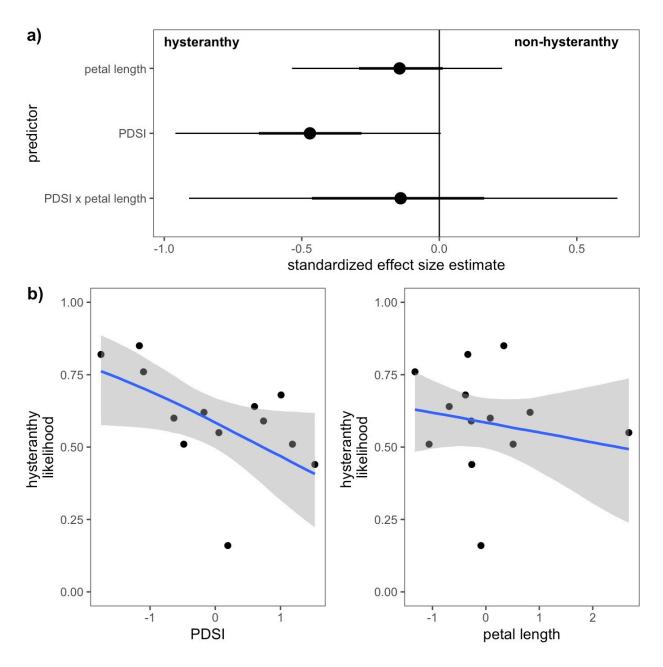


Figure 3: Relationships between hysteranthy and environmental and biological traits for the 13 species of the American Plums. Panel a) shows the estimated effects of each predictor and their interaction on the likelihood hysteranthy. Points indicate the mean effects and the thick and thin bars represent the 50% and 89% uncertainty intervals, respectively. Panel b) depicts the conditional effects of each predictor on hysteranthy likelihood. Blue lines indicate the mean estimate and grey fill the 89% uncertainty intervals. Predictor values (x-axis) have been z-scored to allow direct comparisons between predictors.

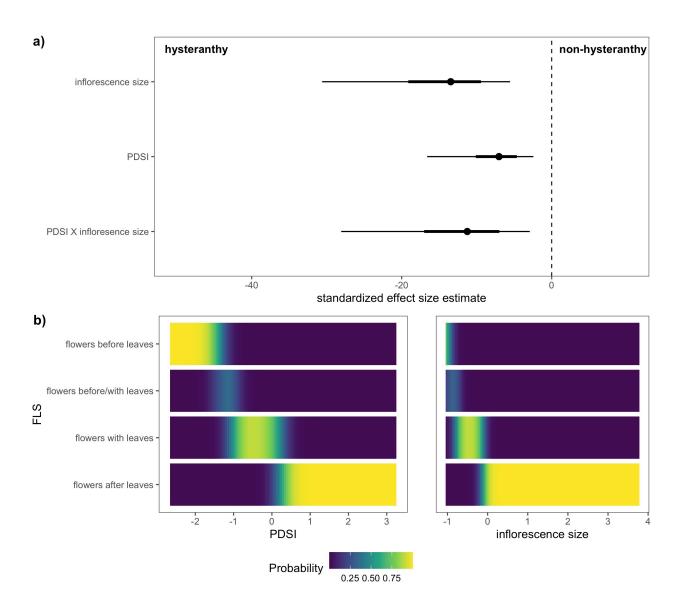


Figure 4: Relationships between the likelihood of hysteranthy and environmental and biological traits for 32 species of the genus Prunus native to, or established in North America. Panel a) shows the estimated effect size of each predictor. Points indicate the mean estimate for each predictor, and thick and thin bars the 50% and 89% uncertainty intervals, respectively. Panel b) depicts the likelihood for each flower-leaf sequence stage (y-axis) at any given values of PDSI or number of flowers/inflorescence (inflorescence size). Predictor values (x-axis) have been z-scored to allow direct comparisons between predictors.