

# Different Alternative Types

David Gerard

2016-05-17

## Abstract

I compare SUCCOTASH to various competitors under the same alternative scenarios in Stephens (2016). SUCCOTASH generally has superior performance in terms of (1) estimating  $\pi_0$ , (2) having higher AUC, and (3) having lower MSE. LEAPP does better under some scenarios in terms of estimating  $\pi_0$  but has far worse AUC and MSE. SUCCOTASH accurately estimates the average sign error rate, though not in as conservative manner as one might hope in some scenarios.

## Simulation Setup

I ran through 200 repetitions of generating data from GTEX muscle data under the following parameter conditions:

- $n \in \{10, 20, 40\}$ ,
- $p = 1000$ .
- $\pi_0 \in \{0.5, 0.9\}$ ,
- The alternative distribution being either spiky, near-normal, flattop, skew, big-normal, or bimodal, where these are the same alternatives defined in Stephens (2016) and the following table. New alternatives are generated every iteration.

Scenario	Alternative Distribution
Spiky	$0.4N(0, 0.25^2) + 0.2N(0, 0.5^2) + 0.2N(0, 1^2), 0.2N(0, 2^2)$
Near Normal	$2/3N(0, 1^2) + 1/3N(0, 2^2)$
Flattop	$(1/7)N(-1.5, .5^2) + N(-1, .5^2) + N(-.5, .5^2) + N(0, .5^2) + N(0.5, .5^2) + N(1.0, .5^2) + N(1.5, .5^2)$
Skew	$(1/4)N(-2, 2^2) + (1/4)N(-1, 1.5^2) + (1/3)N(0, 1^2) + (1/6)N(1, 1^2)$
Big-normal	$N(0, 4^2)$
Bimodal	$0.5N(-2, 1^2) + 0.5N(2, 1^2)$

I extracted the most expressed  $p$  genes from the GTEX muscle data and  $n$  samples are chosen at random. Half of these samples are randomly given the “treatment” label 1, the other half given the “control” label 0. Of the  $p$  genes,  $\pi_0 p$  were chosen to be non-null. Signal was added by a Poisson-thinning approach, where the log-2 fold change was sampled from one of five the alternative models above. That is

$$A_1, \dots, A_{p/2} \sim f \tag{1}$$

$$B_i = 2^{A_i} \text{ for } i = 1, \dots, p/2, \tag{2}$$

where  $f$  is from the table above. If  $A_i > 0$  then we replace  $Y_{[1:(n/2), i]}$  with  $\text{Binom}(Y_{[j, i]}, 1/B_i)$  for  $j = 1, \dots, n/2$ . If  $A_i < 0$  then we replace  $Y_{[(n/2+1):n, i]}$  with  $\text{Binom}(Y_{[j, i]}, B_i)$  for  $j = n/2 + 1, \dots, n$ .

I now describe the justification for this. Suppose that

$$Y_{ij} \sim \text{Poisson}(\lambda_j). \quad (3)$$

Let  $x_i$  be the indicator of treatment vs control for individual  $i$ . Let  $\Omega$  be the set of non-null genes. Let  $Z$  be the new dataset derived via the steps above. That is

$$Z_{ij}|Y_{ij} = \begin{cases} \text{Binom}(Y_{ij}, 2^{A_j x_i}) & \text{if } A_j < 0 \text{ and } j \in \Omega \\ \text{Binom}(Y_{ij}, 2^{-A_j(1-x_i)}) & \text{if } A_j > 0 \text{ and } j \in \Omega \\ Y_{ij} & \text{if } j \notin \Omega. \end{cases} \quad (4)$$

Then

$$Z_{ij}|A_j, A_j < 0, j \in \Omega \sim \text{Poisson}(2^{A_j x_i} \lambda_j) \quad (5)$$

$$Z_{ij}|A_j, A_j > 0, j \in \Omega \sim \text{Poisson}(2^{-A_j(1-x_i)} \lambda_j), \quad (6)$$

and

$$E[\log_2(Z_{ij}) - \log_2(Z_{kj})|A_j, A_j < 0, j \in \Omega] \approx A_j x_i - A_j x_k, \text{ and} \quad (7)$$

$$E[\log_2(Z_{ij}) - \log_2(Z_{kj})|A_j, A_j > 0, j \in \Omega] \approx -A_j(1-x_i) + A_j(1-x_k). \quad (8)$$

if individual  $i$  is in the treatment group and individual  $k$  is in the control group, then this just equals  $A_j$ . I treat the  $A_j$ 's as the true coefficient values when calculating the MSE below.

## Methods

I first normalized the counts by  $\log_2(COUNTS + 1)$ . The number of hidden confounders was estimated using the methods of Buja and Eyuboglu (1992) implemented in the `num.sv()` function in the `sva` package in R.

The confounder adjustment methods I look at in this write-up are:

- OLS + qvalue.
- OLS + ASH
- SUCCOTASH using normal mixtures and heteroscedastic PCA as the factor-analysis method.
- The robust regression version of CATE using PCA as the factor analysis method + qvalue.
- SVA + qvalue.
- Negative control version of CATE using PCA as the factor analysis method + qvalue.
- RUV2 + qvalue.
- RUV4 + qvalue.
- Sparse version of LEAPP. Since this is a sparsity-inducing procedure, I used the proportion of zeros as the estimate of  $\pi_0$ .
- Ridge version of LEAPP + qvalue.

## Results

Note that in the plots below,  $n$  refers to the size of each group, not the total size.

## Estimates of $\pi_0$

- SUCCOTASH has slightly anti-conservative estimates of  $\pi_0$  in the Flat-top and bimodal Scenarios. It does well for every other scenario for larger  $n$ .
- LEAPP does amazingly well in the Big-normal and bimodal scenarios, even for  $n = 10$ . However, it is far too conservative in every other non-null scenario. This seems to indicate that LEAPP functions best if there is a separation of the alternative signal from zero.
- No method using  $q$ -value ever performed as well as succotash. Indeed, none exhibited this “conservative with high-probability” behavior that is desirable.

## Succotash lfsr behavior.

The next set of plots show the relationship between lfsr and the false sign proportion when declaring all genes below said lfsr “significant”. False sign proportion here means the proportion of declared significant genes that have the incorrect sign. The shaded region contains the point-wise 95% bounds on the observed curves.

The same type of plots follow these, except using  $s$ -values instead of lfsr’s. In general, the  $s$ -values for  $n = 20$  or 40 reasonably estimate the average error rate. It plateaus because the maximum  $s$ -value is often near 0.5 when  $\pi_0 = 0.5$  and near 0.9 when  $\pi_0 = 0.9$ . So it might make more sense for me not to include those values above the max  $s$ -value.

## AUC performance.

SUCCOTASH always has higher AUC, even in the bimodal and big-normal scenarios where LEAPP estimated  $\pi_0$  more accurately.

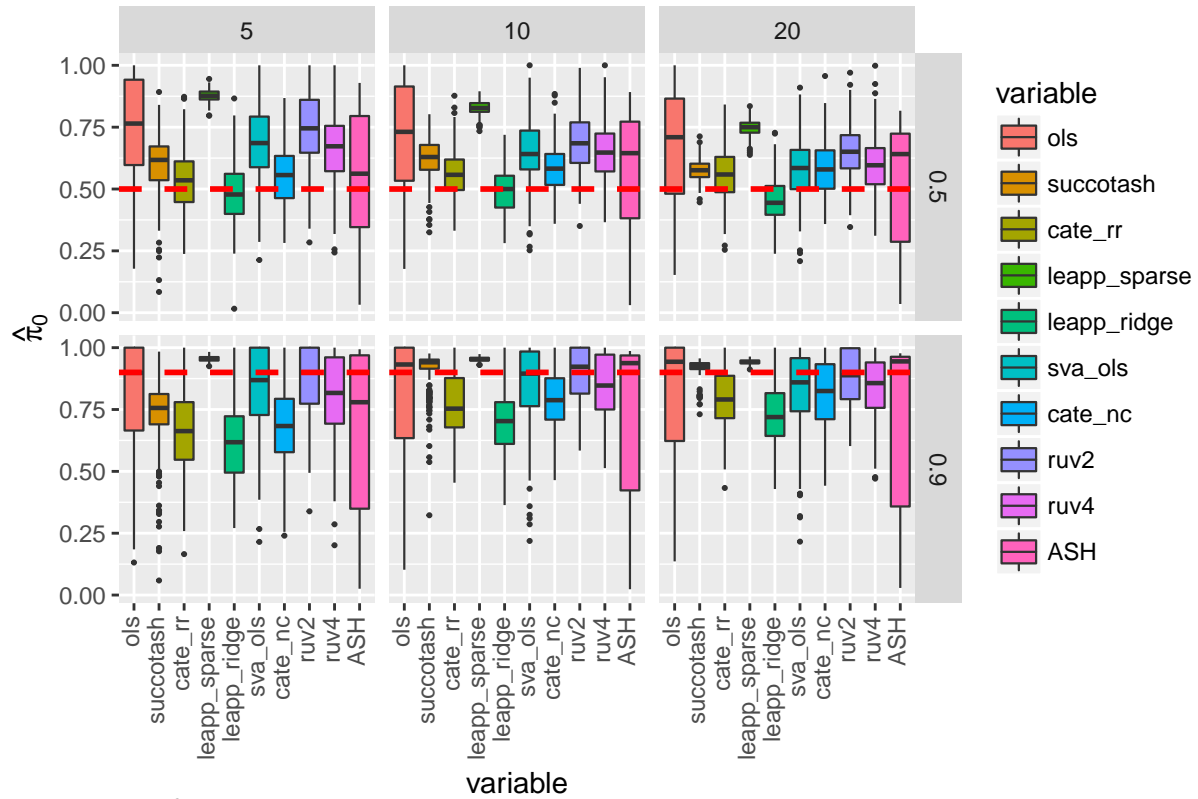
## ROC

I calculated the point-wise mean true positive rate (TPR) and false positive rate (FPR) at each ordered position under each scenario. These are plotted after the AUC boxplots. SUCCOTASH always has higher and further left point-wise average ROC curves, even under the bimodal scenario.

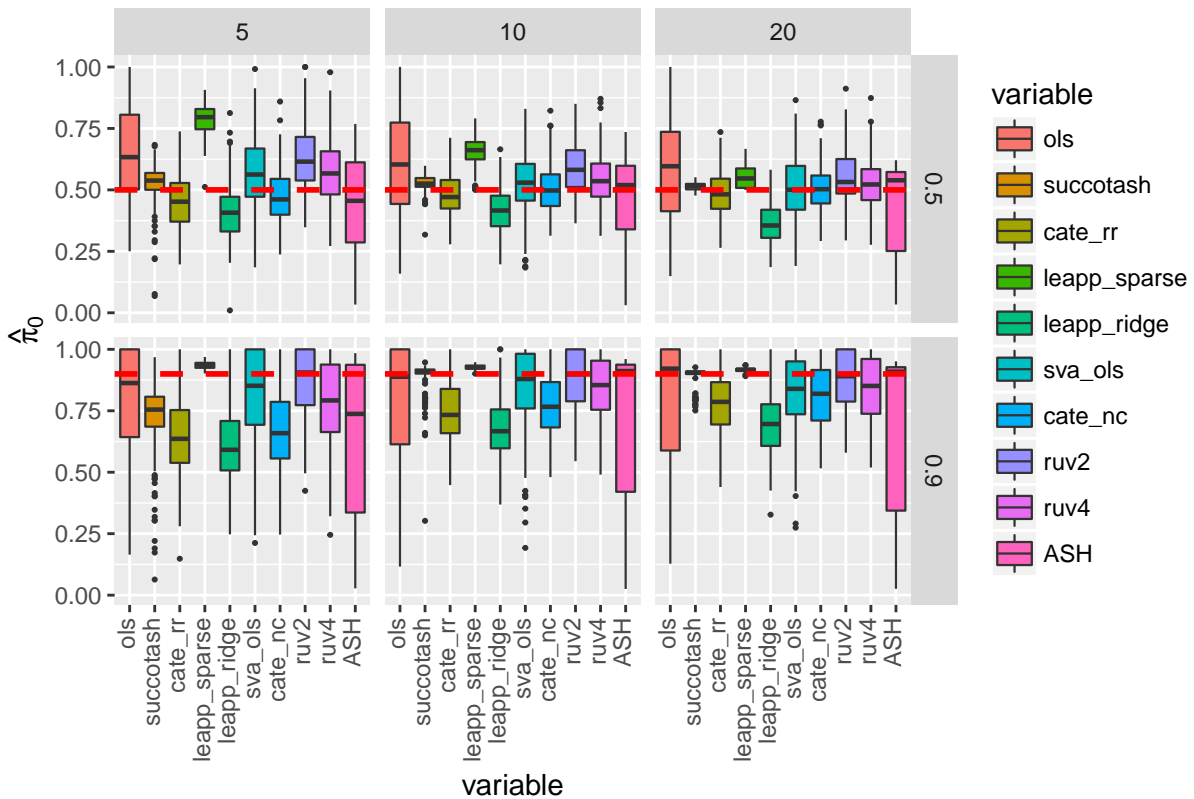
## MSE

SUCCOTASH has superior performance in term of AUC compared to other methods. LEAPP has terrible MSE performance in all cases except the all-null setting. Even then, SUCCOTASH has better performance.

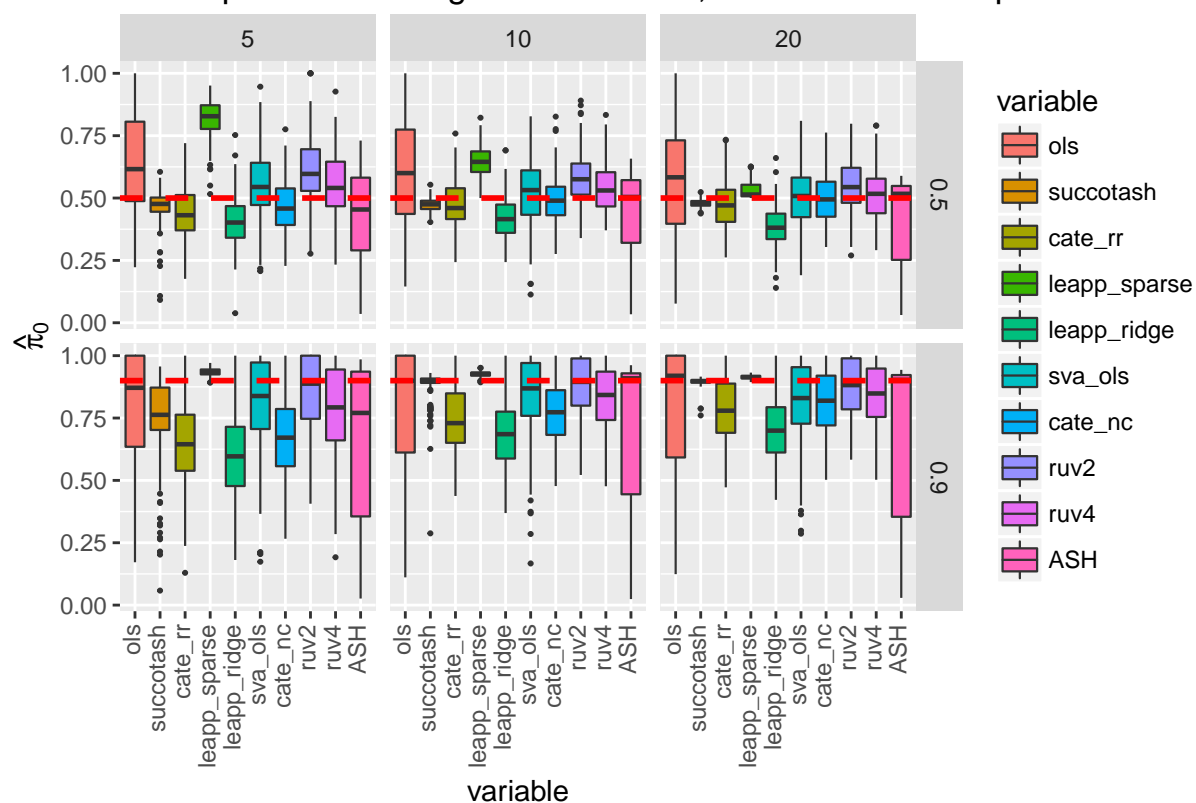
Estimates of  $\pi_0$  When Using Muscle Tissue, Alternative = spiky



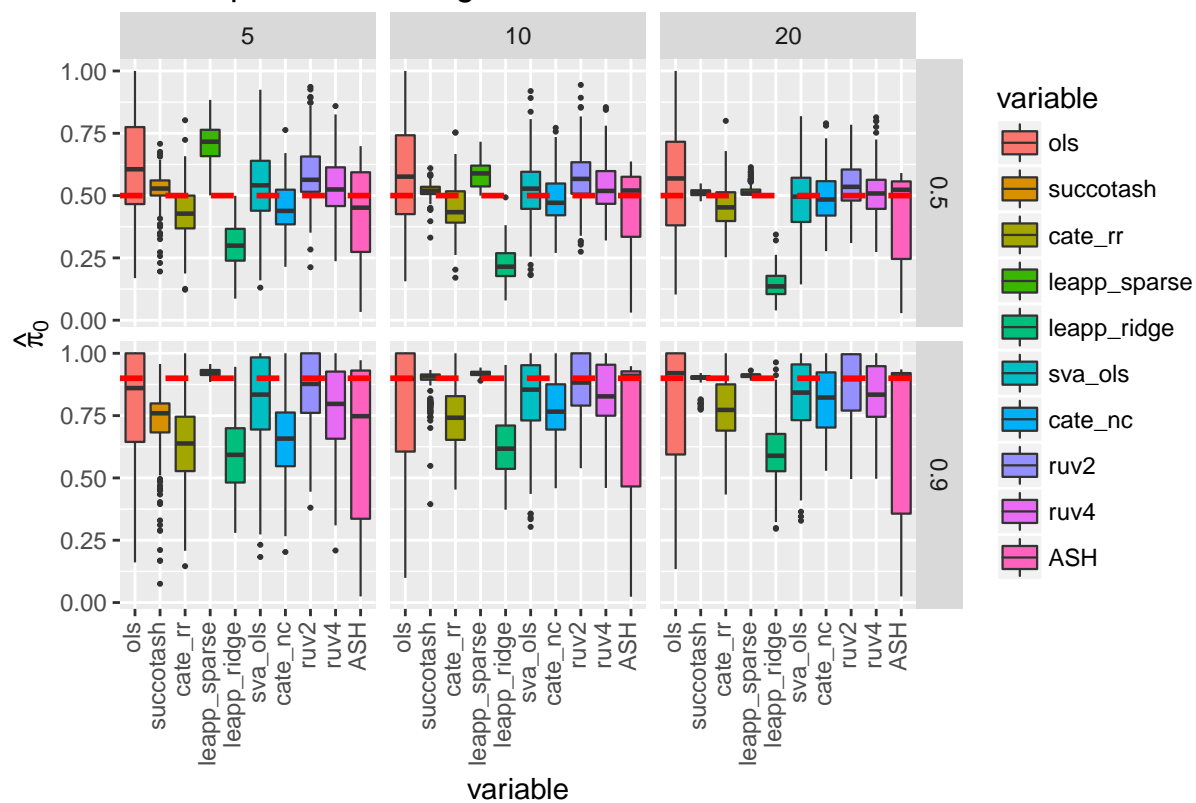
estimates of  $\pi_0$  When Using Muscle Tissue, Alternative = near\_normal



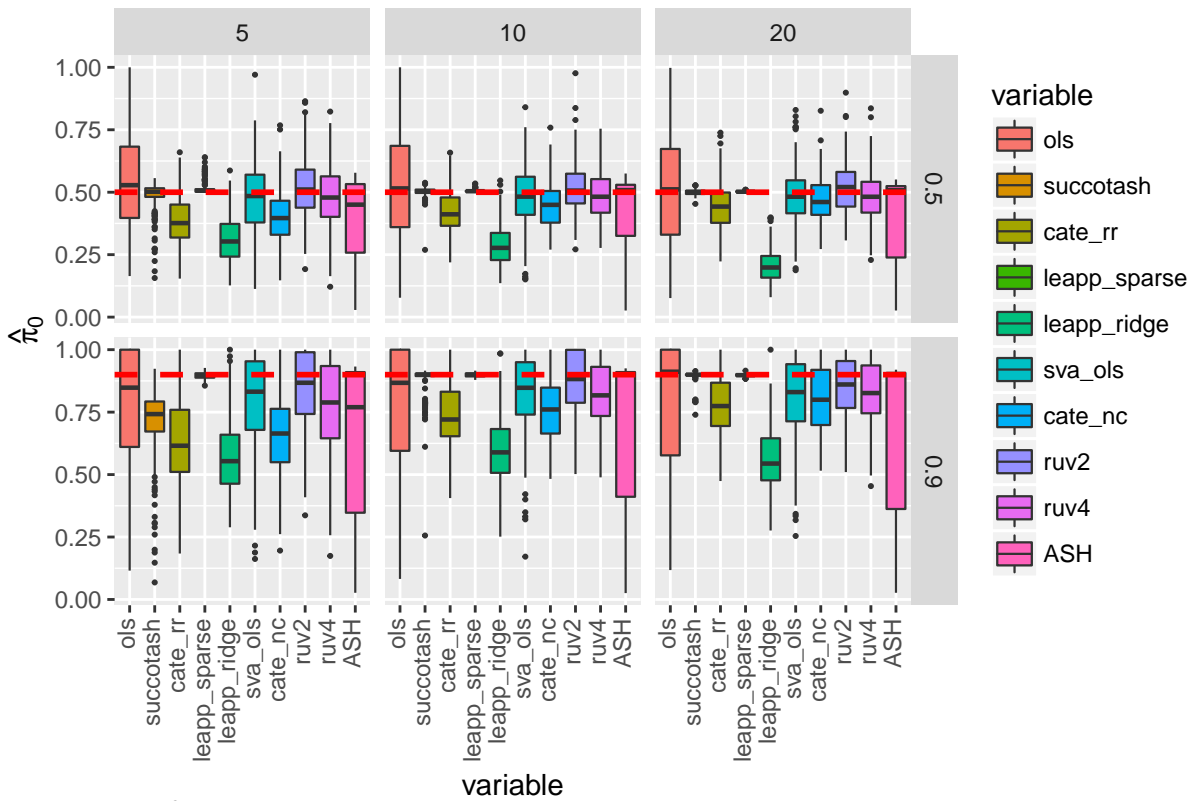
Estimates of  $\pi_0$  When Using Muscle Tissue, Alternative = flattop



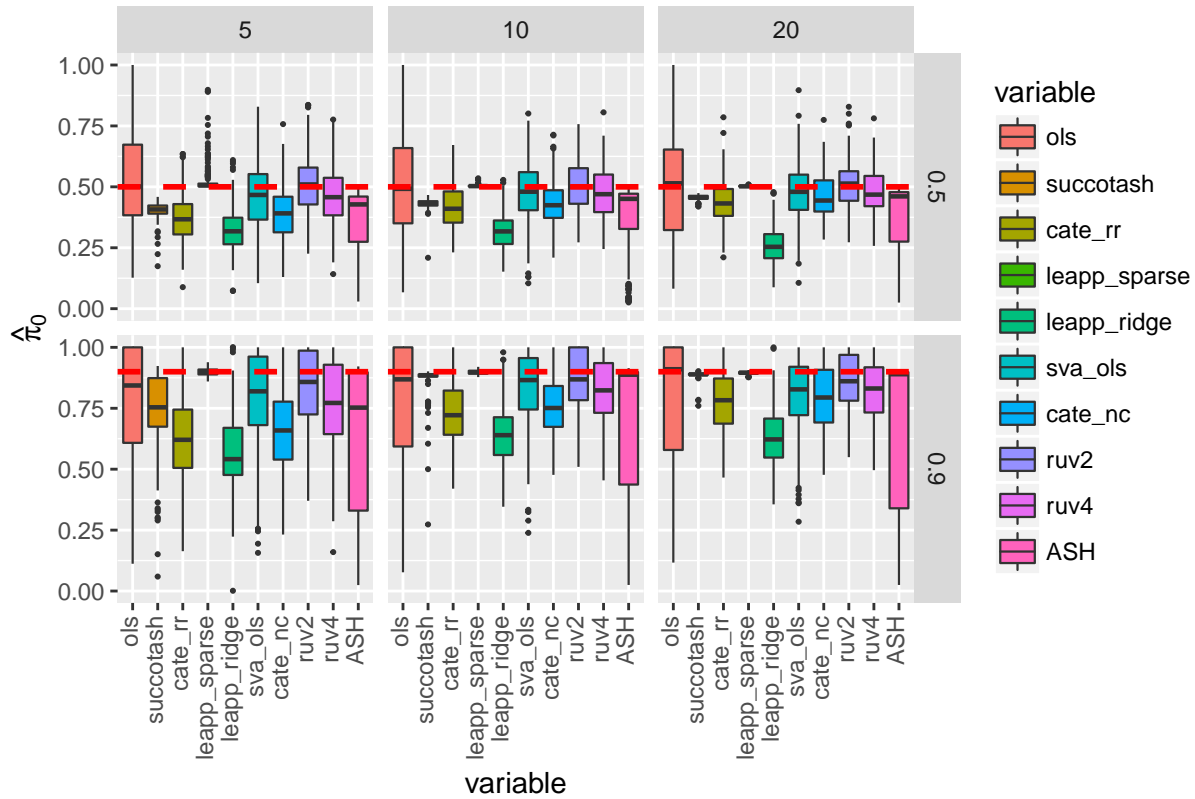
Estimates of  $\pi_0$  When Using Muscle Tissue, Alternative = skew



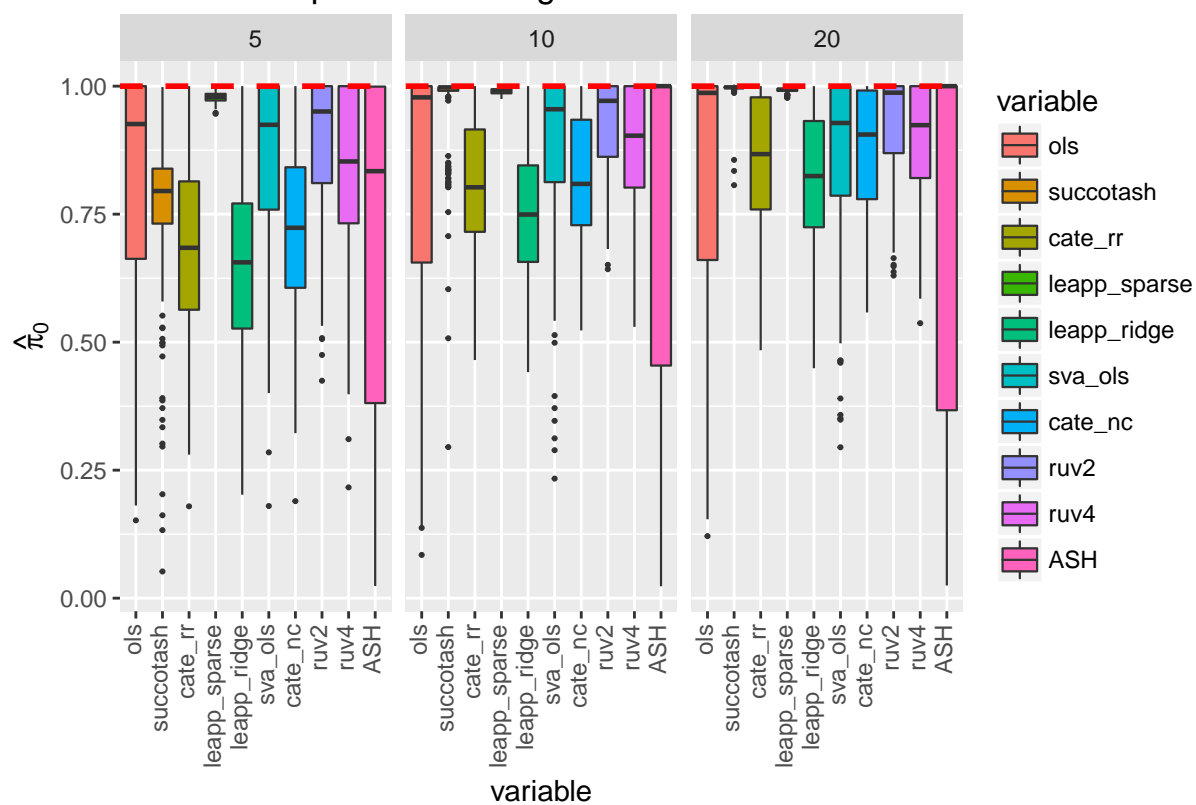
Estimates of  $\pi_0$  When Using Muscle Tissue, Alternative = big\_normal



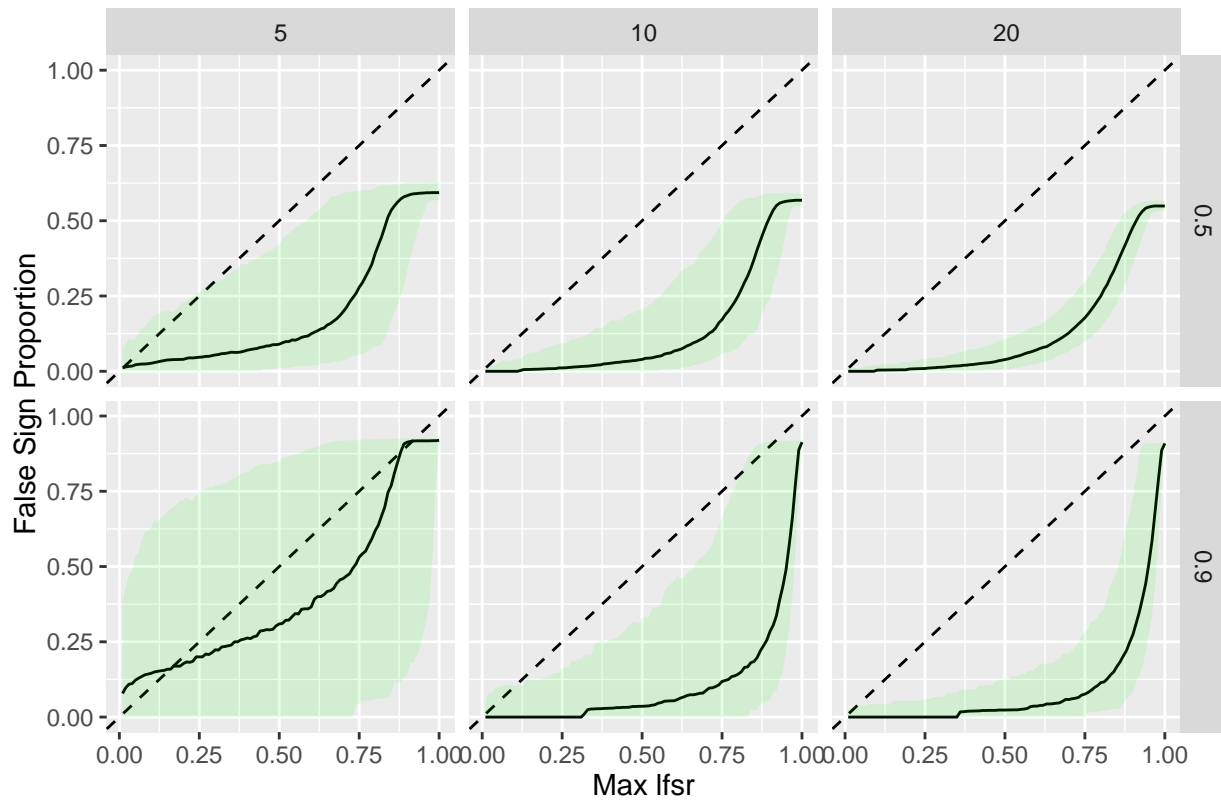
Estimates of  $\pi_0$  When Using Muscle Tissue, Alternative = bimodal



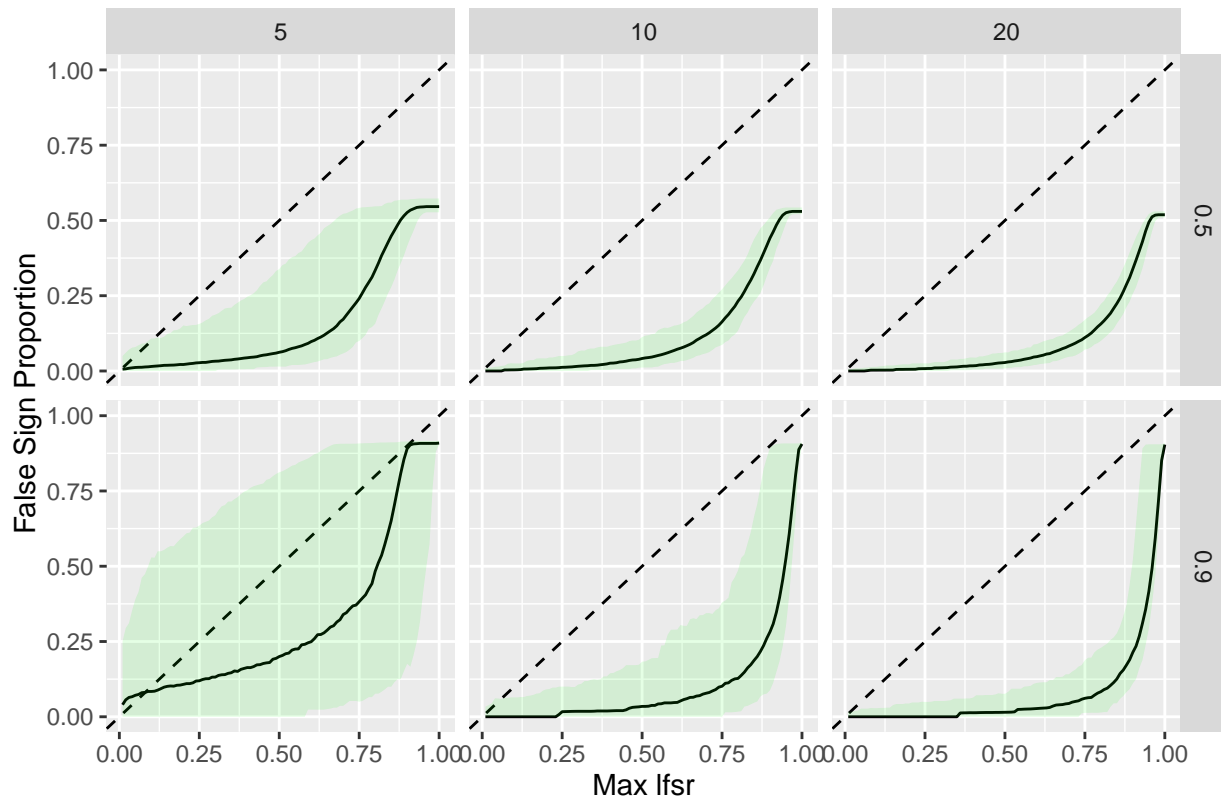
Estimates of  $\pi_0$  When Using Muscle Tissue and All Null



FSP Below Max LFSR when Alt Type = spiky

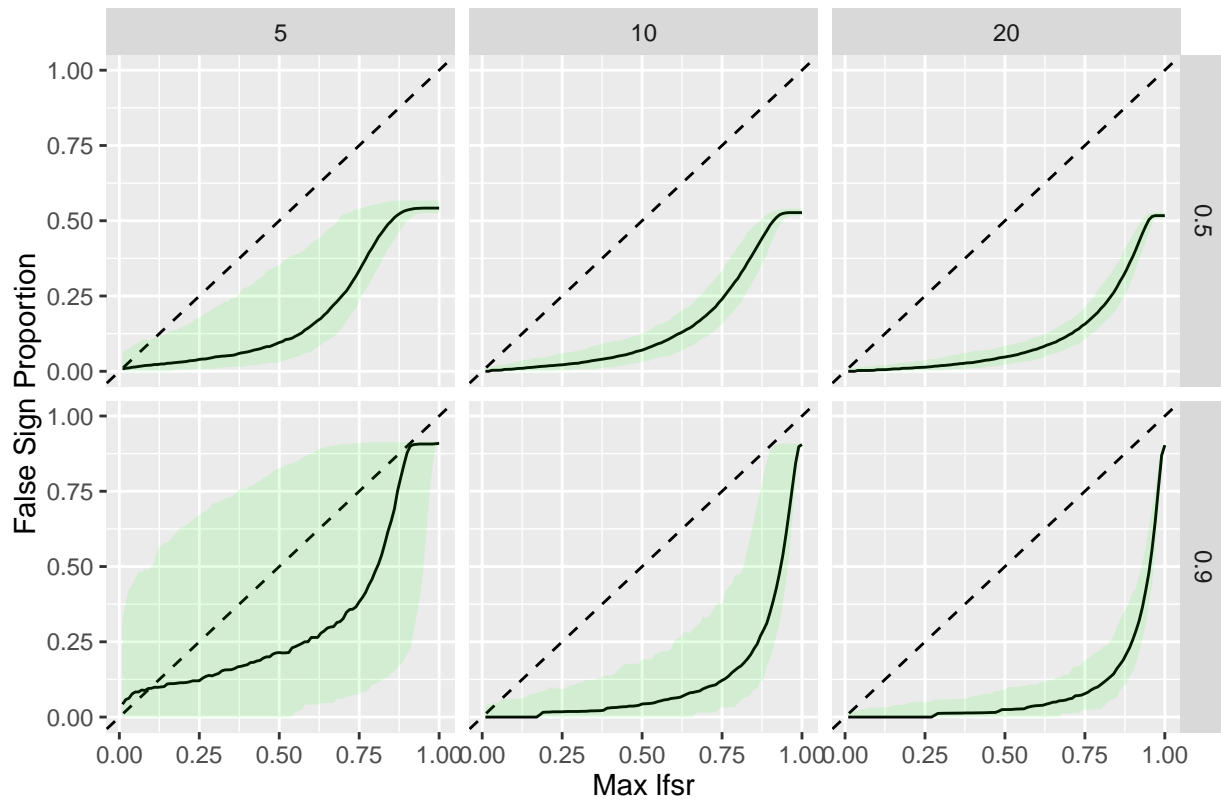


FSP Below Max LFSR when Alt Type = near\_normal

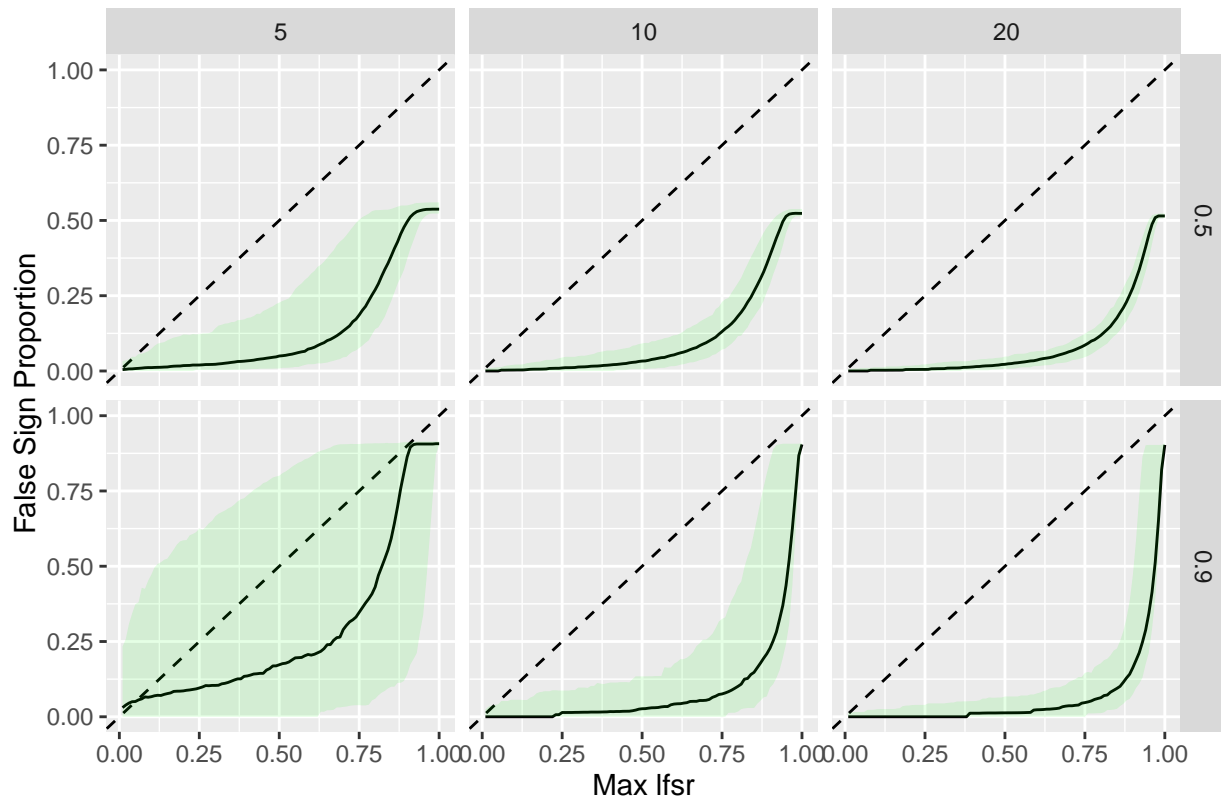




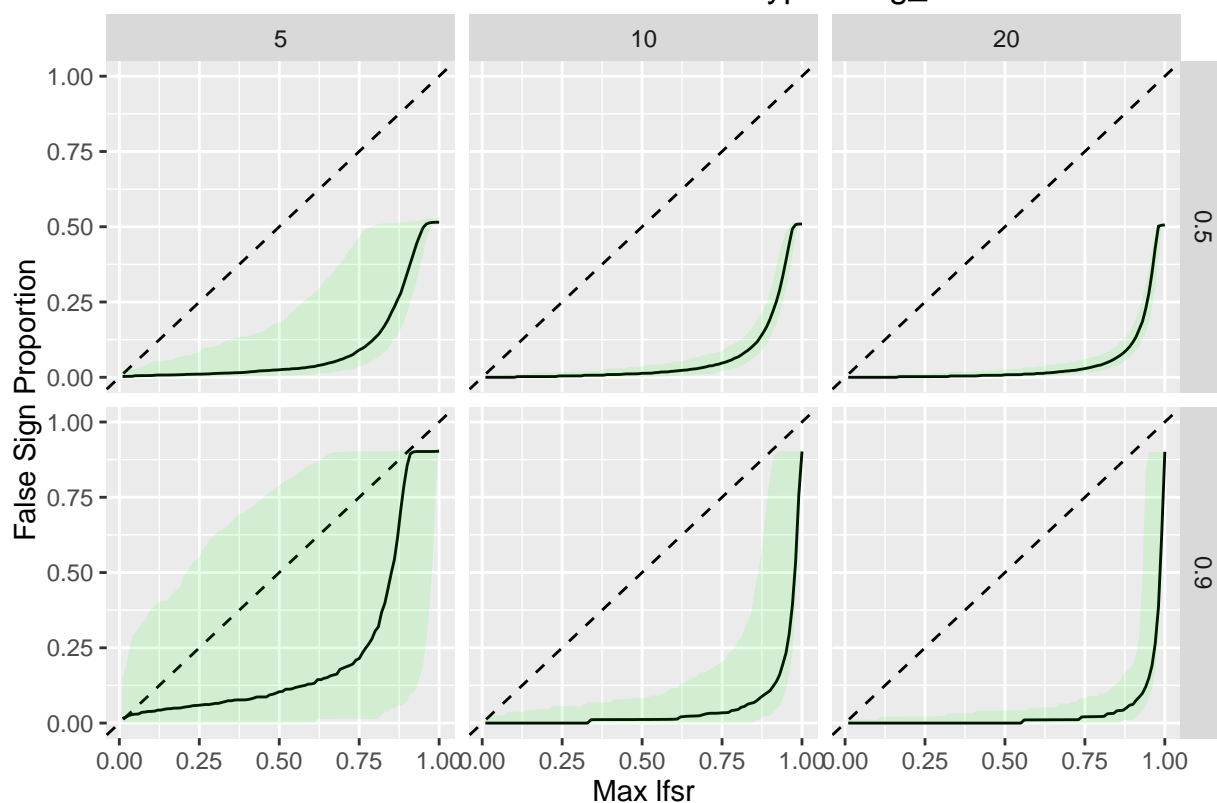
FSP Below Max LFSR when Alt Type = flattop



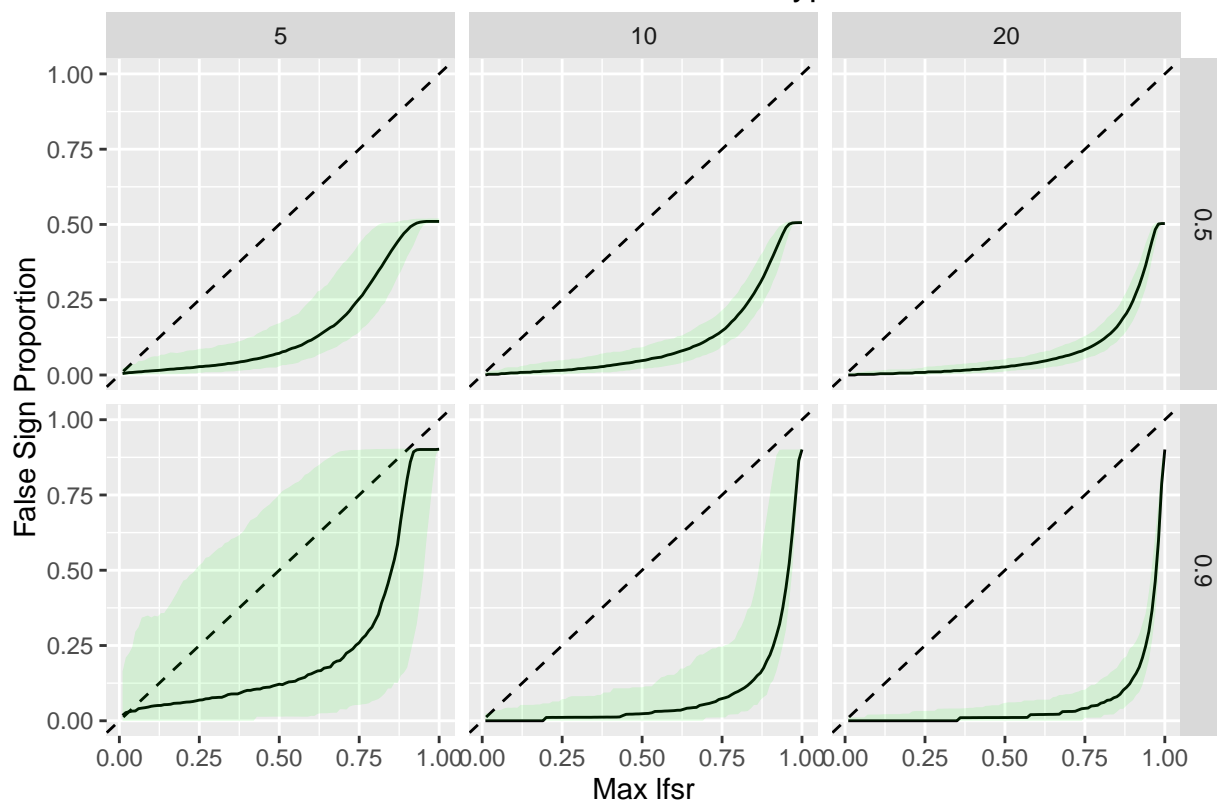
FSP Below Max LFSR when Alt Type = skew



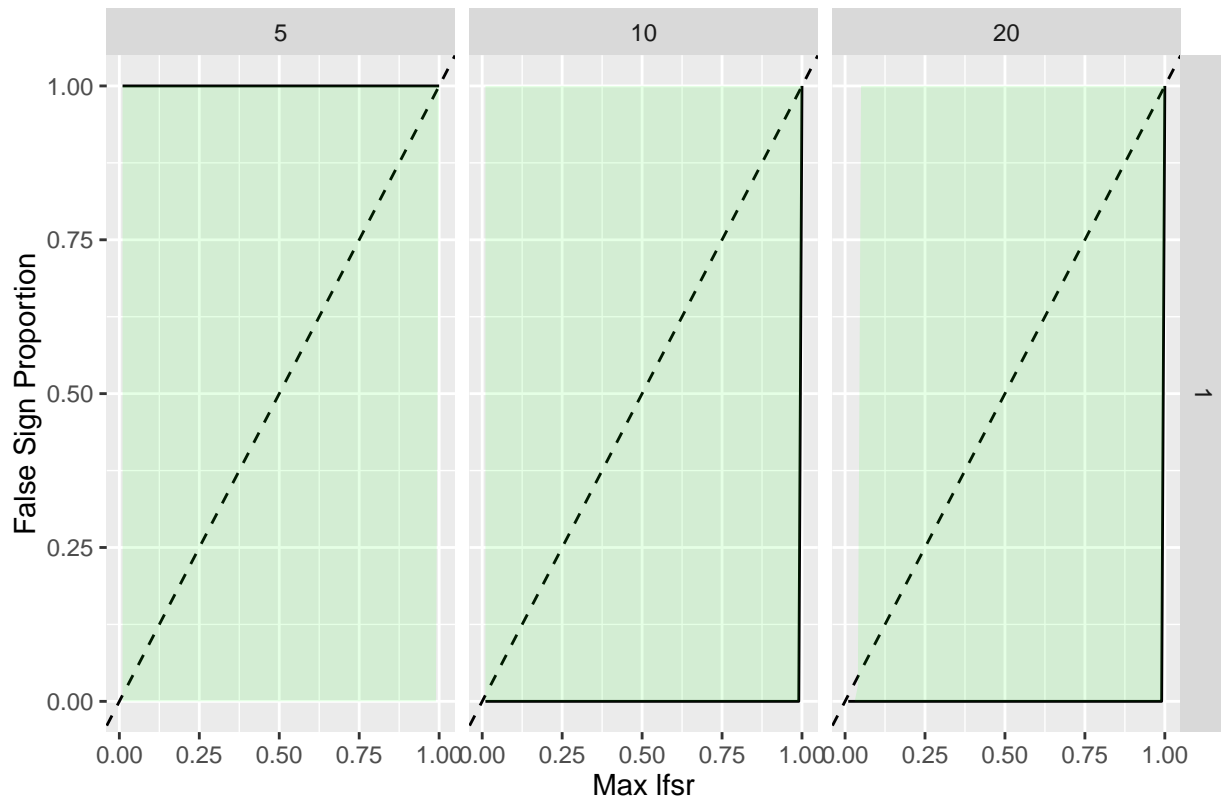
FSP Below Max LFSR when Alt Type = big\_normal



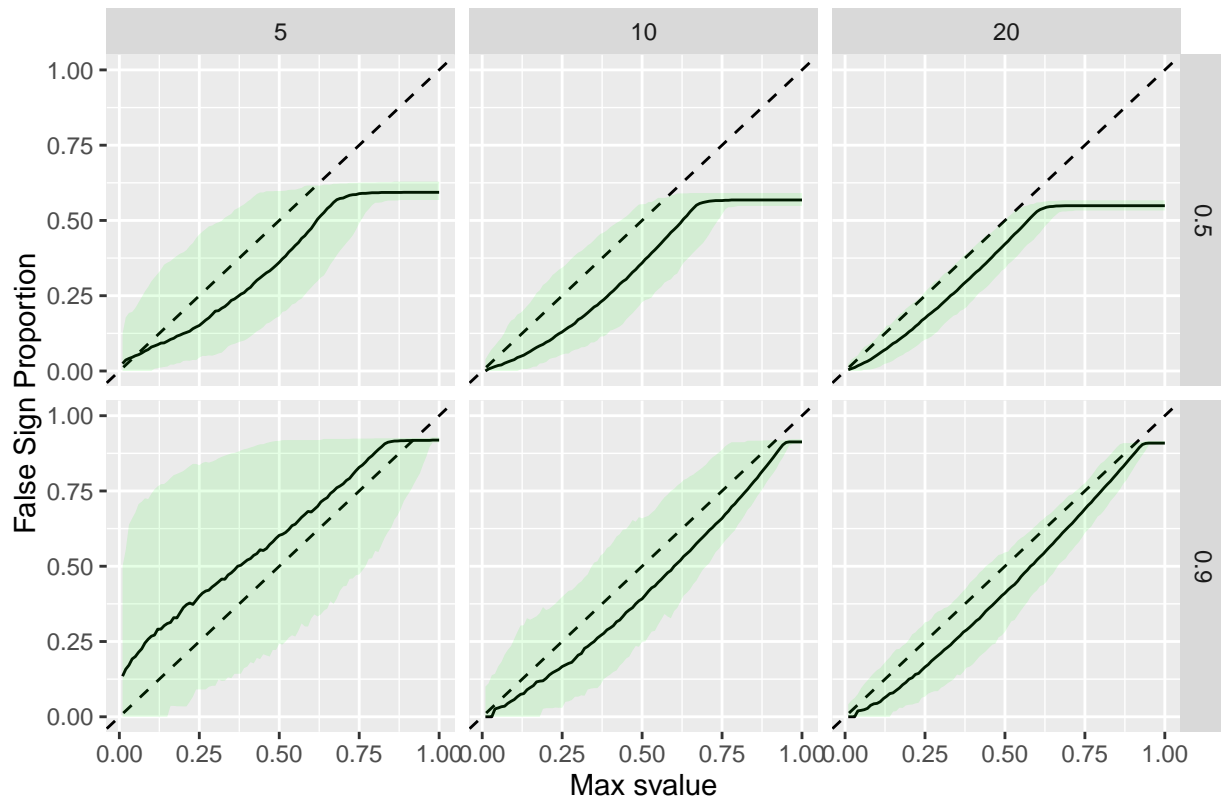
FSP Below Max LFSR when Alt Type = bimodal



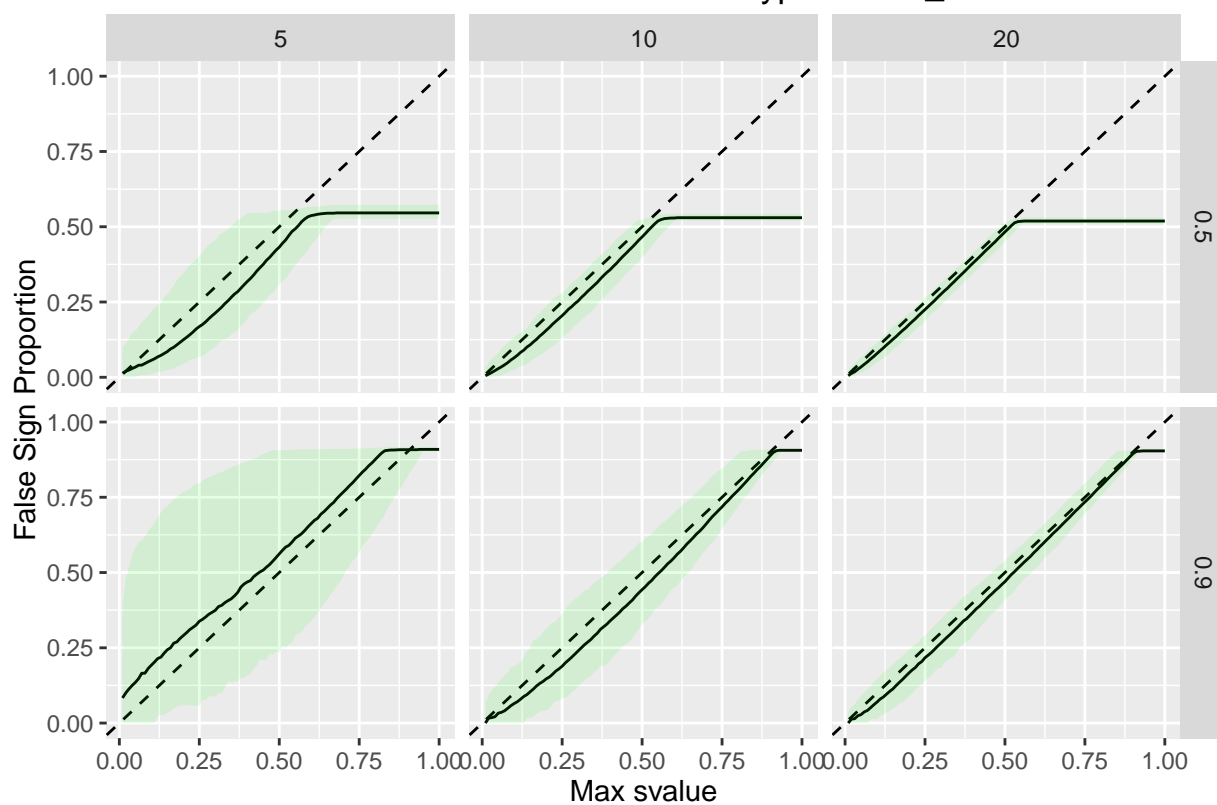
FSP Below Max LFSR when Alt Type = all\_null



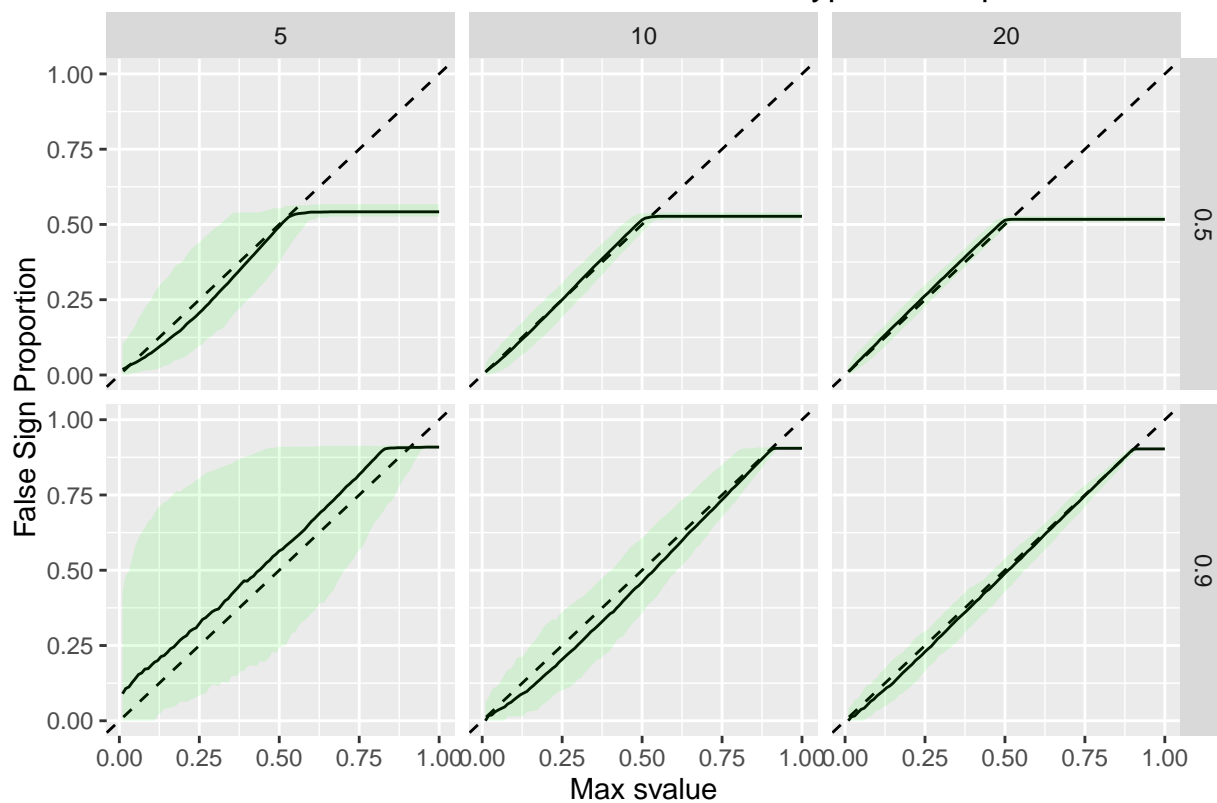
FSP Below Max svalue when Alt Type = spiky

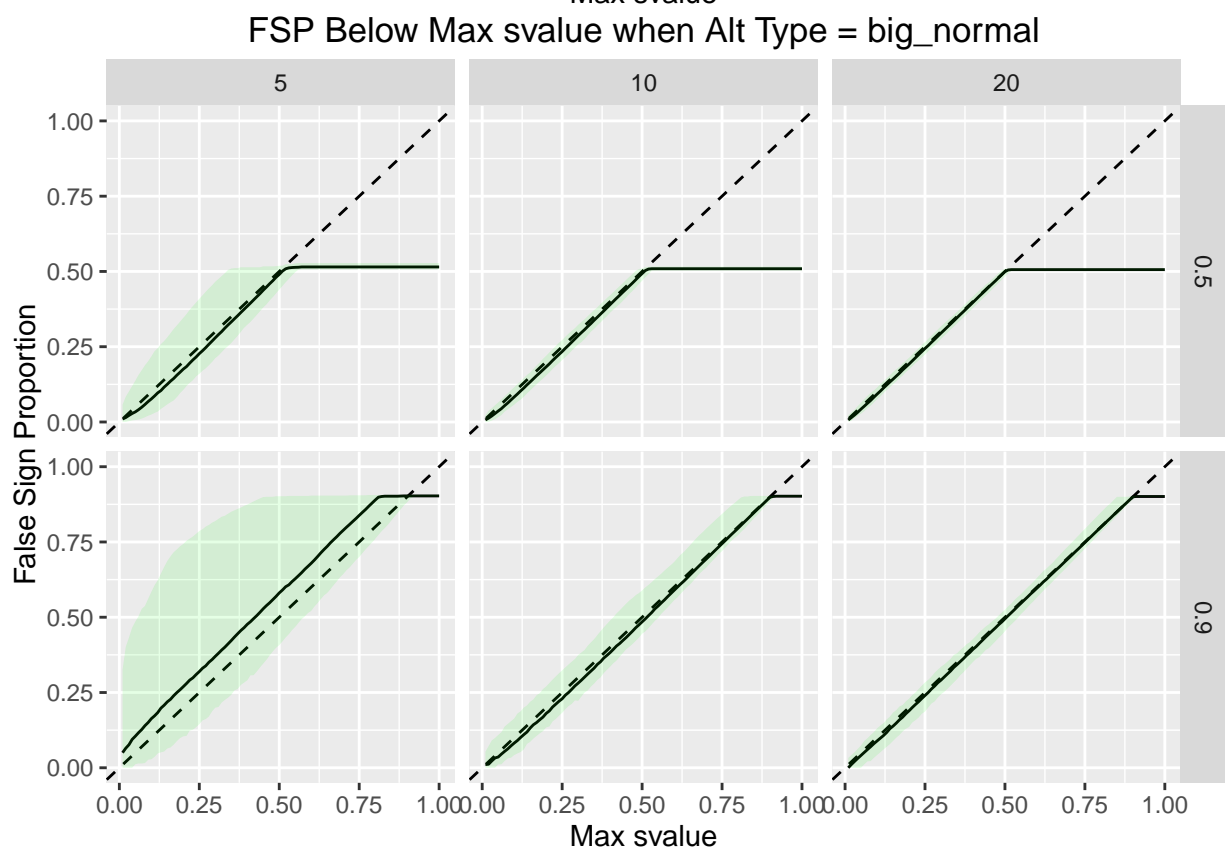
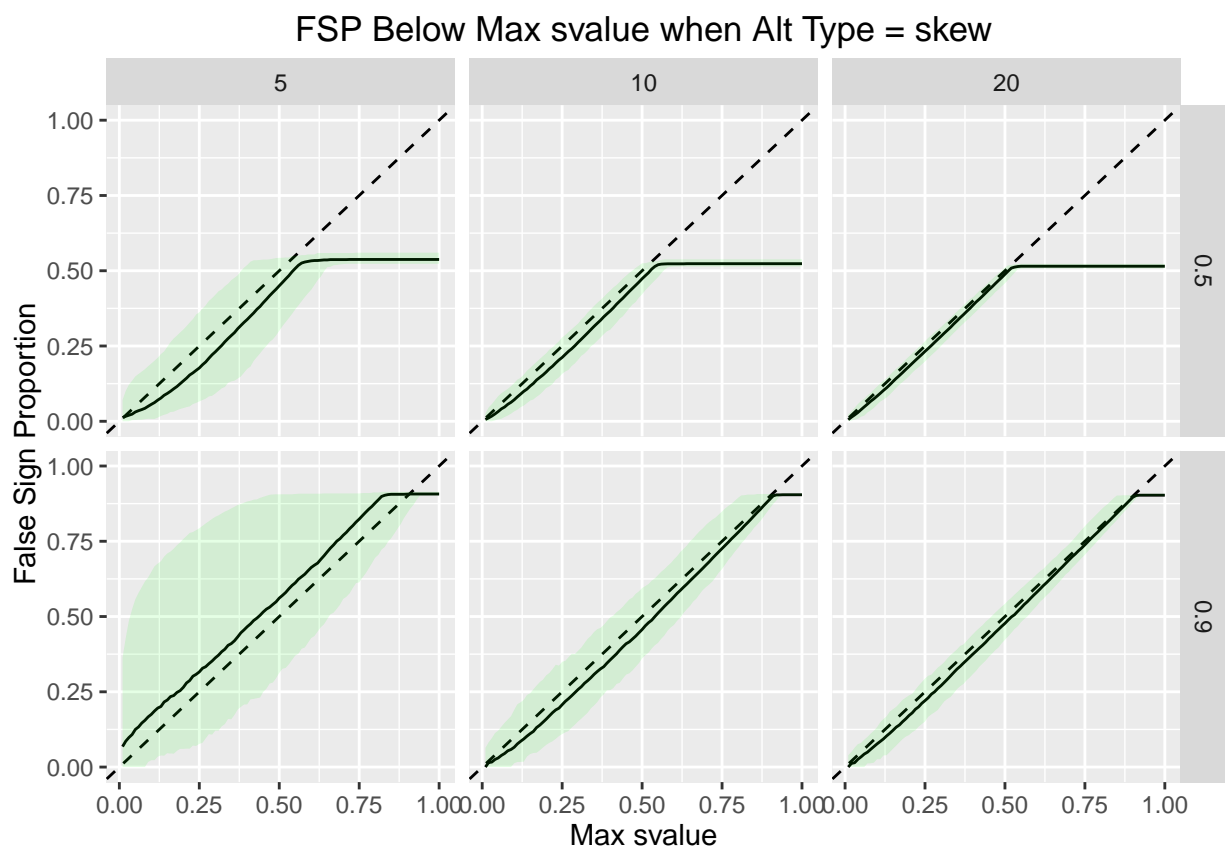


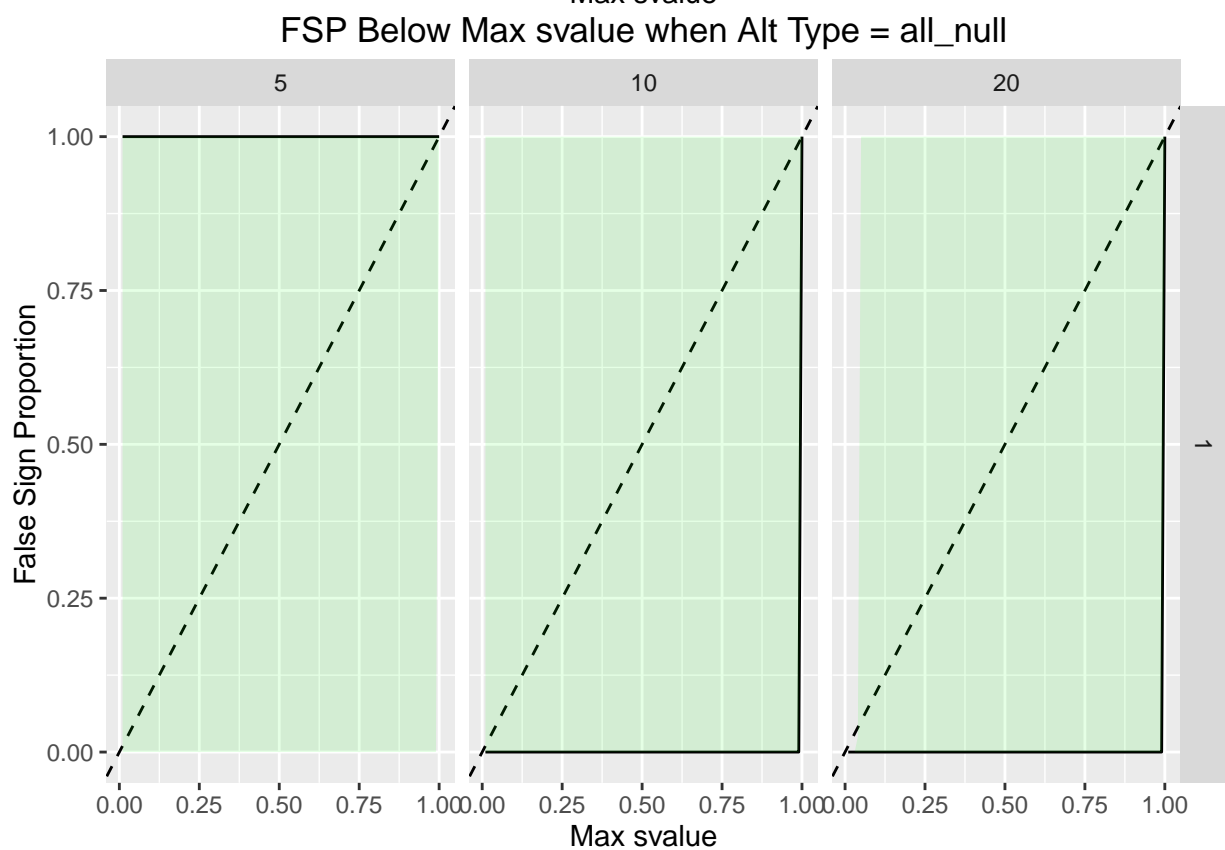
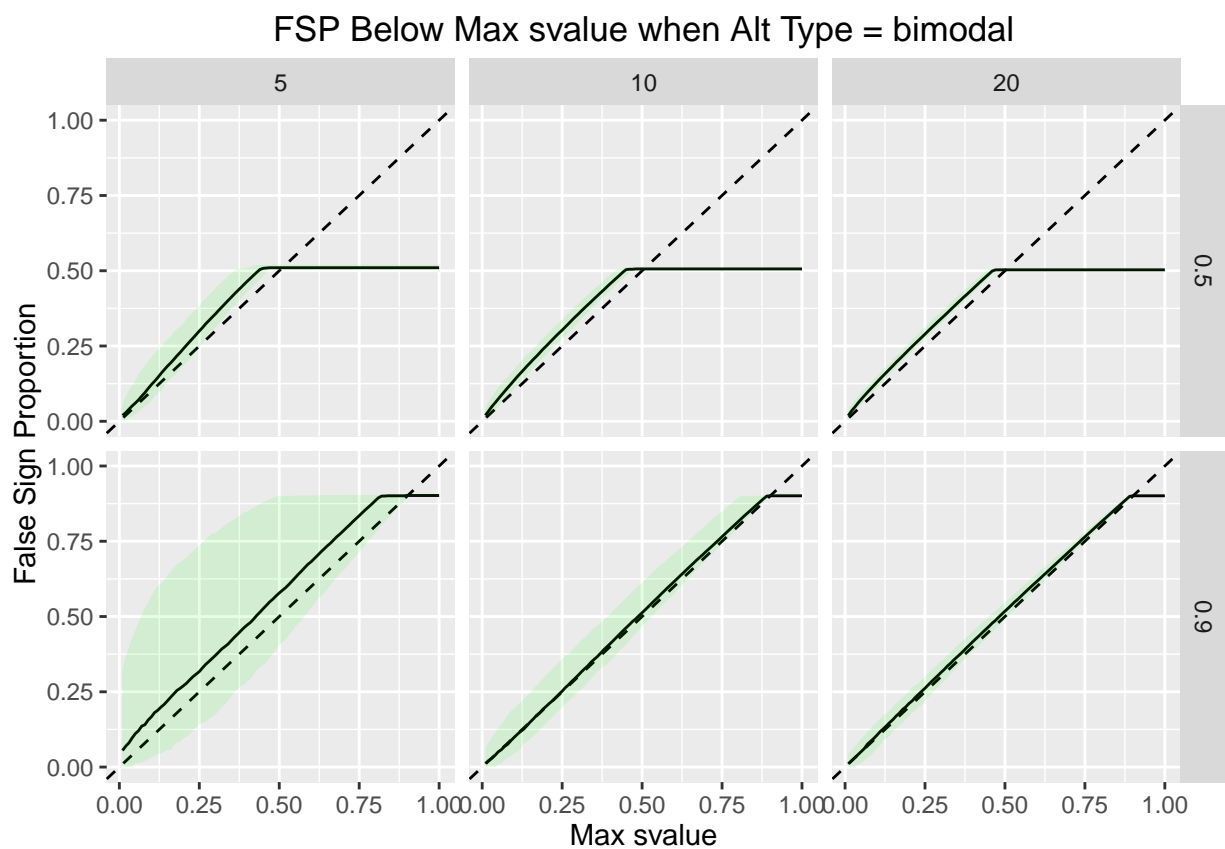
FSP Below Max svalue when Alt Type = near\_normal



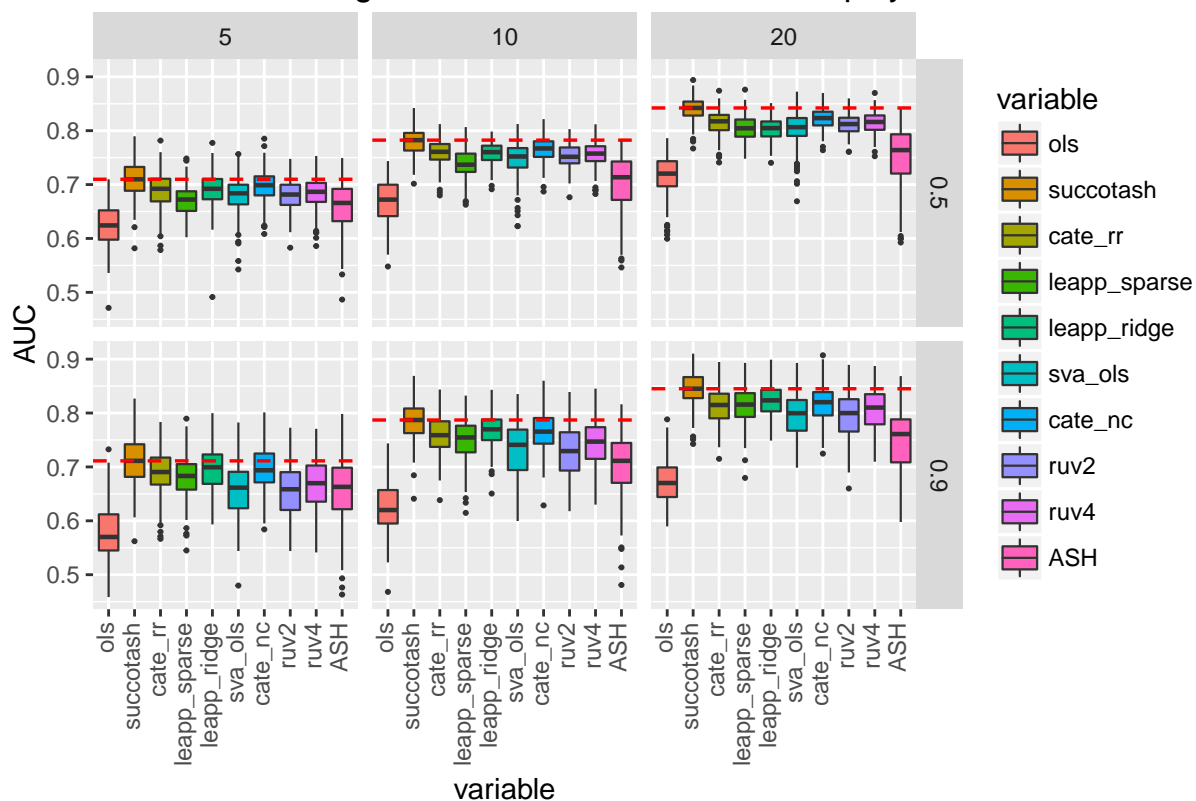
FSP Below Max svalue when Alt Type = flattop



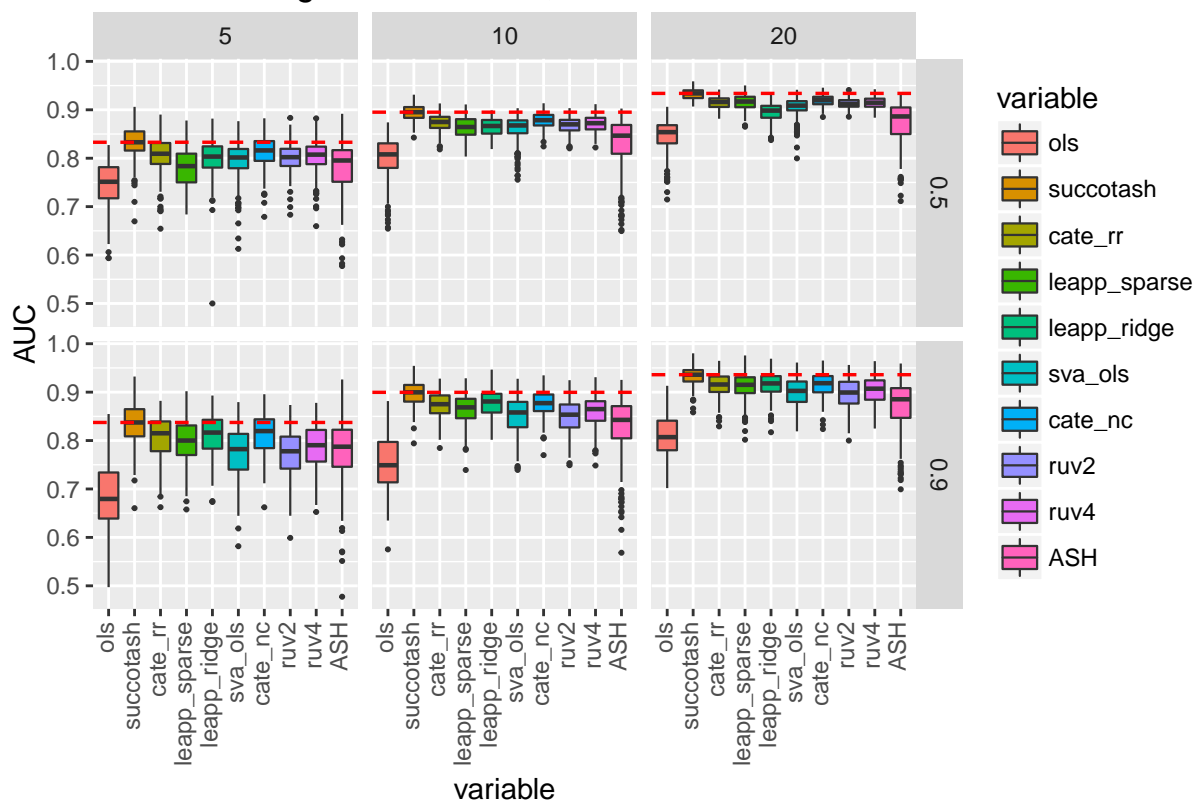




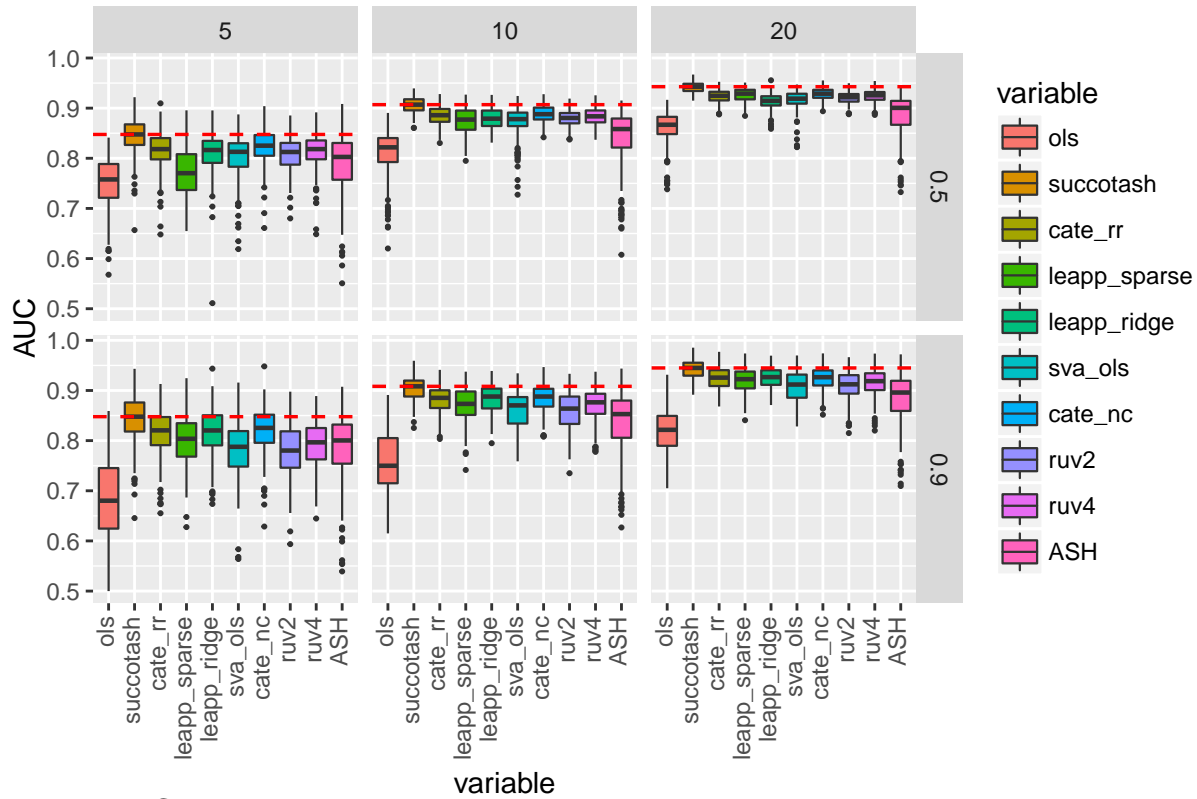
AUC When Using Muscle Tissue, Alternative = spiky



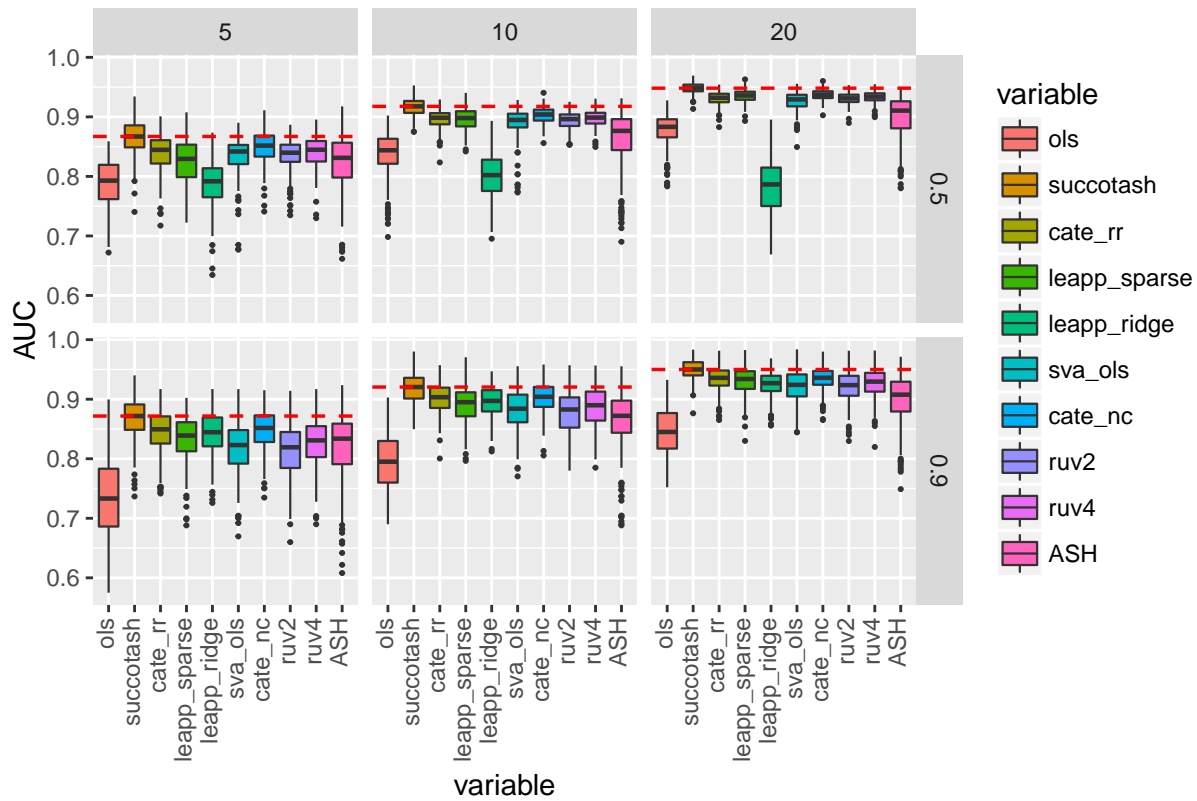
AUC When Using Muscle Tissue, Alternative = near\_normal



AUC When Using Muscle Tissue, Alternative = flattop

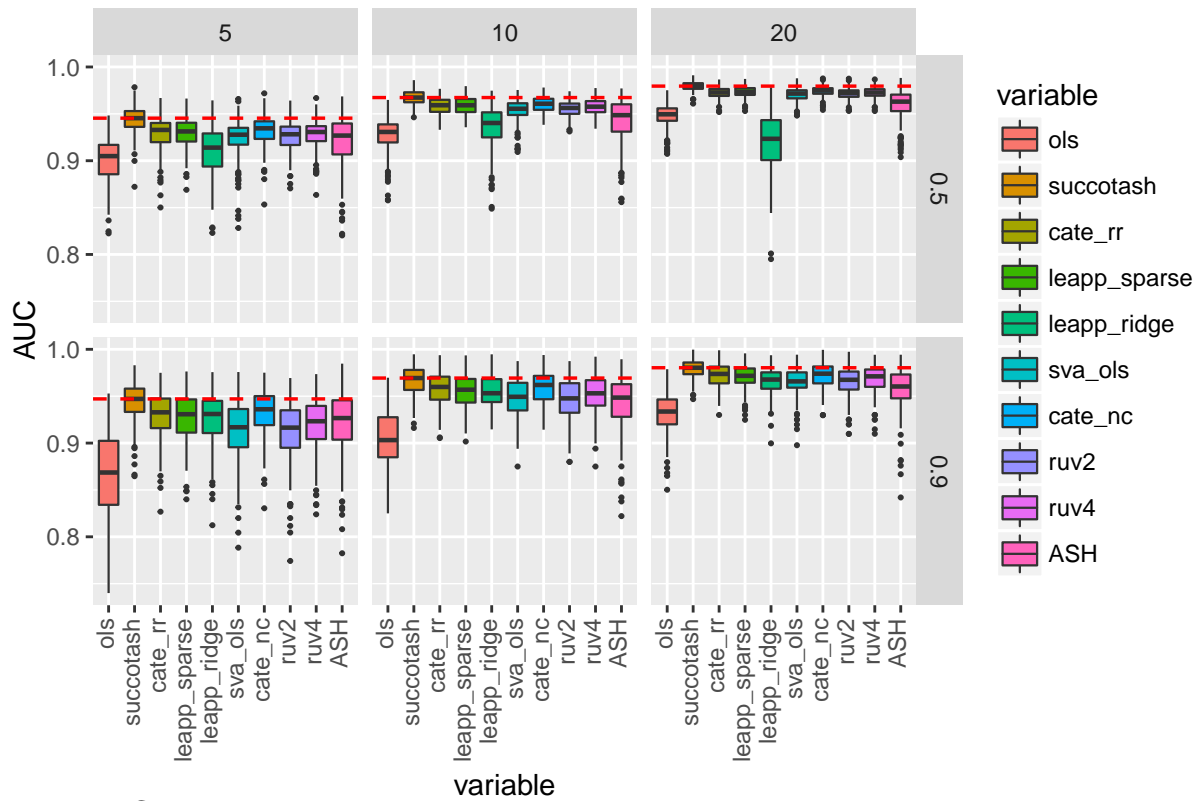


AUC When Using Muscle Tissue, Alternative = skew

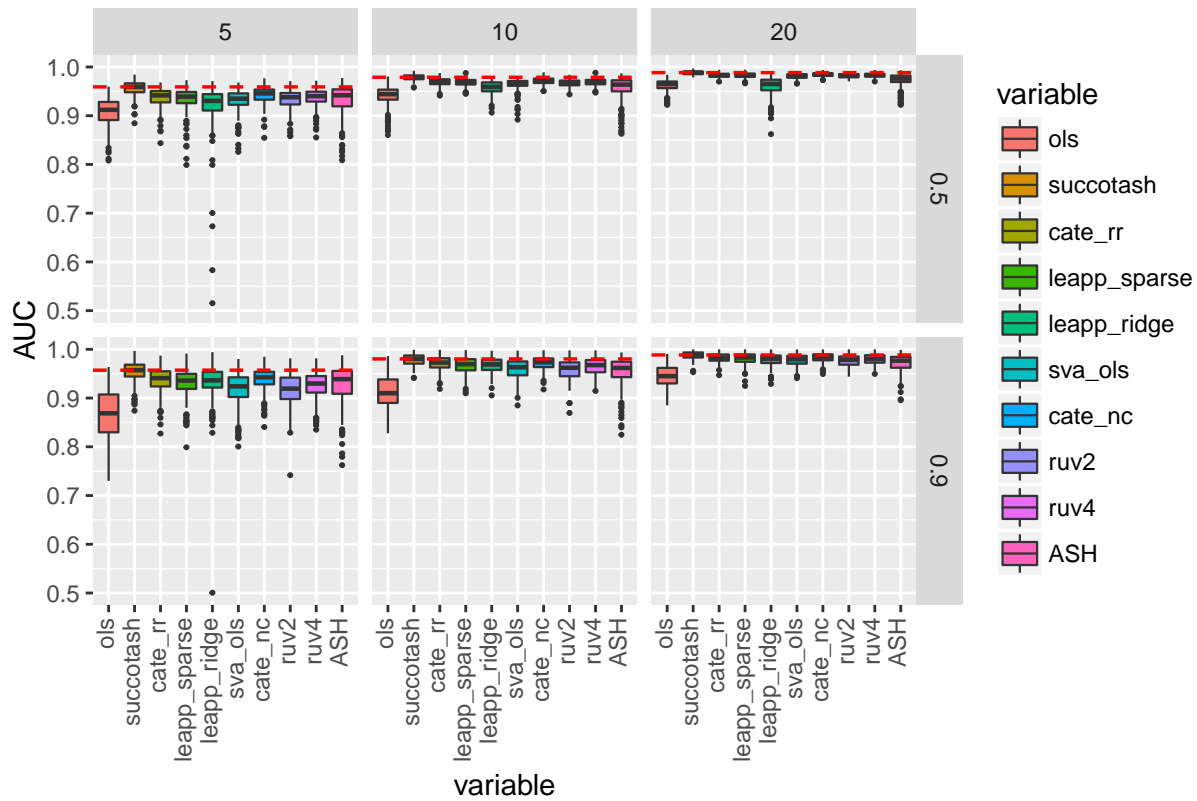


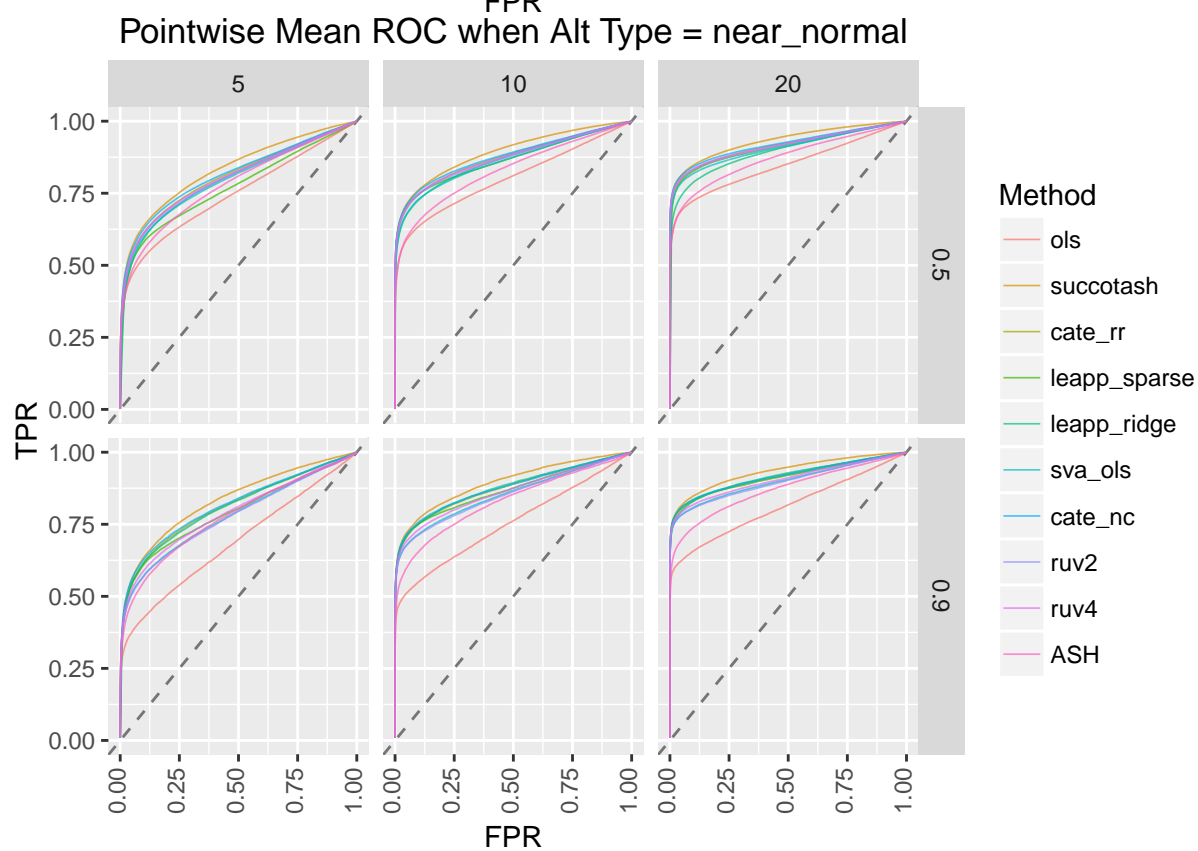
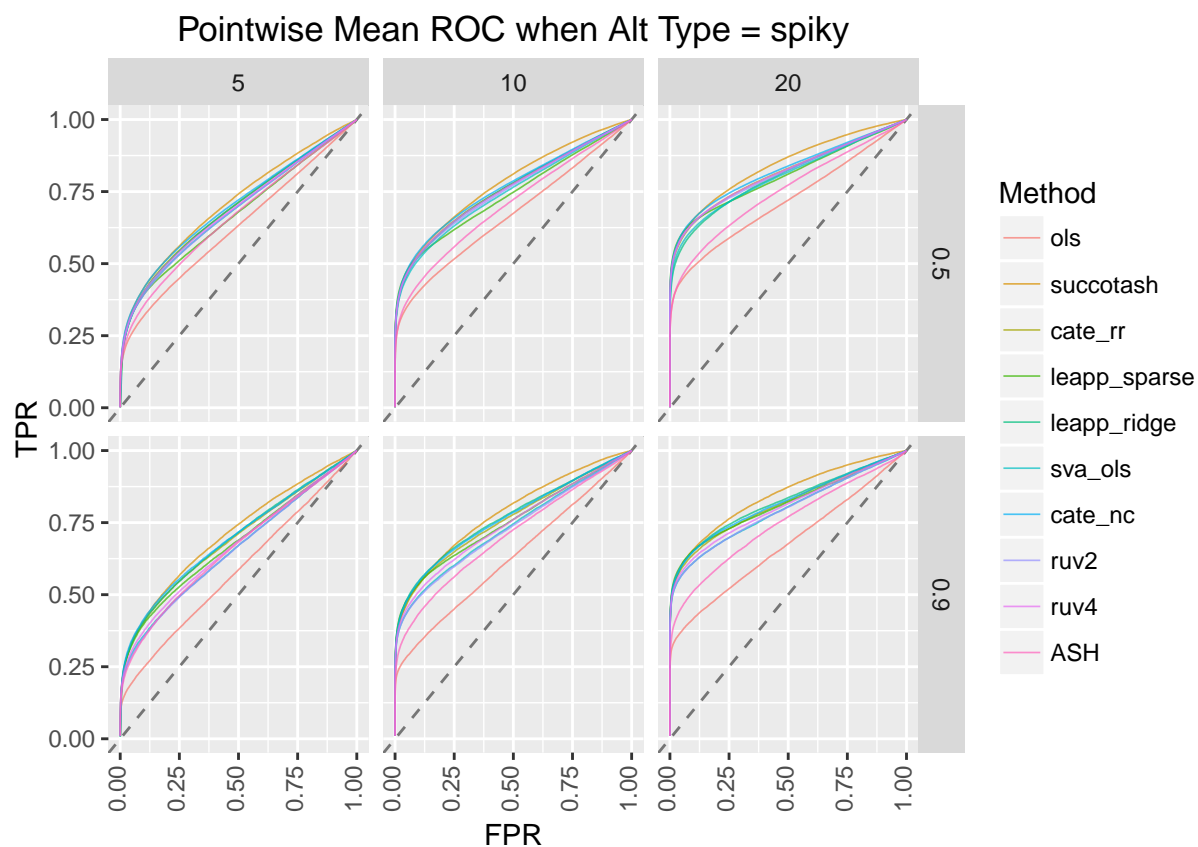


AUC When Using Muscle Tissue, Alternative = big\_normal

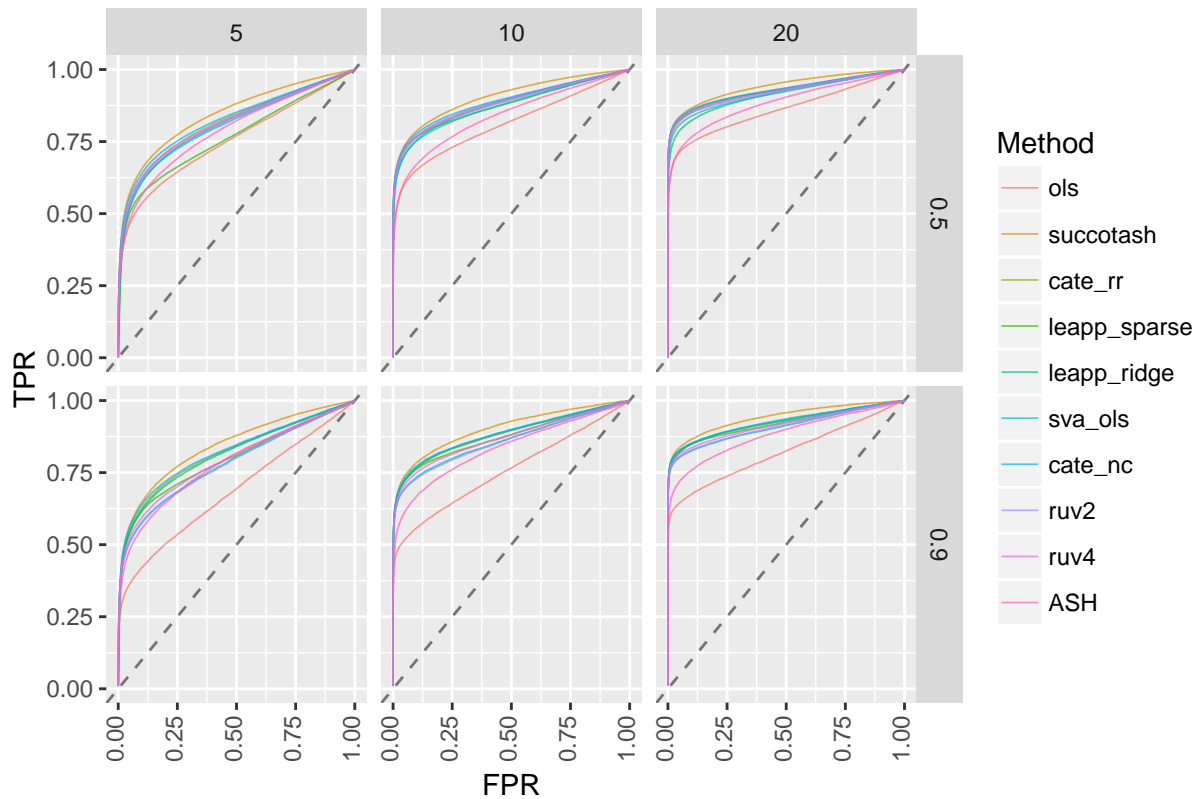


AUC When Using Muscle Tissue, Alternative = bimodal

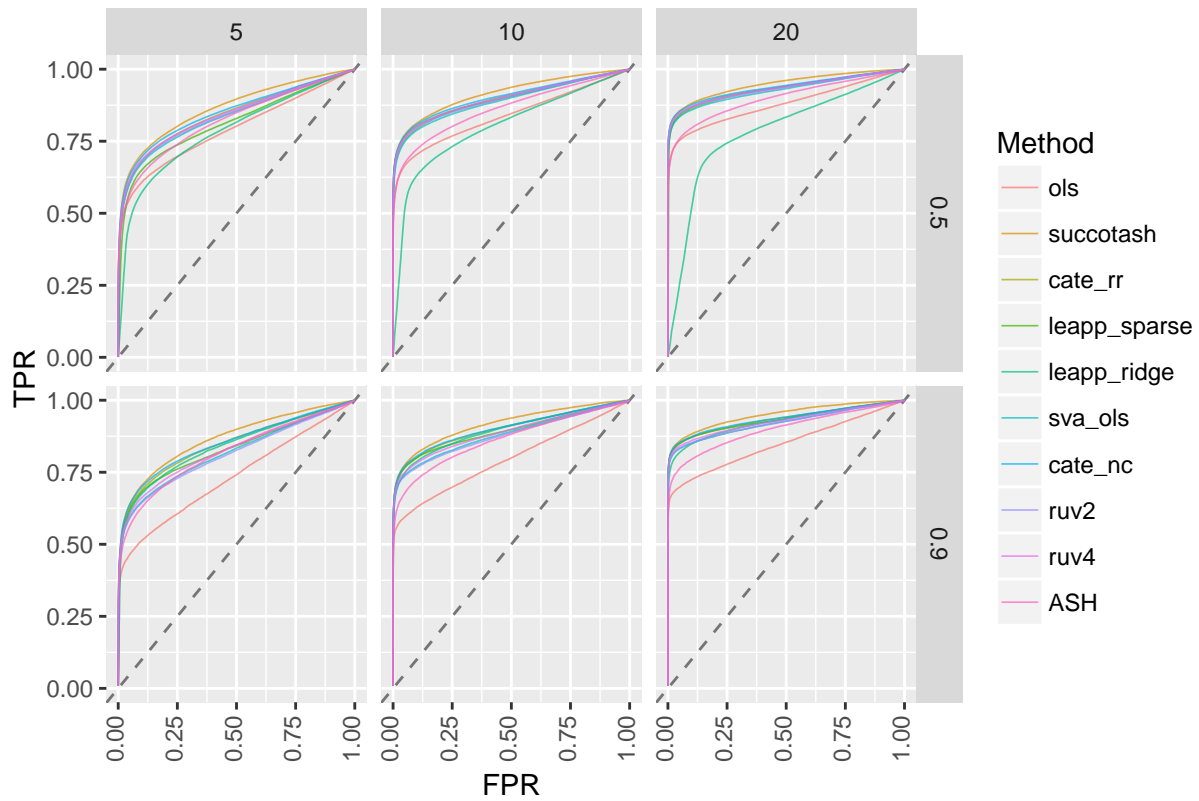




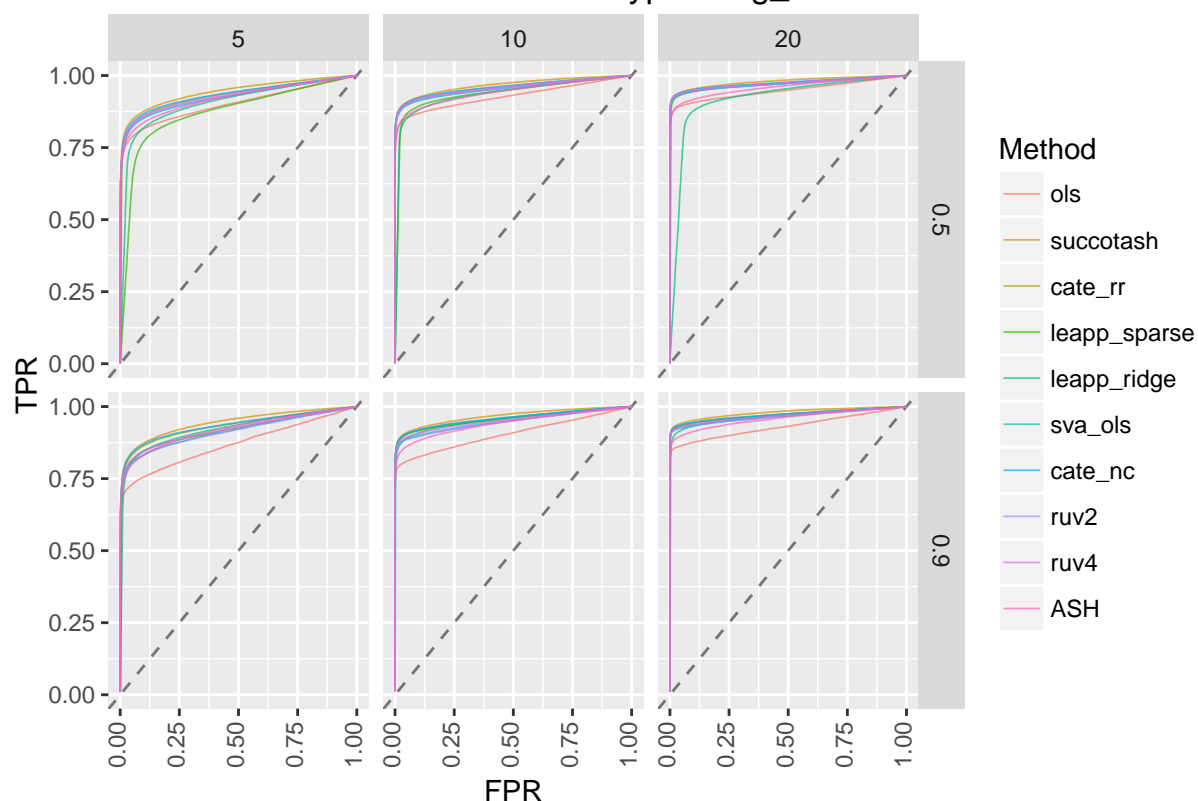
Pointwise Mean ROC when Alt Type = flattop



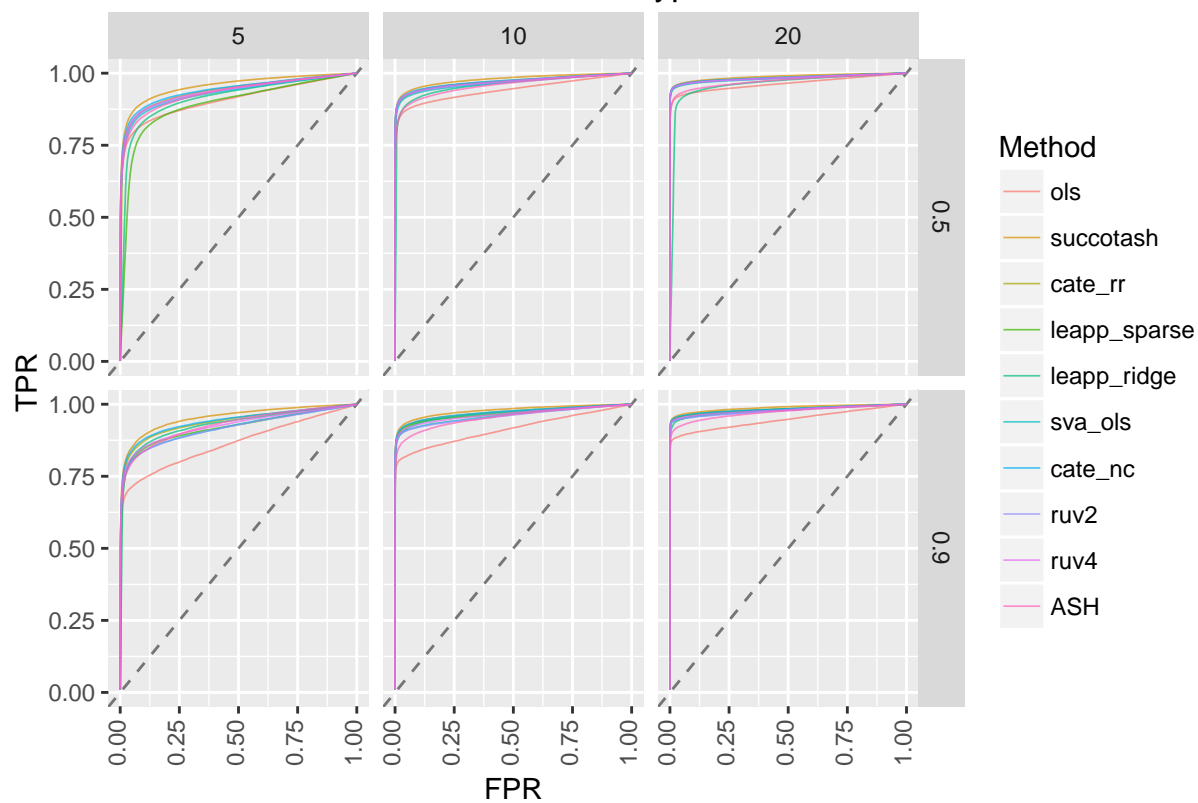
Pointwise Mean ROC when Alt Type = skew



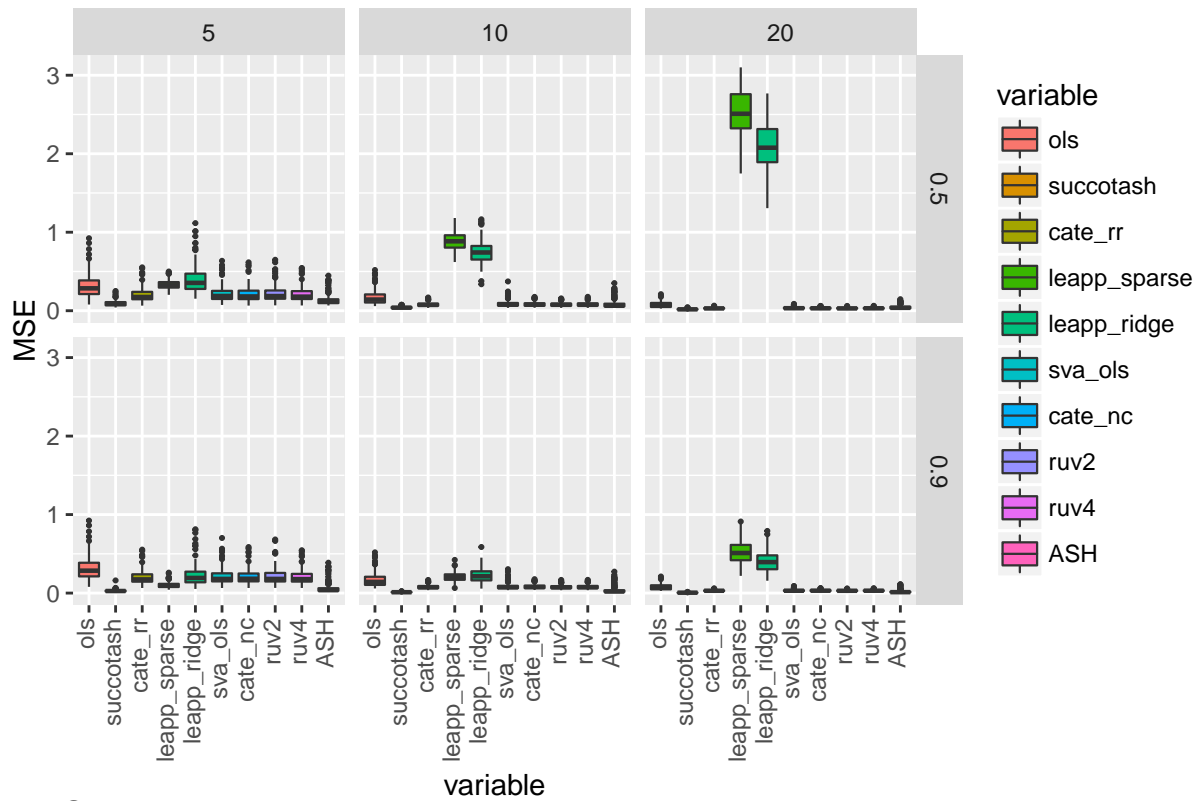
Pointwise Mean ROC when Alt Type = big\_normal



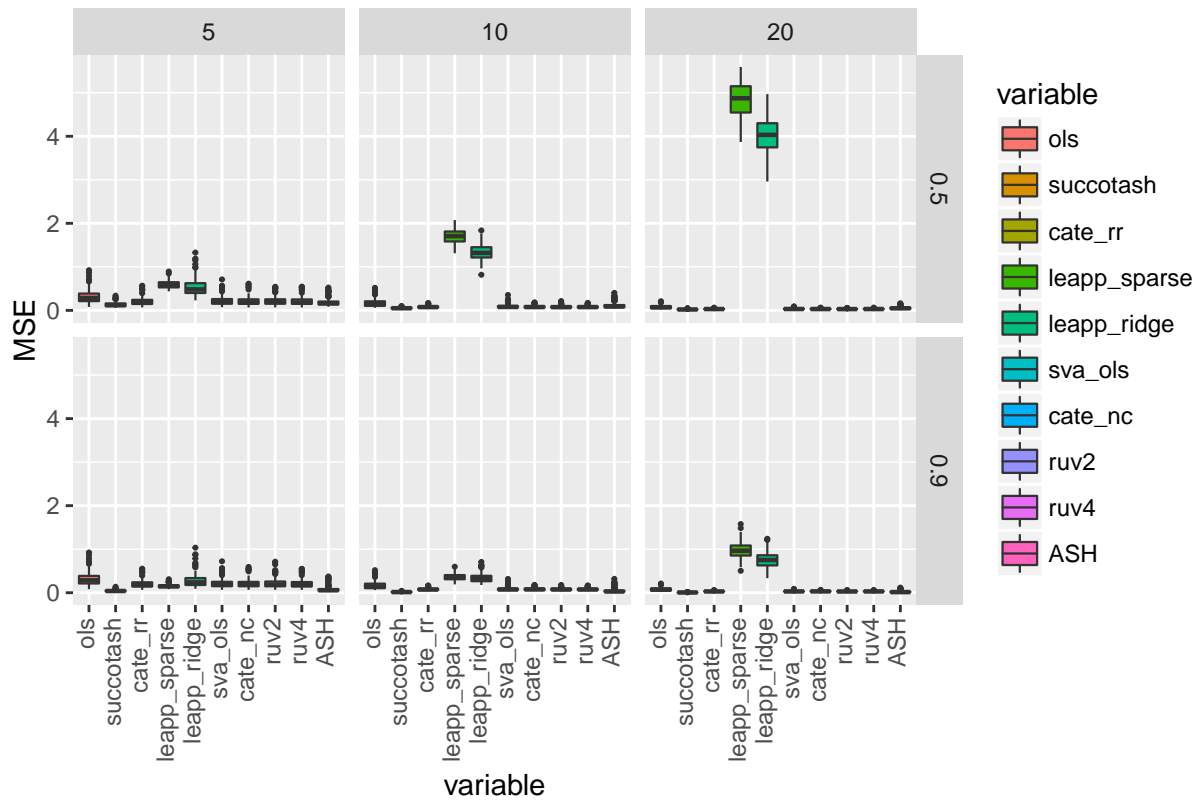
Pointwise Mean ROC when Alt Type = bimodal



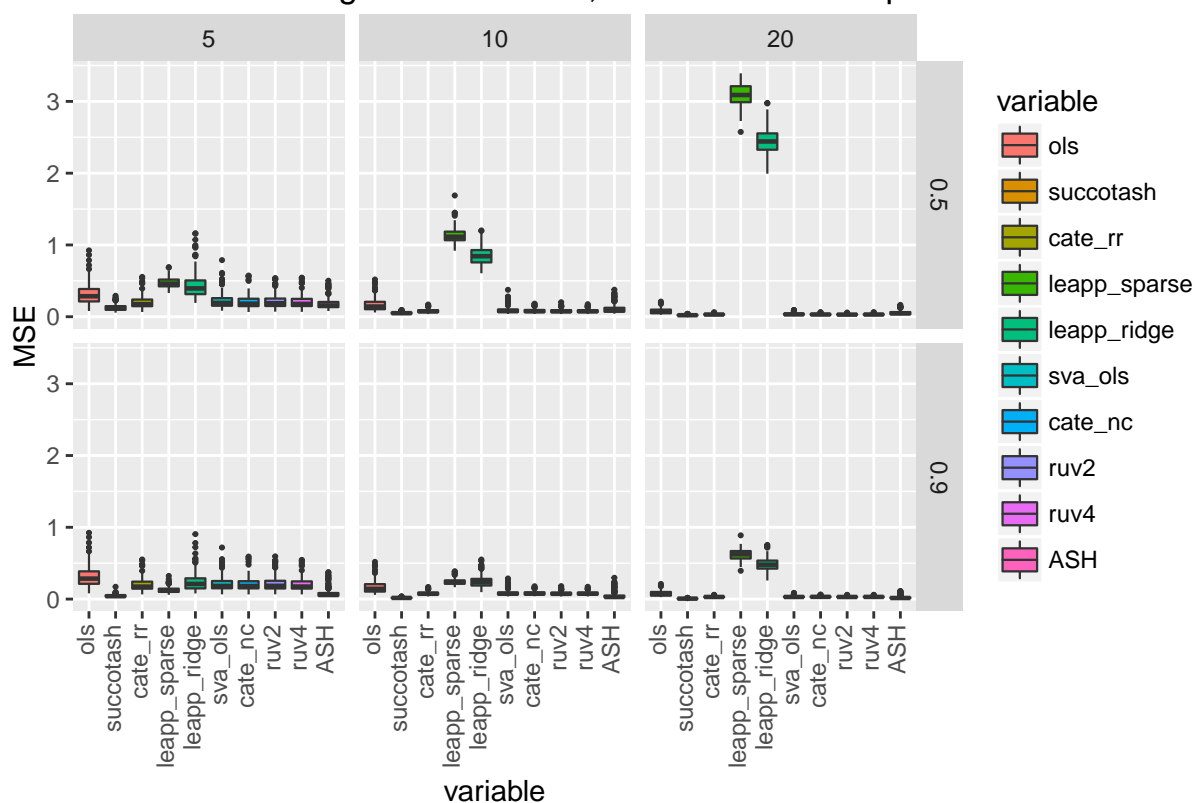
MSE When Using Muscle Tissue, Alternative = spiky



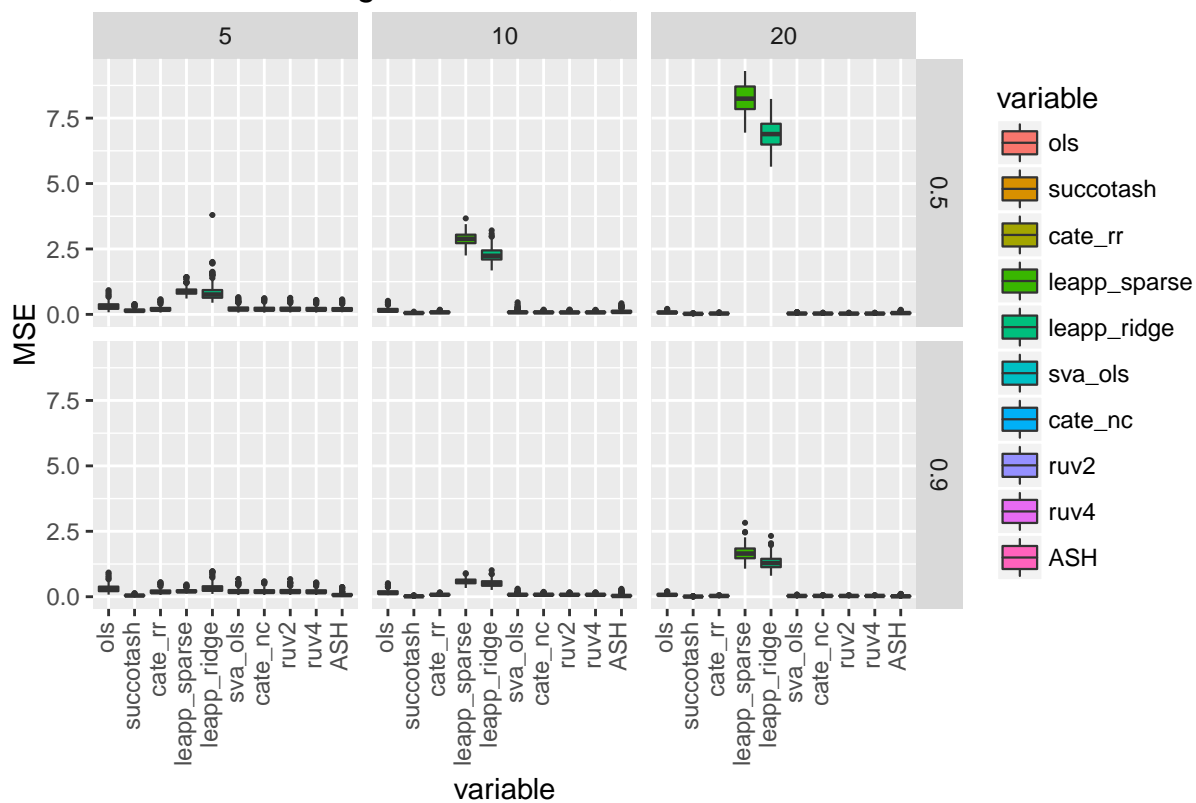
MSE When Using Muscle Tissue, Alternative = near\_normal



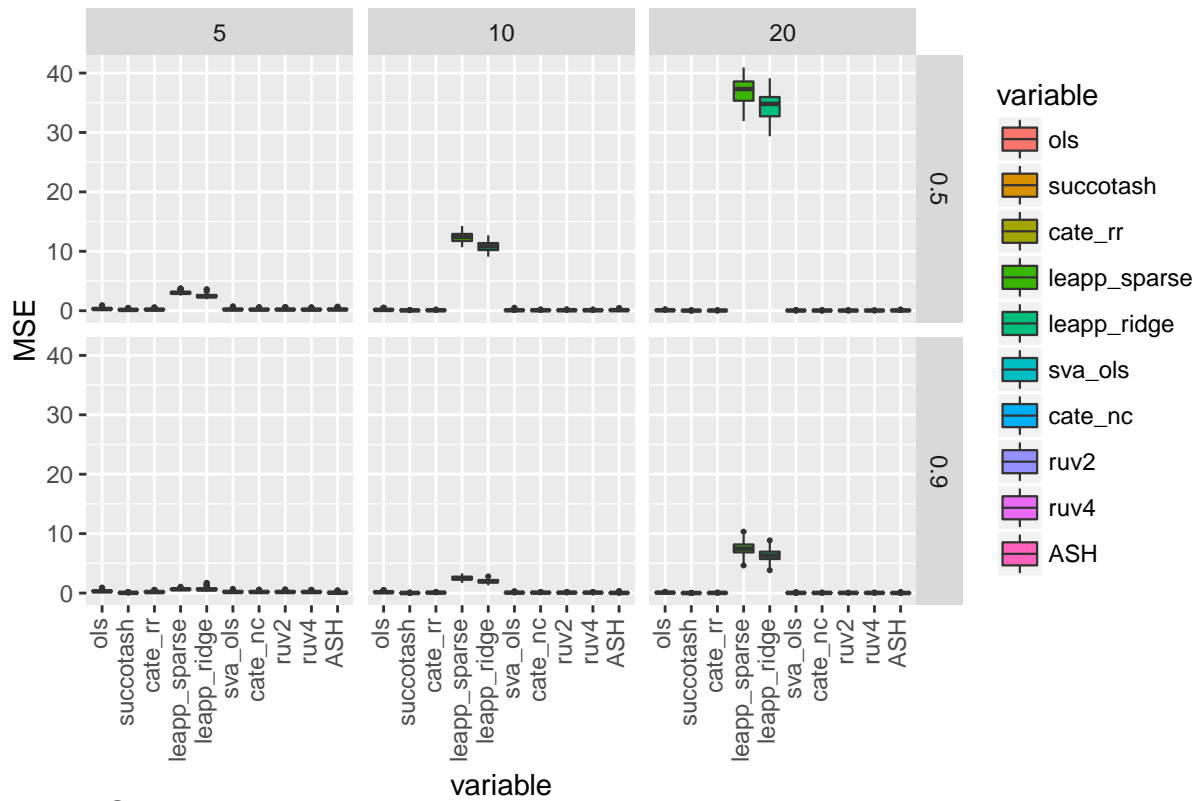
MSE When Using Muscle Tissue, Alternative = flattop



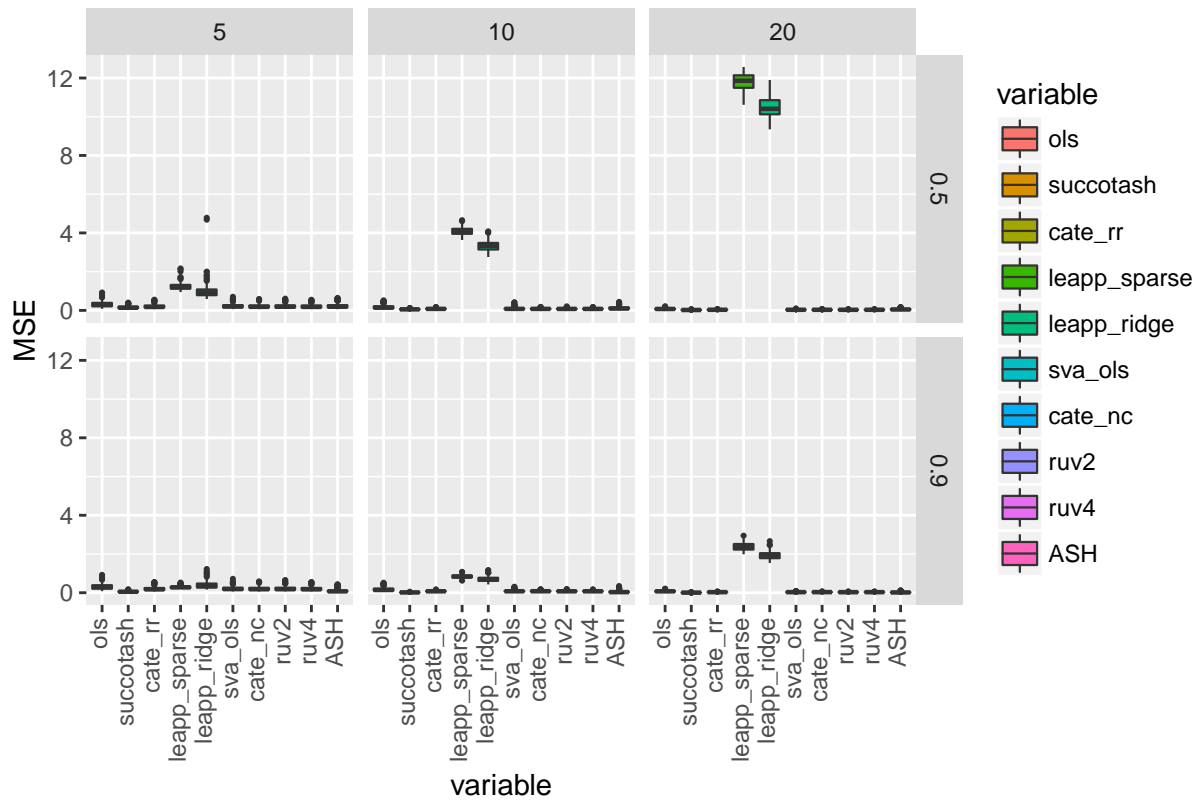
MSE When Using Muscle Tissue, Alternative = skew



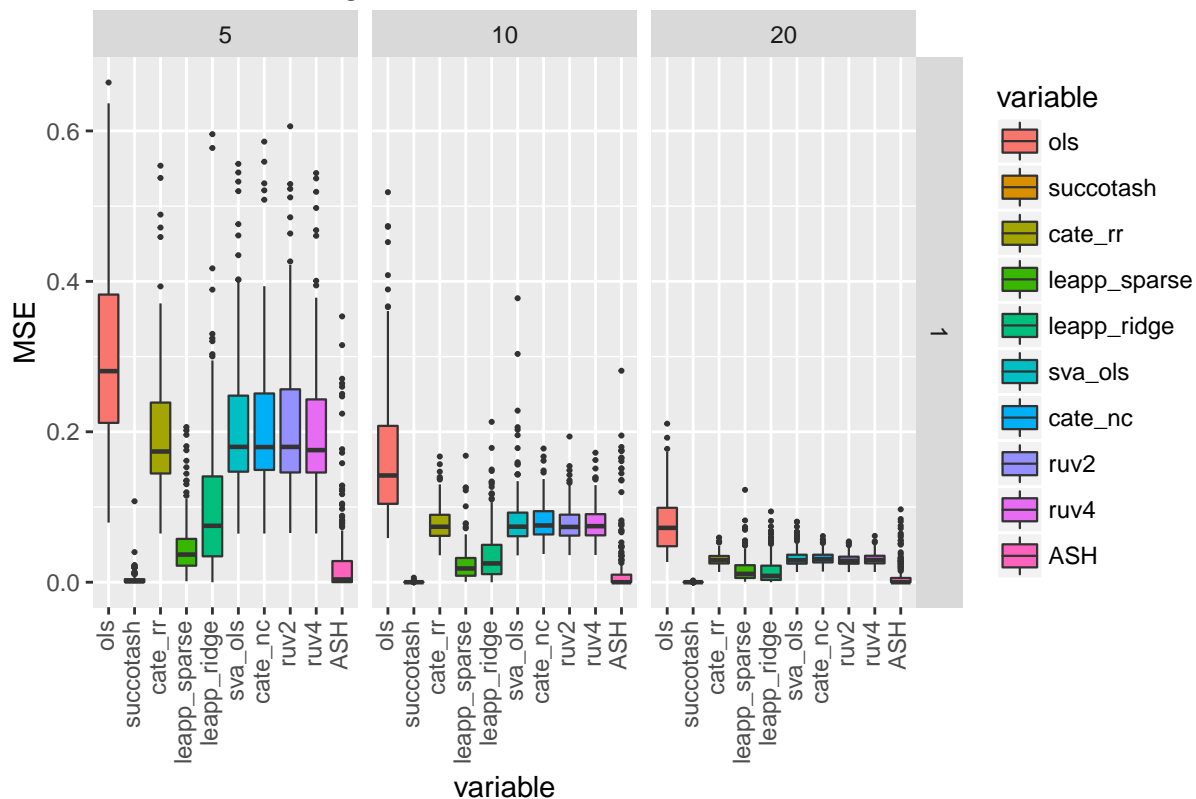
MSE When Using Muscle Tissue, Alternative = big\_normal



MSE When Using Muscle Tissue, Alternative = bimodal



## MSE When Using Muscle Tissue, Alternative = all\_null



`sessionInfo()`

```
## R version 3.3.0 (2016-05-03)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Ubuntu 14.04.4 LTS
##
## locale:
##  [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
##  [3] LC_TIME=en_US.UTF-8      LC_COLLATE=en_US.UTF-8
##  [5] LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
##  [7] LC_PAPER=en_US.UTF-8     LC_NAME=C
##  [9] LC_ADDRESS=C             LC_TELEPHONE=C
## [11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods    base
##
## other attached packages:
## [1] pROC_1.8      dplyr_0.4.3  reshape2_1.4.1 ggplot2_2.1.0
##
## loaded via a namespace (and not attached):
##  [1] Rcpp_0.12.4    knitr_1.12.28  magrittr_1.5    munsell_0.4.3
##  [5] colorspace_1.2-6 R6_2.1.2      stringr_1.0.0   plyr_1.8.3
##  [9] tools_3.3.0    parallel_3.3.0 grid_3.3.0      gtable_0.2.0
## [13] DBI_0.4        htmltools_0.3.5 yaml_2.1.13     lazyeval_0.1.10
## [17] assertthat_0.1 digest_0.6.9   formatR_1.3     codetools_0.2-14
## [21] evaluate_0.9   rmarkdown_0.9.6 labeling_0.3     stringi_1.0-1
```



## [25] compiler\_3.3.0 scales\_0.4.0

Buja, Andreas, and Nermin Eyuboglu. 1992. “Remarks on Parallel Analysis.” *Multivariate Behavioral Research* 27 (4). Taylor & Francis: 509–40.

Stephens, Matthew. 2016. “False Discovery Rates: A New Deal.” *BioRxiv*. Cold Spring Harbor Labs Journals, 038216.