Competitors when Non-null, Varying Sample Size, log2-fold standard deviation, and π_0.

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Abstract

I compare MOUTHWASH to various competitors. I look at AUC and estimates of π_0. I also look at the Kendall's Tau between the p-values or lfdr's for the different methods. This file runs the same analysis as varyNsampNullpiLog2sd.pdf but with smaller sample sizes and smaller log2sd.

1 Competitors

For each competitor, I performed two methods. The first method consisted of a two-step procedure:

1. Estimate $\hat{\beta}_{[2,i]}$ and it's corresponding standard error $\hat{s}_i$.
2. Run ASH on $\hat{\beta}_{[2,i]}$ and $\hat{s}_i$.

The second method was to just calculate a normal (or t, where appropriate) p-values from $\hat{\beta}_{[2,i]}/\hat{s}_i$.

The ASH methods provide an estimate of π_0. I obtained an estimate of π_0 from the p-values by the `qvalue` package in `R` [Storey, 2002]. In some cases for the quasi-binomial methods, the largest p-values were less than 0.9 and `qvalue` would return an error (because it uses the largest p-values to estimate the proportion of nulls). For these, I used the upper quartile of p-values to estimate the proportion of nulls. Maybe a bad idea.

The methods that Mengyin and I have coded to get $\hat{\beta}_{[2,i]}$ and $\hat{s}_i$ were

- VOOM [Law et al., 2014].
- RUVseq [Risso et al., 2014] followed by VOOM [Law et al., 2014] with the estimated confounding factors. Half of the factors were used as control genes. RUVseq is RUV2 on the log(counts + 1) matrix.
- SVAseq [Leek, 2014] followed by VOOM [Law et al., 2014] with the estimated confounding factors. SVAseq is SVA on the log(counts + 1) matrix.
- Quasi-binomial glm.
- RUVseq + quasi-binomial glm.
- SVAseq + quasi-binomial glm.
- MYRNA, which is just a quasi-binomial glm using the 75th percentile of the samples' counts as covariates [Langmead et al., 2010].
- MYRNA offset, which is just a quasi-binomial glm using the 75th percentile of the samples' counts as offsets [Langmead et al., 2010].
- DESeq2glm [Love et al., 2014].
- The robust regression version of CATE [Wang et al., 2015] on the log(counts + 1).

- The negative controls version of CATE [Wang et al., 2015] on the log(counts + 1).
- SVA [Leek and Storey, 2007] with the number of confounders estimated using the method of Buja and Eyuboglu [1992] on the log(counts + 1), followed by OLS.
- RUV2 [Gagnon-Bartsch et al., 2013] with 50% of the observations being control genes with the number of confounders estimated using the method of Buja and Eyuboglu [1992] on the log(counts + 1), followed by OLS.
- OLS on the log(counts + 1).
- The ridge-regression version of LEAPP [Sun et al., 2012] on the log(counts + 1).
- The soft-thresholding version of LEAPP [Sun et al., 2012] on the log(counts + 1).

Notes:

- "VOOM" means using VOOM [Law et al., 2014] to find weights for each observations, then fitting a linear model using LIMMA [Smyth, 2005].
- LEAPP does not easily provide standard errors, so I excluded it from the ASH analysis. But I still use it for the `qvalue` analysis.
- EdgeR was giving me trouble, so I excluded it.

The factor analysis part of MOUTHWASH was done with the quasi-mle approach of Bai et al. [2012] with the number of hidden confounders using the methods of Buja and Eyuboglu [1992] implemented in the `num.sv()` function in the `sva` package in `R`.

In summary, there are 31 methods that I compared in estimating π_0 and in their AUC:

1. MOUTHWASH
2. voom + ASH
3. Quasi-binomial GLM + ASH
4. MyrnaQB + ASH
5. Myrnaoffqb + ASH
6. RUVseq + voom + ASH
7. SVAseq + voom + ASH
8. RUVseq + Quasi-binomial GLM + ASH
9. SVAseq + Quasi-binomial GLM + ASH
10. DESeq2glm + ASH
11. OLS on log(counts + 1) + ASH
12. RUV2 on log(counts + 1) + ASH
13. SVA on log(counts + 1) + ASH
14. Robust Regression Cate + ASH
15. Negative Control CATE + ASH
16. voom + `qvalue`
17. Quasi-binomial GLM + `qvalue`
18. Myrnaqb + `qvalue`
19. Myrnaoffqb + `qvalue`
20. RUVseq + voom + `qvalue`
21. SVAseq + voom + `qvalue`
22. RUVseq + Quasi-binomial GLM + `qvalue`
23. SVAseq + Quasi-binomial GLM + `qvalue`
24. DESeq2glm + `qvalue`
25. OLS on log(counts + 1) + `qvalue`
26. RUV2 on log(counts + 1) + `qvalue`

27. SVA on log(counts + 1) + `qvalue`
28. Robust Regression CATE + `qvalue`
29. Negative Control CATE + `qvalue`
30. Soft-thresholding version of LEAPP+ `qvalue`
31. Ridge version of LEAPP+ `qvalue`

2 Simulation Study

I ran through 100 repetitions of generating data from GTEX lung data under the following parameter conditions:

- $n \in \{3, 5\}$,
- $p = 1000$,
- $\pi_0 \in \{0.5, 0.9\}$,
- $\sigma_{log2} \in \{1, 5\}$.

I extracted the most expressed p genes (excluding the top 5 expressed genes) from the GTEX lung data and n samples are chosen at random. Half of these samples are randomly given the "treatment" label 1, the other half given the "treatment" label 0. Of the p genes, $\pi_0 p$ were chosen to be non-null. Signal was added by the Poisson-thinning approach in Mengyin's code with a mean log2-fold change of 0 and a standard deviation log2-fold change of σ_{log2}. That is

$$A_1, \ldots, A_{p/2} \sim N(0, \sigma^2_{log2}) \tag{1}$$

$$B_i = 2^{A_i} \text{ for } i = 1, \ldots, p/2. \tag{2}$$

If $A_i > 0$ then we replace $Y_{[1:(n/2),i]}$ with $Binom(Y_{[j,i]}, 1/B_i)$ for $j = 1, \ldots, n/2$. If $A_i < 0$ then we replace $Y_{[(n/2+1):n,i]}$ with $Binom(Y_{[j,i]}, B_i)$ for $j = n/2 + 1, \ldots, n$.

For each iteration, I calculated three things:

1. The pairwise Kendall's tau between the methods' lfdr's or p-values.
2. The AUC using either the lfdrs or p-values.
3. The estimates of π_0.

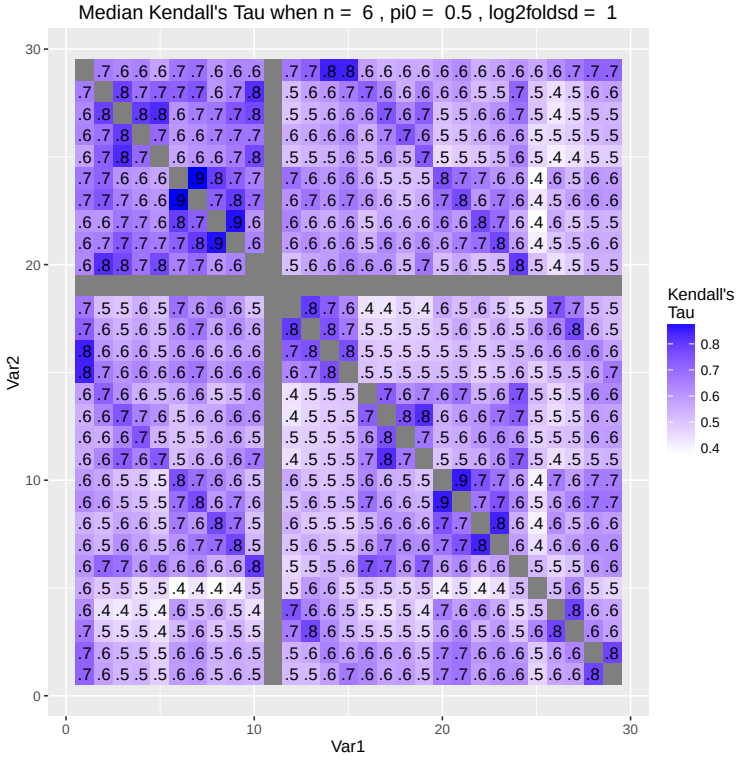
3 Results

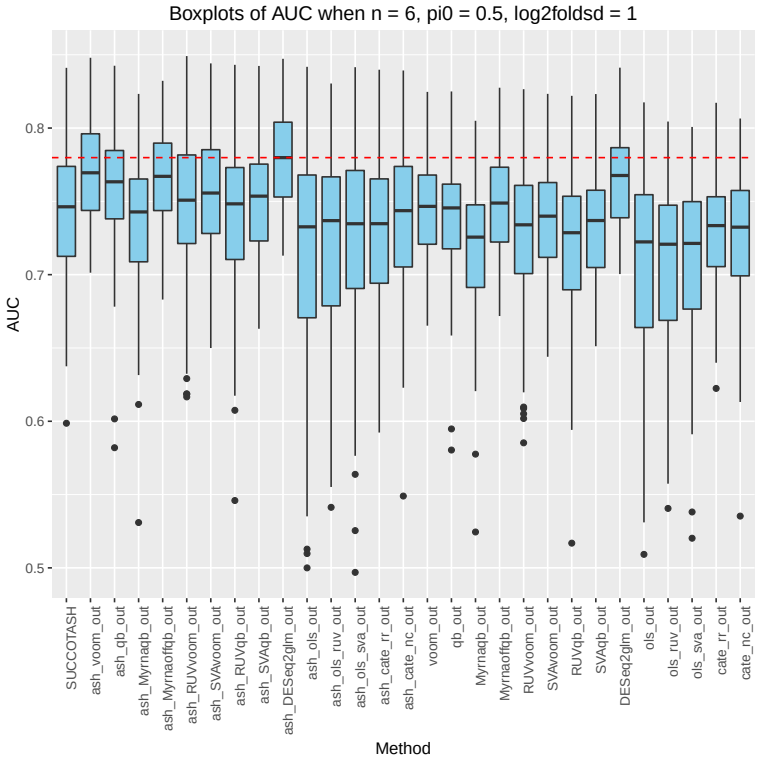
For the frequentist procedures, I used the vector of p-values as the predictions and I used the vector of lfdr's from the ASH-like procedures for prediction. These were used to create ROC curves and calculate AUCs. In general, the AUC's were all very similar with the ash-like methods having slightly higher AUC. DESeq2glm (with or without ASH) is the winner when the sample size is 3 vs 3. The results are less clear for 5 vs 5.

Graphical representations of the median Kendall's taus are presented below. The median Kendall's tau can get quite low between the separate groups — as small as 0.2. This indicates that for many datasets, the rankings can be quite different.

From the p-values, I used the `qvalue` package [Storey, 2002] to estimate π_0. Estimates of π_0 are given from `ashr` for the ASH-like methods. MOUTHWASH (SUCCOTASH) performs the worst in estimating π_0, usually underestimating it. The ASH-like methods usually estimate π_0 to be smaller

than their non-ASH counterparts. Notably, the negative controls version of CATE does not excel in the small sample size scenario.


Median Kendall's Tau when n = 6 , pi0 = 0.5 , log2foldsd = 1
Var2
Var1
Kendall's Tau
0.8
0.7
0.6
0.5
0.4
0
10
20
30


Boxplots of AUC when n = 6, pi0 = 0.5, log2foldsd = 1
AUC
Method
0.8
0.7
0.6
0.5
SUCCOTASH
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ash_qb_out
ash_Mymaqb_out
ash_Myrnaoffqb_out
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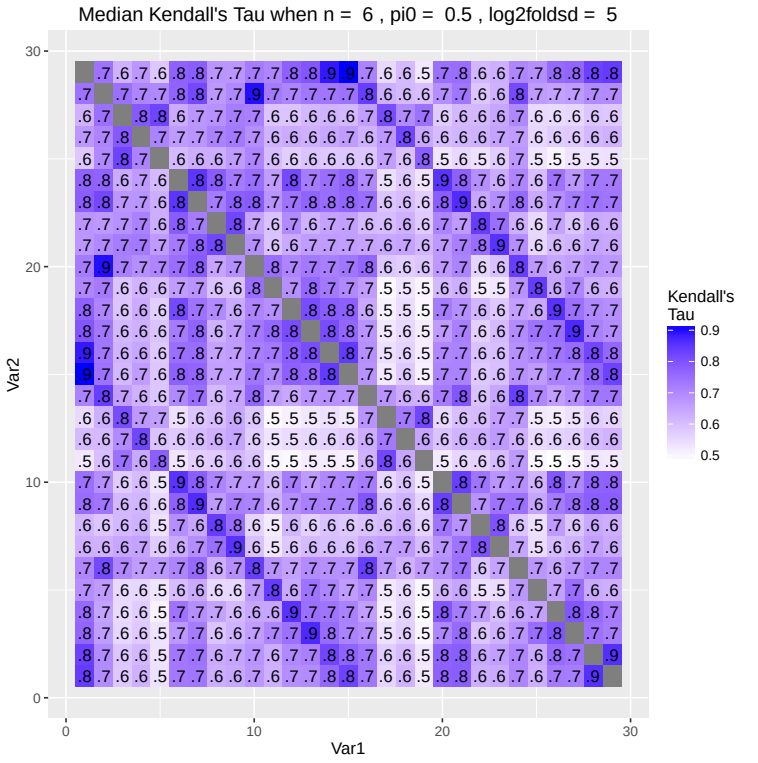
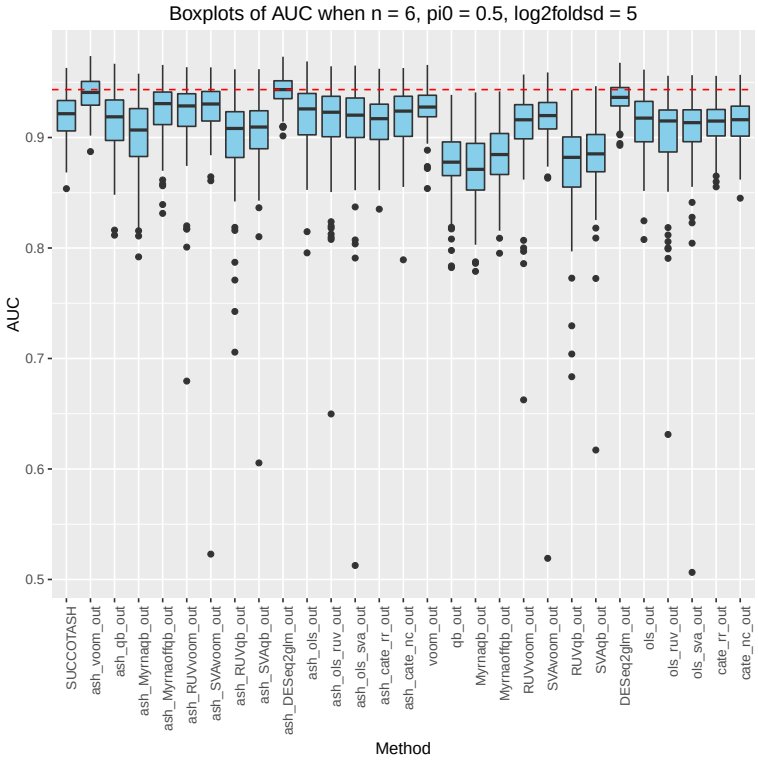

Median Kendall's Tau when n = 6 , pi0 = 0.5 , log2foldsd = 5
Var2
Var1
Kendall's Tau
0.9
0.8
0.7
0.6
0.5
0
10
20
30


Boxplots of AUC when n = 6, pi0 = 0.5, log2foldsd = 5
AUC
Method
0.9
0.8
0.7
0.6
0.5
SUCCOTASH
ash_voom_out
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ash_Mymadb_out
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Mymadb_out
Myrnaoffqb_out
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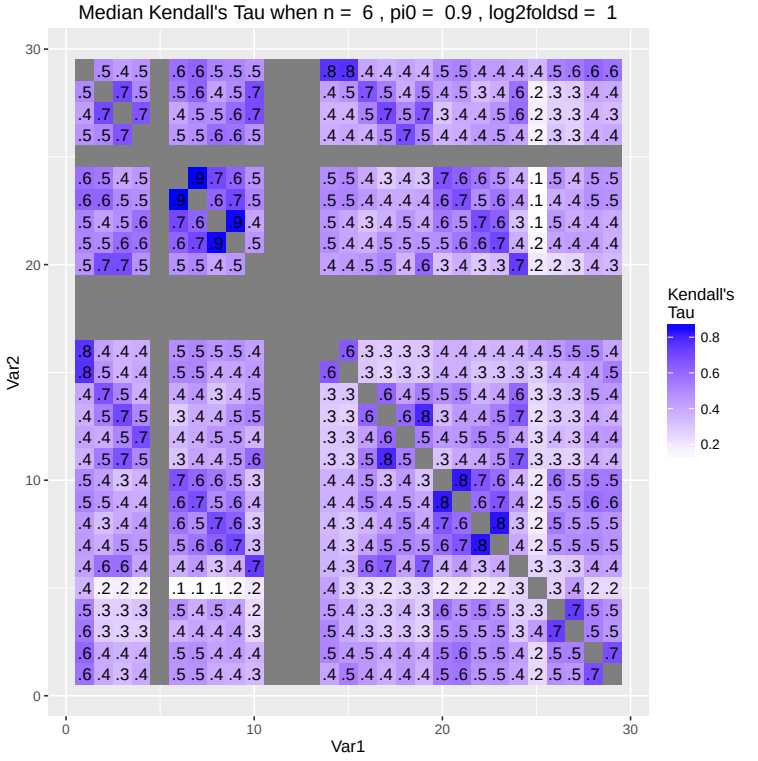
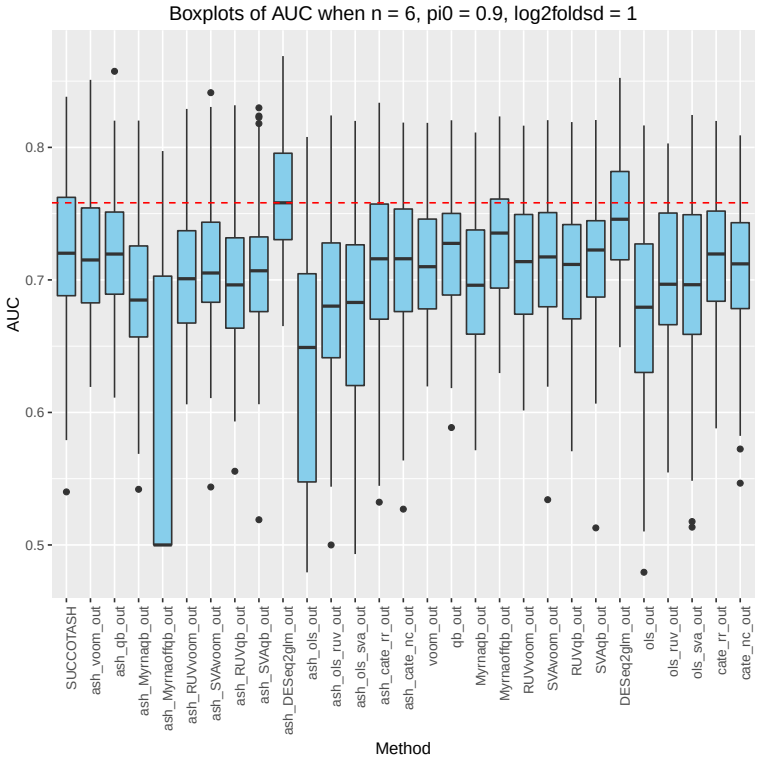

Median Kendall's Tau when n = 6 , pi0 = 0.9 , log2foldsd = 1
Var2
Var1
Kendall's Tau
0.8
0.6
0.4
0.2
0
10
20
30


Boxplots of AUC when n = 6, pi0 = 0.9, log2foldsd = 1
AUC
Method
0.8
0.7
0.6
0.5
SUCCOTASH
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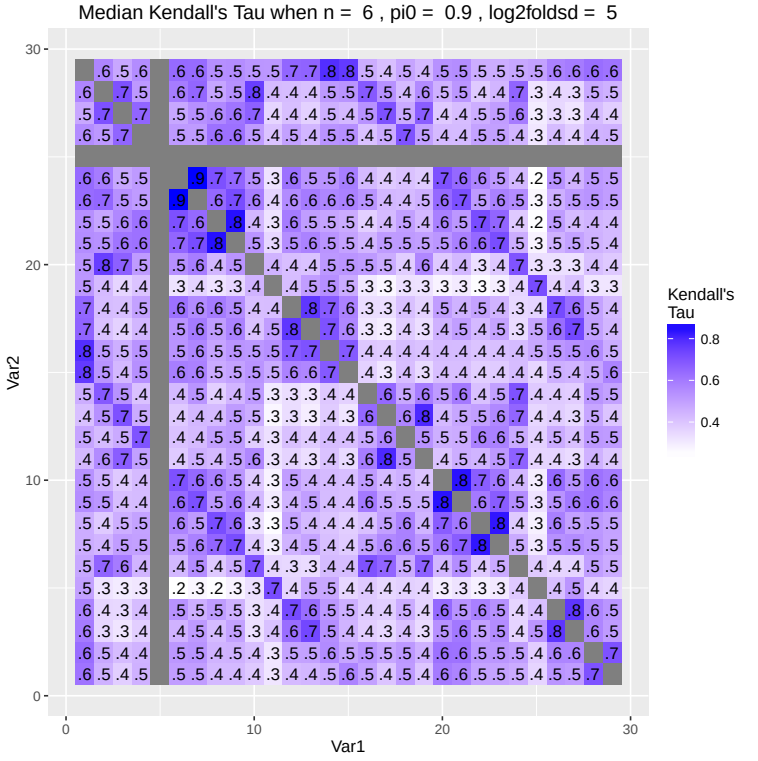
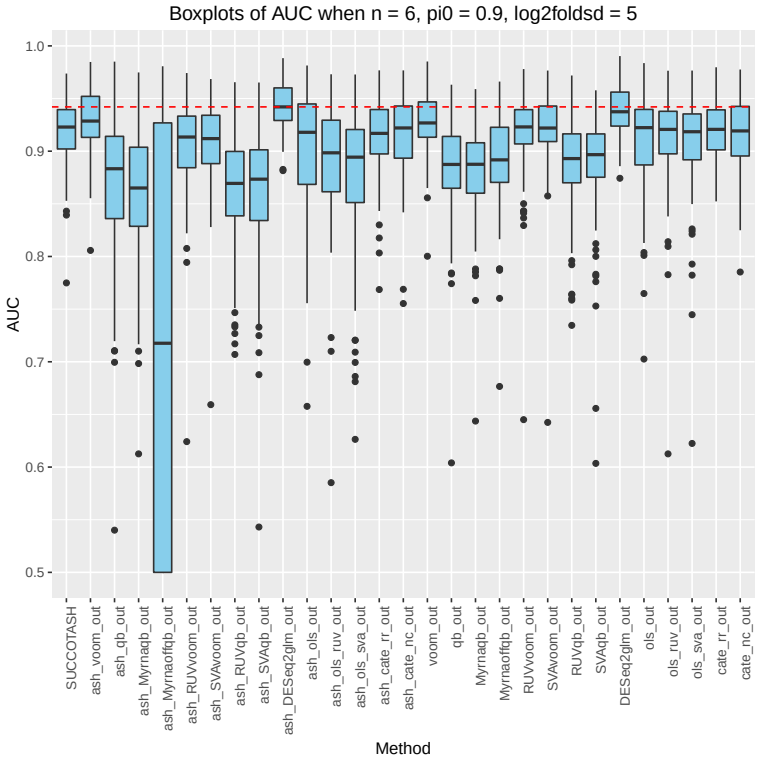

Median Kendall's Tau when n = 6 , pi0 = 0.9 , log2foldsd = 5
Var1
Var2
Kendall's Tau
0.8
0.6
0.4
0
10
20
30


Boxplots of AUC when n = 6, pi0 = 0.9, log2foldsd = 5
AUC
Method
1.0
0.9
0.8
0.7
0.6
0.5
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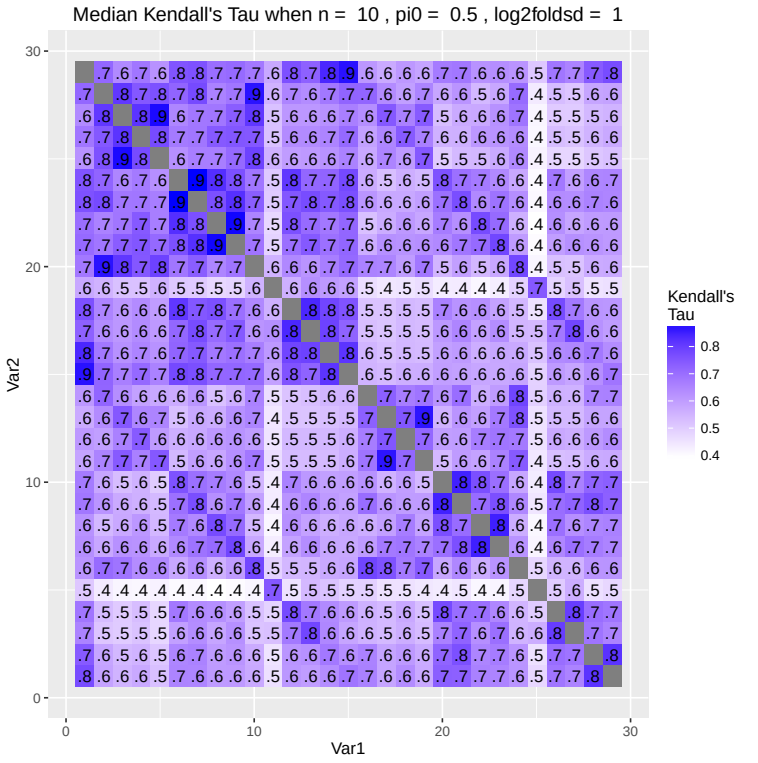
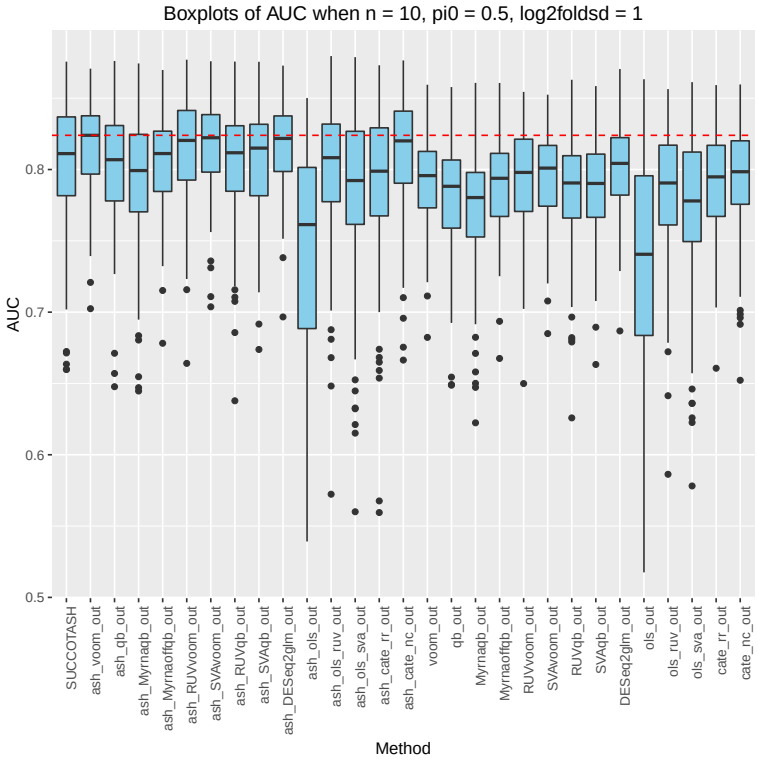
cate_nc_out


Median Kendall's Tau when n = 10 , pi0 = 0.5 , log2foldsd = 1
Var1
Var2
Kendall's Tau
0.8
0.7
0.6
0.5
0.4
0
10
20
30


Boxplots of AUC when n = 10, pi0 = 0.5, log2foldsd = 1
AUC
Method
0.5
0.6
0.7
0.8
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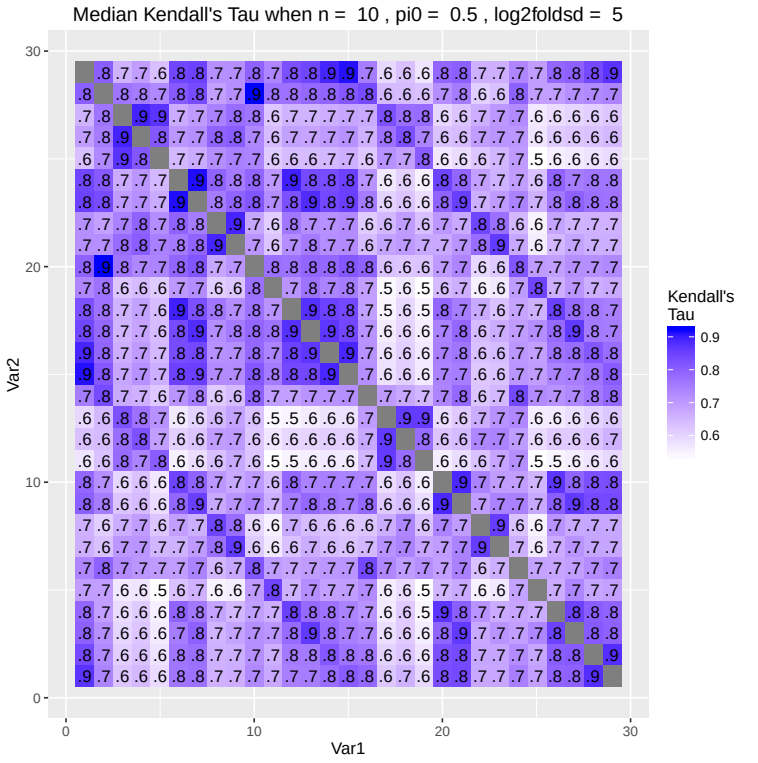
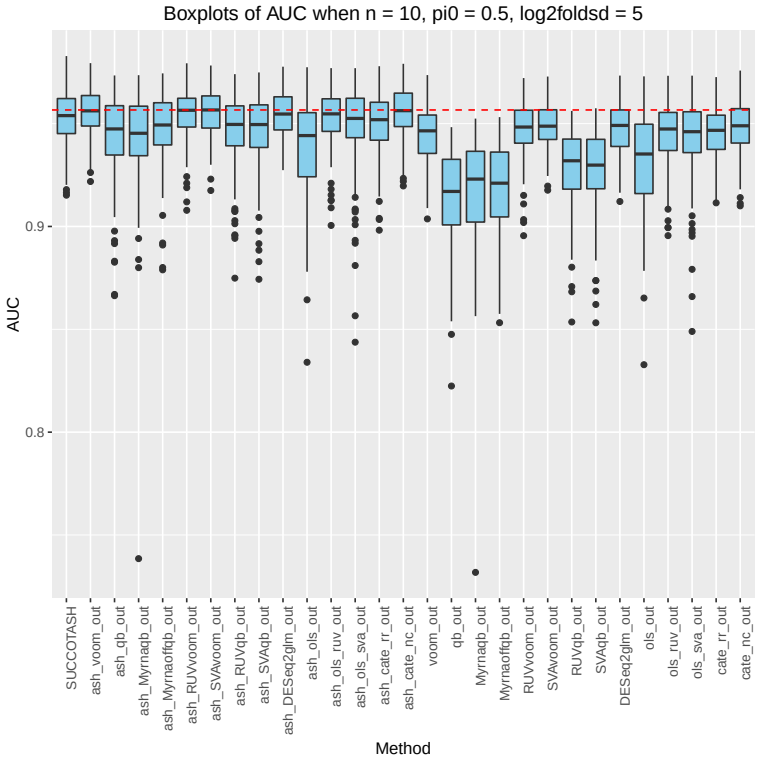

Median Kendall's Tau when n = 10 , pi0 = 0.5 , log2foldsd = 5
Var1
Var2
Kendall's Tau
0.9
0.8
0.7
0.6


Boxplots of AUC when n = 10, pi0 = 0.5, log2foldsd = 5
AUC
Method
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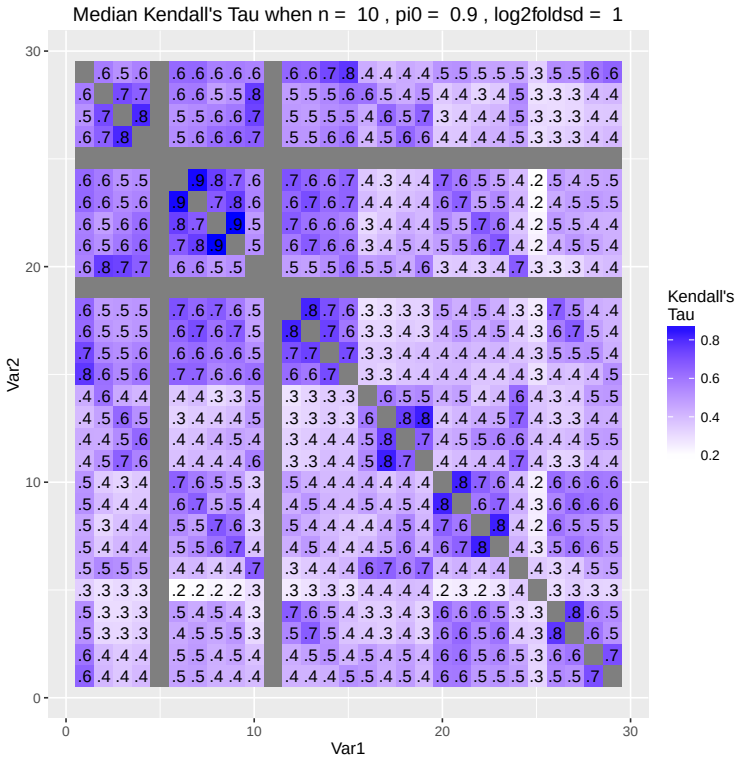
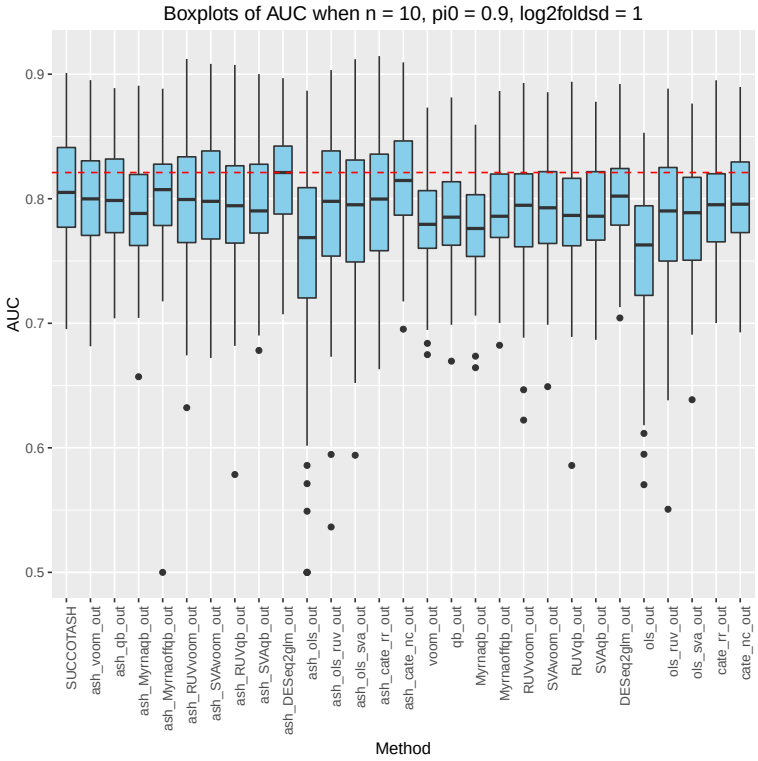

Median Kendall's Tau when n = 10 , pi0 = 0.9 , log2foldsd = 1
Var2
Var1
Kendall's Tau
0.8
0.6
0.4
0.2
0
10
20
30


Boxplots of AUC when n = 10, pi0 = 0.9, log2foldsd = 1
AUC
Method
0.9
0.8
0.7
0.6
0.5
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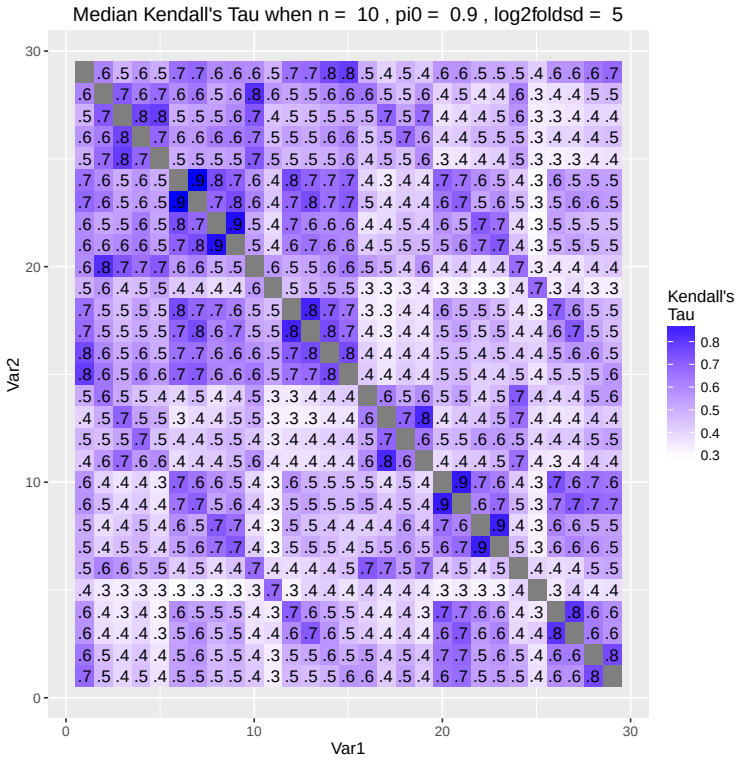
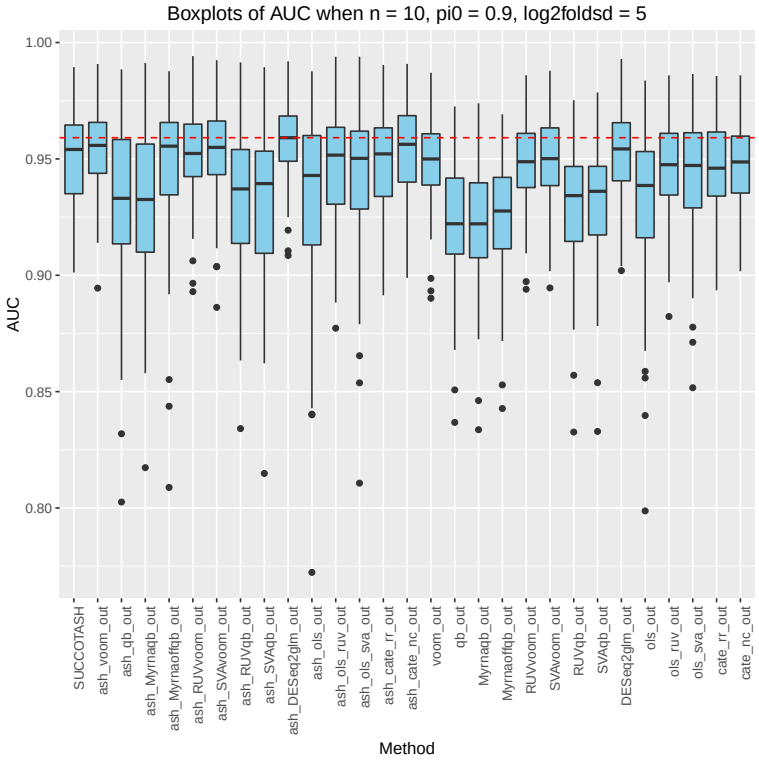

Median Kendall's Tau when n = 10 , pi0 = 0.9 , log2foldsd = 5
Var2
Var1
Kendall's Tau
0.8
0.7
0.6
0.5
0.4
0.3
0
10
20
30


Boxplots of AUC when n = 10, pi0 = 0.9, log2foldsd = 5
AUC
1.00
0.95
0.90
0.85
0.80
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ash_SVAqb_out
ash_DESeq2glm_out
ash_ols_out
ash_ols_ruv_out
ash_ols_sva_out
ash_cate_rr_out
ash_cate_nc_out
voom_out
qb_out
Myrnaqb_out
Myrnaoffqb_out
RUVvoom_out
SVAvoom_out
RUVqb_out
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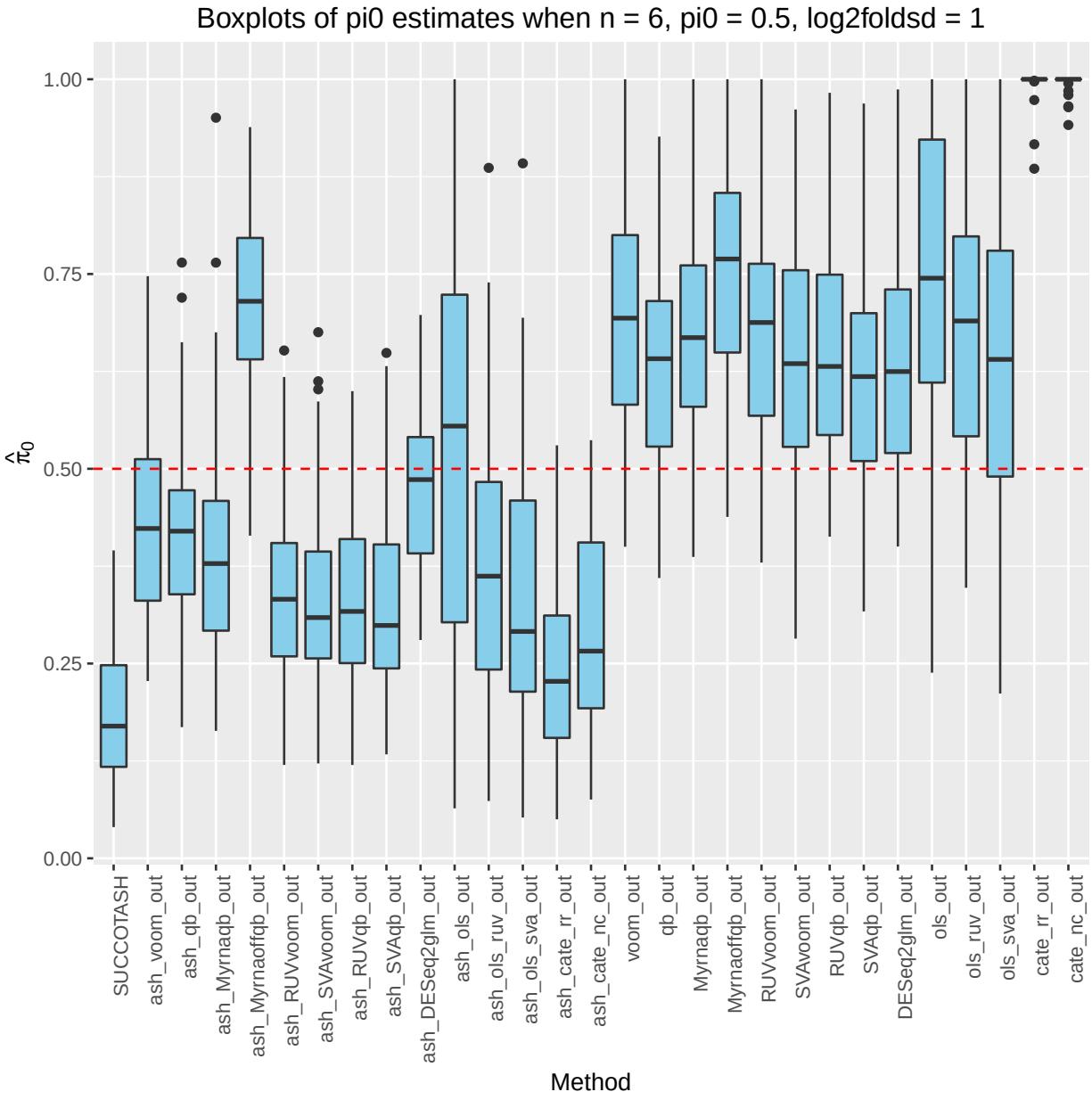
Method


Boxplots of pi0 estimates when n = 6, pi0 = 0.5, log2foldsd = 1
1.00
0.75
0.50
0.25
0.00
$\hat{\pi}_0$
SUCCOTASH
ash_voom_out
ash_qb_out
ash_Myrnaqb_out
ash_Myrnaoffqb_out
ash_RUVvoom_out
ash_SVAvoom_out
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ash_ols_out
ash_ols_ruv_out
ash_ols_sva_out
ash_cate_rr_out
ash_cate_nc_out
voom_out
qb_out
Myrnaqb_out
Myrnaoffqb_out
RUVvoom_out
SVAvoom_out
RUVqb_out
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ols_out
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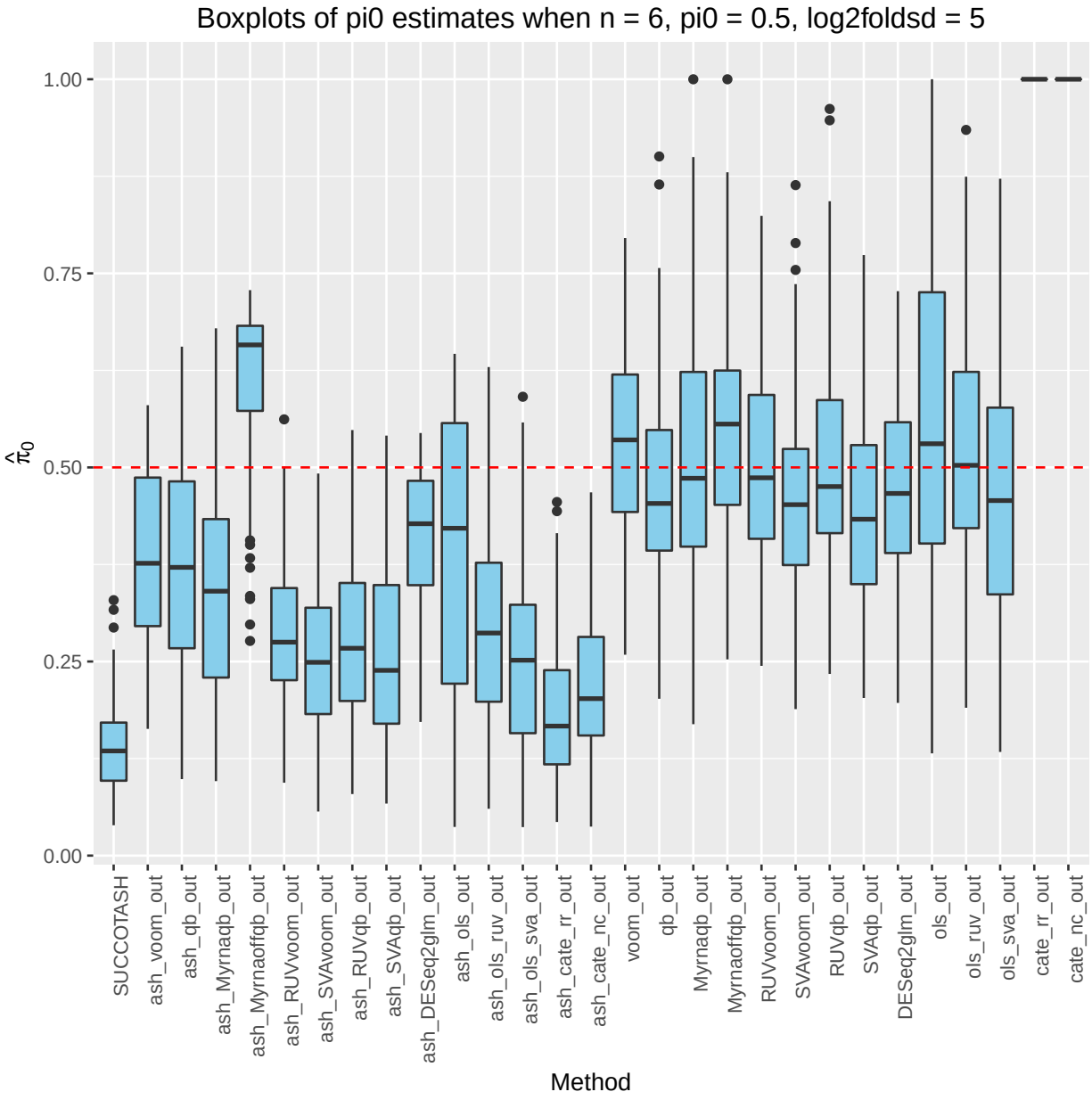
ols_sva_out
cate_rr_out
cate_nc_out
Method


Boxplots of pi0 estimates when n = 6, pi0 = 0.5, log2foldsd = 5
1.00
0.75
0.50
0.25
0.00
$\hat{\pi}_0$
SUCCOTASH
ash_voom_out
ash_qb_out
ash_Myrnaqb_out
ash_Myrnaoffqb_out
ash_RUVvoom_out
ash_SVAvoom_out
ash_RUVqb_out
ash_SVAqb_out
ash_DESeq2glm_out
ash_ols_out
ash_ols_ruv_out
ash_ols_sva_out
ash_cate_rr_out
ash_cate_nc_out
voom_out
qb_out
Myrnaqb_out
Myrnaoffqb_out
RUVvoom_out
SVAvoom_out
RUVqb_out
SVAqb_out
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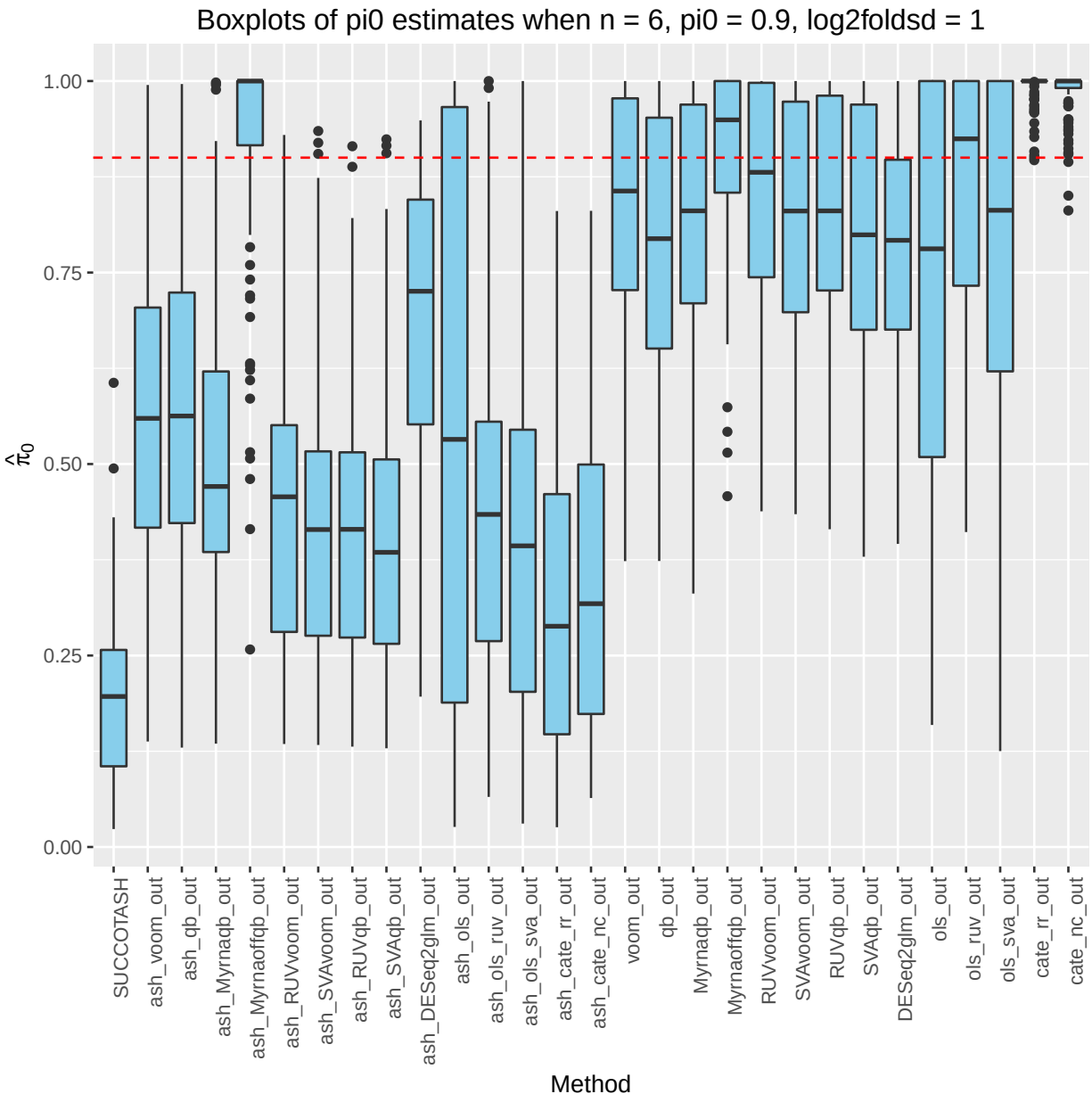
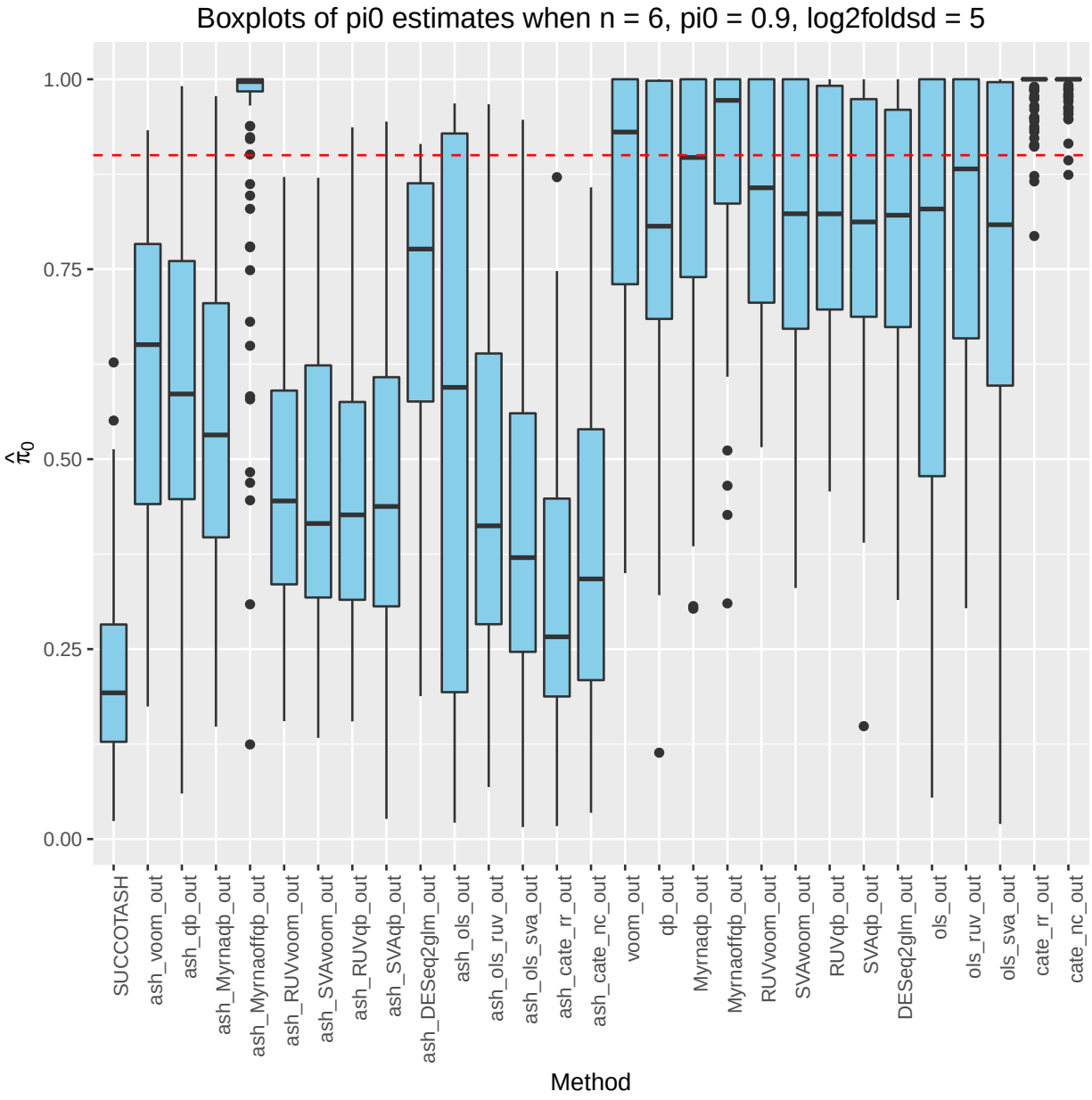
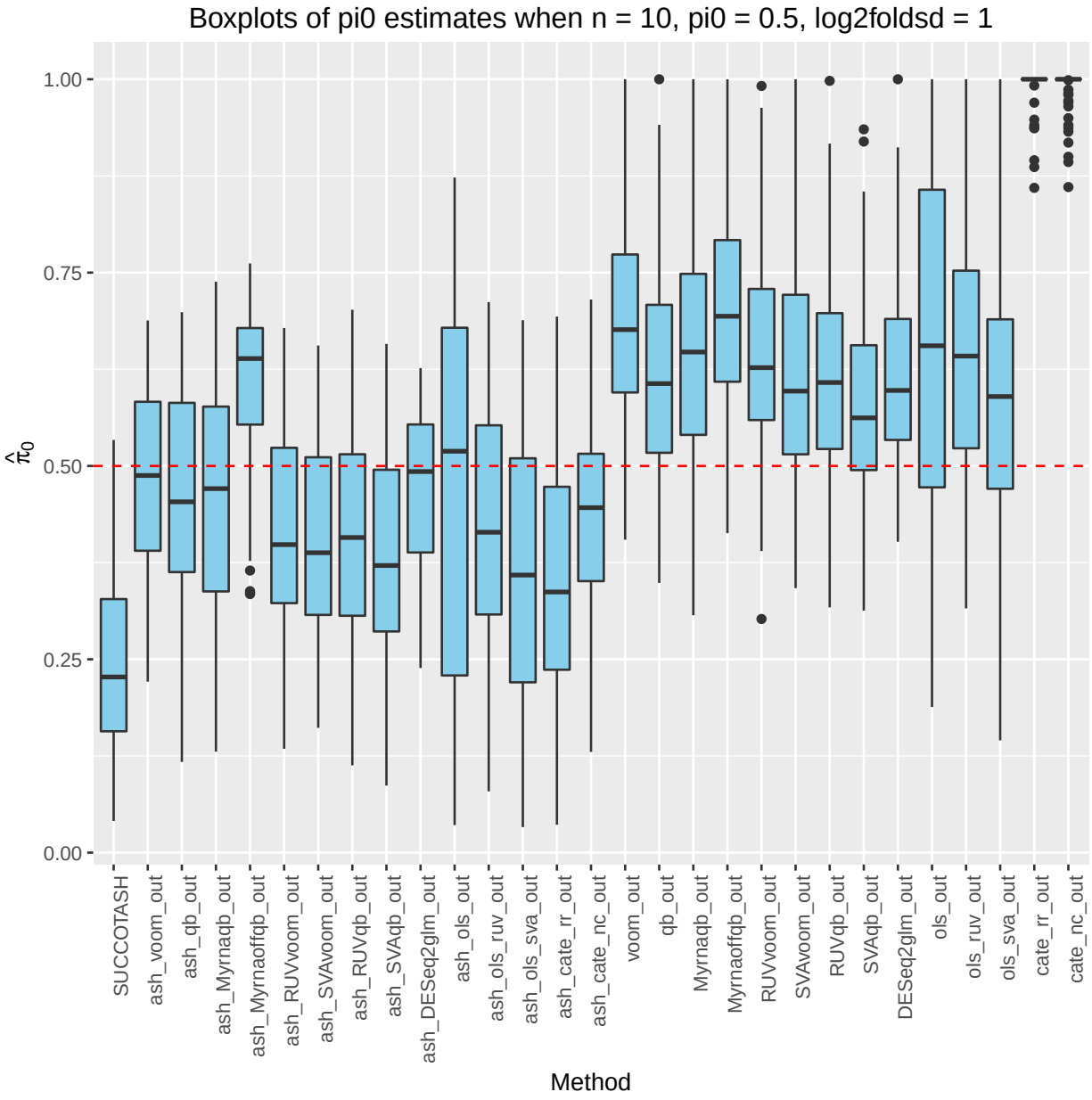
ols_out
ols_ruv_out
ols_sva_out
cate_rr_out
cate_nc_out
Method


Boxplots of pi0 estimates when n = 6, pi0 = 0.9, log2foldsd = 1
1.00
0.75
0.50
0.25
0.00
$\hat{\pi}_0$
SUCCOTASH
ash_voom_out
ash_qb_out
ash_Myrnaqb_out
ash_Myrnaoffqb_out
ash_RUVvoom_out
ash_SVAvoom_out
ash_RUVqb_out
ash_SVAqb_out
ash_DESeq2glm_out
ash_ols_out
ash_ols_ruv_out
ash_ols_sva_out
ash_cate_rr_out
ash_cate_nc_out
voom_out
qb_out
Myrnaqb_out
Myrnaoffqb_out
RUVvoom_out
SVAvoom_out
RUVqb_out
SVAqb_out
DESeq2glm_out
ols_out
ols_ruv_out
ols_sva_out
cate_rr_out
cate_nc_out
Method


Boxplots of pi0 estimates when n = 6, pi0 = 0.9, log2foldsd = 5
1.00
0.75
0.50
0.25
0.00
$\hat{\pi}_0$
SUCCOTASH
ash_voom_out
ash_qb_out
ash_Myrnaqb_out
ash_Myrnaoffqb_out
ash_RUVvoom_out
ash_SVAvoom_out
ash_RUVqb_out
ash_SVAqb_out
ash_DESeq2glm_out
ash_ols_out
ash_ols_ruv_out
ash_ols_sva_out
ash_cate_rr_out
ash_cate_nc_out
voom_out
qb_out
Myrnaqb_out
Myrnaoffqb_out
RUVvoom_out
SVAvoom_out
RUVqb_out
SVAqb_out
DESeq2glm_out
ols_out
ols_ruv_out
ols_sva_out
cate_rr_out
cate_nc_out
Method


Boxplots of pi0 estimates when n = 10, pi0 = 0.5, log2foldsd = 1
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0.75
0.50
0.25
0.00
$\hat{\pi}_0$
SUCCOTASH
ash_voom_out
ash_qb_out
ash_Myrnaqb_out
ash_Myrnaoffqb_out
ash_RUVvoom_out
ash_SVAvoom_out
ash_RUVqb_out
ash_SVAqb_out
ash_DESeq2glm_out
ash_ols_out
ash_ols_ruv_out
ash_ols_sva_out
ash_cate_rr_out
ash_cate_nc_out
voom_out
qb_out
Myrnaqb_out
Myrnaoffqb_out
RUVvoom_out
SVAvoom_out
RUVqb_out
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DESeq2glm_out
ols_out
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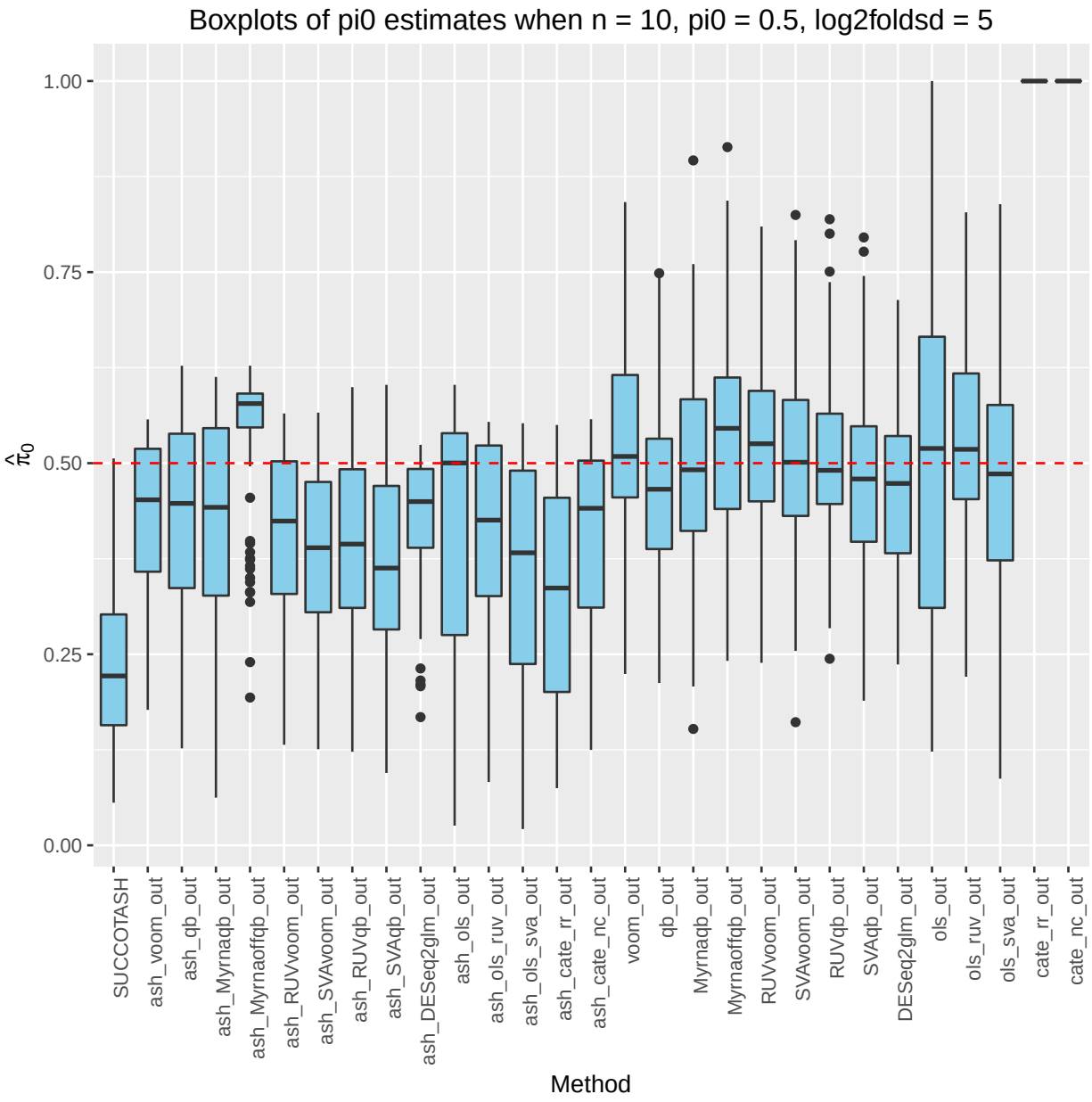
cate_rr_out
cate_nc_out
Method


Boxplots of pi0 estimates when n = 10, pi0 = 0.5, log2foldsd = 5
1.00
0.75
0.50
0.25
0.00
$\hat{\pi}_0$
SUCCOTASH
ash_voom_out
ash_qb_out
ash_Myrnaqb_out
ash_Myrnaoffqb_out
ash_RUVvoom_out
ash_SVAvoom_out
ash_RUVqb_out
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ash_ols_out
ash_ols_ruv_out
ash_ols_sva_out
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voom_out
qb_out
Myrnaqb_out
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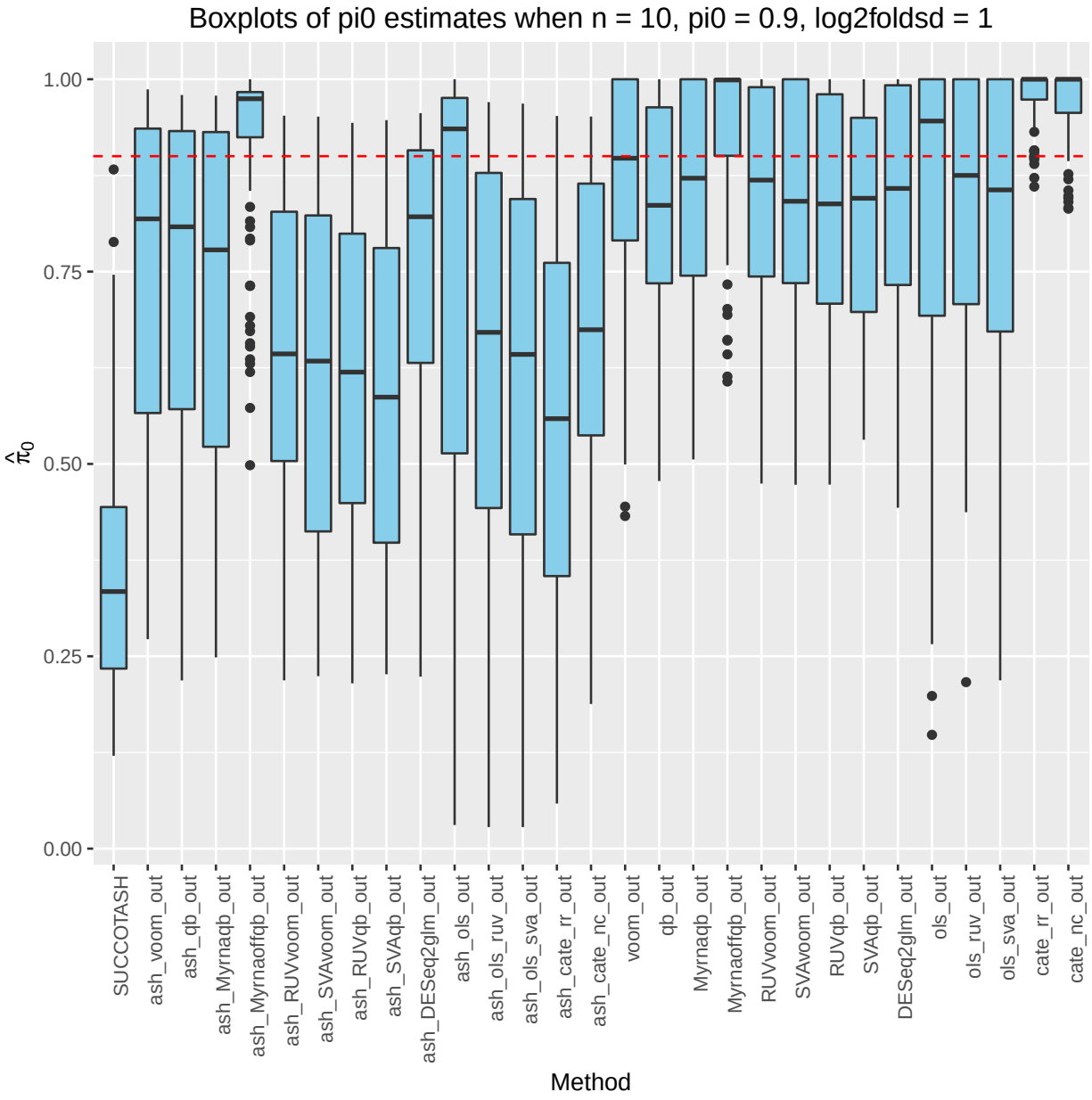
Boxplots of pi0 estimates when n = 10, pi0 = 0.9, log2foldsd = 1
1.00
0.75
0.50
0.25
0.00
$\hat{\pi}_0$
SUCCOTASH
ash_voom_out
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ash_Myrnaqb_out
ash_Myrnaoffqb_out
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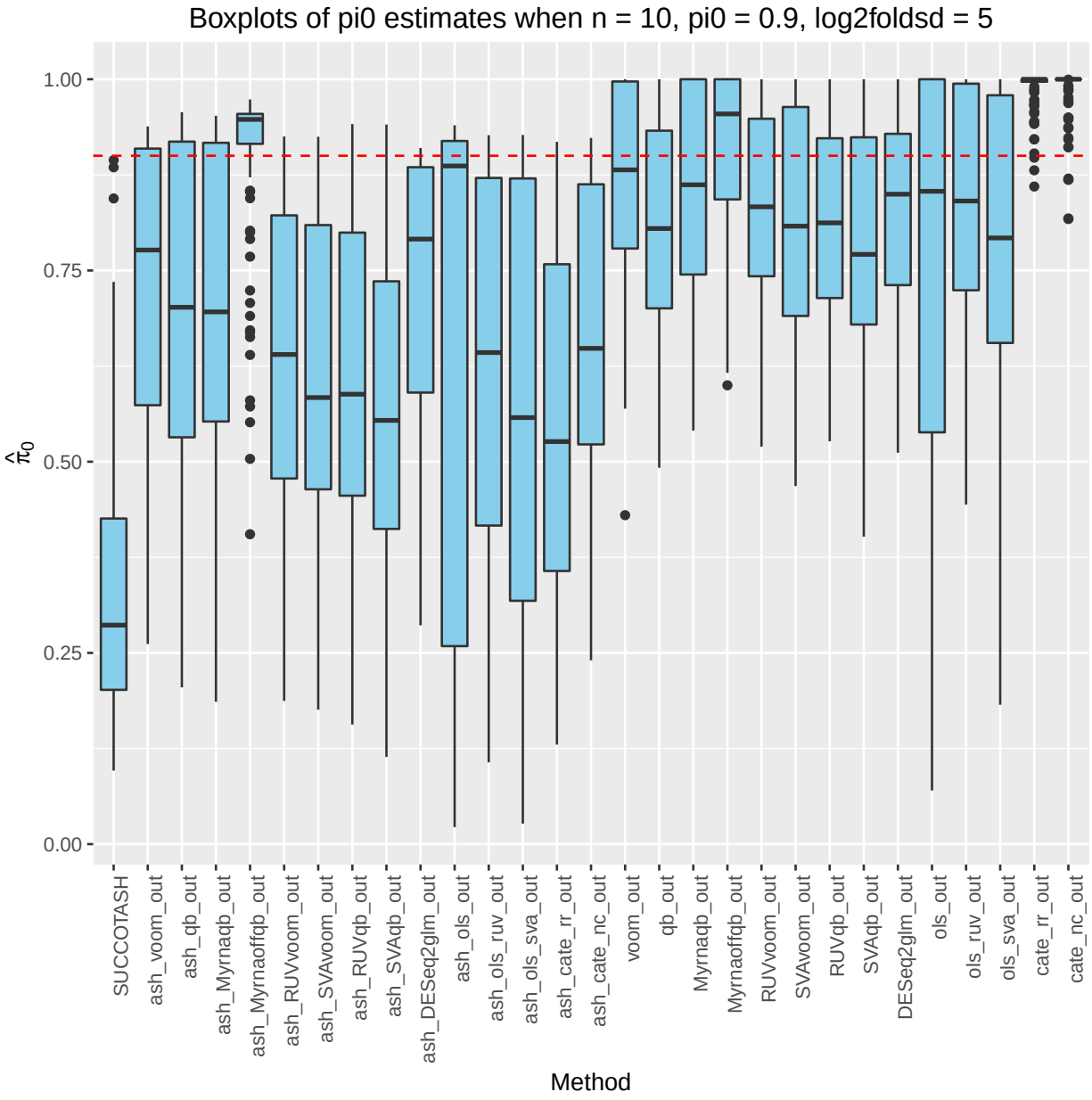
Method



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