# Measuring Energy Intake via Energy Balance Equation while Accounting for Measurement Error

### Daniel Ries

#### November 15, 2016

# 1 Introduction

Accurately measuring one's energy intake (EI measured in kcals consumed) is a very difficult task [1],[2],[3]. This is especially true for measuring energy intake in free living conditions and not in inpatient, controlled settings [4]. It has been said that our inability to measure energy intake accurately is the one of the biggest pitfalls of obesity research [5], [6]. According to the National Institute of Diabetes and Digestive and Kidney Diseases, 66.8% of adults in the US are overweight or obese, and 35.7% are obese [7]. Obesity has been linked to a multitude of negative health effects like cardiovascular disease [8], high blood pressure, Type 2 diabetes, some cancers, stroke, osteoarthritis, and mental illness [9], [10], [11]. It has been found that the number of chronic conditions due to obesity is larger than that of smoking or drinking [12]. Because of this, obesity has been a hot topic in legislation and the health care industry. Being able to accurately measure energy intake would be extremely useful to monitor patient compliance with set diets [13]. The so called 6 month plateau, which is the typical flattening of weight loss seen by those on weight loss programs after 6 months, has recently been attributed to noncompliance rather than metabolic adaptation [14], [15]. This is important because validated models show this weight loss should continue for between one and two years. This is an issue because the patients still report diet adherence [15]. It has been argued that mathematical modeling could help in assessment of adherence in diet interventions [16], [17]. Dynamic mathematical models to predict an individual's energy intake have started to gain attention, partly due to the now widespread knowledge that Wishnofsky's assertion that one pound

of body mass is equivalent to 3500 calories [18] is false. These models have shown the benefits of collaboration across multiple disciplines by advancing the field of obesity research greatly. However, these models use as an input data that is measured with error. In this dissertation, we develop statistical methods to assess the extent of measurement error and bias and calibrate the noisy measurements that in turn will make these new mathematical models even more useful.

In this chapter we review methods for measuring EI and after introducing the energy balance principle, methods for measuring energy expenditure (EE) and changes in energy stores  $\Delta$ ES. We then discuss modeling energy balance as a whole. We then explore statistical methods for analyzing data with measurement error.

#### 1.1 Measurement Instruments

### 1.1.1 Measuring Energy Intake

There are several ways one can estimate the energy intake of a free living human. One of the more common methods, a self-report via a food frequency questionnaire (FFQ), is known to be biased and an unreliable measure [19], [20], [21]. This method, however, continues to be popular due to its low cost and ease to implement on a large sample. A FFQ is a checklist that asks one to mark how many times one ate a specified food during a specified period of time. Portion size may or may not be collected. 24 hour recalls are another popular option [?]. These are typically conducted over the phone or sometimes in person. A trained dietician using administers these recalls. These are popular because they are relatively cheap compared to other methods, and they only require remembering 24 hours of food consumption. Additionally, if the recalls are done randomly, then the diets of individuals being surveyed are not affected. However, there still remains a considerable amount of within person variability unless many 24 hour recalls are taken on the same individual. This is also still a form of self report meaning individuals can lie about what they are or forget, although the multiple pass method helps aleviate this issue [22]. Doubly labeled water (DLW) has been called the gold standard for estimating energy intake when individuals are weight stable [6], [13], [23]. Patients drink a small amount of liquid that contains an isotope of hydrogen and oxygen. Two weeks later a urine sample is taken and the concentrations are measured to get an estimate of energy expenditure that is estimated based on washout kinetics [23]. However, we are not only interested in weight stable individuals, but also those who are gaining and losing weight as well. Moreover, DLW is expensive and requires trained personnel to administer and analyze, therefore it is not reasonable to give DLW to all study participants [24], [23]. DLW has found to have an average error of between 2-8%, which depends on the loading dose, length of metabolic period and number of samples [25]. Direct calorimetery is based on production of heat by a human, and it provides accurate estimates of energy expenditure, but it must be administered in a special lab setting, and the patient must be in energy balance [26]. Indirect calorimetry is similar, except the patient has a mask that collects the air exhaled and gives an accurate measure of energy expenditure, but like direct calorimety, the patient must be in energy balance for this to give a measure of energy intake [26]. Neither of these options provide realistic measures if we are talking about free living humans because clearly, these instruments would affect one's everyday life. Several other new methods of measuring energy intake accurately and cheaply have been proposed: food photography [27], bite count devices [28], and devices that measure chewing [29], however, for obvious reasons such as influencing food intake, these methods still don't meet the need for an accurate, noninvasive, and relatively cheap measure of energy intake [30].

#### 1.1.2 Energy Balance Principle

Another approach to measuring EI that has gained popularity more recently is through what's called the energy balance principle. The energy balance equation is given in (1). This principle is based off of the first law of thermodynamics, that is, energy can neither be created or destroyed. Looking at (1), it is easy to see that we can rewrite EI as a simple function of energy expenditure (EE) and the change in energy storage ( $\Delta$ ES). In other words, instead of trying to measure energy intake directly via the methods of the previous section (which were either notoriously inaccurate, or unreasonably expensive), we can try to measure EE and  $\Delta$ ES and calculate EI. The energy balance model has been found to be an accurate and precise assessment of EI, at least during weight gain [6]. Many models have shown up in the epidemiology and nutrition journals using this approach [14], [23], [31], [30], [13], [32]. Although it may seem like we are complicating the problem by having to estimate two quantities instead of one, it turns out EE and  $\Delta$ ES are easier to measure accurately and cheaply.

$$\Delta ES = EI - EE \tag{1}$$

### 1.1.3 Measuring Energy Expenditure

From (1), if we want to estimate the EI for an individual, we need to have estimates for both EE and  $\Delta$ ES. We will focus on EE in this section, and  $\Delta$ ES in the next section. We refer to EE as the amount of energy used by an individual in a day, measured in kcals. There are a number of different ways our bodies burn calories, so we can break EE into four components: dietary induced thermogensis (DIT), volitional physical activity (PA), resting metabolic rate (RMR), and spontaneous physical activity (SPA) [23].

$$EE = DIT + PA + RMR + SPA$$
 (2)

DIT is related to the energy used when digesting, metabolizing, and transporting food [23]. If one eats more food, then his or her body must use more energy to process that food and vice versa. Thomas et al. assumes that DIT is proportional to EI and thus is estimated by a single value  $\beta$ . PA represents all fitness related activities such as walking, running, playing sports, etc [23]. We refer to these as voluntary activity, as one could technically live without burning calories this way. Thomas et al. measures PA as being proportional to body mass. RMR is the energy expended in order to keep the body functioning [23]. Thomas et al. uses the Livingston-Kohlstadt equation to calculate RMR. SPA is the energy used through changes in posture and fidgeting [23]. Thomas et al. notes that measuring SPA is very difficult, but use that the ratio of change in SPA to change in EE is constant. Thus, they use an integration along with the relationship in (2) to get an estimate.

The beauty of many of the EE measurement methods is that we don't need to measure each of those four components separately. We already discussed one of the available methods for measuring EE in the previous section. DLW is considered the gold standard measurement for EE [33],[34],[?],[13],[6],[30], [32], and it provides an estimate of EI if an individual is weight stable, otherwise by

(1) it doesn't tell the whole story. As discussed before DLW does give us the best estimate but at a very high cost. Alternatives to DLW to measure EE exist. Consumer based physical activity monitors with accelerometers have become popular in recent years. Products such as Fitbit, Nike Fuel Band, Jawbone UP band, and Basis B1 Band that measure steps taken, calories burned, heart rate, sleep, GPS, etc. are available to the general public at reasonable cost. They have become more popular in the research area as providing a measure of EE [35], and their reliability has been tested [36],[37],[38],[39],[40]. The results are promising, the physical activity monitors seem to give a relatively cheap, accurate, and objective measure of EE.

The SenseWear Pro3 armband (SWA) and SenseWear Mini (mini) specifically have been shown to be relatively good measures of energy expenditure [38]. The SWA has a biaxial accelerometer while the mini has a triaxial accelerometer. Some of what makes these standard apart from popular alternatives like Fitbit, is they also have sensors to measure heat flux, temperature, and galvanic skin response. One of the concerns with these wearables is assessing compliance. It is often hard to tell if someone has been following the researcher's directions of using the device. The SWA and mini don't have this problem as they provide estimates of wear time directly [38]. These two specific devices are not without flaws though, as both tend to underestimate EE from more vigorous levels of activity compared to DLW.

Self reports are also commonly used to measure EE largely due to their ease and cost. They have the benefit of providing details in terms of type of exercise as well as duration, intensity, etc. that an activity monitor would provide. Like with EI, we have the same issue with reporting bias and are not an accurate measure of EE [41]. Respondents typically overestimate their activity and its intensity. However, this cheap method can provide a rough activity level estimate and allow one to categorize participants by an approximate activity level if desired.

Pedometers are another cheap and easy method of estimating EE. Like self-report, they contain a lot of variability. Since they only count steps, pedometers do not tell us anything about intensity or duration of exercise. This can be an issue, for instance, 3000 steps walking in a day versus 3000 steps sprinting in a day will result in the same predicted EE, when in reality, the EE would be vastly

different. Like self-reports, pedometers can at least give an approximate population measure of activity. Heart rate monitors are another cheap and easy way to measure EE. Heart rate monitors provide accurate measures of heart rate, but factors other than exercise alone can affect heart rate. Temperature, emotional state, hydration status, fitness level, and age can all affect the heart rate of an individual. Direct observation is when trained personnel observe and record participant activities. This way, all activities can be monitored and objectively recorded. Of course, this method is extremely time consuming because someone must track a single participant. This can be difficult in large sample studies. These can be used to validate activity monitors, pedometers, or heart rate monitors.

# 1.1.4 Energy Storage

The other term we must estimate in order to use the energy balance principle is the change in energy stores over a specified period of time. We will deal with ES in terms of relative storage, that is, the change in energy stores since the last body composition measurement. For body composition, we will deal with fat mass (FM) and fat free mass (FFM) and assume total body mass = FM + FFM. The validated equation [42] below (3) shows how we can further break down equation (1).

$$\Delta ES = c_{FFM} \frac{\Delta FFM}{\Delta t} + C_{FM} \frac{\Delta FM}{\Delta t}$$
(3)

Where the estimates  $c_{FFM} = 1020$  and  $c_{FM} = 9500$  represent the energy densities of FFM and FM respectively. These numbers are estimates that are prone to measurement error, as in a different study, same author, [31],  $c_{FFM} = 1100$  and  $c_{FM} = 9300$  was used.

Setting aside the issue of measuring the energy densities of FM and FFM, we are left with the issue of measuring the change in FM and change in FFM over a specified period of time. This boils down to: how to we measure FM and FFM for an individual? Dual-energy X-ray absorptiometry (DXA) is considered the gold standard to measure fat mass (FM) and fat free mass (FFM) [23],[?],[30],[6]. DXA scans are often used to measure bone mineral density, often to diagnose or track osteoporosis, but it also gives measures of body composition. As one might imagine, DXA

scans are not cheap due to the specialized equipment and personnel needed. [43] shows the precision of the new GE Lunar iDXA machine. Coefficient of variation for both FM and FFM were under 1%.

Air displacement plethysmography (ADP) is another measure of body composition. ADP measures body volume through Boyle's Law, which describes the inverse relationship between volume and pressure [44]. After an intial up front cost, ADP machines can be used quite easily and cheaply as no highly trained personnel is needed. It is also very quick to get a measurement and can be used for a wide variety of the population including obese and elderly. Bod Pod is one of the well known measurement tools that performs this.

Bioimpedance spectroscopy (BIS) also provides a relatively accurate and cheap way to measure body composition. In short, BIS instruments have sensors at one's extremities (or a subset of them), and it sends a shock at one end and measures the resistance to reach the other end(s). The results are dependent upon body water levels, which can then be a proxy for FM versus FFM. There are various different levels of quality for these machines. One can buy a cheap and easy to use one at a local nutrition store that is not particularly accurate, and much more expensive, research quality devices are also available. One of the issues with this approach is hydration levels. Since this method provides a measure of body composition that is based on how a shock passes through the body as a function of body water, one's hydration status could heavily affect the results.

Calipers are one of the cheapest and easiest tools to measure body composition. The caliper method takes measurements of skinfolds at certain locations to estimate % body fat (and then can be used to calculate FM and FFM). The tool itself costs less than \$100, and someone can be quickly trained to use a caliper. Of course with this comes lots of error.

#### 1.1.5 Estimating Energy Intake via Energy Balance Equation

Based on the availability of alternative measures of self report for EE and  $\Delta$ ES, we prefer to calculate EI using the energy balance equation. To achieve the goal of measuring EI, we simply would add

the estimates of EE and  $\Delta$ ES. However, care must still be taken, since (1) holds in principle, that is, if we know EE and  $\Delta$ ES exactly for a specified period of time, but even with DXA and DLW measurements we don't have perfect data. Thomas took this approach in her 2010 paper [31], but noted that even with DXA and DLW measurements, the energy balance equation wasn't in equality, and she hypothesized this was largely due to measurement error and biological variation.

## 1.2 Modeling Energy Balance

We seek to develop a joint model for energy balance measurements that accounts for all forms of measurement error and biases. Ultimately, we would like to be able to accurately measure EI using cheaper and easier to administer devices compared to DXA and DLW. In the end, we want to calibrate these cheaper alternatives to the gold standards DXA and DLW to estimate EI. Those who have developed and tested models based on the energy balance principle have not taken measurement error into consideration, and as we've discussed, it shows up in a multitude of locations.

Here we will give a brief introduction into what we mean by dealing with the measurement error inherent in this problem. Our ultimate objective is to estimate true EI, and of course if were able to observe this, our problem would be done. Instead, we are using the energy balance principle, and thus if we were able to observe EE and  $\Delta$ ES, our problem would be done as well. However, instead of observing these two quantities exactly, we gather information on EE by, for instance DLW, and  $\Delta$ ES by, for instance, DXA. But as we've said, these are not perfect measurements and thus have noise involved that we aim to quantify. Moreover, using DXA and DLW as a final product to estimate EI is unreasonable. We would like to measure EI accurately using cheap and easy tools like armband monitors and calipers. Armband monitors give an estimate of EE for a fixed period of time, but they are prone to even more error than DLW, and we want to measure this error. In the end, we want to calibrate the cheaper tools using our results from the measurement error model. The main benefit is future studies requiring measurements of EI, EE or  $\Delta$ ES would be able to use our model and calibrate cheaper tools to get data that is closer to the truth without having to use gold standard measurements.

# 1.3 Measurement Error Modeling

Here we give a brief overview of measurement error modeling in statistics. We will go over basic considerations, simple models and classical methods. Then we will extend to more complicated models that are nonlinear and involve heterogeneous variance. We will examine Frequentist and Bayesian methods, and parametric, nonparametric, and semiparametric models. Complete overviews can be found in [45], [46], [47].

## 1.3.1 Dangers of Ignoring Measurement Error

For the purposes of this report, measurement error refers to not being able to measure a covariate, or explanatory variable **X** precisely. That is, instead of observing **X**, we actually observe another quantity **W**, that is not always (and rarely ever is) equal to **X** even though our interest still lies in the relationship between the response variable **Y** and the true explanatory variable with no error, **X**. We still allow the possibility of observing some covariates without error, and we denote these with **Z**. Recall, one of the assumptions made in linear regression using OLS is that the explanatory variables are measured without error.

Data collected in the epidemiology and kinesiology fields often have measurement error due to the nature of the studies. For example, if one was trying to understand the relationship between obesity and heart disease, one would be interested in how much that person eats on average. But determining someone's average daily caloric intake is incredibly difficult, and so we could use a Food Frequency Questionnaire (FFQ). This might seem like a reasonable way to determine how much someone eats on average, that is randomly picking a week or month and have the person of interest record what they consume. The time was chosen randomly, but does a week or month really describe that person's usual consumption patterns? The time of the year, weather, social desirability, etc. are all going to affect what the person actually eats and actually records eating. Therefore an estimate of EI based on the FFQ for usual daily consumption is not likely going to be the true value, or sometimes not even close due to bias (see pg. 8 [46]). Then this mismeasured explanatory variable will surely affect our estimated relationship between obesity and heart disease.

As a result, when measurement error is present in covariates, it can have a major impact on the statistical analysis performed if it is ignored. For one, it will bias estimates. This means the estimated effects will be biased in one direction or another and therefore not giving the whole story. Another problem is the loss of power in determining relationships between variables. Having measurement error in covariates essentially clouds the data with extra noise making it harder to see the true relationships and thus miss meaningful conclusions.

#### 1.3.2 Differential Error

We say the error in  $\mathbf{W}$  with respect to  $\mathbf{X}$  is non-differential if  $\mathbf{Y}$  is conditionally independent of  $\mathbf{W}$ . In other words,  $f_{Y|X,Z,W} = f_{Y|X,Z}$ . If this doesn't hold, then we say the error is differential If the error is non-differential,  $\mathbf{W}$  is called a *surrogate* or *proxy* for  $\mathbf{X}$ . Clearly, assuming non-differential error can be used as a simplifying assumption, and it is often the case that this is a valid assumption. Continuing on the obesity/heart disease example above, if we knew the true average daily EI of someone, we wouldn't have reason to have them fill out a FFQ, and it probably wouldn't give us any more information about the chances of heart disease by having him/her fill one out. This would be the rational for assuming non-differential error. The distinction is not always easy, and Buzas et al. [47] gave a nice way to think about it: Consider whether  $\mathbf{W}$  would have been measured had  $\mathbf{X}$  been available. If it wouldn't have, non-differential error is reasonable.

#### 1.3.3 Classical Measurement Error Model

Here we will go over the basic measurement error model. All these standard results can be found in [45] with more details there. The classical measurement error model is the simple case in which the true value of the covariate  $\mathbf{X}$  is measured with additive error. That is:

$$\mathbf{W} = \mathbf{X} + \mathbf{U} \tag{4}$$

where **U** is an error term and  $E(\mathbf{U}|\mathbf{X})=0$ , meaning **W** is an unbiased estimate of **X**. It is also

assumed that **U** is independent of **X**, therefore the value of the true covariate doesn't have any effect on the error in measurement. The variance of **U** need not be constant, but it is often assumed that  $\mathbf{U} \sim N(0, \sigma_u^2)$ .

There are various ways to check the assumptions that are used for the classical measurement error model. Common additional assumptions are that the distribution of **U** is symmetric and has constant variance given (**Z**,**X**). Carroll et al. [46] gives plot diagnostic checks for these assumptions for the classical model.

#### 1.3.4 Linear Regression with Classical Measurement Error

Recall from before the Classical Measurement Error model assumed additive measurement error (4) and that the observed value  $\mathbf{W}$  was an unbiased, surrogate for unobserved  $\mathbf{X}$ . Now, suppose we are interested in the relationship between response variable  $\mathbf{Y}$  and explanatory variable  $\mathbf{X}$  and we assume the form of a simple linear model:

$$\mathbf{Y} = \beta_0 + \beta_1 \mathbf{X} + \epsilon \tag{5}$$

where  $E(\epsilon|\mathbf{X}) = 0$  and each  $\epsilon$  is uncorrelated with all the others. The problem is we don't know  $\mathbf{X}$ , but we know its *surrogate*  $\mathbf{W}$  where from (4) we know  $\mathbf{W} = \mathbf{X} + \mathbf{U}$ . Because we don't know  $\mathbf{X}$ , assume it has mean  $\mu_x$  and variance  $\sigma_x^2$ . Again, we are assuming  $E(\mathbf{U}|\mathbf{X}) = 0$  and all  $\mathbf{U}$ 's are uncorrelated with all other  $\mathbf{U}$  and all  $\epsilon$ . Let  $var(\mathbf{U}) = \sigma_u^2$ . Plugging this into (5) we get:

$$\mathbf{Y} = \beta_0 + \beta_1(\mathbf{W} - \mathbf{U}) + \epsilon \tag{6}$$

$$= \beta_0 + \beta_1 \mathbf{W} + \epsilon - \beta_1 \mathbf{U} \tag{7}$$

$$= \beta_0 + \beta_1 \mathbf{W} + \epsilon^* \tag{8}$$

then we can see that the normal OLS estimator for  $\beta_1$  is biased by what is often called the attenuation

factor,  $\lambda$ .

$$\frac{cov(\mathbf{Y}, \mathbf{W})}{var(\mathbf{W})} = \frac{cov(\beta_0 + \beta_1 \mathbf{X} + \epsilon, \mathbf{X} + \mathbf{U})}{var(\mathbf{X} + \mathbf{U})} = \frac{cov(\beta_1 \mathbf{X}, \mathbf{X})}{\sigma_x^2 + \sigma_u^2}$$
(9)

$$= \beta_1 \frac{\sigma_x^2}{\sigma_x^2 + \sigma_u^2} = \beta_1 \lambda = \beta_1^* \tag{10}$$

Based on our model, the attenuation factor will always be less than one, meaning our OLS estimate for  $\beta_1$  using **W** directly in place of **X** will result in an estimate flattened towards 0. Also, the residual variance increases:

$$var(\mathbf{Y}|\mathbf{W}) = \sigma_e^2 + \frac{\beta_1^2 \sigma_u^2 \sigma_x^2}{\sigma_x^2 + \sigma_u^2}$$
(11)

As an example of the attenuation of the slope under this model, we simulated data under the following parameter values:  $\beta_0 = 0$ ,  $\beta_1 = 1$ ,  $\sigma_e^2 = 0.5$ ,  $\sigma_u^2 = 1$ , where  $\epsilon$  and  $\mathbf{U}$  are normally distributed and  $\mathbf{X} \sim \text{Unif}(0,5)$ . OLS was applied to the data with true  $\mathbf{X}$  and with noisy  $\mathbf{W}$ . Figure 1 shows the results.

From (10), one can see the usual OLS estimator for  $\beta_1$  using **W** instead of **X** produces a biased estimate for the true relationship between **Y** and **X**. Looking at (10), one can note if we knew  $\lambda$ , then we could get an unbiased estimator of  $\beta_1$  by dividing by  $\lambda$ . Of course we would not expect this to be the case very often. Instead, we could find a consistent estimator for  $\lambda$ , then divide our estimate of  $\beta_1$  by that estimate, and we will have an unbiased estimate for  $\beta_1$ . Using  $\sigma_x^2 = \sigma_w^2 - \sigma_u^2$ , we get  $\hat{\lambda} = \frac{\sigma_w^2 - \sigma_u^2}{\sigma_w^2}$ .  $\hat{\sigma}_w^2$  and  $\hat{\sigma}_u^2$  is the sample variance of the observed **W**'s and the estimated variance of the measurement error, respectively. As long as both of these are consistent (which we know  $\hat{\sigma}_w^2$  is), then  $\hat{\lambda}$  is consistent for  $\lambda$ . Carroll et al. gives an estimate for  $\sigma_u^2$  on page 71. It turns out in small sample sizes, this estimator for  $\lambda$  has a a very skewed distribution, and Fuller [45] gives a modified version of the above approach to give a more symmetric sampling distribution. The above method is a method of moments estimator because it only depends on moments based on the data and no distributional assumptions are made.

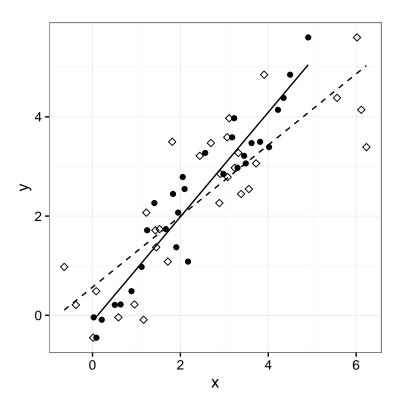


Figure 1: Simulated data with observed X (solid dots) and W (diamonds) along with fitted OLS lines. Solid is for the true X, and dashed is for the W.

#### 1.4 Energy Balance Measurement Error Models

To the best of our knowledge, no one has attempted to model the energy balance error structure jointly. While there have been measurement error models constructed for both physical activity and various aspects of nutrition, no one has used the joint relationships that exist between physical activity and food consumed (or equivalently physical activity and body composition changes) to assess the measurement error present in today's devices. We will construct our model based on univariate methods developed within the field as well as models constructed with no intent of application in exercise science or nutrition.

## 1.5 Bayesian Measurement Error Models

Richardson et al. [48] presents a general approach to Bayesian modeling of measurement error problems with particular focus on epidemiology applications. This work was done when MCMC methods were becoming popular, and thus this provided a new way to approach these types of problems compared to frequentist methods. The authors propose hierarchical modeling that utilizes

conditional independence. The state that the classical measurement error problem can be broken into the three components below:

Disease Model = 
$$Y_i | X_i, Z_i, \theta_y$$
 (12)

Measurement Model = 
$$W_i | X_i, \theta_w$$
 (13)

Exposure Model = 
$$X_i | Z_i, \theta_x$$
 (14)

where  $Y_i$  is the response, often representing the presence of a disease.  $W_i$  is a surrogate for  $X_i$ , ie a covariate measured with error.  $Z_i$  represent other covariates that are assumed to be error free. The interest in the problem is the influence of the covariates measured with error  $(W_i)$  and without error  $(Z_i)$  on the response which is some sort of health outcome, whether it be binary or not. Conditional on the truth covariates  $X_i$ ,  $Z_i$ , the response is conditionally independent of the surrogate  $W_i$ . The joint distribution can then be easily written as the product over i of the above model components. Richardson et al. give more details and more complex dependence structures, but this is their general framework. They presented a linear and generalized linear (binomial) measurement error model with normal distributions for errors and exposure, but this general framework can easily be expanded upon.

The paper by Berry et al. [49] introduced a fully Bayesian approach to the problem of nonparametric regression when there is measurement error in the covariates. This extends the traditional linear measurement error by allowing a non-linear response of unknown form. They explored the following situation:

$$Y_i = m(X_i) + \epsilon_i \tag{15}$$

$$W_{ij} = X_i + U_{ij} \tag{16}$$

Berry et al. used penalized splines to estimate the function m(), and opt to use a relatively large number of knots (20-40). Assuming normal distributions for the error terms as well as the latent variables  $X_i$ , they present an MCMC algorithm to sample from the posterior. In a simulation study, they showed their model had better frequentist properties than some competeing frequentist methods. They also did some rbobustness checks to their normal assumptions, and although there were some losses of efficiency, the authors note "the results are at least encouraging in the case of small model deviations". The analysis was also not sensitive to priors.

Sinha et al. [50] presents a semiparametric Bayesian analysis of nutritional data. In this paper the authors are interested in a binary response variable as unknown function of a variable measured with error plus a linear function of error-free covariates. The authors assume normal errors for the measurement error terms, but place a Dirichlet Process on the latent variables  $X_i$  in an attempt to lighten distributional assumptions. The authors used B-splines instead of P-splines and also opted to use a large number of fixed knots.

Sarkar et al. [51] presents a semiparametric model that attempts to make as few distributional assumptions as possible. Their setup is nearly identical to Berry et al. They proposed using a Dirichlet Process for the distribution of the latent variables, measurement error, and regression error terms to avoid the standard normality assumption. They also allow for non-constant variance. Sarkar et al. proposes using splines to model the unknown regression function, but they chose to use B-splines largely because they are more stable. The number and location of knots are chosen beforehand. Simulation studies show their model outperforms the model of Berry et al. under various situations. The authors note however, that the model of Berry et al. is competitive when using B-splines instead of the originally proposed P-splines. The authors attributed this to the numerical stability of the B-splines.

# 1.6 Summary

# References

- [1] Adrian Cook, Jane Pryer, and Prakash Shetty. The problem of accuracy in dietary surveys. analysis of the over 65 uk national diet and nutrition survey. *Journal of Epidemiology and Community Health*, 54:611–16, 2000.
- [2] Rachel K. Johnson. Dietart intake-how do we measure what people are really eating? *Obesity*, 10:63S–68S, 2002.

- [3] Rupert W. Jakes, E. Day, Nicholas, Robert Luben, Ailsa Welch, Sheila Bingham, Jo Mitchell, Susie Hennings, Kirsten Rennie, and Nicholas J. Wareham. Adjusting for energy intake-what measure to use in nutritional epidemiological studies? *International Journal of Epidemiology*, 33:1382–6, 2004.
- [4] CK Martin, JB Correa, H Han, HR Allen, JC Rood, CM Champagne, BK Gunturk, and GA Bray. Validity of the remote food photography method (rfpm) for estimating energy and nutrient intake in near real-time. *Obesity*, 20:891–9, 2012.
- [5] J. T. Winkler. The fundamental flaw in obesity research. Obesity Reviews, 6:199–202, 2005.
- [6] L Anne Gilmore, Eric Ravussin, George Bray, Hongmei Han, and Leanne Redman. An objective estimate of energy intake during weight gain using the intake-balance method. *American Journal of Clinical Nutrition*, 100:806–12, 2014.
- [7] NIDDK. Niddk website. http://www.niddk.nih.gov/health-information/health-statistics/Pages/overweight-obesity-statistics.aspx. Accessed: 2016-02-16.
- [8] Peter W.F. Wilson, Ralph B. D'Agostino, Lisa Sullivan, Helen Parise, and William B. Kannel. Overweight and obesity as determinants of cardiovascular risk. Archives of Internal Medicine, 162:1867–72, 2002.
- [9] CDC. Cdc website. http://www.cdc.gov/healthyweight/effects/. Accessed: 2016-02-05.
- [10] Stephanie Kasen and et al. Obesity and psychopathology in women: a three decade prospective study. *International Journal of Obesity*, 2008.
- [11] Floriana S Luppino and et al. Overweight, obesity, and depression: a systematic review and meta-analysis of longitudinal studies. *Archives of general psychiatry*, 2010.
- [12] Roland Sturm. The effects of obesity, smoking, and drinking on medical problems and costs.

  Health Affairs, 21:245–253, 2002.
- [13] Kevin D Hall and Carson C Chow. Estimating changes in free-living energy intake and its confidence interval. *American Journal of Clinical Nutrition*, 94:66–74, 2011.

- [14] Kevin D. Hall. Predicting metabolic adaptation, body weight change, and energy intrake in humans. Am J Physical Endocrinol Metab, 298:449–66, 2010.
- [15] Diana M. Thomas, Corby K. Martin, Leanne M. Redman, Steven B. Heymsfield, Steven Lettieri, James A. Levine, Claude Bouchard, and Dale A. Schoeller. Effect of dietary adherence on the body weight plateau: a mathematical model incorporating intermittent compliance with energy intake prescription. American Journal of Clinical Nutrition, 100:787–95, 2014.
- [16] Ignatius Brady and Kevin D. Hall. Dispatch from the field: Is mathematical modeling applicable to obesity treatment in the real world? *Obesity*, 22:1939–41, 2014.
- [17] Diana Thomas. Modeling in clinical nutrition: does it add to patient care? European Journal of Clinical Nutrition, 67:555–7, 2013.
- [18] Diana M. Thomas, Christina M. Gonzalez, Andrea Z. Pereira, Leanne M. Redman, and Steven B. Reymsfield. Time to correctly predict the amount of weight loss with dieting. *Journal* of the Academy of Nutrition and Dietetics, 114:857–61, 2014.
- [19] DA Schoeller, D Thomas, E Archer, SB Heymsfield, SN Blair, MI Goran, JO Hill, RL Atkinson, BE Corkey, J Foreyt, and et al. Self-report-based estimates of energy intake offer an inadequate basis for scientific conclusions. American Journal of Clinical Nutrition, 97:1413–5, 2013.
- [20] LS Freedman, JM Commins, JE Moler, L Arab, DJ Baer, V Kipnis, D Midthune, AJ Moshfegh, ML Neuhouser, Rl Prentice, and et al. Pooled results from 5 validation studies of dietary selfreport instruments using recovery biomarkers for energy and protein intake. American Journal of Epidemiology, 180:172–88, 2014.
- [21] Ernst J. Schaefer, Joi L. Augustin, Mary M. Schaefer, Helen Rasmussen, Jose M. Ordovas, Gerard E. Dallal, and Johanna T. Dwyer. Lack of efficacy of a food-frequency questionnaire in assessing dietary macronutrient intakes in subjects consuming diets of known composition. American Journal of Clinical Nutrition, 71:746–51, 2000.
- [22] Nancy Raper, Betty Perloff, Linda Ingwersen, Lois Steinfeldt, and Jaswinder Anand. An overview of usda's dietary intake data system. *Journal of Food Consumption and Analysis*, 17:545–55, 2004.

- [23] Diana M. Thomas, Corby K. Martin, Steven Heymsfield, Leanne M. Redman, Dale A. Schoeller, and James A. Levine. A simple model predicting individual weight change in humans. *Journal* of Biological Dynamics, 5:579–599, 2011.
- [24] Kevin Hall. Estimating human energy intake using mathematical models. *American Journal of Clinical Nutrition*, 100:744–5, 2014.
- [25] Dale A. Schoeller. Measurement of energy expenditure in free-living humans by using doubly labeled water. *Journal of Nutrition*, 118:1278–89, 1988.
- [26] Ronald Laporte, Henry J. Montoye, and Carl J. Caspersen. Assessment of physical activity in epidemiologic research: Problems and prospects. *Public Health Reports*, 100:131–46, 1985.
- [27] BL. Daugherty, TE Schap, R Ettienne-Gittens, FM Zhou, M Bosch, EJ Delp, DS Ebert, DA Kerr, and CJ Boushey. Novel technologies for assessing dietary intake: evaluating the usability of a mobile telephone food record among adults and adoloescents. J Med Internet Res, 14, 2012.
- [28] JM Fontana, JA Higgins, SC Schuckers, F Bellisle, Z Pan, EL Melanson, MR Neuman, and E Sazonov. Energy intake estimation from counts of chews and swallows. Appetite, 85:14–21, 2015.
- [29] JL Scisco, ER Muth, and AW Hoover. Examining the utility of a bite-count-based measure of eating activity in free-living human beings. *J Acad Nutr Diet*, 114:464–9, 2013.
- [30] Arjun Sanghvi, Leanne M. Redman, Corby K. Martin, Eric Ravussin, and Kevin D. Hall. Validation of an inexpensive and accurate mathematical method to measure long-term changes in free-living energy intake. American Journal of Clinical Nutrition, 102:353–8, 2015.
- [31] Diana M. Thomas, Dale A. Schoeller, Leanne A. Redman, Corby M. Martin, James A. Levine, and Steven B. Heymsfield. A computational model to determine energy intake during weight loss. American Journal of Clinical Nutrition, 92:1326–31, 2010.
- [32] Susan B. Racette, Sai Krupa Das, Manjushri Bhapkar, Evan C. Hadley, Susan B. Robers, Eric Ravussin, Carl Pieper, James P. DeLany, William E. Kraus, James Rochon, and Leanne M. Redman. Approaches for quantifying energy intake and %calorie restriction during calorie

- restriction interventions in humans: the multicenter calerie study. Am J Physiol Endorinol Metab, 302:441–8, 2011.
- [33] Y.T. Lagerros and P. Lagiou. Assessment of physical activity and energy expenditure in epidemiological research of chronic diseases. *European Journal of Epidemiology*, 205:353–62, 2007.
- [34] C. Bouten, W. Verboeket-van de Venne, K.R. Westerterp, M. Verduin, and J.D. Janssen. Daily physical activity assessment: comparison between movement registration and doubly labeled water. *Journal of Applied Physiology*, 81:1019–26, 1996.
- [35] G.J. Welk, J.A. Schaben, and J.R. Jr Morrow. Reliability of accelerometry-based activity monitors: a generalizability study. *Med Sci Sports Exerc*, 36:1637–45, 2004.
- [36] D.R et al. Bassett. Validity of four motion sensors in measuring moderate intensity physical activity. Med Sci Sport Exer, 32:S471–S480, 2000.
- [37] C. Drenowatz and J.C. Eisenmann. Validation of the sensewear armband at high intensity exercise. Eur J Appl Physiol, 111:883–7, 2011.
- [38] Johannsen D.L. et al. Accuracy of armband monitors for measuring daily energy expenditure in healthy adults. *Medicine and Science in Sports and Exercise*, 42:2134–40, 2010.
- [39] K. L. et al. Dannecker. A comparison of energy expenditure estimation of several physical activity monitors. *Med Sci Sports Exerc*, 45:2105–12, 2013.
- [40] Lee Jung-Min, Youngwon Kim, and Gregory J. Welk. Validity of consumer-based physical activity monitors. *Medicine and Science in Sports and Exercise*, 46:1840–8, 2014.
- [41] Heather K. Neilson, Paula J. Robson, Christine M. Friedenreich, and Ilona Csizmadi. Estimating activity energy expenditure: how valid are physical activity questionnaires? American Journal of Clinical Nutrition, 87:279–91, 2008.
- [42] D. M. Thomas, C. Bouchard, T. Church, C. Slentz, W.E. Kraus, Redmanm L.M., C.K. Martin, A.M. Silva, M. Vossen, K. Westererp, and S.B. Heymsfield. Why do individuals not lose more weight from an exercise intervention at a defined dose? an energy balance analysis. *Obesity Reviews*, 2012.

- [43] K. Hind, B. Oldroyd, and J. G. Truscott. In vivo precision of the ge lunar idxa densitometer for the measurement of total body composition and fat distribution in adults. *European Journal* of Clinical Nutrition, 65:140–2, 2011.
- [44] Vickie et al. Baracos. Advances in the science and application of body composition measurement. *Journal of Parenteral and Enteral Nutrition*, 36:96–107, 2011.
- [45] Wayne A. Fuller. Measurement Error Models. New York: John Wiley and Sons, 1987.
- [46] Raymond J. Caroll, David Ruppert, Leonard A. Stefanski, and Ciprian M. Crainiceani. *Measurement Error in Nonlinear Models*. Chapman and Hall/CRC, 2006.
- [47] J.S. Buzas, T.D Tosteson, and L.A. Stefanski. Measurement error. Unpublished Report, Institue of Statistics Mimeo Series No. 2544, 2003.
- [48] Sylvia Richardson and Walter R. Gilks. A bayesian approach to measurement error problems in epidemiology using conditional independence models. American Journal of Epidemiology, 138:430–42, 1993.
- [49] Scott M. Berry, Raymond J. Carroll, and David Ruppert. Bayesian smoothing and regression splines for measurement error problems. *Journal of the American Statistical Association*, 97:160–69, 2002.
- [50] Samiran Sinha, Bani K. Mallick, Victor Kipnis, and Raymond J. Carroll. Semiparametric bayesian analysis of nutritional epidemiology data in the presence of measurement error. *Bio*metrics, 66:444–54, 2010.
- [51] Abhra Sarkar, Bani K. Mallick, and Raymond J. Carroll. Bayesian semiparametric regression in the presence of conditionally heteroscedastic measurement and regression errors. *Biometrics*, 70:823–34, 2014.