

# HOW TO USE ZOLA

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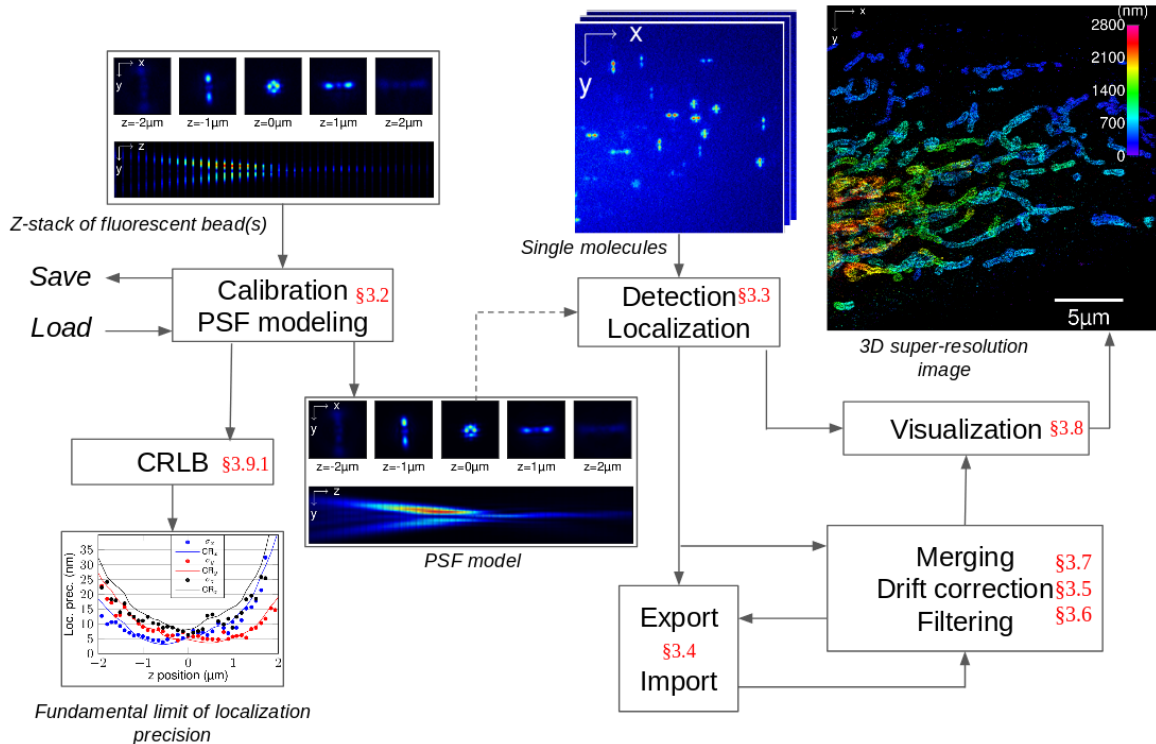
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## Contents

<b>1</b>	<b>Introduction</b>	<b>1</b>
<b>2</b>	<b>Requirement</b>	<b>2</b>
<b>3</b>	<b>ZOLA plugin</b>	<b>2</b>
3.1	ZOLA plugin installation . . . . .	2
3.2	Nvidia GPU . . . . .	2
3.3	Source code . . . . .	2
3.4	Camera photon count . . . . .	3
3.5	Calibration - PSF modeling . . . . .	3
3.6	Single molecule Localization . . . . .	4
3.7	Import/Export . . . . .	6
3.8	Drift correction . . . . .	6
3.9	Filtering . . . . .	7
3.10	Merging . . . . .	7
3.11	Visualization . . . . .	8
3.12	Additional tools . . . . .	9
3.12.1	Theoretical localization precision limit: CRLB (Cramér Rao Lower Bound) . . . . .	9

## 1 Introduction

Thank you for downloading ZOLA. ZOLA is an ImageJ/Fiji plugin that allows you to determine the PSF model corresponding to your optical system, and use this model to reconstruct 3D single molecule localization microscopy (SMLM) images. ZOLA also provides tools to correct drifts, merge consecutive localizations or render images. The following scheme presents an overview of methods featured ZOLA:



## 2 Requirement

Our plugin requires the 64 bit version of ImageJ or Fiji software available here: <https://imagej.net/Fiji/Downloads>. The calibration, localization or drift correction tools are highly computationally expensive. To increase efficiency, the computations can be performed on GPU (Graphical Process Unit). Currently, these functions can be launched on Cuda compatible NVIDIA GPU.

Please, go to <http://www.nvidia.fr/page/home.html> and <http://docs.nvidia.com/cuda/index.html> to install your NVIDIA GPU and Cuda-toolkit.

## 3 ZOLA plugin

Plugin installation requires 10 minutes. If you plan to benefit from GPU acceleration, plugin installation requires 20 minutes more. Follow these instructions to install the plugin:

### 3.1 ZOLA plugin installation

Moving ZOLA-0.1.X-SNAPSHOT.jar to your *ImageJ/plugin* folder is enough to install the plugin. However, you can not benefit from the possibility to update new releases of the plugin. Using Fiji, you can follow these instructions to install the plugin with the last version:

1. Launch Fiji.
2. Select **Help** → **Update**.
3. Select **Manage update sites**.
4. Select **IMOD-ZOLA** from the list.
5. Clic **Close**.
6. Clic **Apply changes**.
7. Restart Fiji.
8. ZOLA should now appear in the **Fiji** → **Plugins** dropdown menu

### 3.2 Nvidia GPU

Computation using GPU requires the use of some libraries. Follow these instructions to add them:

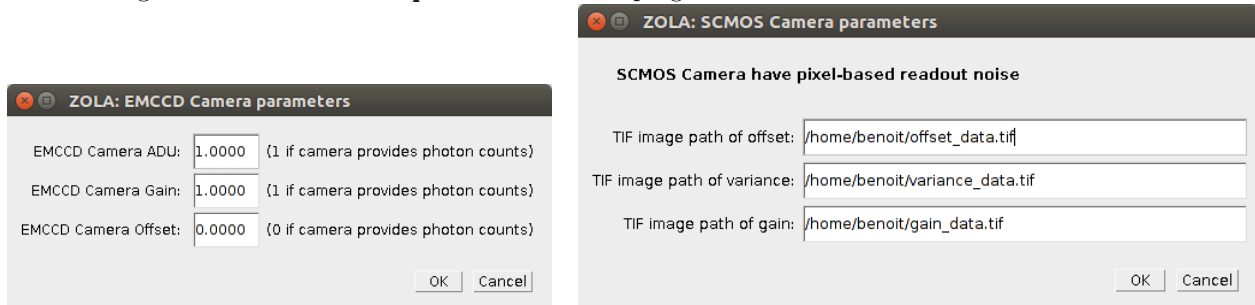
1. Open a terminal.
2. Check your version by executing `nvcc -version` in the terminal. We provide libraries for cuda 7.5, 8 and 9.
3. Go to <https://github.com/imodpasteur/ZOLA-3D> website and download **JCuda\_libraries.zip**.
4. Extract **JCuda\_libraries.zip** file.
5. Open *JCuda\_libraries* folder and open the subfolder corresponding to your operating system and your Cuda version. If your Cuda version older than 7.5, the libraries can be found here:  
<http://www.jcuda.org/downloads/downloads.html>.
6. Paste the *lib* folder into your *ImageJ* folder.
7. Copy and paste the content of the *jars* folder into your *ImageJ/jars* folder (or *ImageJ/plugin/jars* folder depending on your version of ImageJ).

### 3.3 Source code

The source code of ZOLA is also available in *zola\_source\_code* folder. Please, check the *zola\_source\_code/README.md* file to compile it.

### 3.4 Camera photon count

Zola's algorithm requires camera pixel values to be in photon counts. New EMCCD camera provide photon counts. If this is not the case, Zola can manage both EMCCD and SC MOS camera specific noise. please, set camera parameters using ZOLA → Camera setup → EMCCD/SC MOS plugin.

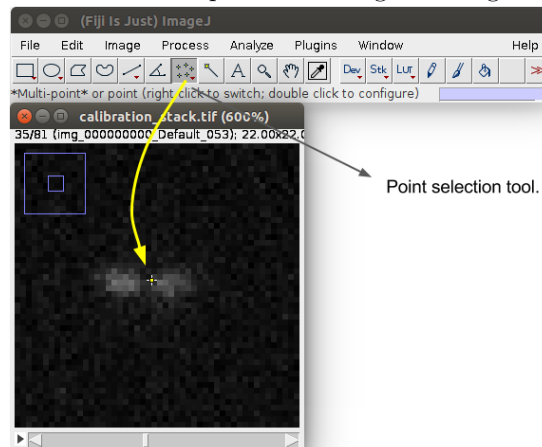


If you use EMCCD camera, pixel photon conversion needs three parameters: ADU and Offset that are given in your Camera specifications, and the Gain choosen in the image acquisition software. If you use SC MOS camera, pixel photon conversion is a bit more complicated since each pixel has its own statistic. We propose the photon conversion proposed by Huang et al. *Nat. Methods, (10) 2013*. It uses three maps that have to be calibrated (Offset, Variance and Gain) allowing to convert each pixel in photon count in ZOLA. The three maps correspond to TIF files whose width and height have to be the same as images used for calibration and localization.

### 3.5 Calibration - PSF modeling

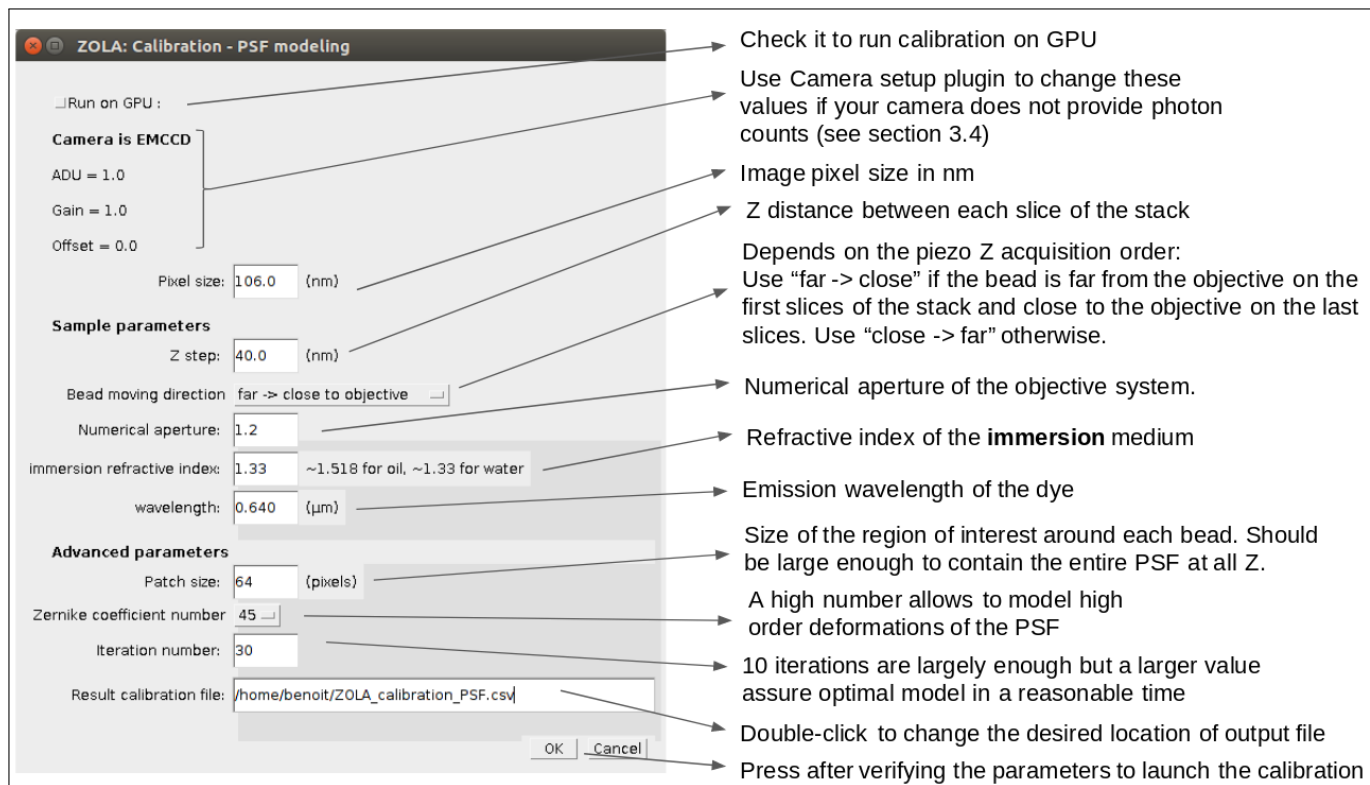
ZOLA's calibration tool allows to model the PSF from a stack of images of fluorescent beads at different Z positions. The algorithm assumes that there is no drift during the acquisition. Thus, it is recommended to acquire the stack of images fast with stable temperature in the room. Usually, the axial distance between slices is set to 50 nm and the acquisition time per slice is 50 ms. Thus, the acquisition time for a 2 μm Z range is 2 seconds.

First, open a stack of images and select one or multiple beads using the ImageJ Point selection tool:

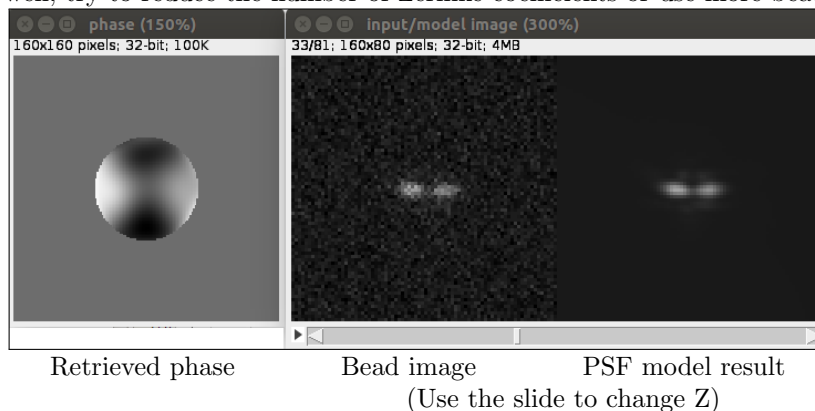


A single bead is usually enough to accurately model the PSF. If your image contains overlapping beads or has a low signal to noise ratio, it is still possible to model the PSF by selecting multiple beads.

Next, open the ZOLA → Calibration PSF modeling plugin. The following window pops up:



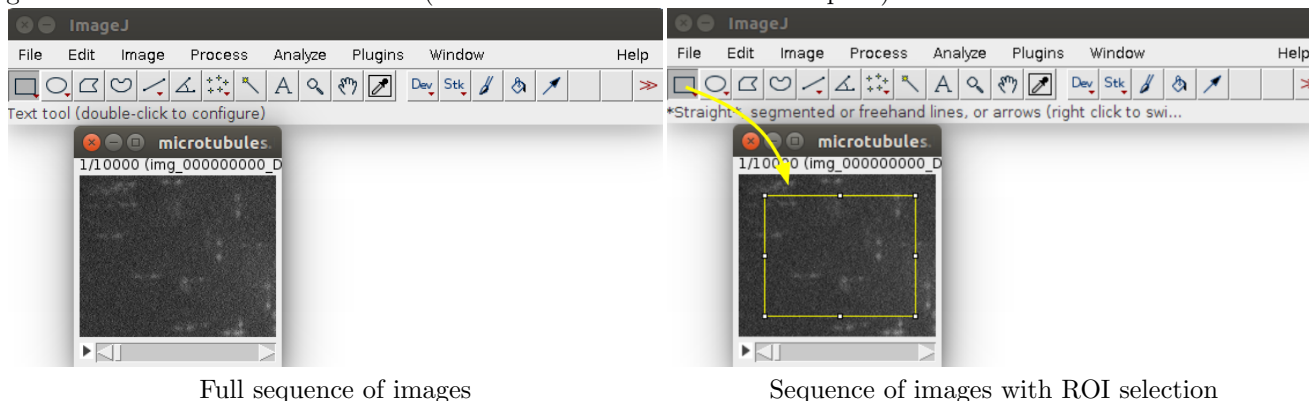
After verifying the parameters, then press OK. Phase retrieval begins. This can take few minutes. At the end of the phase retrieval, the plugin shows the phase and the model of the PSF as shown below. If the modeled PSF does not match the bead image well, try to reduce the number of Zernike coefficients or use more beads.



### 3.6 Single molecule Localization

ZOLA's localization plugin allows to detect and precisely localize single molecules in a sequence of diffraction limited images as obtained in SMLM experiments. The process is the following:

First, open the sequence of images. If desired, a region of interest (ROI) can be drawn to reduce the size of the region to reconstruct as shown below (this increases the reconstruction speed).



Then, open the ZOLA → Localization plugin. The following window pops up:

**ZOLA: Localization**

☐ Run on GPU :

**Camera is EMCCD**

ADU = 1.0

Gain = 1.0

Offset = 0.0

PSF calibration file: /home/benoit/ZOLA\_calibration\_PSF.csv

**Sample parameters**

Mounting medium refractive index: 1.330 1.33 for water

Distance focus to coverslip: 0 (μm)

**Advanced parameter**

Patch size: 32 (pixels)

Expected axial range: 3.5 (μm)

Min number of photons: 500

Localization table: /home/benoit/ZOLA\_localization\_table.csv

OK Cancel

Check it if you want to run localization on GPU  
Same as on § 3.2

Input calibration file path (this is the output of the calibration tool from § 3.2). Double-click to pop up a selection window.

Refractive index of the **mounting** medium (different from § 3.2)

Distance between the focal plane and the coverslip. It can be measured before single molecule image acquisition by computing the distance between the focal plane and the focal position of the coverslip (using beads stucked on it). If refractive index between immersion (§ 3.2) and mounting mediums are different, please, set this value properly to deal with this mismatch.

Size of the windows surrounding molecules. This value should be as small as possible but large enough to contain the entire PSF for all Z positions.

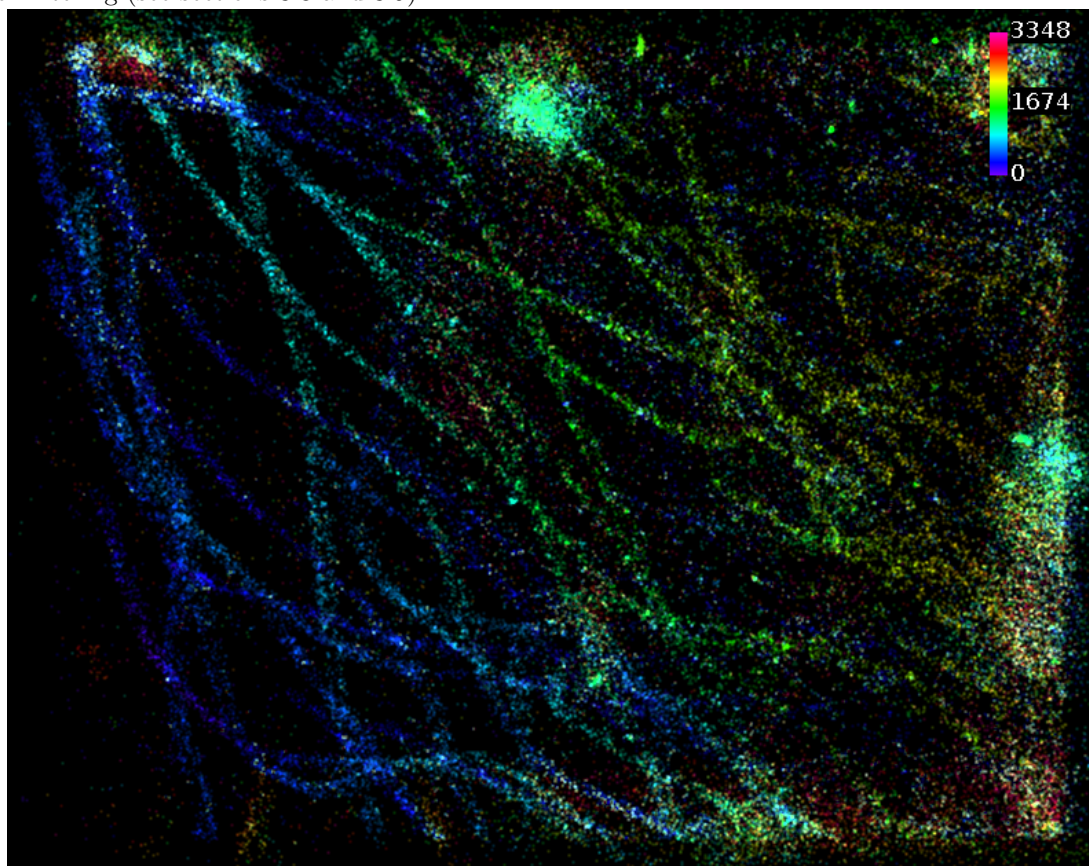
The axial range that can be reconstructed. This depends on the PSF (Astigmatism: ~2 μm, Saddle point: ~4 μm, tetrapod: >4 μm)

Only molecules with a higher number of photon than this value are kept

Output localization table. Double-click to pop up a selection window.

Press after verifying the parameters to launch the localization

The localization starts after clicking on OK. On test\_data we provide, localization requires around 10 minutes. During localization, a colored super-resolution image (whose color encodes Z position) is updated every 30 seconds. The following figure shows the result obtained at the end of the localization. Other visualizations can be obtained using the ZOLA visualization tool (see section 3.11). Note that the quality of this image can be improved applying drift correction or filtering (see sections 3.8 and 3.9).





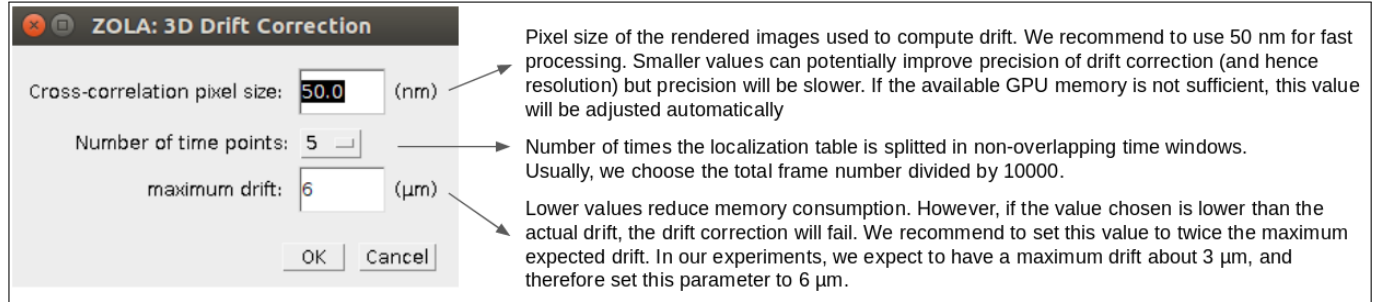
### 3.7 Import/Export

ZOLA allows to load a localization table in memory, or save it on disk.

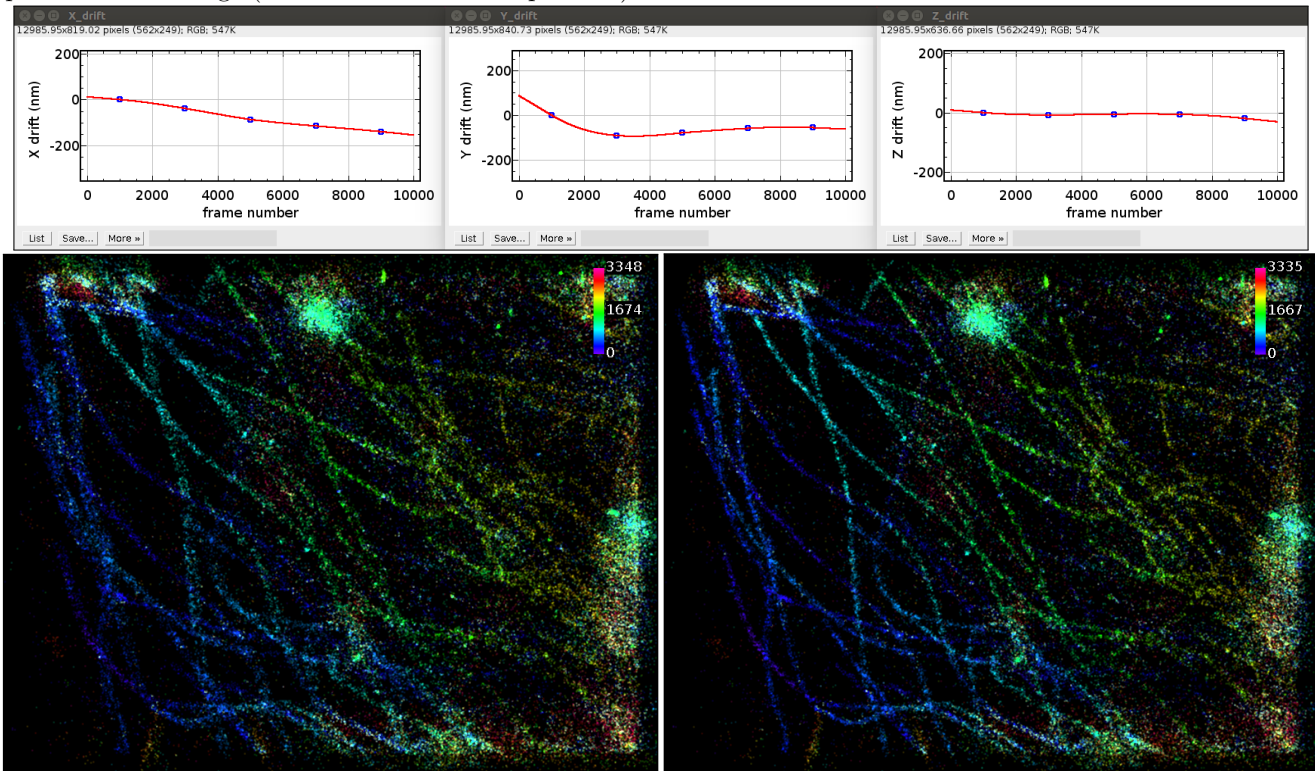
- Import: load a localization table.
- Append: Append another localization table to the existing table (useful when consecutive image sequences were processed separately).
- Export: Save a localization table (useful to save data after post-processing, *e.g.* after drift correction, filtering or merging (see sections 3.8, 3.9 and 3.10)).

### 3.8 Drift correction

ZOLA features a 3D drift correction method based on redundant cross correlation. The drift correction window is as follows:



As output, the drift correction plugin shows three curves corresponding to X, Y and Z drifts. A drift-corrected super-resolution image (whose color encodes Z position) is also shown:



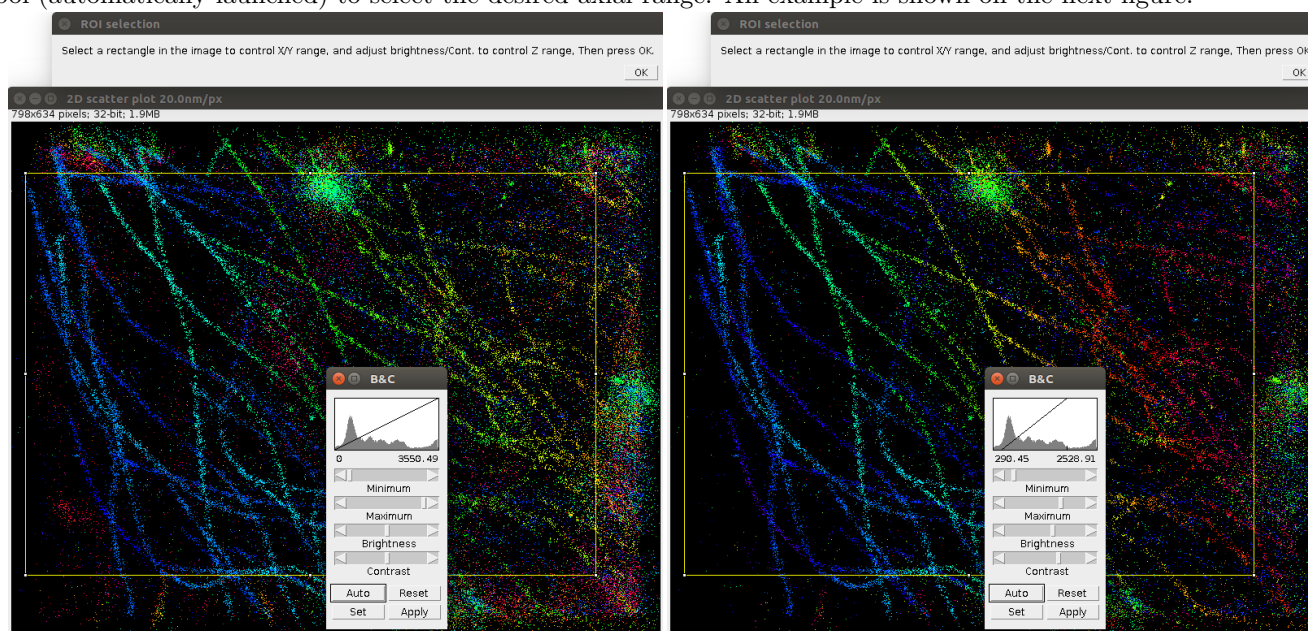
Super-resolution image before drift correction

Drift-corrected super-resolution image

After applying drift correction, use ZOLA→Export plugin to save the localization table (see section 3.7).

### 3.9 Filtering

Filtering is useful to define a sub region of a 3D image, or to remove localizations with low precision or unsatisfying goodness-of-fit. First, the user selects a region of interest in the image (optional), and can use **Brightness&Contrast** tool (automatically launched) to select the desired axial range. An example is shown on the next figure:

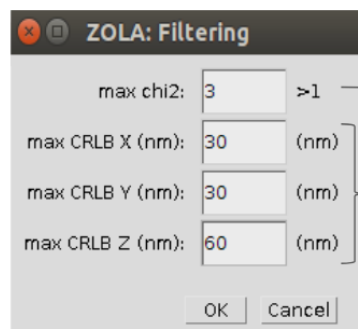


Full axial range: 3.5  $\mu\text{m}$

Clipped axial range: 2.3  $\mu\text{m}$

Then, after pressing OK, the following panel is opened allowing to filter localizations according to their good goodness-of-fit and their precision.

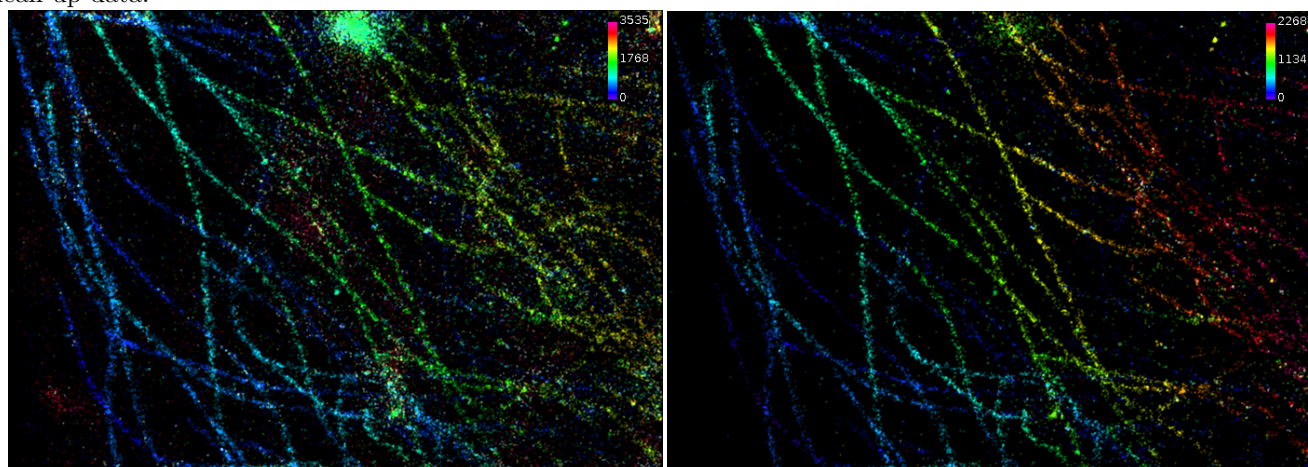
2



Maximum  $\chi$  bounds. This goodness-of-fit parameter is used to remove localizations that correspond to overlapping molecules. Usually, we set min. and max. bounds to 0.8 and 3 resp.

Maximum precision limit (given by Cramér Rao Lower Bound) according to X, Y and Z. This depends on the PSF (Astigmatism: ~20 to 40, saddle point: ~30 to 60, tetrapode: >40)

After pressing OK, the super-resolution resulting image pops up as shown below. As we can see, filtering allows to clean up data.



Super-resolution image before filtering

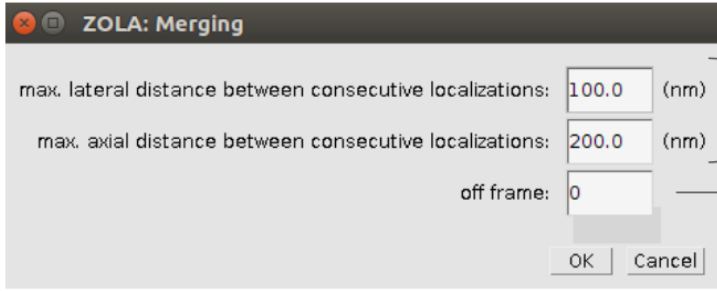
Super-resolution image after filtering

After applying merging, use ZOLA→Export plugin to save the localization table (see section 3.7).

### 3.10 Merging

This plugin is useful to merge localizations that are in close proximity in consecutive frames and are likely to originate from the same molecule. Open ZOLA→Merging and the following window pops up:





Maximum distance between localizations to be merged

Maximum number of frames a molecule is allowed to disappear before reappearing. Usually, we set it to 0, meaning that only molecules that appear in consecutive frames will be merged.

Then, click OK to launch the merging. As output, the plugin gives the number of localization that are merged and show the resulting super-resolution image.

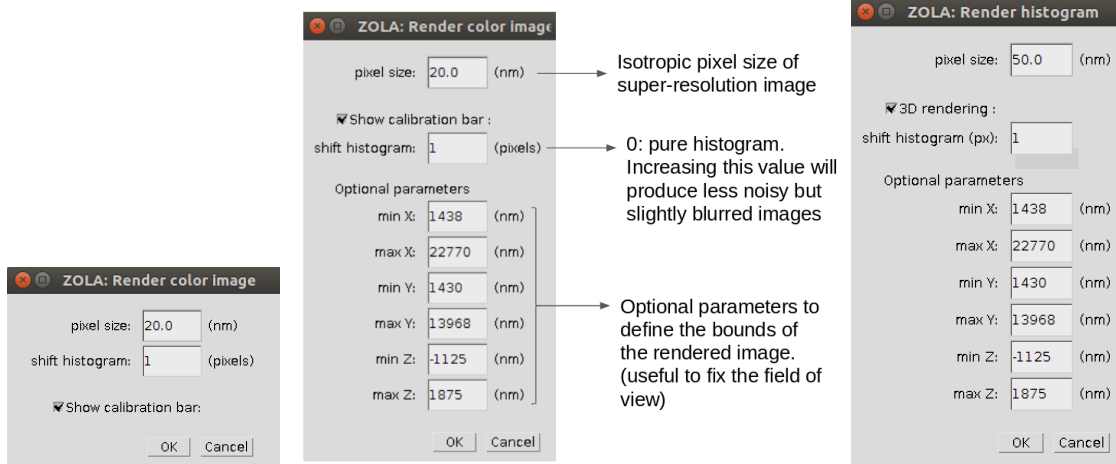
After applying merging, use ZOLA→Export plugin to save the localization table (see section 3.7).

### 3.11 Visualization

ZOLA visualization plugin allows to render super-resolution images. The visualization can be launched during or after localization or after importing a localization table. Three rendering methods are proposed based on histograms:

- Automatic 2D color histogram: Renders a 2D color histogram adjusting automatically the axial range and localizations that have a good precision (This method is launched automatically during localization and after import, drift correction, filtering and merging functions).
- 2D color histogram: Renders a 2D color histogram.
- 2D/3D histogram: Render a 2D or 3D histogram.

The three visualization windows are as follow:

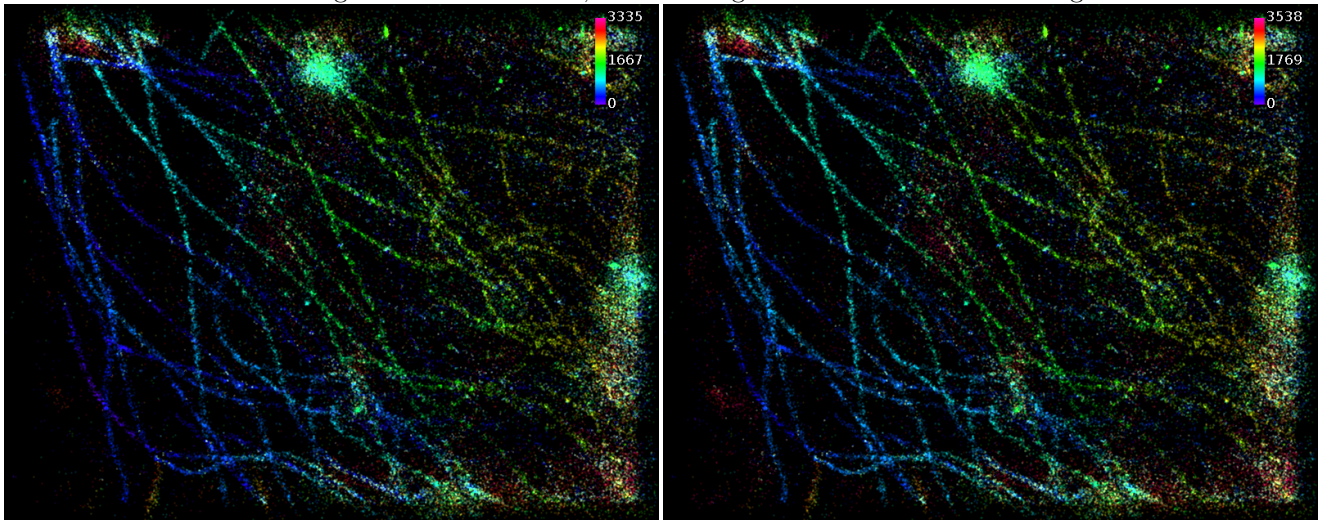


automatic color histogram

Color histogram

2D/3D Histogram

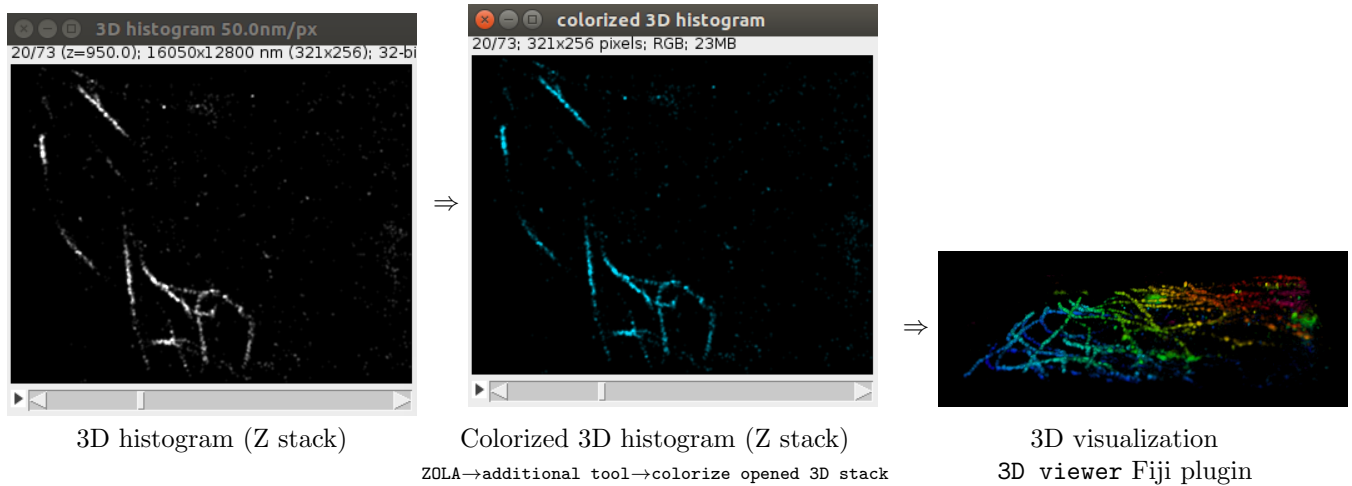
According to the method used, the resulting visualization is the following:



automatic color histogram (clipped axial range)

Color histogram (full axial range)

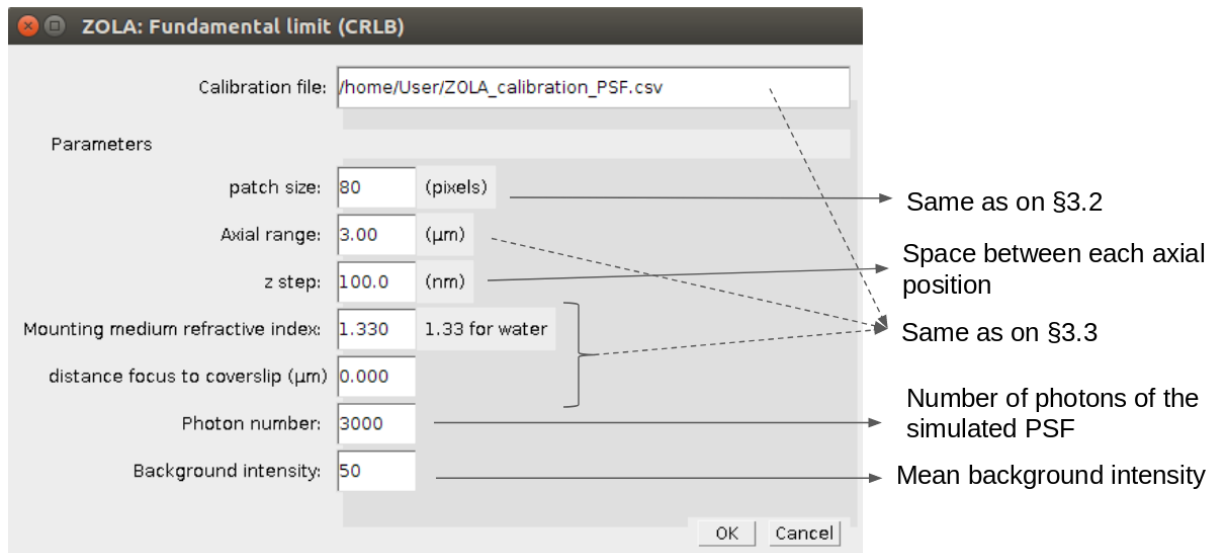




## 3.12 Additional tools

### 3.12.1 Theoretical localization precision limit: CRLB (Cramér Rao Lower Bound)

Zola's CRLB plugin allows to compute the fundamental localization precision limit for a given PSF and an expected molecule number of photon and background. The CRLB of X, Y and Z is calculated as a function of axial position. Open the ZOLA → CRLB plugin. The following window pops up:



As a result, the tool shows the PSF and 3 plots showing the precision limit (the standard deviation of localization errors) for X, Y and Z as a function of axial position Z:

