

Tables

There are several options for making tables in R Markdown.

- just print a dataframe or matrix
 - easy - most popular format
 - better: `knitr::kable` - not OODS friendly
- `gt`
 - tidyverse compatible syntax, highly flexible - only text for now
- `stabilize::stabilize`
 - highly customizable - only for paths (a little biased)
- `janitor::cleaner`
 - default tables for different choices
- `stargazer::stargazer`

Figures

Figures produced by R code will be placed after the code chunk from which they were generated.

- Most common options are `fig.asp` and `fig.asp.cmt.viz.opts`.
- `fig.asp.cmt`

```
---
title: VZC01 neutralization report
author: David Beckhouse
date: "x format (fig.data(), 'yy-mm-dd') "
output: html_document
---
[{"x, load-data, echo = FALSE}
# replace with your own
shs_file_path <-
~/Documents/teaching/DSTR/intro_repro_workfile/lectures/04
neutralization/"
full_file_path <- paste0(
shs_file_path, "hiv_project/data/vzco1_data.csv"
)
data <- read.csv(full_file_path, header = TRUE)

# Descriptive Analysis
## Viral analysis
Characteristics of the 'x row(data)' viruses from the CATMAP
database are displayed in the table below.
[{"x, table}]
# labelled - grammar package for making pretty html tables
library(typosummary)
# label variables
var_label(data) <- list(
id = "ID",
sh_resistance = "Antibody resistance",
shield_glycans = "Shield glycans",
region = "Region",
env_length = "Length of Env protein"
)
data$number_glycans <- ifelse(data$shield_glycans < 4, "< 4", ">= 4")
# make the table
data |>
select("region", "env_length", "sh_resistance",
"number_glycans") |>
tbl_summary(by = number_glycans) |>
modify_spanning_header(c("stat_1", "stat_2") ~ "***P<0.05 in
glycan shield***") |>
add_overall() |>
add_p()

## Graphical analysis
Below we report a scatter plot of resistance as a function of
PMS in the glycan shield.
[{"x, plot=detail}
# fig.asp = "Scatter plot with linear model smoother",
# fig.align = "center",
# plot.width = "600px"
library(ggplot2)
ggplot(data, aes(x = shield_glycans, y = sh_resistance)) +
geom_point() +
geom_smooth(method = lm) +
theme_bw()

# Regression Analysis
## Primary model
[{"x, fit=primary-model}
mod <- glm(
sh_resistance ~ shield_glycans + region + env_length,
data = data
)
tbl_regression(mod) |>
add_global_p()

## Secondary model
In this analysis, we analyzed a binary endpoint where level 1 means
antibody resistance score was greater than 1.
[{"x, binary-model}
binary_mod <- glm(
1(sh_resistance > 1) ~ shield_glycans + region + env_length,
data = data,
family = binomial()
)
tbl_regression(binary_mod, exponentiate = TRUE) |>
add_global_p()

From <https://raw.githubusercontent.com/beckhouse/teach_repro_workfile/lectures/04
neutralization_analysis_notebook_to_read>
```

Code chunk options

Change options for all chunks using code chunk at the top of the .Rmd.

```
---
title: VZC01 neutralization report
author: David Beckhouse
date: "x format (fig.data(), 'yy-mm-dd') "
output: html_document
---
[{"x, chunk=opts, include = FALSE, message = FALSE, warning = FALSE,
echo = TRUE, eval = TRUE, message = TRUE, warning = TRUE}
Note that you still need to suppress the code chunk that includes the call to
optr, chunk(opts).
```

```
---[{"x, chunk=opts, include = FALSE, message =
FALSE, warning = FALSE}
library(hill)
optr_chunk(opts)
echo = FALSE, warning = FALSE, message = FALSE
}

```

From <https://raw.githubusercontent.com/beckhouse/teach_repro_workfile/lectures/04
neutralization_analysis_notebook_to_read>

Code chunk options

Supporter we would like to use the same first code for two purposes

- setting display code messages for debugging purposes
- production - ignore code messages for sharing purposes

- Option values can be any valid R expression
- Below a `knitr::eval` variable
- `knitr::code_chunk`
- `FALSE` display of code and messages

```
---[{"x, production=eval, include = FALSE}
# we will learn a smarter way to do this
production <- TRUE
eval = production()
# if production == FALSE, then this code will not
# be evaluated and thus code chunks will appear in
# the rendered document
library(hill)
optr_chunk(opts)
echo = FALSE, warning = FALSE, message = FALSE
}

```

From <https://raw.githubusercontent.com/beckhouse/teach_repro_workfile/lectures/04
neutralization_analysis_notebook_to_read>

Rendering from the command line

Option 1

• From the directory that contains `hiv_report.Rmd`, run the following:

```
knitr::render("hiv_report.Rmd")
```

• This will render the code in the command line.

Option 2

• From the directory that contains `hiv_report.Rmd`, run the following:

```
knitr::render("hiv_report.Rmd", output = "hiv_report.html")
```

See the `knitr` documentation for options.

Parameterized reports

The code chunk defining the product in `write_hiv` is not that

• `write_hiv` is a function that takes a parameter to change the output.

In `write_hiv`, define `param` option to not default.

• `param` option is available in a session to check parameter settings

when document is rendered

```
---
title: VZC01 neutralization report
author: David Beckhouse
date: "x format (fig.data(), 'yy-mm-dd') "
output: html_document
param:
production: FALSE
output:
output:
output:
---
[{"x, chunk=opts, include = FALSE, message = FALSE, warning = FALSE,
eval = param$production}
# if production == FALSE, then this code will not
# be evaluated and thus code chunks will appear in
# the rendered document
library(hill)
optr_chunk(opts)
echo = FALSE, warning = FALSE, message = FALSE
}
[{"x, load-data, echo = FALSE}
shs_file_path <-
~/Documents/teaching/DSTR/intro_repro_workfile/lectures/04
neutralization/"
full_file_path <- paste0(
shs_file_path, "hiv_project/data/vzco1_data.csv"
)
data <- read.csv(full_file_path, header = TRUE)

```

```
# Descriptive Analysis
## Tabular analysis
Characteristics of the 'x row(data)' viruses from the CATMAP
database are displayed in the table below.
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# label variables
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sh_resistance = "Antibody resistance",
shield_glycans = "Shield glycans",
env_length = "Length of Env protein"
)
data$number_glycans <- ifelse(data$shield_glycans < 4, "< 4", ">= 4")
# make the table
data |>
select("region", "env_length", "sh_resistance", "number_glycans") |>
tbl_summary(by = number_glycans) |>
modify_spanning_header(c("stat_1", "stat_2") ~ "***P<0.05 in glycan
shield***") |>
add_overall() |>
add_p()

```

```
# Graphical analysis
Below we report a scatter plot of resistance as a function of PMS in
the glycan shield.
[{"x, plot=detail}
# fig.asp = "Scatter plot with linear model smoother",
# fig.align = "center",
# plot.width = "600px"
library(ggplot2)
ggplot(data, aes(x = shield_glycans, y = sh_resistance)) +
geom_point() +
geom_smooth(method = lm) +
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```

```
# Regression Analysis
## Primary model
[{"x, fit=primary-model}
mod <- glm(
sh_resistance ~ shield_glycans + region + env_length,
data = data
)
tbl_regression(mod) |>
add_global_p()
## Secondary model
In this analysis, we analyzed a binary endpoint where level 1 means
antibody resistance score was greater than 'x param$endpoint'.
[{"x, binary-model}
binary_mod <- glm(
1(sh_resistance > param$endpoint) ~ shield_glycans + region +
env_length,
data = data,
family = binomial()
)
tbl_regression(binary_mod, exponentiate = TRUE) |>
add_global_p()

```

```
From <https://raw.githubusercontent.com/beckhouse/teach_repro_workfile/lectures/04
neutralization_analysis_notebook_to_read>
```

