WR.causal (cWR) R Documentation

Causal inference on Win Ratio for composite endpoints of semi-competing risk time-to-event data

This function uses Win Ratio (WR) as a summary statistic to compare the composite endpoints of semi-competing risk time-to-event data between two groups, in observational study setting.

Analysis can be done for the following scenarios: (1) Causal inference of independent subjects with confounders (2) Causal inference of cluster-dependent subjects with confounders (observational study with one-level cluster structure). This cluster structure assumes that all clusters include both comparison groups For instance, patients are nested within hospitals, and each hospital has patients in treatment group and patients in control group. Clusters with only one memberships will be automatically excluded in analysis.

WR.causal(treatment, cluster, v1, v2, delta1, delta2, x.con, x.char, null.WR=1, alpha.sig=0.05, control=NA, n.boot=200)

Arguments

treatment Integer vector with code 0 as control group and 1 as treatment group for each subject cluster Integer vector with unique cluster ID for each cluster. When subjects are independent, the cluster ID is unique for each subject. Numeric vector with min(T_H, T_D, T_C) for each subject, where T_H, T_D and T_C are time to non-fatal event, time to fatal event and censoring time, respectively y1 Numeric vector with min(T_D, T_C) for each subject Integer vector with code 1 indicating that T_H is observed, 0 oth Integer vector with code 1 indicating that T_D is observed, 0 otherwise delta2 Continuous covariate matrix with observations as rows and variables as columns. All columns have to be numeric type x.con x.char Categorical covariate matrix with observations as rows and variables as columns. All columns have to be character type null. WR Null hypothesis of the WR statistic. The default is H0: WR=1 or log(WR)=0. alpha.sig Significance level, with default value 0.05

control
List of control options for the estimation of the lagrange multipliers in the estimation of calibration weights. The control options are the same as the ones in R package "nleqsly". The default value is control=NA, and the algorithm uses all default settings in "nleqsly". When specify the options, option names are needed. For instance, control=list(xtol=1e-05, flot=1e-06).

Number of bootstrap replications for variance estimation. It's only used for analysis of cluster-dependent data. Default value is 200.

Details

The function "WR.causal" conducts causal inference and significance testing comparing two composite time-to-event outcomes between groups, accounting for confounders. The Win Ratio summary statistic is built on the "unmatched" approach described by Pocock et al. (2011). We assume that the composite endpoints can be formulated as semi-competing risk data. Each individual in the study is measured on time to non-fatal (non-terminal) event (e.g. hospitalization) and time to fatal (terminal) event (e.g. death). Specifically, the fatal event is considered clinically more important compared to the non-fatal event. Censoring is allowed, but time to censor needs to be observed.

This function can handle independent data, as well as clustered data. The inference of clustered data is based on the calibration weighted stratified estimator studied by Zhang and Jeong (2019). The construction of the causal inference for Win Ratio is based on the causal U-statistics by Mao (2017), and the calibration weighted stratified estimator accounts for potential correlations among subjects within a cluster and confounder effects. The estimation of the calibration weights requires the estimation of lagrange multipliers through numeric iterations. We adopted the R package "nleqsly" for the estimation.

Note: The option "treatment", "cluster", "y1", "y2", "delta1", "delta2" are required and no defaults are provided. These options have to be numeric vectors with the same length. No missing values are all

Warning: The convergence becomes difficult when there are many multi-level categorical variables. We recommend to dichotomize multi-level variables to increase the probability of convergence

Test name Number of patients and number of clusters in analysis info clusters Clusters that are included in analysis (excluded those with only one membership within clusters) U1 First estimated clustered U-statistic U2 Second estimated clustered U-statistic Estimated WR on log scale logWR Estimated standard error of the WR on log scale Test statistic 100(1-alpha.sig)% confidence interva p-value P-value of the significance testing

convergence Info of convergence for the estimation of lagrange multipliers. Values are the same as the ones in R package "nleqsiv"

Author(s)

Di Zhang <diz11@pitt.edu>; Jong H. Jeong <jjeong@pitt.edu>

Maintainer: Di Zhang <diz11@pitt.edu>

References

Pocock, S. J., Ariti, C. A., Collier, T. J., andWang, D. (2011). The win ratio: a new approach to the analysis of composite endpoints in clinical trials based on clinical priorities. European heart journal 33, 176-182.

Mao, L. (2017). On causal estimation using-statistics. Biometrika, 105(1), 215-220.

Yang, S. (2018). Propensity score weighting for causal inference with clustered data. Journal of Causal Inference, 6(2),

Zhang, D. and Jeong, H. J. Causal Inference on Win Ratio for Clustered Semi-Competing Risk Data. (In draft, 2019)

Examples

```
# load library
library
library(gumbel)
library(copula)
```

download and install package through Github install_github("dee1008/cWR")

Or you may download the cNR_1.0.tar.gz file from https://github.com/deel008/cNR, and install the package in R using command "install.packages("Your_Path/cNR 1.0.tar.gz",repos=NULL)".

library(cWR)

```
#-------1. Data generation for independent semi-competing risk data with confounders----
# joint survival: bivariate exponential with Gumbel-Hougaard copula
# define functions
gumbel_causal_individual<-function(n.sub,dim,alpha,lambdaH,lambdaD,etaH,etaD,cov.H, cov.D)
  exprand <- matrix(rexp(dim * n.sub), c(n.sub, dim))
unifpirand <- runif(n.sub, 0, pi)
exprand2 <- rexp(n.sub)
beta <- l/alpha
stablerand <- sin((1 - beta) * unifpirand)^((1 - beta)/beta) *
(sin(beta * unifpirand))/(sin(unifpirand))^(l/beta)
stablerand <- stablerand/(exprand2^alpha - 1))
unifrand <- invphigumbel(exprand3*tablerand, alpha) # generating bivariate uniform random variables for marginal survival funtions--(*)
     natrix(c(-log(unifrand[,1])/(lambdaH*exp(-etaH+cov.H)),-log(unifrand[,2])/(lambdaD*exp(-etaD+cov.D))),c(n.sub,dim)) # inverting specific forms of survival functions in (*) to create true bivariate event times adjusted for event types and trt groups
qen_causal_individual<-function(N,dim,alpha,lambdaH,lambdaD,lambdaC,etaH,outcome.H.eta,etaD,outcome.D.eta,etaC,ps.model.phi){
   # generate covariates z1 and z2
    z1 < rnorm(N)

z2 < -sample(c(1,0), N, prob=c(0.5, 0.5), replace=TRUE)
```

```
# generate treatment status
# generate Leatment status
# design matrix X
X<-cbind(rep(l,N), z1, z2)
Ka-cbind(rep(l,N), z1, z2)
# calculate probability of getting treatment for each obs
pc-exp(X8*8ps.model.phi) /(1+exp(X8*8ps.model.phi))
ptrtc-sapply(p, function(x) sample(c(l,0), 1, prob-c(x, (1-mt-sup)))
# number of treatment obs
mc-sum(trt)
# number of control obs</pre>
                                                                                                                          )
=c(x, (1-x))))
       -length(trt)-sum(trt)
 # percentage of treatment and control obs
percent<-c("control%"=n/N, "trt%"=m/N)</pre>
# combine data trt, z1, z2
temp<-cbind(trt, z1, z2)
# separate treatment temp data
temp.trtx-temp[(temp[,1]==1),]
temp.con<-temp[(temp[,1]==0),]</pre>
     generate time to events: time to non-fatal and time to fatal events
roupO<-gumbel_causal_individual(n,dim,alpha,lambdaH,lambdaH,0,0,temp.con[,2:3]%*%outcome.H.eta, temp.con[,2:3]%*%outcome.D.eta)
roupI<-gumbel_causal_individual(m,dim,alpha,lambdaH,lambdaD,etaH,etaD, temp.trt[,2:3]%*%outcome.H.eta, temp.trt[,2:3]%*%outcome.D.eta)
           mbine time to events and time to censoring
 true.t<-rbind(group0,group1)
t=mo.data<-cbind(true.t,c(rexp(n,lambdaC),rexp(m,lambdaC*exp(-etaC))))
t.obs<-apply(temp.data,1,min)
```

```
delD-rep(0,dim(true.t)[1])
delH[temp.data[,1]==t.obs]<-1
delD[temp.data[,2]==t.obs]<-1
      my.data<-cbind(temp.data,t.obs,delH,delD,rbind(temp.con, temp.trt))
yl<-rep(0,n+m)
y2<-rep(0,n+m)</pre>
       \label{eq:continuity} $$\inf_{x\in\mathbb{R}^2}(my.data.f$c < my.data.f$t) $$ (my.data.f$c < my.data.f$t2) $$my.data.f$y1[indi.1]<-my.data.f$c[indi.1] $$ my.data.f$v2[indi.1]<-my.data.f$v2[indi.1] <-my.data.f$v2[indi.1] $$
      \label{eq:continuity} $$\inf_3<-(my.data.f$t1 < my.data.f$c) $$ (my.data.f$c < my.data.f$t2) $$my.data.f$y1[indi.3]<-my.data.f$t1[indi.3] $$my.data.f$y2[indi.3]<-my.data.f$c[indi.3] $$
      \label{eq:continuous} $$\inf_{x \in \mathbb{R}^2 \times \mathbb{R}^2} (my.data.f$t2 < my.data.f$c) $$my.data.f$y1[indi.4]<-my.data.f$y1[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.f$y2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.fy2[indi.4]<-my.data.f
      my.data.f$delD[indi.4]<-1
      names (my.data.f) <-c("time_Non_Fatal", "time_Fatal", "time_censor", "t.obs", "deltal", "delta2", "treatment", "z1", "z2", "y1", "y2")
      output<-list(my.data.f, n, m, percent)
names(output)<-c("data", "#control", "#trt", "assignment
return(output)</pre>
 # generate independent data data|
data|<-gen_causal_individual(N=1000,dim=2, alpha=2, lambdaH=0.1, lambdaD=0.08, lambdaC=0.09, etaH=0, outcome.H.eta=c(0.1, 0.3), etaD=0, outcome.D.eta=c(0.2, 0.4), etaC=0.1, ps.model.phi=c(-0.2, 0.5, 0.5))$data
 # generate continuous covariate matrix
x.con<-as.matrix(data1[, c("z1","z2")])</pre>
 # independent win ratio ind.wr<-with(datal, WR.causal(treatment-treatment, cluster-cluster, y1-y1, y2-y2, deltal-delta1, delta2-delta2, x.con-x.con, n.boot-2))
# logWR
ind.wr$logWR
# se of logWR
ind.wr$se
# 95% CI of logWR
ind.wr$ci
 #-----2. Data generation for cluster-dependent semi-competing risk data with confounders-
 gumbel_causal_PScluster<-function(n.sub,dim,alpha,lambdaH,lambdaD,etaH,etaD,cov.H, cov.D, frail)</pre>
     exprand <- matrix(rexp(dim * n.aub), c(n.sub, dim))
unifpirand <- runif(n.sub, 0, pi)
exprand2 <- rexp(n.aub)
beta <- l/alpha
stablerand <- sin((1 - beta) * unifpirand)^((1 - beta)/beta) *
(sin(beta * unifpirand))/(sin(unifpirand))^(l/beta)
stablerand <- stablerand(- stablerand(- exprand2/ealpha - 1))
unifrand <- invphigumbel(exprand/stablerand, alpha) * generating bivariate uniform random variables for marginal survival funtions--(*)
       \text{matrix}(c(\cdot|\log(\text{unifrand}[,1])/(\text{frail*lambdaH*exp}(\cdot-\text{etaH+cov.H})),-\log(\text{unifrand}[,2])/(\text{frail*lambdaD*exp}(\cdot-\text{etaD+cov.D}))),c(\text{n.sub,dim})) \ \text{# inverting specific forms of survival functions in (*) to create $\text{# true bivariate event times adjusted for event types and trt groups} 
gen_causal_PScluster<-function(N, n.cluster, shape, rate, dim, alpha, lambdaH, lambdaD, lambdaC, etaH, outcome.H.eta, etaD, outcome.D.eta, etaC, ps.model.phi){
       # generate covariates z1 and z2, z3=z1*z2
        # generate Covariates 21 and 22, 23=21-22
z1<-rnorm(N)
#z2<-sample(c(1,0), N, prob=c(0.5, 0.5), replace=TRUE)
z2<-rnorm(N, mean=1, sd=2)</pre>
       # generate clusters
cluster<-rep(1:n.cluster, each=N/n.cluster)</pre>
       )
reffect <- rMvdc(n.cluster, myMvd)
rl<-rep(reffect[,1], each=M/n.cluster) # cluster effect for treatment
r2<-rep(reffect[,2], each=M/n.cluster) # cluster effect for outcome
        # generate treatment status
# design matric "
       # generate treatment status
# design marrix X
X<-cbind(rep(1,N), z1, z2)
K<-cbind(rep(1,N), z1, z2)
# calculate probability of getting treatment for each obs
pc-exp(%**ps.model.phi*r1) (/1+exp(%**ps.model.phi*r1))
trt<-sapply(p, function(x) sample(c(1,0), 1, prob~c(x, (1-x))))
# number of treatment obs
mc-sum(trt)
# number of control obs
x<-color="block">**Color="block" block block" block
x<-color="block" block block
x<-color="block" block
x<-color="
        # number of control obs
n<-length(trt)-sum(trt)
# percentage of treatment and control obs
percent<-c("control%"-n/N, "trt%"-m/N)</pre>
      # combine data trt, z1, z2, z3, cluster ID, cluster effect
temp<-cbind(trt, z1, z2, cluster, r1, r2)
# separate treatment temp data
temp.trt<-temp[(temp[,1]=-1),]
temp.con<-temp[(temp[,1]=-0),]</pre>
      # combine time to events and time to censoring
true.t<-rbind(group0,group1)
true.t<-rbind(true.t<,c(rexp(n,lambdaC),rexp(m,lambdaC*exp(~etaC))))</pre>
       t.obs<-apply(temp.data,1,min)
delH<-rep(0,dim(true.t)[1])
delD<-rep(0,dim(true.t)[1])
delH[temp.data[,1]==t.obs]<-1
delD[temp.data[,2]==t.obs]<-1
        my.data<-cbind(temp.data,t.obs,delH,delD,rbind(temp.con, temp.trt))
yl<-rep(0,n+m)
y2<-rep(0,n+m)</pre>
       \label{eq:continuous} $$\inf_{x \in \mathbb{R}^2 \times \mathbb{R}^2}(my.data.f$c < my.data.f$t2)$$ $$my.data.f$y1[indi.1]<-my.data.f$c[indi.1]$$ $$my.data.f$y2[indi.1]<-my.data.f$c[indi.1]$$
       indi.2<-(my.data.f$t2 < my.data.f$t1) & (my.data.f$t1 < my.data.f$c)
```

delH<-rep(0,dim(true.t)[1])

```
india_Comp_data_ffit2 cm_data_ffit2 cm_data_ffit2 cm_data_ffit2]
my_data_ffit[dut_2] india_[2] mid_2] mid_2[1 mid_2] mid_2[2]
my_data_ffit[dut_2] india_[2] mid_2[2] mid_2[2]
```

End(Not run)