

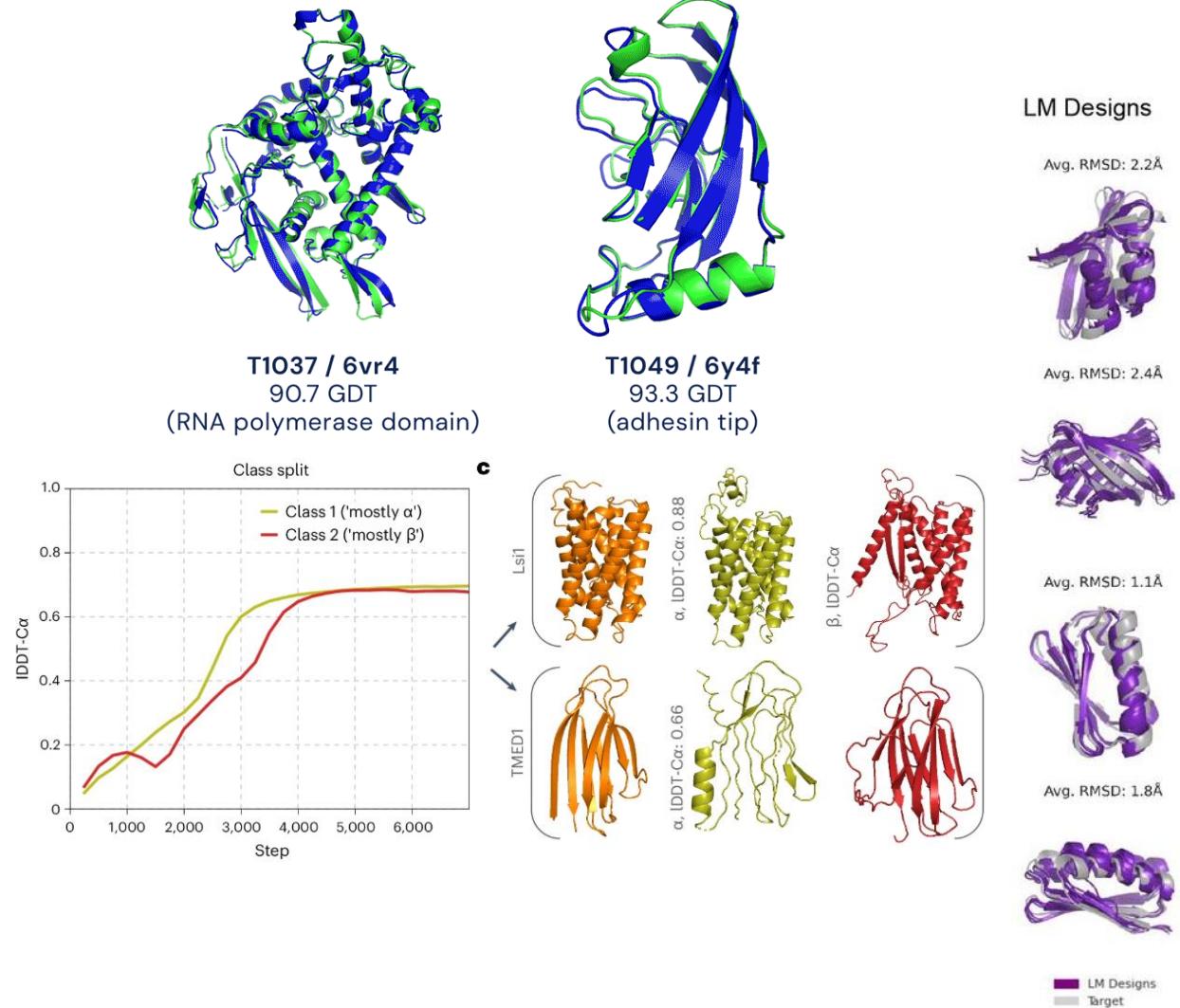
# *De novo prediction of protein structural dynamics*

Diego del Alamo, PhD

Principal Scientist, Protein Design & Informatics, GSK

# Protein structure prediction is solved, but protein folding is not

- State-of-the-art AI methods can reliably predict static structures of monomeric proteins to high accuracy ([Simpkin 2023](#))
  - Includes *de novo* designed proteins ([Verkuil 2022](#))
  - Unseen topologies ([Ahdritz 2024, Frank 2024](#))
- These do not extend to modeling their dynamics: how/when proteins move
- Some questions about dynamics can still be answered by vanilla protein structure prediction neural networks
- Newer architectures can answer others



Ahdritz et al "OpenFold: retraining AlphaFold2 yields new insights into its learning mechanisms and capacity for generalization" *Nat Methods* 2024

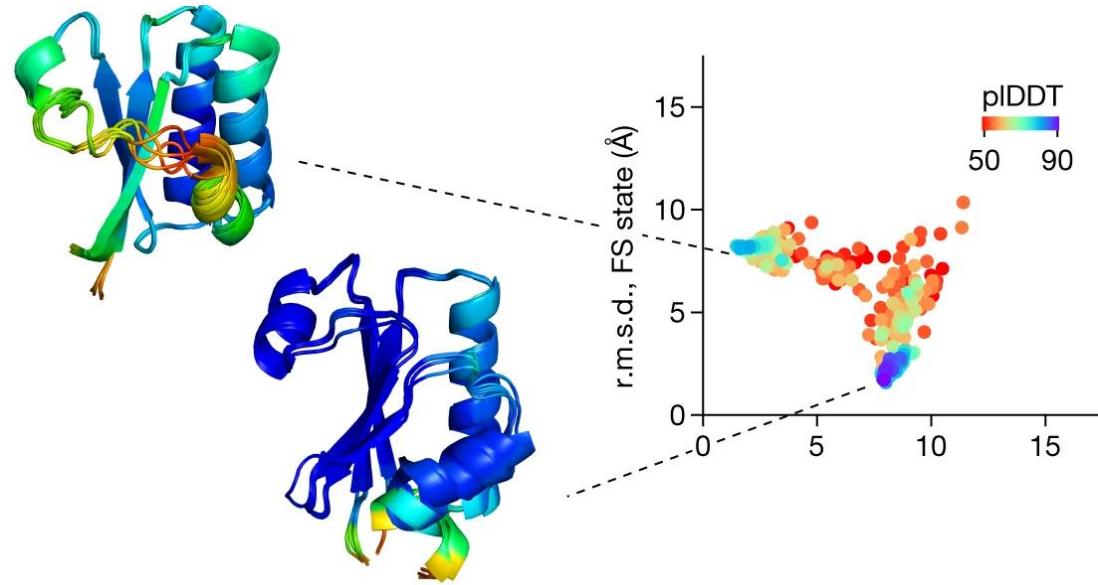
Simpkin et al "Tertiary structure assessment at CASP15" *Proteins* 2025

Verkuil et al "Language models generalize beyond natural proteins" *bioRxiv* 2022

DeepMind "AlphaFold" Github 2021

# Protein structure prediction

- Current ensemble prediction methods can model:
  - User-specified alternate conformations
  - Some aspects of Brownian motion
- They cannot:
  - Model relative proportions of different populations ([Vani 2023, Riccabona 2024](#))
  - Model interconversion dynamics/transition paths ([del Alamo 2023](#))
  - Model the effects of mutations, ligands, or modifications on these populations ([Zheng 2025, Ramasamy 2025](#))
- Requires lots of tinkering with settings, trial & error



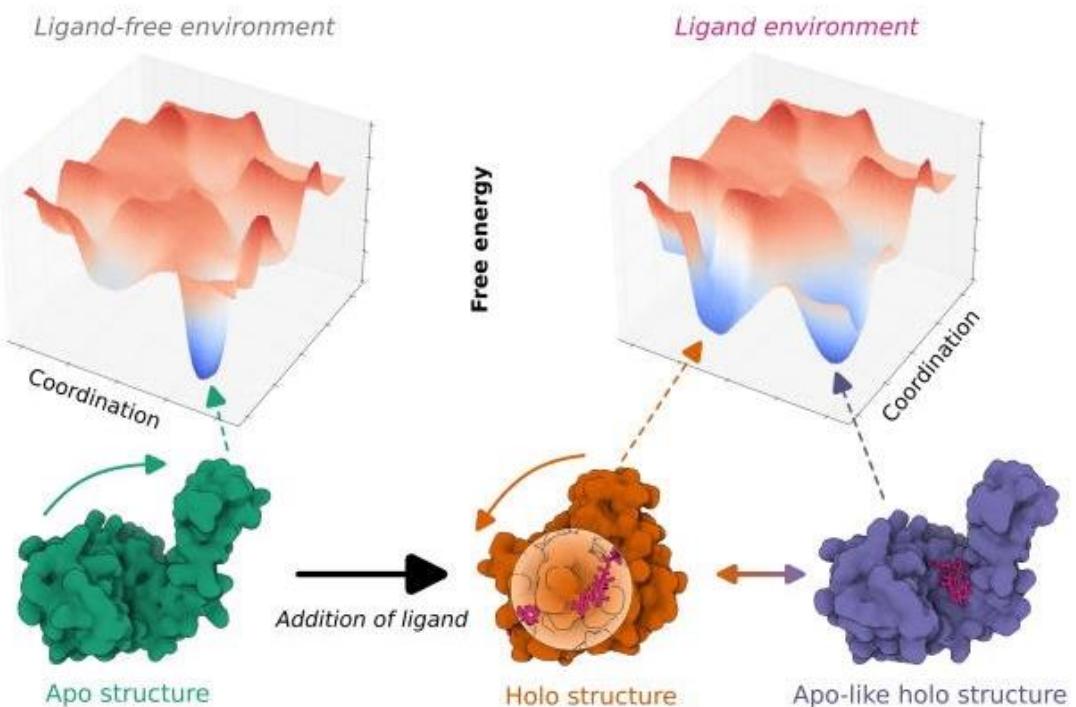
*del Alamo et al "Conformational sampling and interconversion using language-based protein folding neural networks" bioRxiv 2023*  
*Ramasamy et al "Assessing the relation between protein phosphorylation, AlphaFold3 models and conformational variability" bioRxiv 2025*  
*Riccabona et al "Assessing AF2's ability to predict structural ensembles of proteins" Structure 2024*  
*Vani et al "Exploring kinase DFG loop conformational stability with AlphaFold2-RAVE" JCTC 2023*  
*Wayment-Steele et al "Predicting multiple conformations via sequence clustering and AlphaFold2" Nature 2023*  
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# De novo prediction of protein structural dynamics: an outline

- What are protein dynamics?
- What kinds of biological problems are informed by studying dynamics?
- How can vanilla protein folding neural networks be modified to model dynamics?
- What modified and bespoke methods exist to answer these questions?
- Why don't these methods learn protein dynamics?
- Integration of these methods with molecular dynamics
- **Caveat:** this talk will only cover backbone modeling of well-structured proteins, and will ignore innovations in modeling intrinsically disordered proteins (IDPs), oligonucleotides, or sidechain ensembles

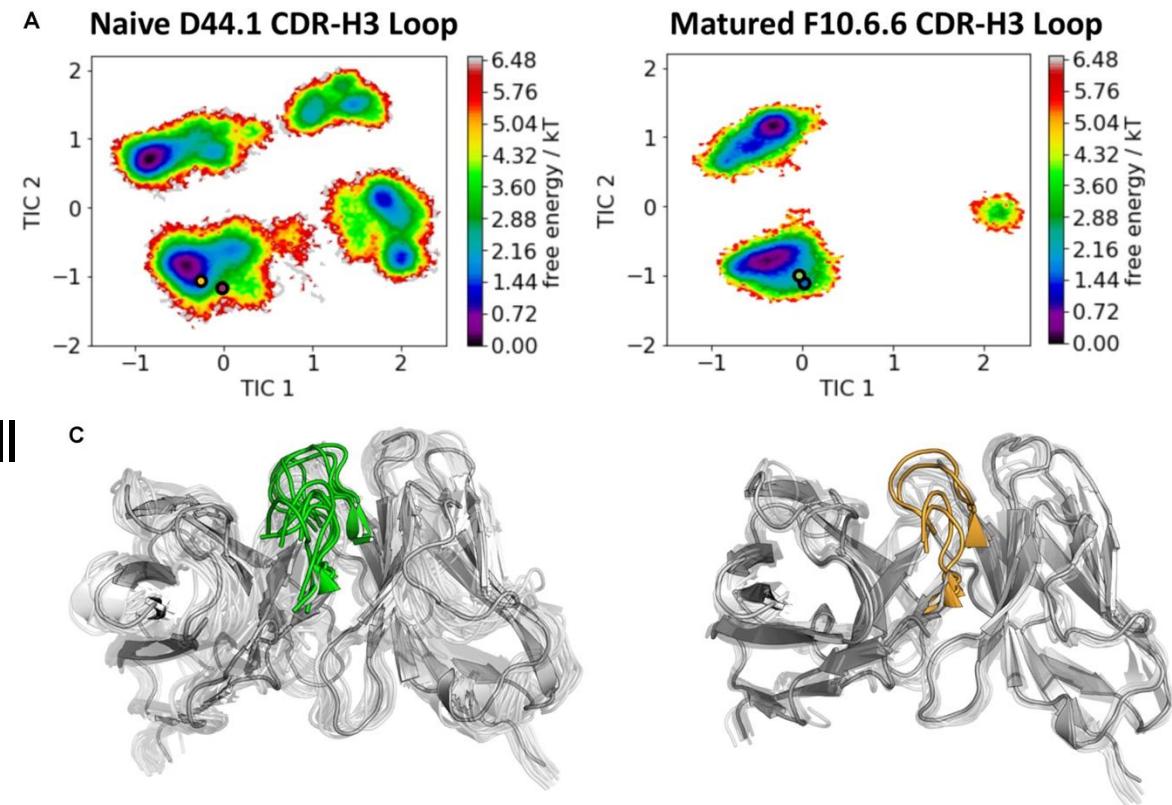
# What are protein dynamics, anyway?

1. Breadth of motion, e.g., Brownian motion (microstates)
2. Structures of distinct low-energy states (macrostates)
3. Boltzmann weights between distinct low energy populations
  - Transition paths and energy barriers separate these populations
  - The whole thing is called the free energy landscape
4. Effect of ligands, mutations, covalent modifications, and combinations thereof on the free energy landscape
  - Metals, phosphoryl groups, waters, protons, etc



# What protein dynamics tell us about biological mechanisms

- Conformational heterogeneity and binding promiscuity (antibodies)
- Active site preorganization (designed and/or engineered enzymes)
- Cryptic pockets and alternative druggable states (drug targets)
- Molecular basis of genetic diseases (almost all human proteins)
- Evolution of functional novelty (most proteins)
- Sometimes, structural modeling isn't the best way to answer biological questions about protein dynamics



Dishman et al, "Evolution of fold switching in a metamorphic protein" Science 2021

Fernandez-Quintero et al, "Local and Global Rigidification Upon Antibody Affinity Maturation" Frontiers in Immunology 2020

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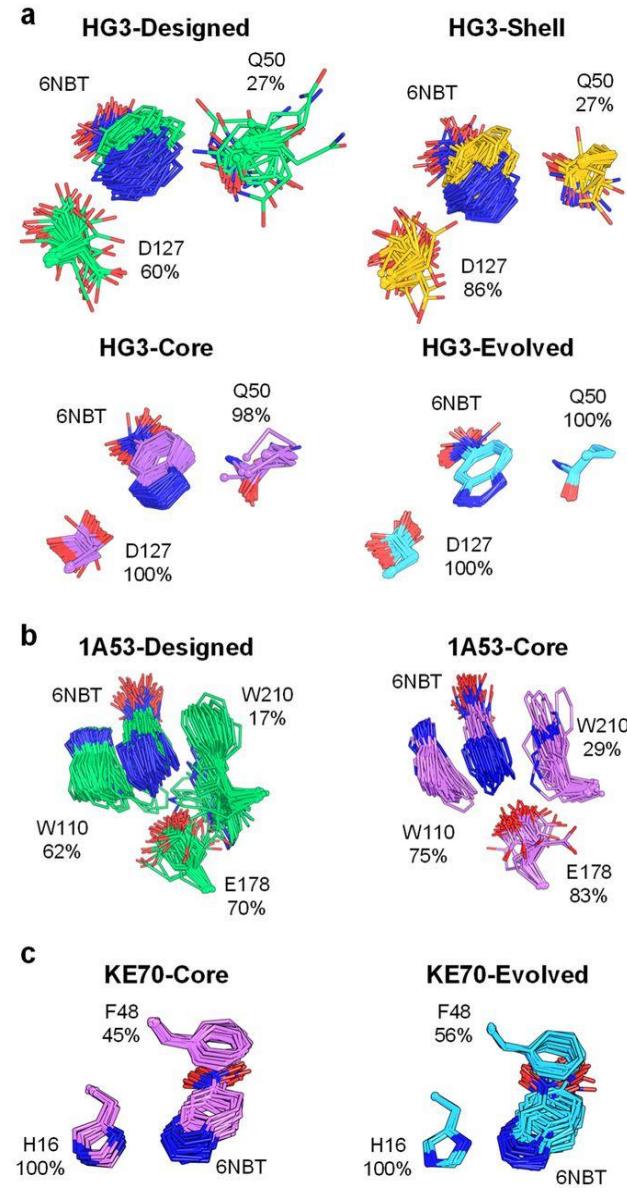
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Yee et al "The full spectrum of SLC22 OCT1 mutations illuminates the bridge between drug transporter biophysics and pharmacogenomics" Mol Cell 2025

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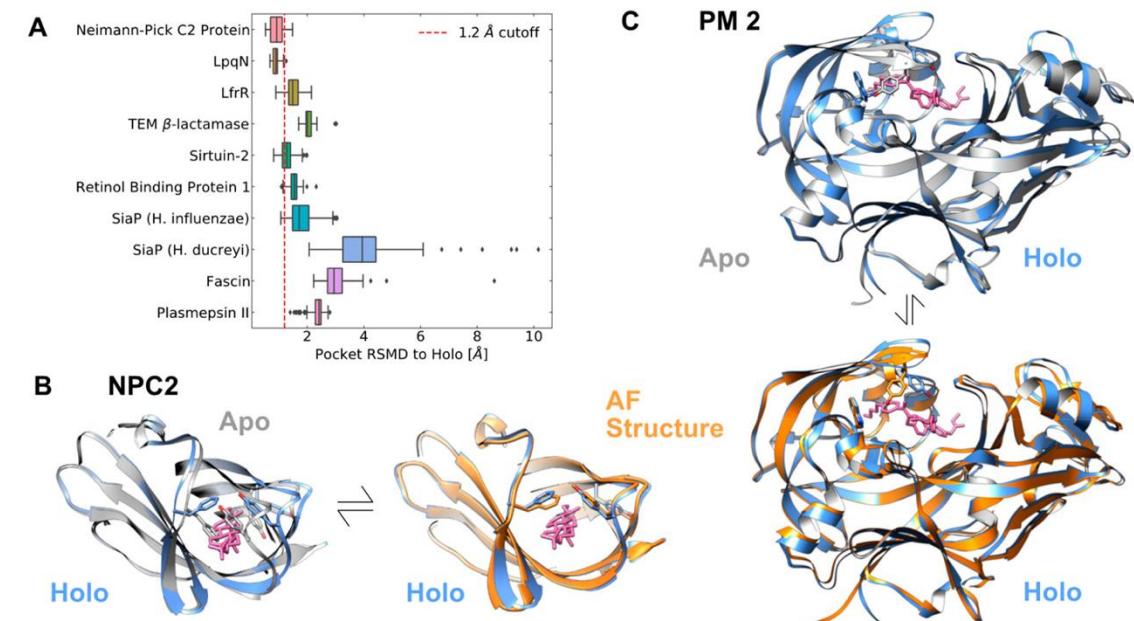
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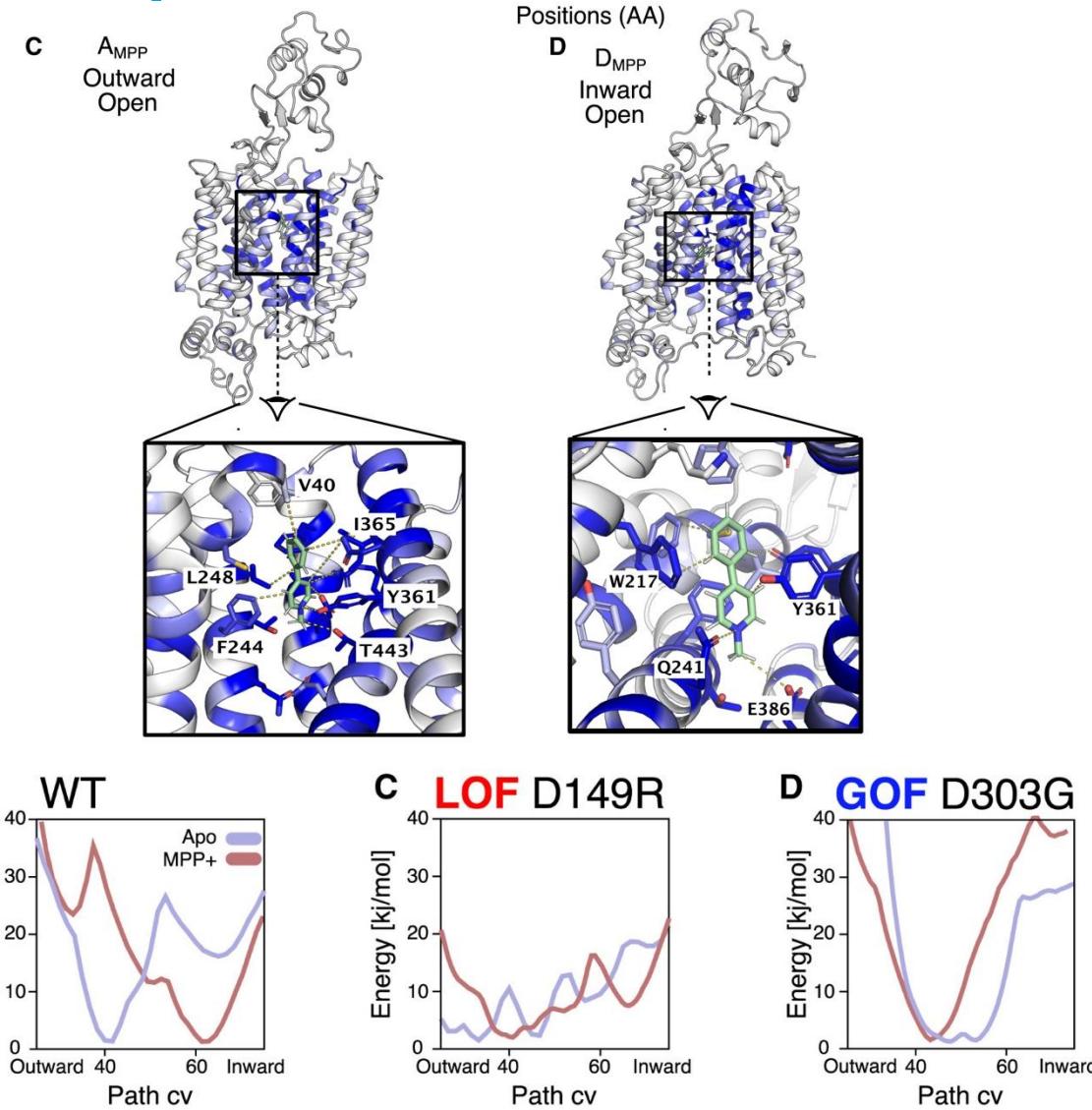
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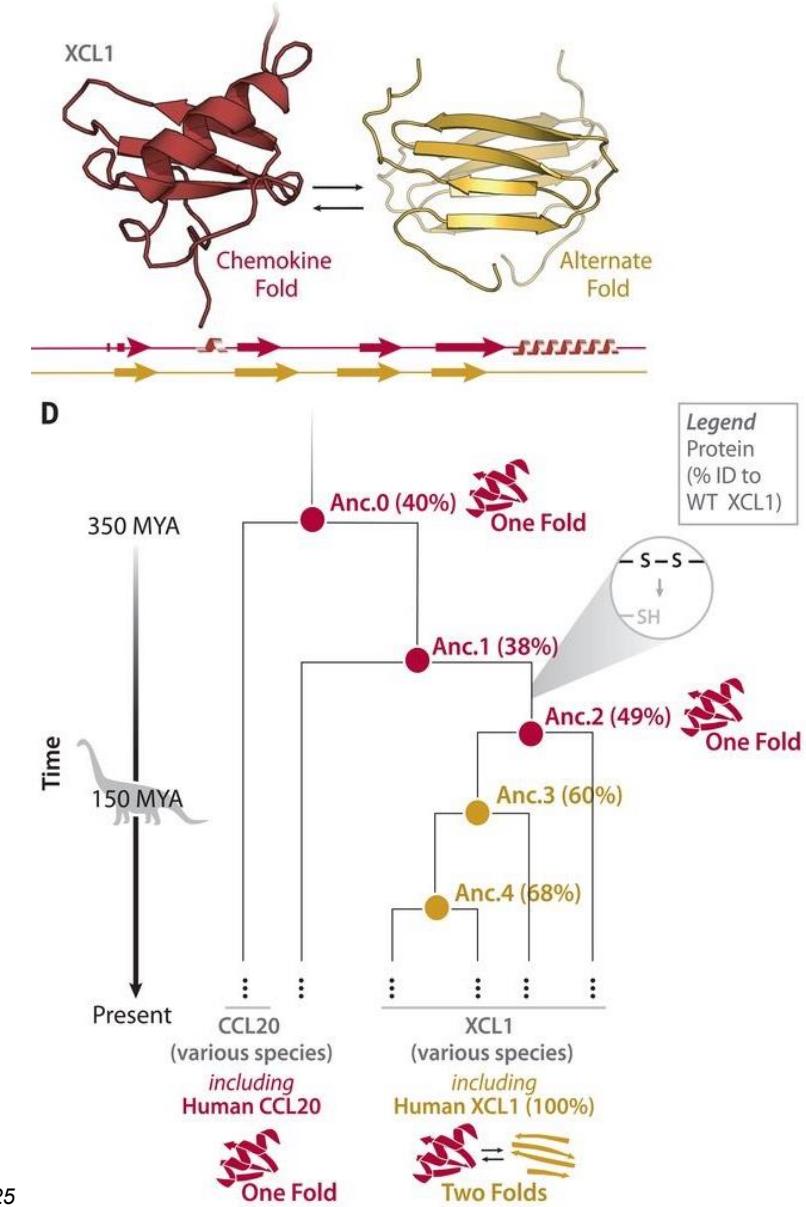
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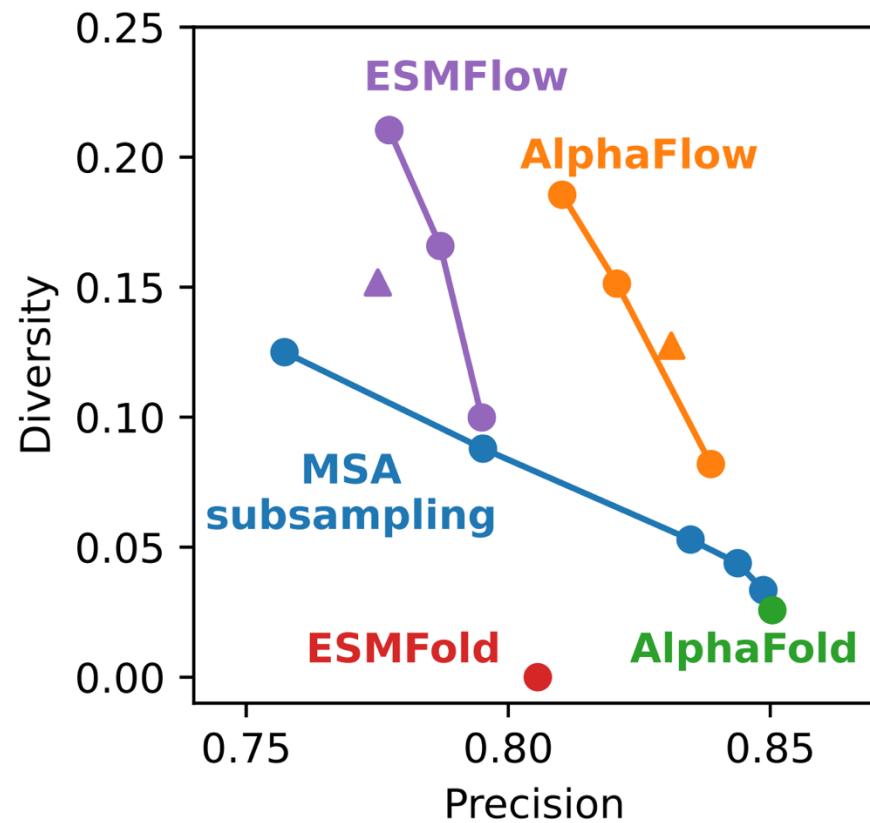
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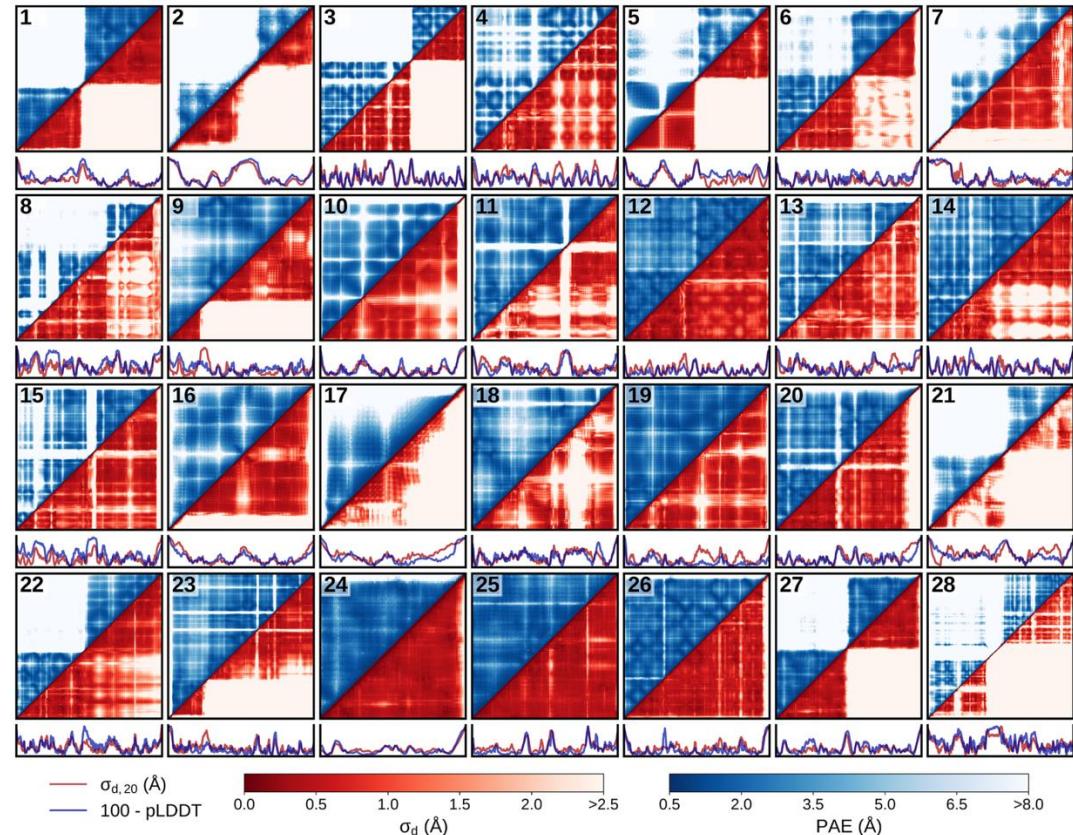
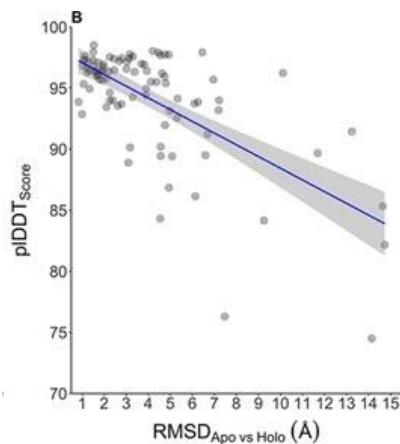
# The state of the art

- Modeling alternative conformations:
  - Vanilla AlphaFold2 with subsampled or modified MSAs, custom template databases, etc
  - Fine-tuned AlphaFold2 (AlphaFlow, DiG, etc), ESMFold (ESMFlow), or RosettaFold2 neural networks
  - Trained from scratch (Str2str, aSAMt, Cfold)
- Modeling entire free energy landscapes
  - Distributional Graphomer (DiG), aSAMt, P2DFlow
  - BioEmu: Prediction of free energy landscapes of monomeric proteins to ~1 kcal/mol
- Precision/diversity trade-off: one highly accurate model vs lots of potentially inaccurate models
- MD is the gold standard, but is exorbitantly expensive



# Protein folding NNs already “know” dynamics

- Error/confidence metrics show some correlation with mobility/conformational flexibility in MD simulations (**Gavaldo-Garcia 2025, Jussupow 2023, Saldaño 2022**)
- Contacts for non-sampled states show up in distograms (**Jumper 2021**)
- (Note that error metrics in AF2 & RosettaFold2 are highly correlated; **Bennett 2023**)



**Figure 2.** Overview of AlphaFold scores and protein dynamics metrics for all studied systems. A fully symmetric matrix indicates a perfect correlation between the AlphaFold and MD data. The figure is divided into 28 subfigures for each of the proteins shown in [Figure 1](#). The top matrices show a comparison between (symmetrized) PAE matrices (blue) against the standard deviation of all C<sub>α</sub> distances σ<sub>d</sub> (red). The PAE scores range between 0.5 and 8.0, while the threshold for σ<sub>d</sub> is set to <2.5 Å to enable better comparisons between the different systems. The bottom graphs compare the pLDDT scores (blue) and average standard deviation of the C<sub>α</sub>-distances to the 20 closest amino acids σ<sub>d,20</sub> (red). See Figures S1–S28 for a detailed analysis of the individual systems.

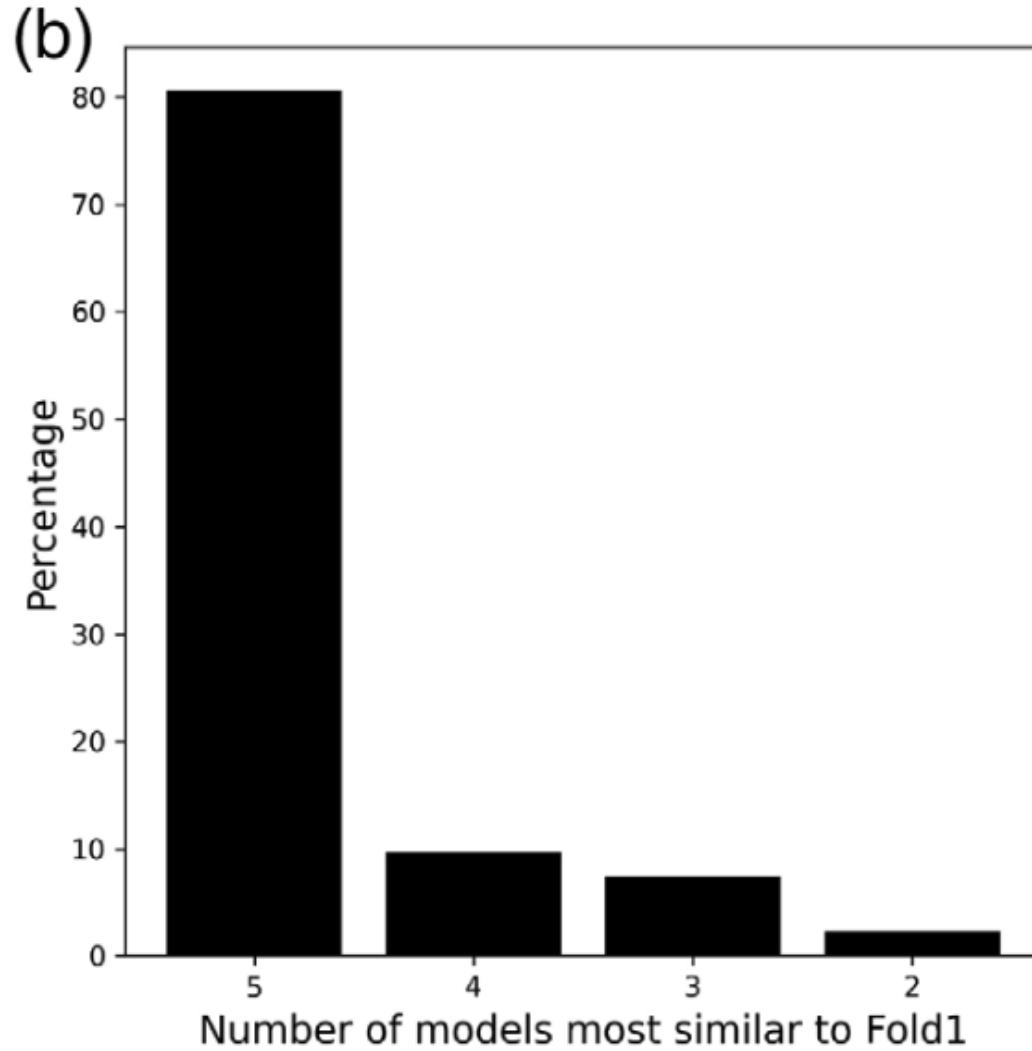
Bennett et al “Improving de novo protein binder design with deep learning” *Nature Comms* 2023

Gavaldo-Garcia et al “Gradations in protein dynamics captured by experimental NMR are not well represented by AlphaFold2 models and other computational metrics” *JMB* 2025

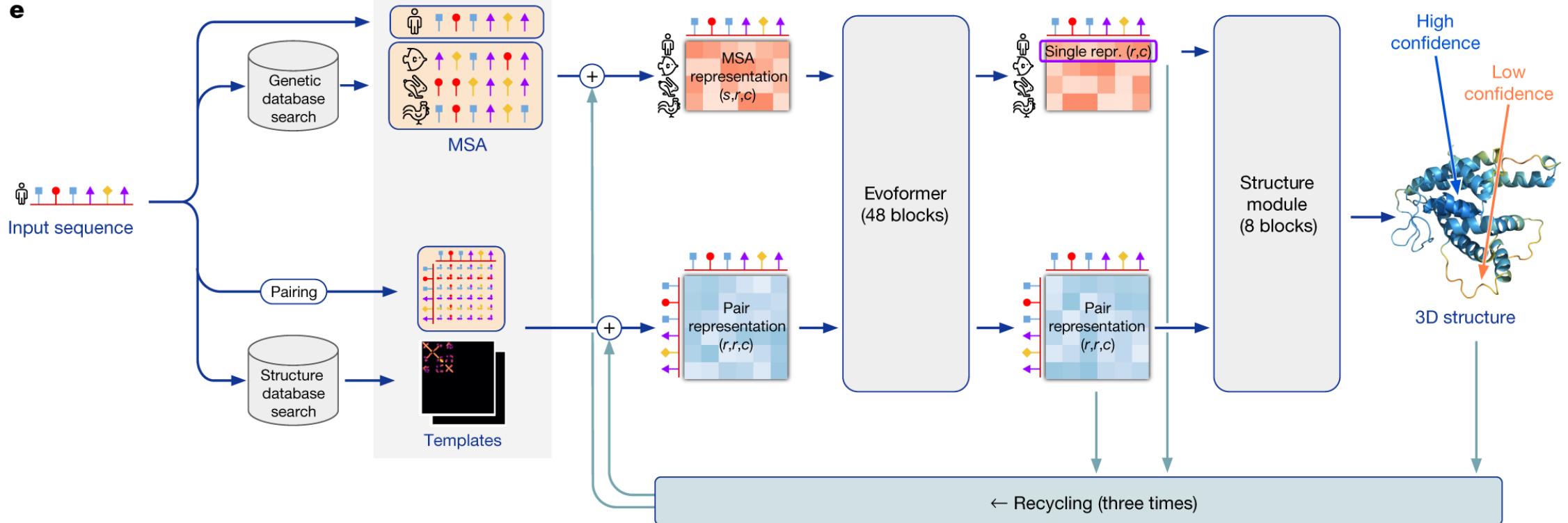
Jussupow & Kaila “Effective Molecular Dynamics from Neural Network-Based Structure Prediction Models” *JCTC* 2023

Saldaño et al “Impact of protein conformational diversity on AlphaFold predictions” *Bioinformatics* 2022

## Protein folding NNs tend to sample the same state repeatedly

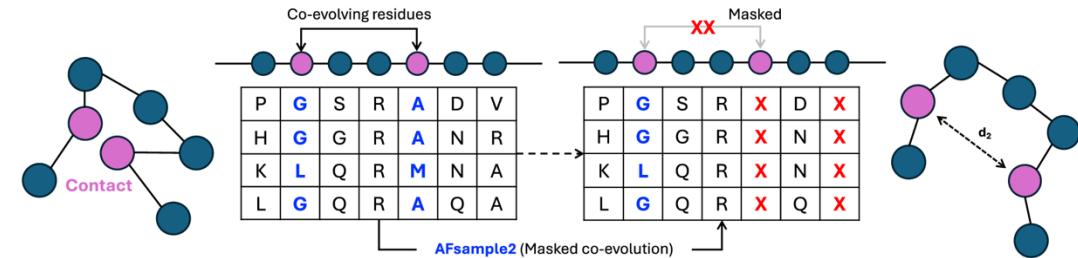


# Review of AF2 architecture



# Predicting alternate conformational states with vanilla AlphaFold2

- Sequence-based methods that work
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  - MSA sourcing ([da Silva 2024, Faezov 2023](#))
  - MSA clustering ([Wayment-Steele 2023](#))
  - MSA subsampling ([del Alamo 2022](#))
  - Truncation of autoinhibitory domains ([del Alamo 2022a](#))
  - Inclusion of conformationally selective binding partners ([Cummins 2022](#))
- Sequence-based augmentations that don't work:
  - Point mutations, including those that work *in vitro* ([Ramsamy 2025, Vani 2023, Zheng 2025](#);  
sometimes many mutations work)



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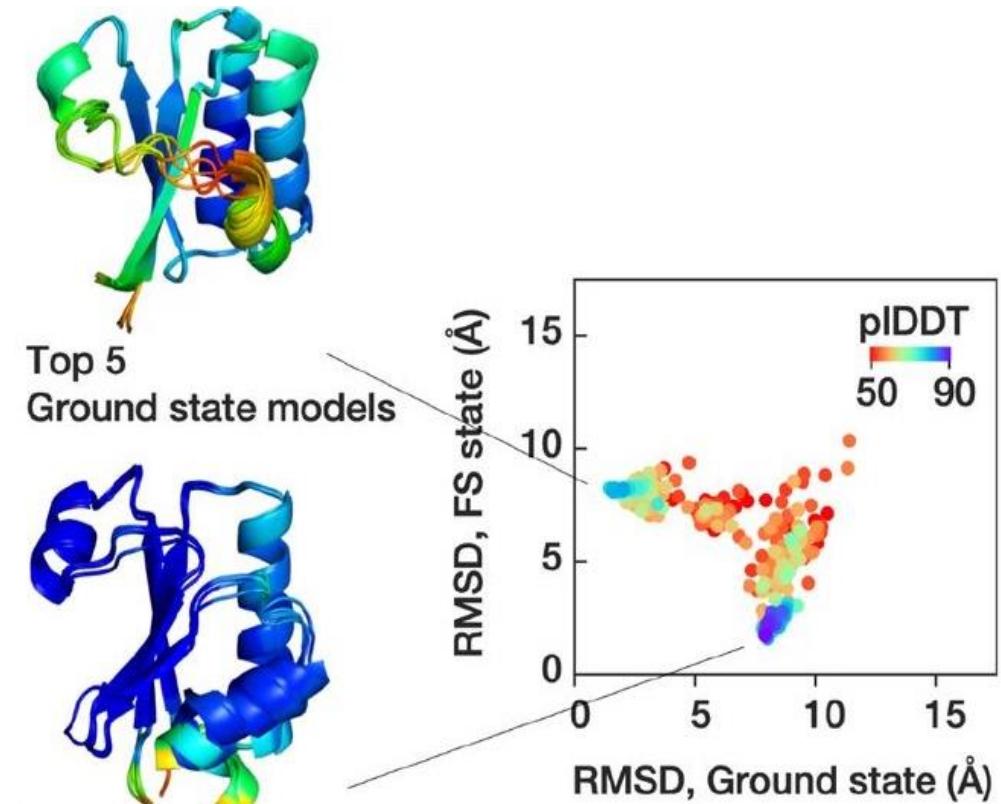
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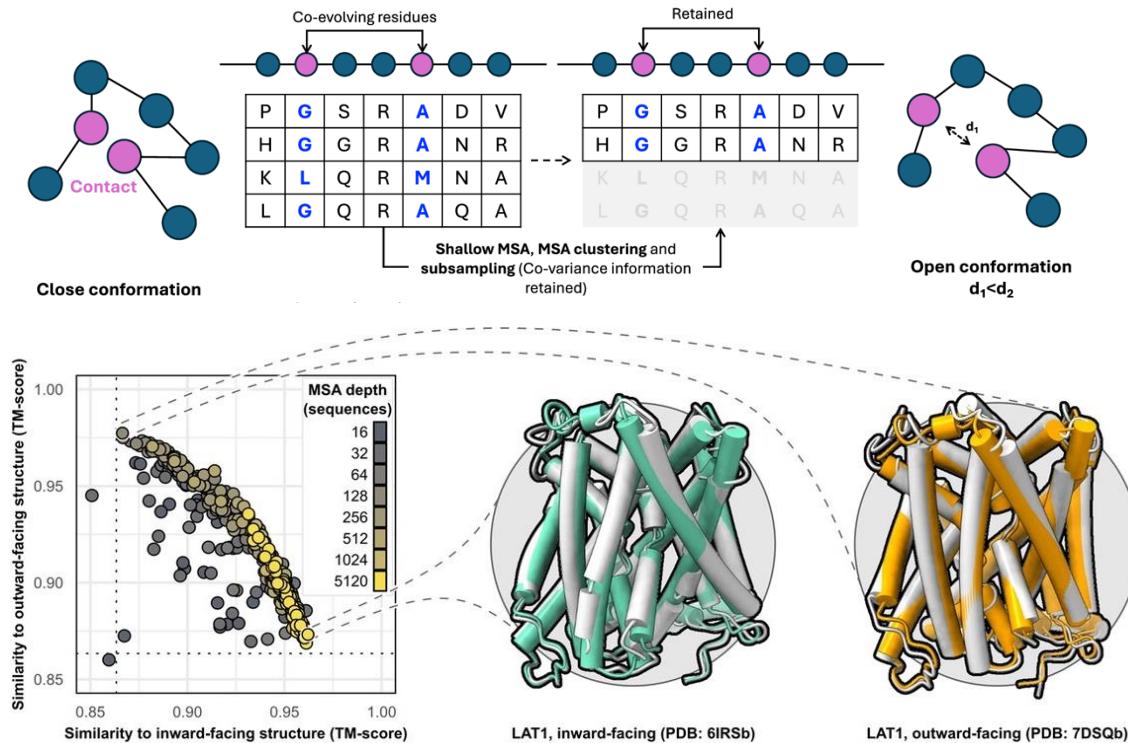
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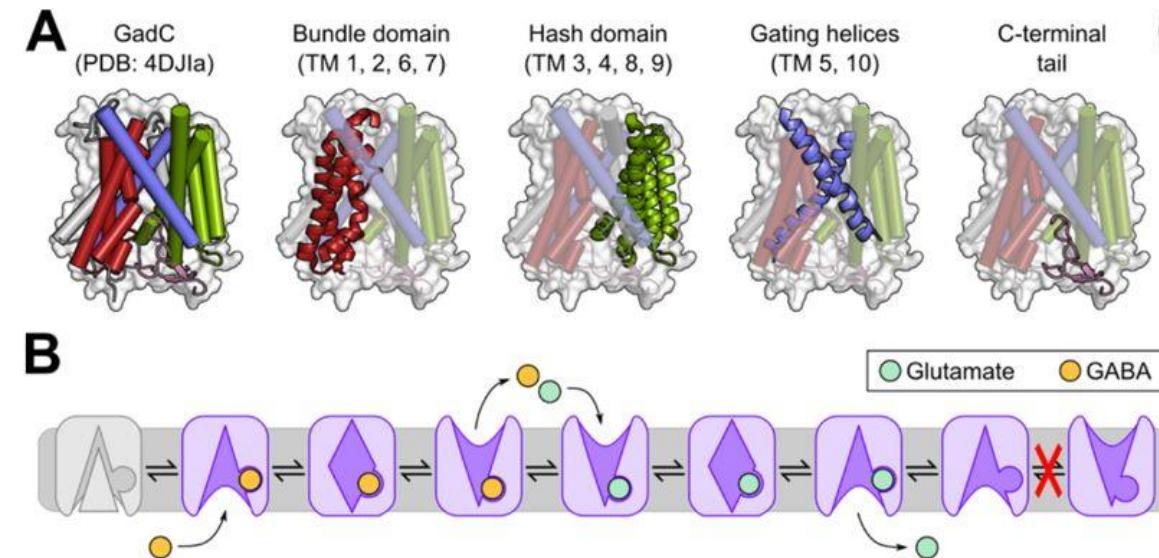


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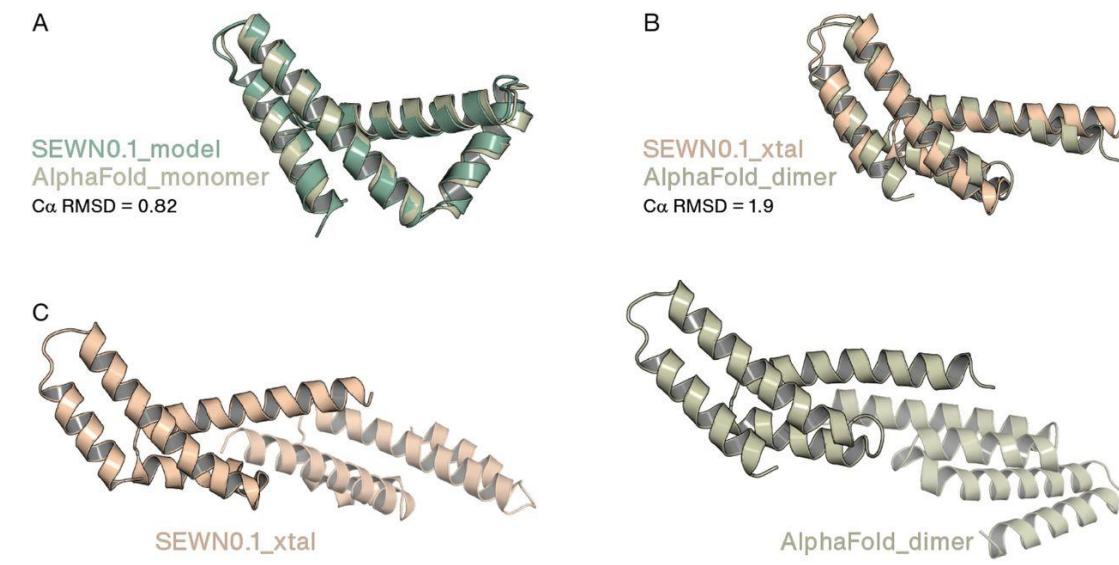


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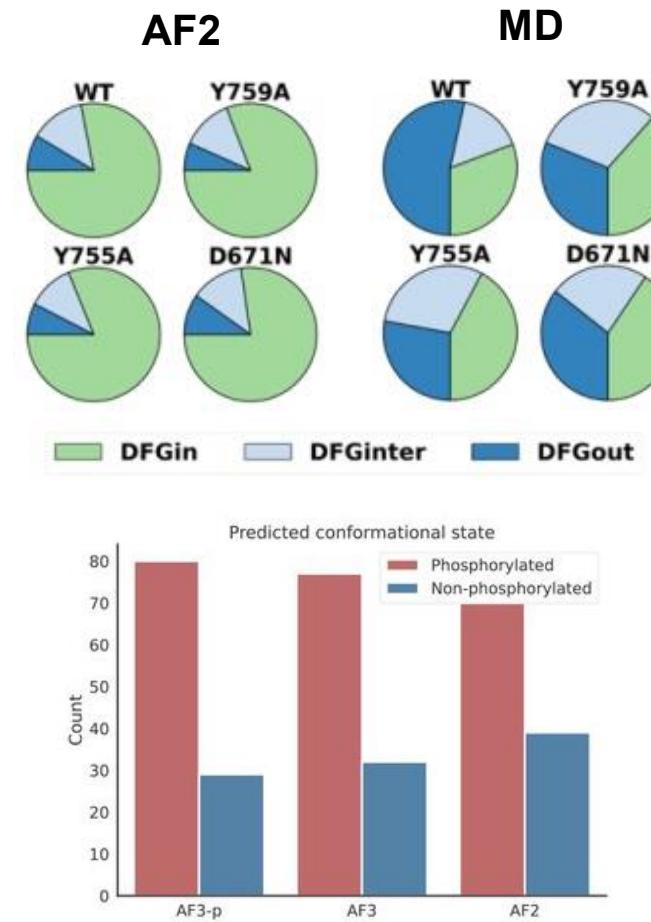
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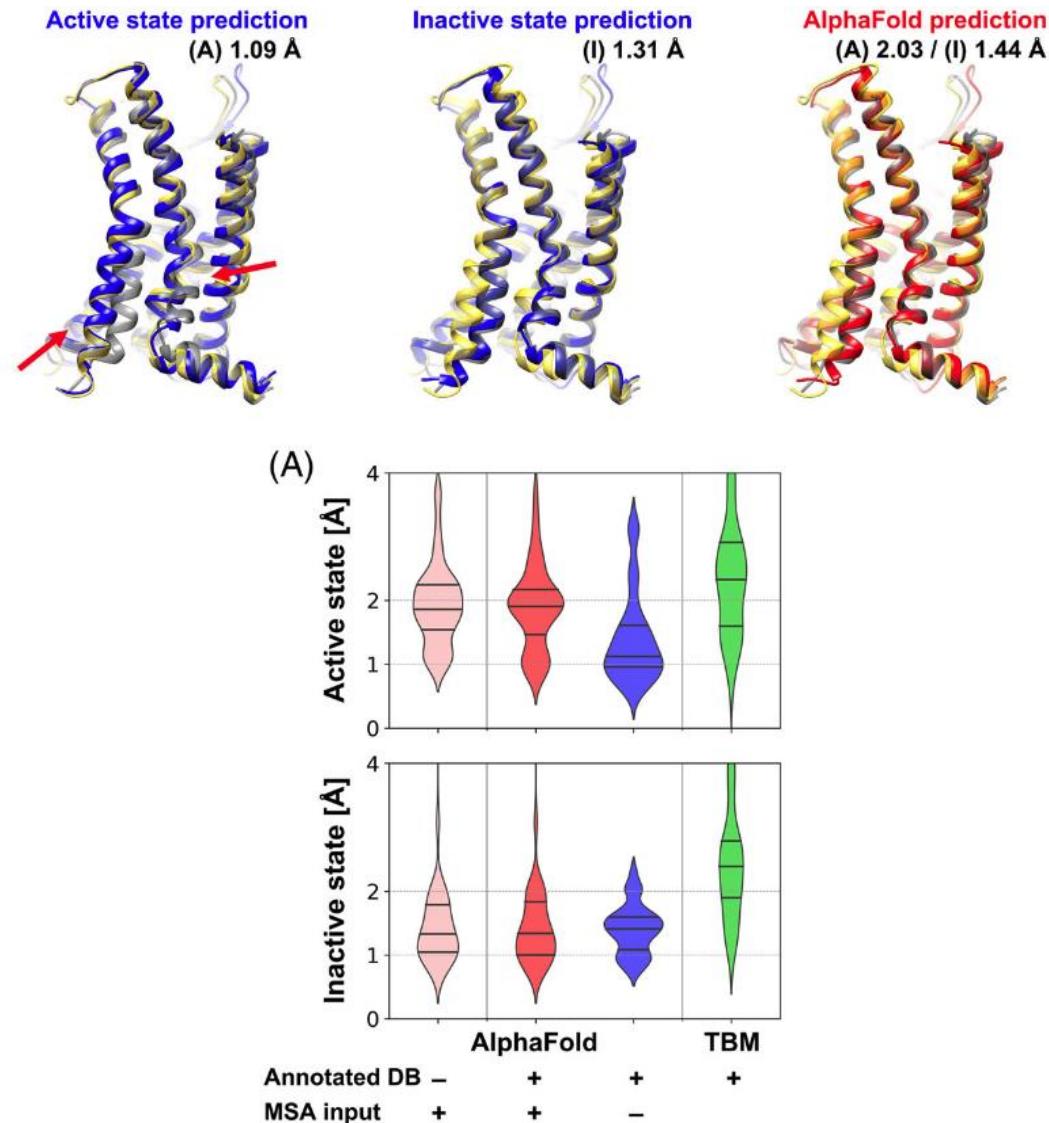
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- Structure-based approaches that work:
  - Template curation** (del Alamo 2022, Faezov 2023, Heo 2022)
  - Caveat:** only two AF2 monomer models can use templates (all multimer models can use templates)

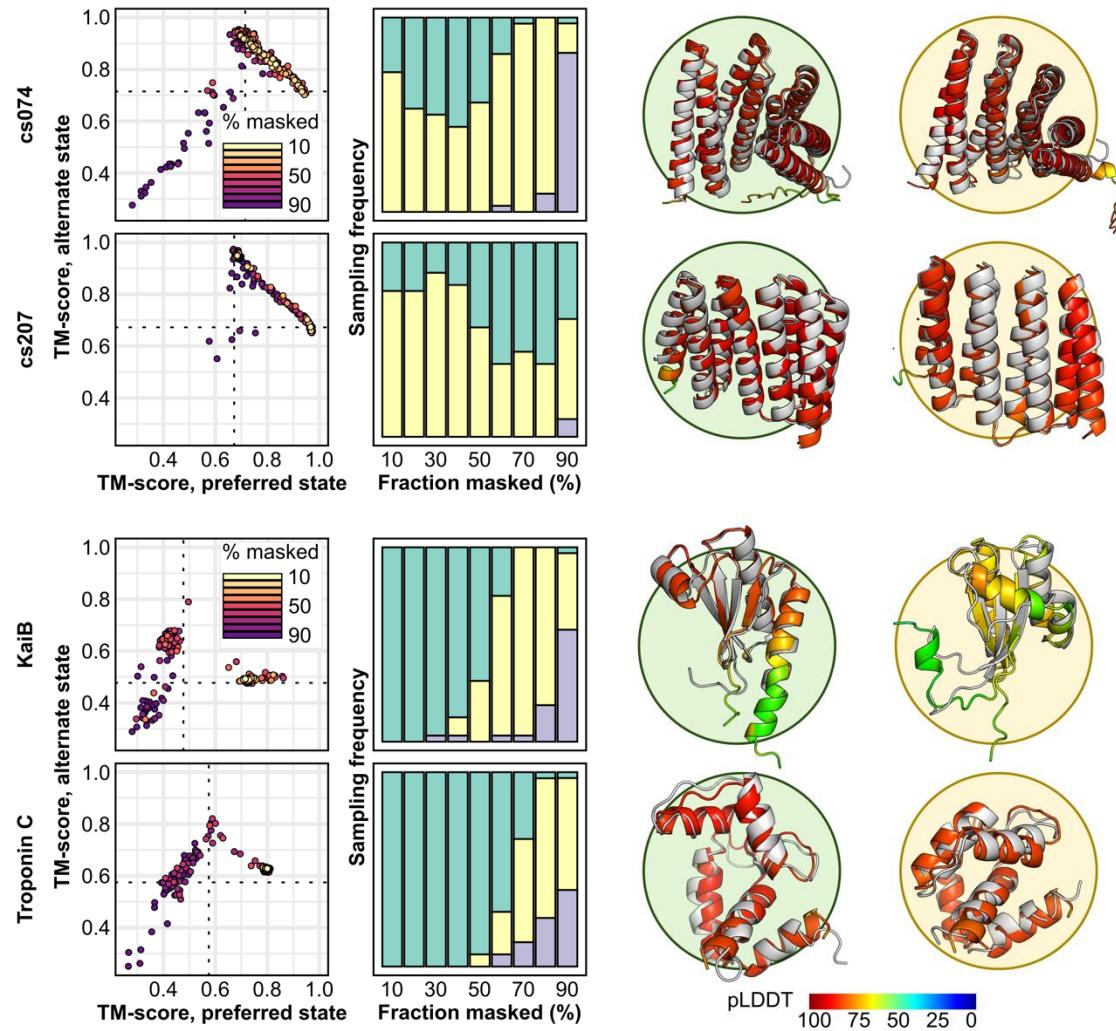


del Alamo et al, "Sampling alternative conformational states of transporters and receptors with AlphaFold2" eLife 2022

Faezov & Dunbrack "AlphaFold2 models of the active form of all 437 catalytically competent human protein kinase domains" bioRxiv 2023

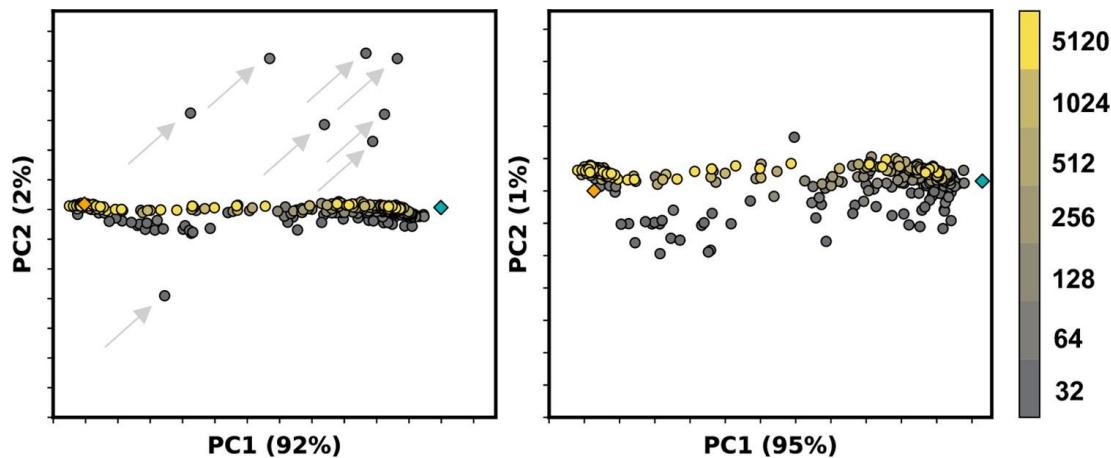
Heo & Feig "Multi-state modeling of G-protein coupled receptors at experimental accuracy" Proteins 2022

# Strategies that work with other NNs



# How to filter bad models

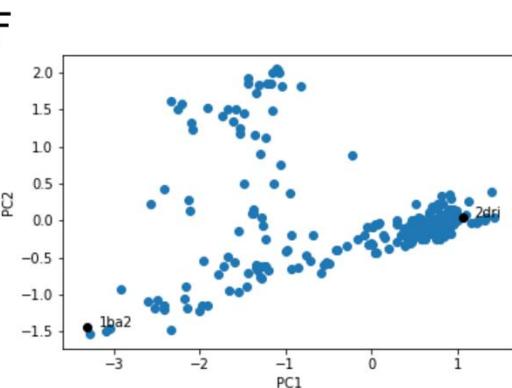
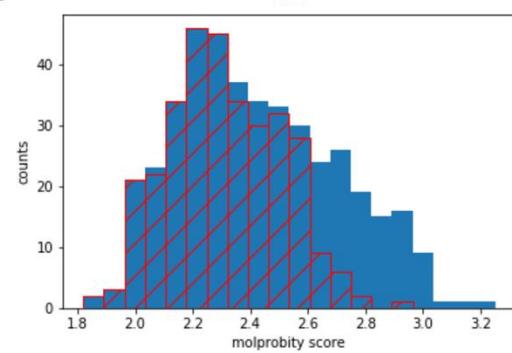
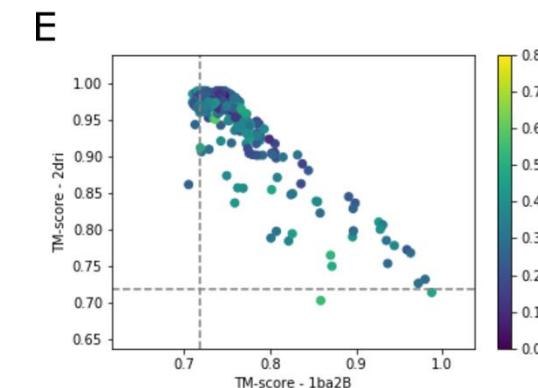
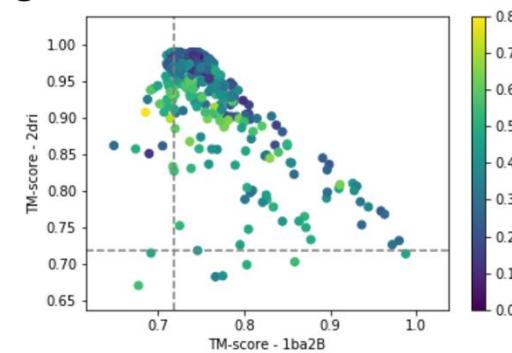
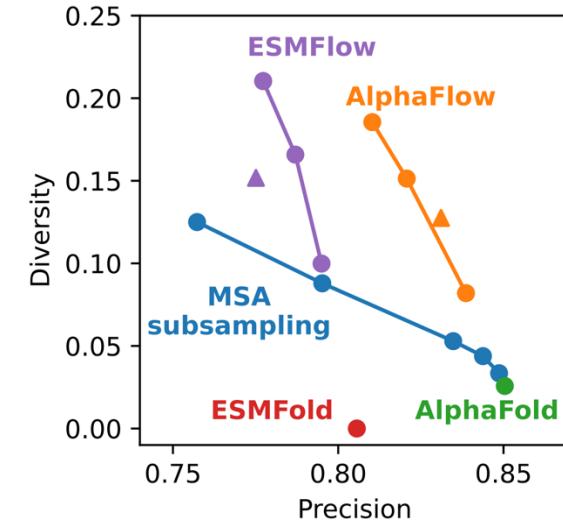
- Iterative PCA: Look for ensembles where predicted motions of models are consistent with one another
- Model quality check: Use MolProbity score to check for residue-scale errors
- Manually



del Alamo et al "Sampling alternative conformational states of transporters and receptors with AlphaFold2" eLife 2022

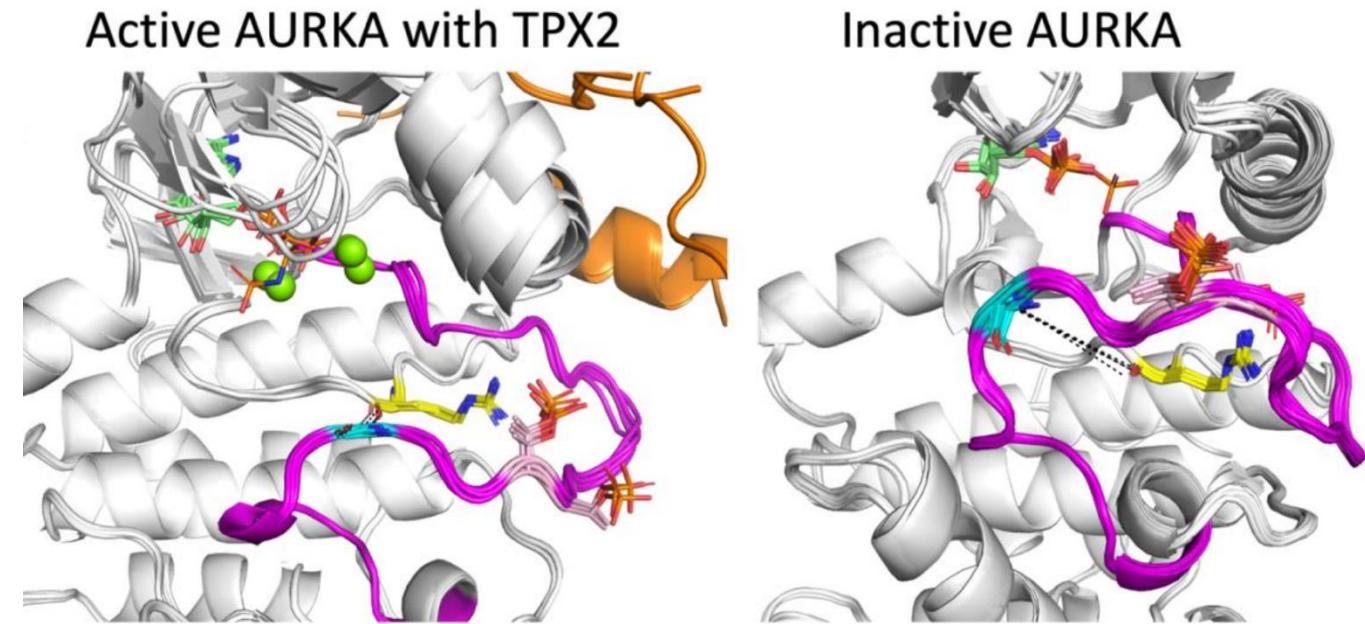
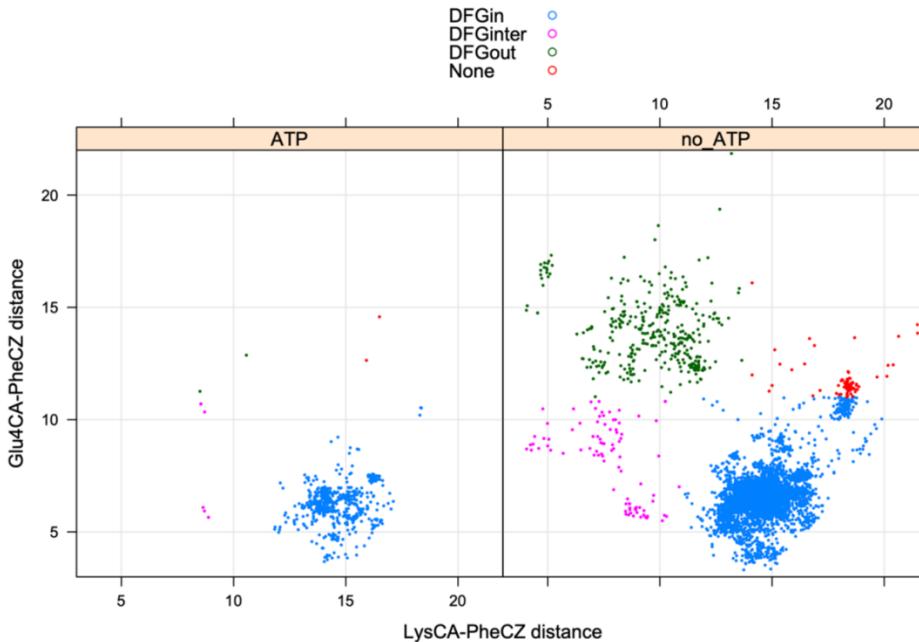
Jing et al "AlphaFold meets Flow Matching for generating protein ensembles" ICLR 2024

Stein & Mchaourab "SPEACH\_AF: Sampling protein ensembles and conformational heterogeneity with AlphaFold2" Plos Comp Biol 2022

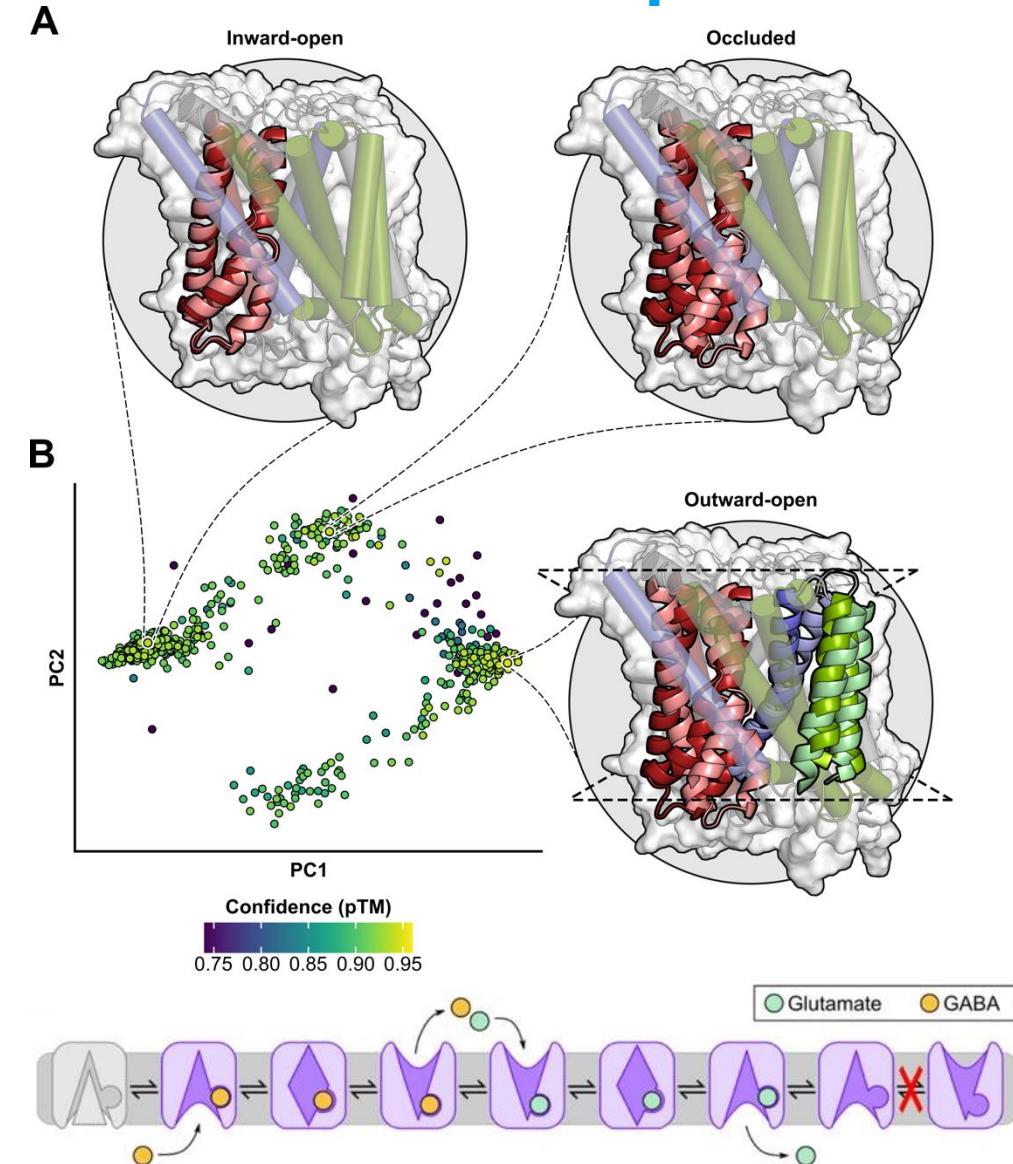
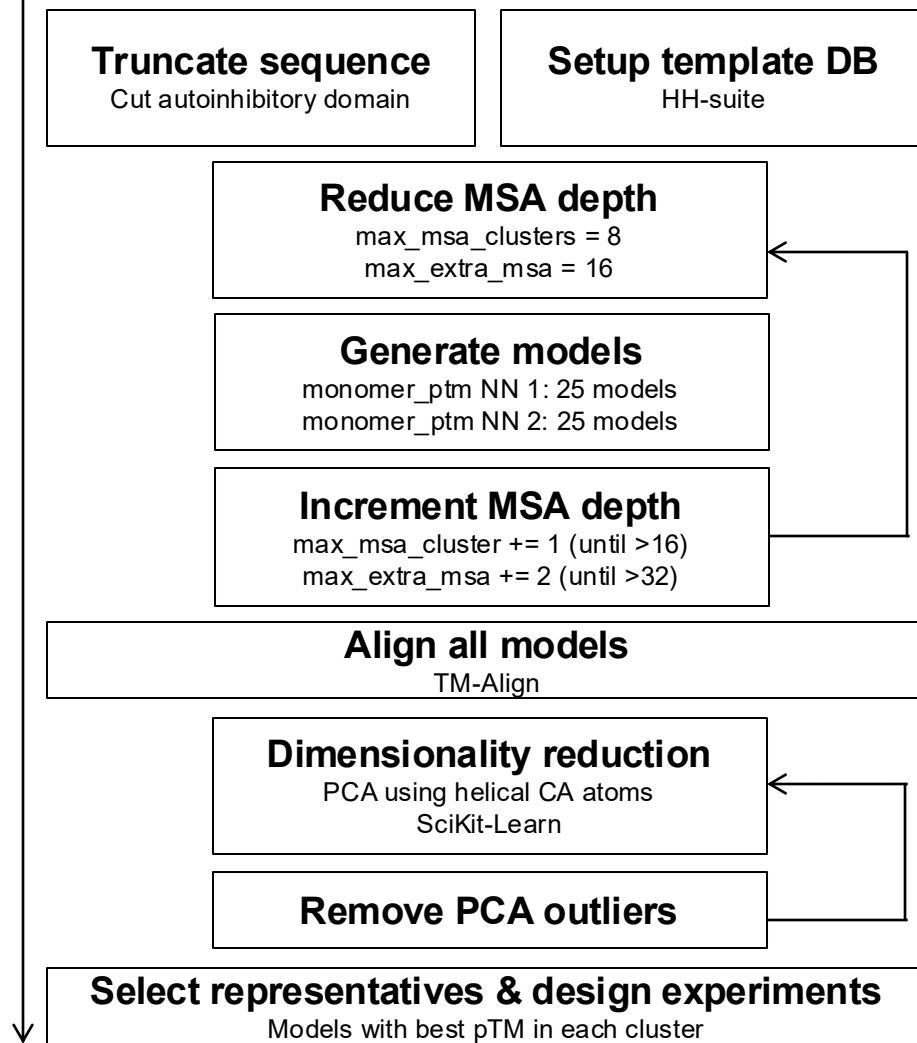


## Example: prediction of all 437 kinases in the human proteome

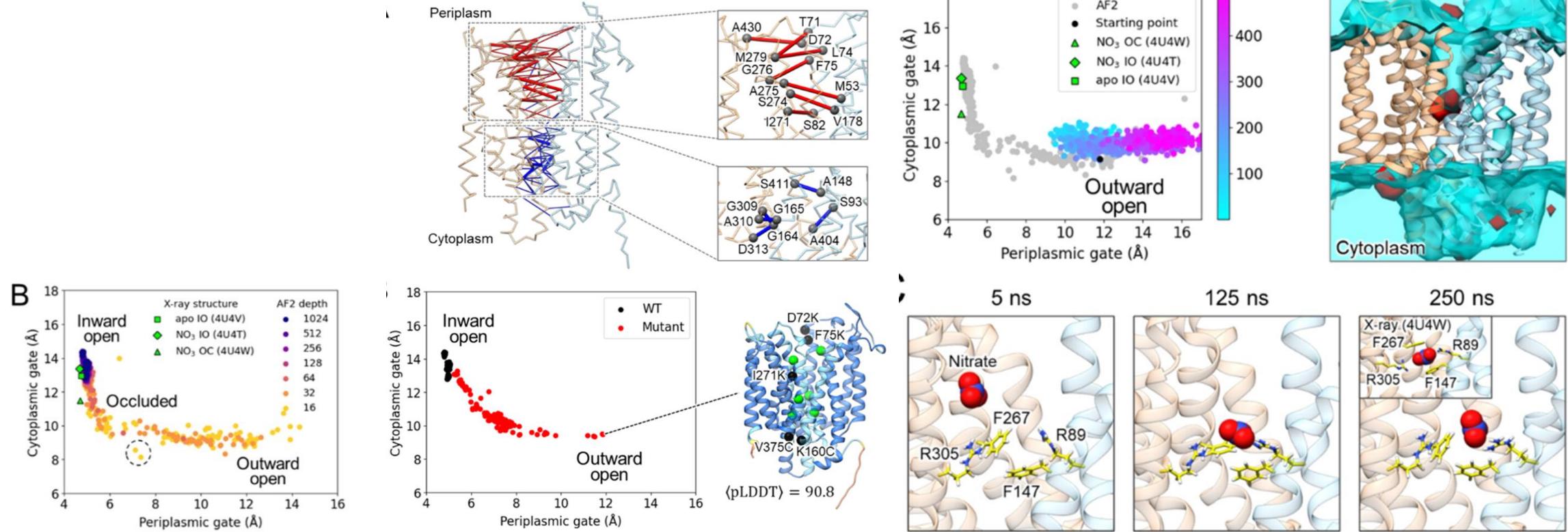
- Faezov & Dunbrack predicted the active states of all kinases using MSA sourcing, MSA subsampling, and custom template DBs
- “*No one set of inputs (MSA source, MSA depth, template database) produces active models of all 437 catalytic targets.*”
- For two cases, AF2 models of active kinases were required as templates



# Example: modeling alternative conformations of a transporter



# Example: modeling alternate conformation of the transporter NarK



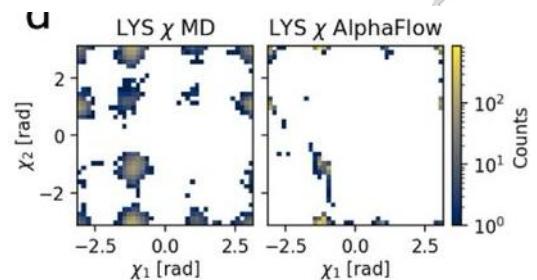
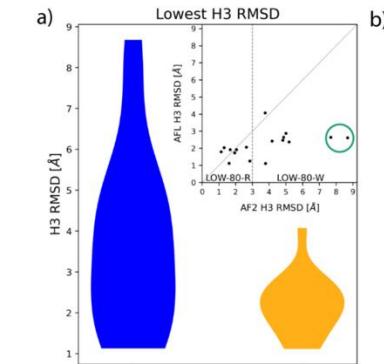
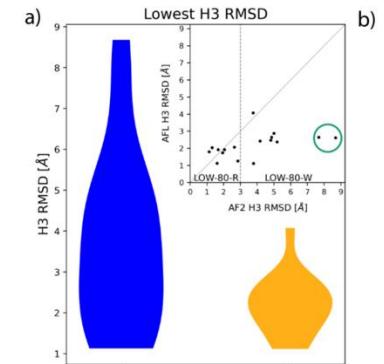
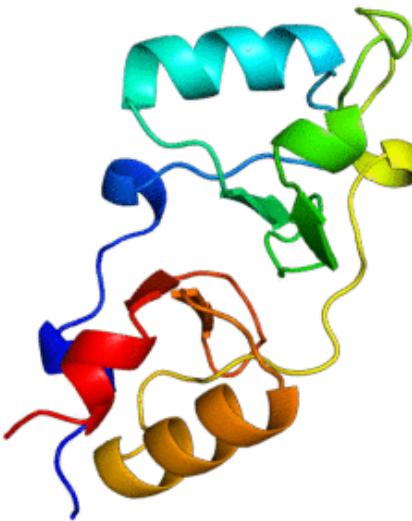
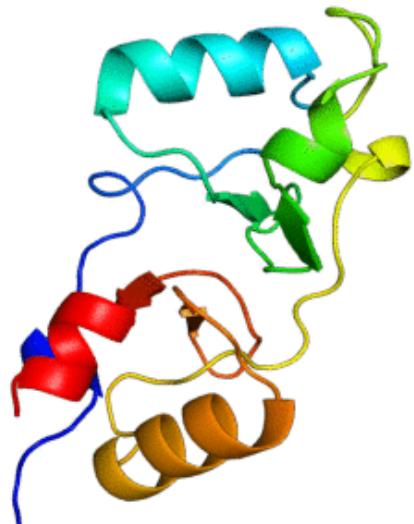
# Custom neural networks for obtaining alternate states

- What makes these special?
  - Custom training data (AlphaFlow, DiG, aSAMt) such as family-specific data (**Xu 2025, Mansoor 2024**)
  - Curated or reweighted training sets (Cfold, UFCConf)
  - Distinct architectures and training schedules (AlphaFlow, BioEmu, DiG, RF-VAE)
  - Noised inputs (Str2str, diffusion- and flow matching-based methods)
- In general, these are:
  - Worse at single-state modeling than dedicated protein folding NNs
  - A bit better at recovering multiple relevant states (although not always)
  - Much better at modeling “Brownian motion”

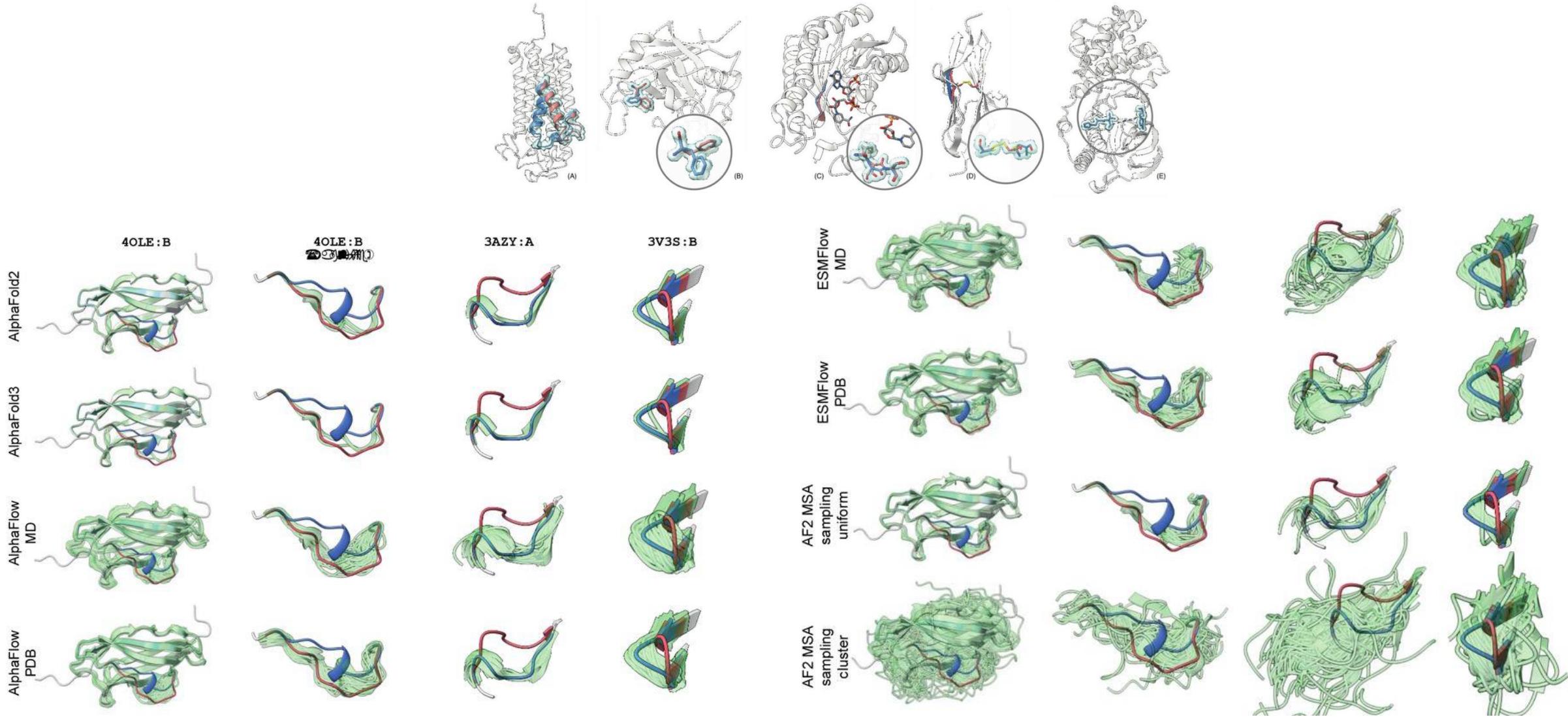
# AlphaFlow and ESMFlow

- Fine-tuned AlphaFold2/ESMFold for prediction of different conformations – trained on short (many  $<1 \mu\text{s}$  MD simulations; **Jing 2024**)
- Anecdotally, doesn't work for all proteins, such as fold-switching proteins
- Still requires MSA for AlphaFold
- In a separate benchmark, AlphaFlow but not AF2 always generated antibody CDRH3 conformations within 3.0 Å RMSD of ground truth (note: monomer only; **Giulini 2025**)
- Limited side chain sampling (**Janson 2025**)

300 ns MD

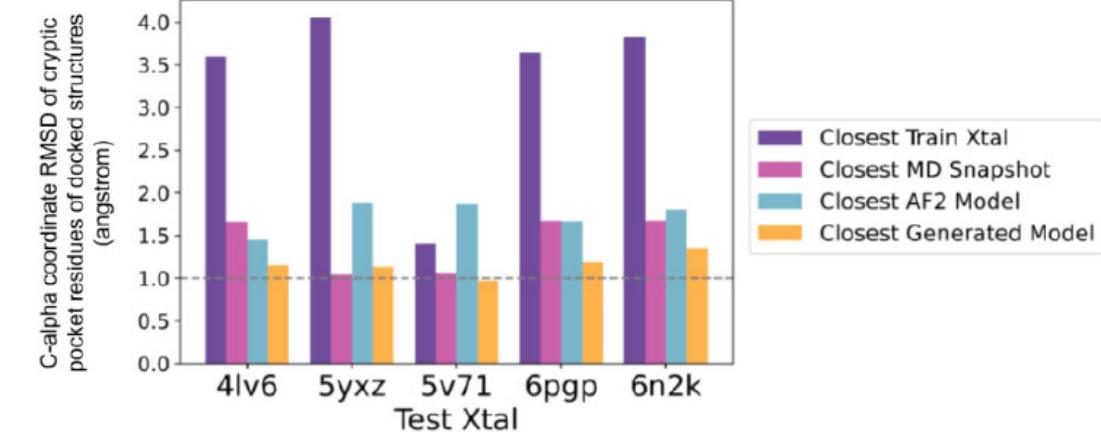
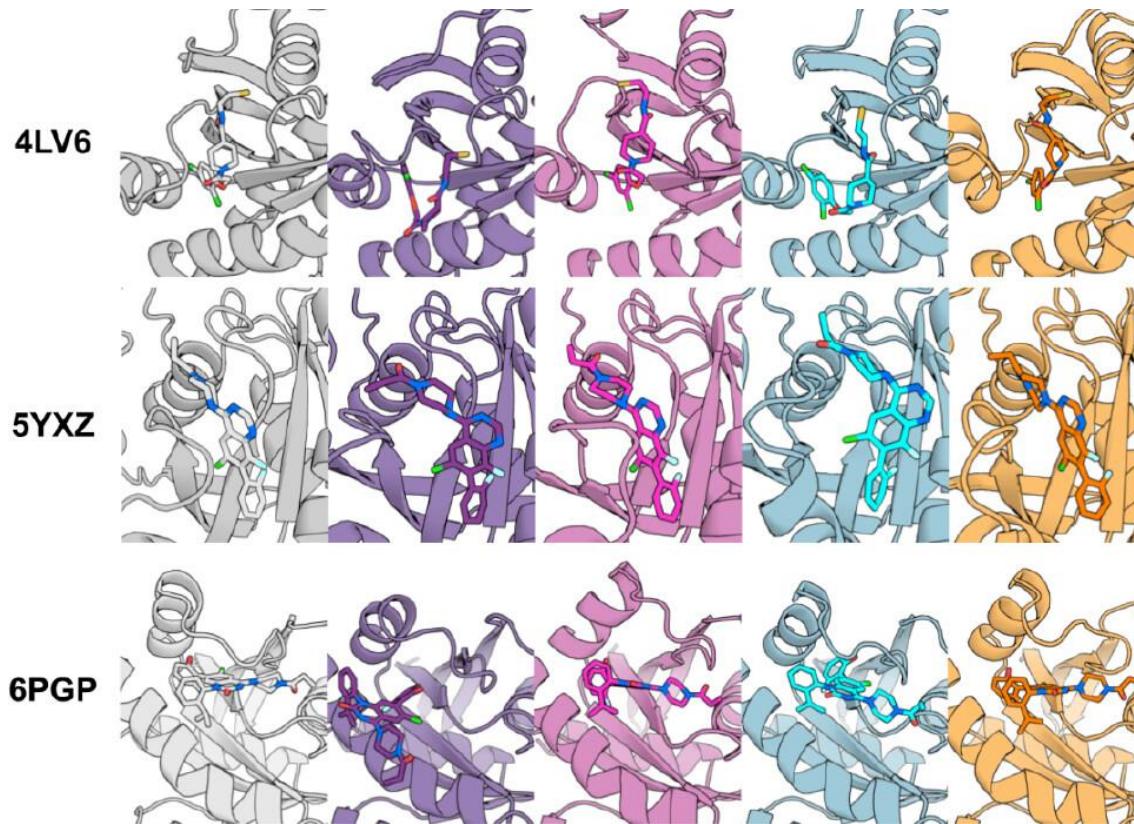


# Examples of these methods failing to predict alternate states



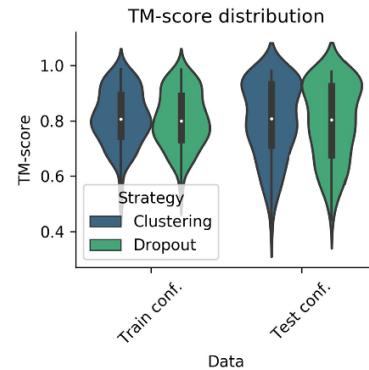
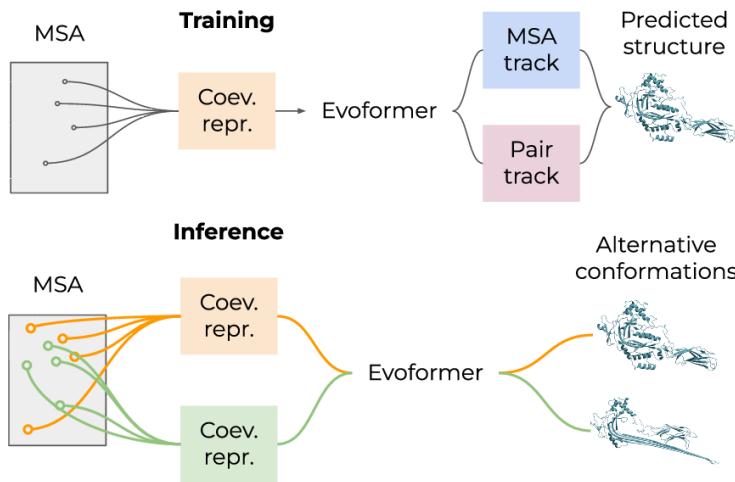
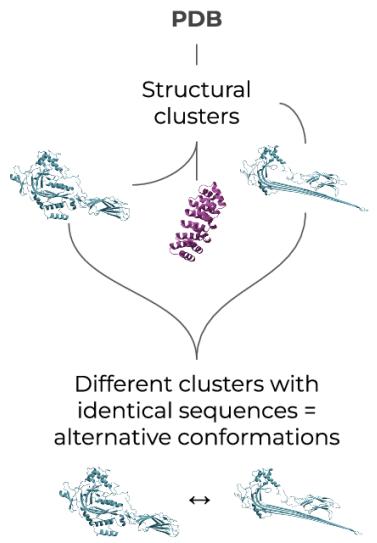
# Fine-tuned RosettaFold+VAE for exploring Ras conformations

- Variational autoencoder added to RosettaFold, fine-tuned on many Ras structures and MD snapshots (**Mansoor 2024**)
- Tested on held-out crystal structures

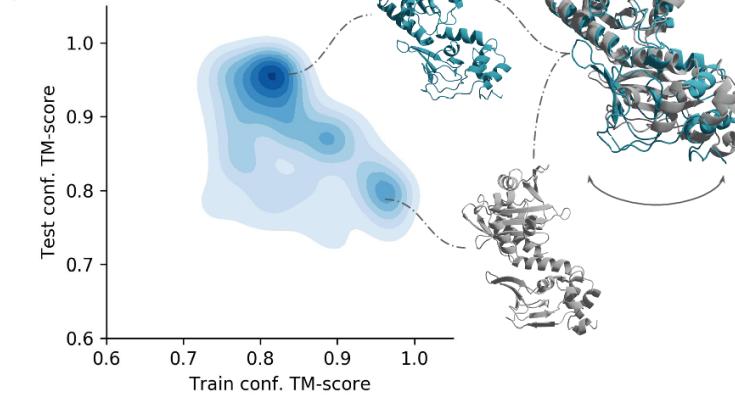


# Cfold: retrained AF2-like NN with a bespoke training set

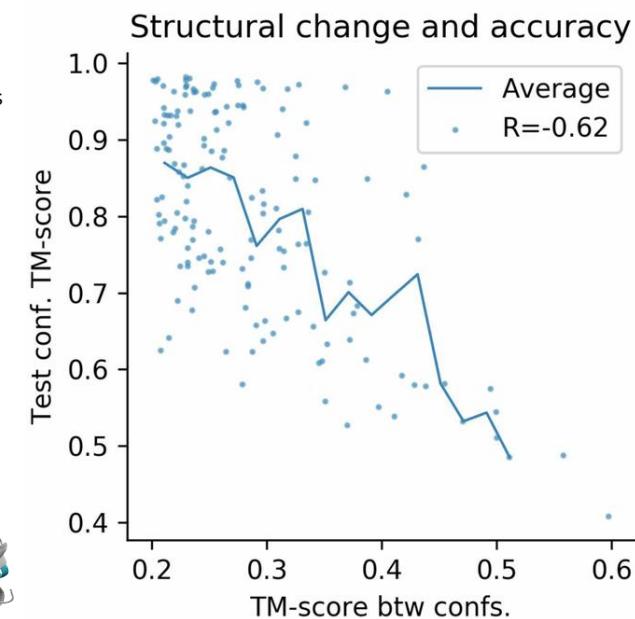
## a Cfold



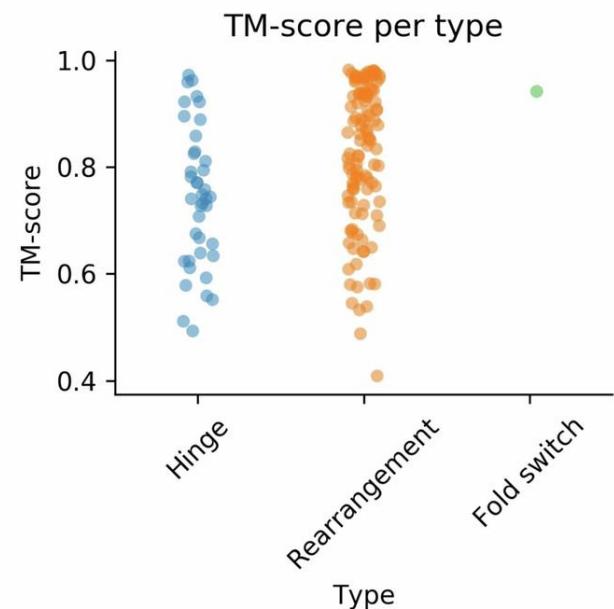
## c



## a

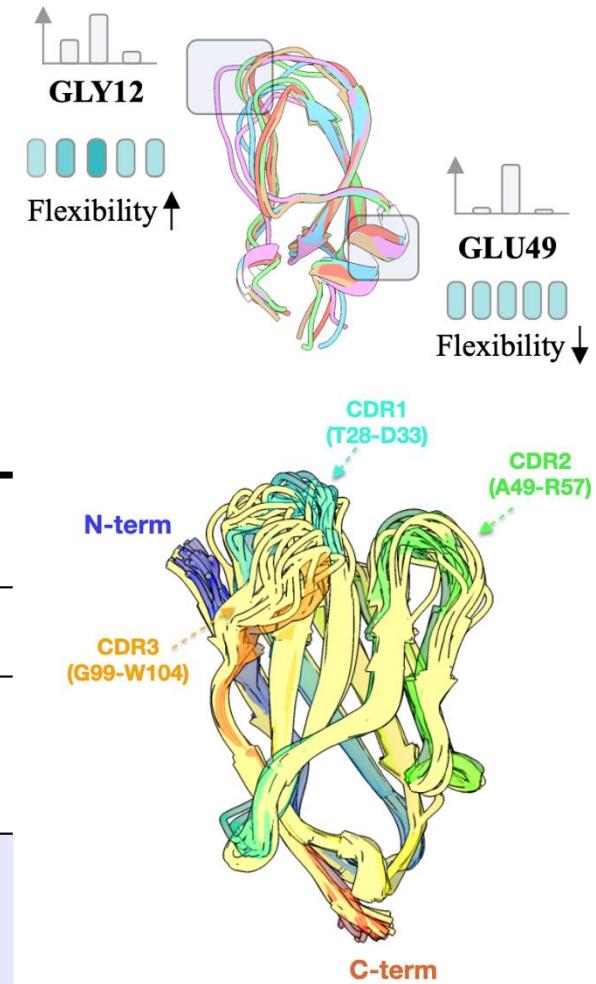


## b



# ESMDiff – Fine-tuned ESM3 for tokenized structure generation

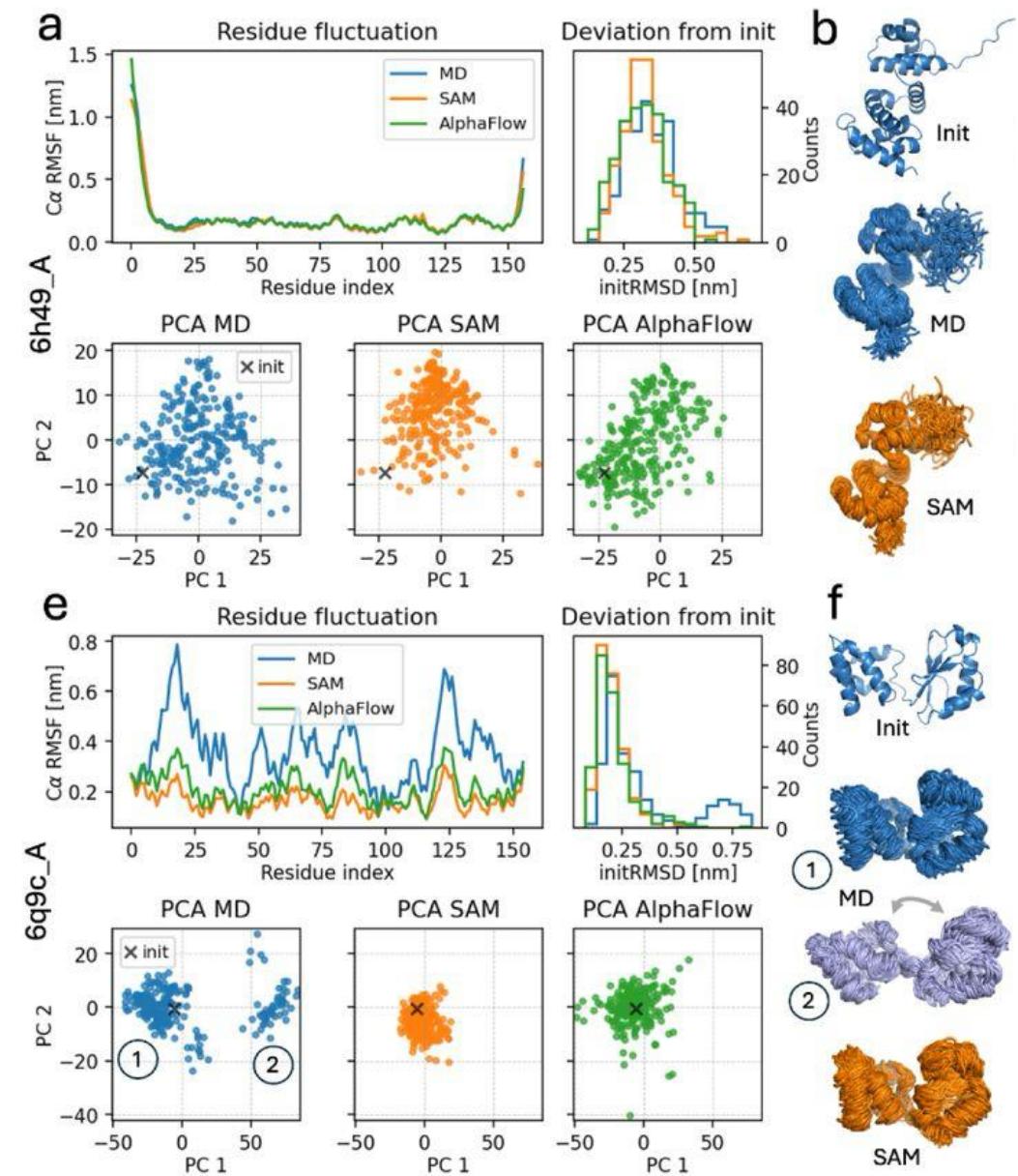
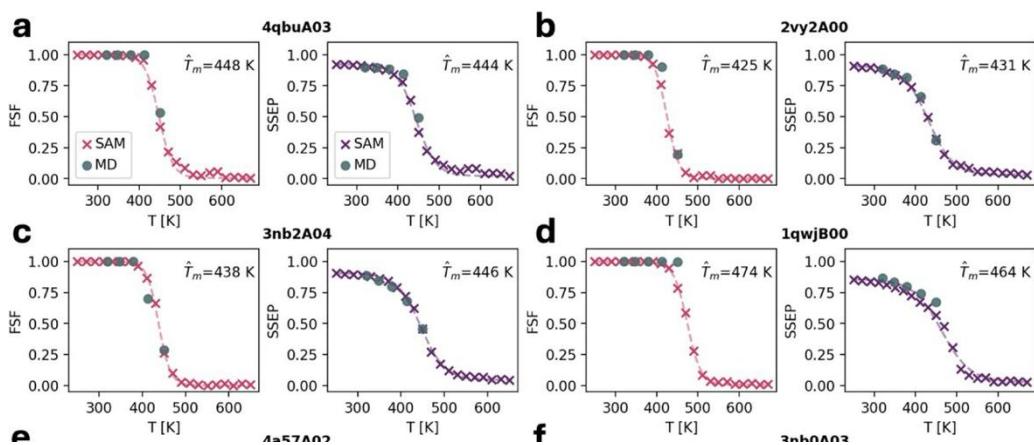
- Relies on structure tokenization, a recent paradigm for encoding structure as a discrete vocabulary via a “codebook” (example: Foldseek)
- Fine-tuned from ESM3, a structure-aware language model
- Promising early results comparable to AlphaFlow



| Method    |                        | Apo/holo              |                                       |                                       | Fold-switch       |                                       |                                       |
|-----------|------------------------|-----------------------|---------------------------------------|---------------------------------------|-------------------|---------------------------------------|---------------------------------------|
|           |                        | ResFlex $r$ (gl.)     | ResFlex $r$ (pt.)                     | TM-ens                                | ResFlex $r$ (gl.) | ResFlex $r$ (pt.)                     | TM-ens                                |
| MSA-based | MSA-Subs.<br>AlphaFlow | 0.398<br><b>0.455</b> | 0.404 / 0.371<br><b>0.527 / 0.527</b> | 0.856 / 0.894<br><b>0.864 / 0.893</b> | 0.350             | 0.320 / 0.303<br><b>0.384 / 0.376</b> | 0.714 / 0.765<br><b>0.730 / 0.788</b> |
| Seq-based | Eigenfold              | 0.126                 | 0.407 / 0.401                         | 0.830 / 0.870                         | 0.225             | 0.279 / 0.255                         | 0.614 / 0.653                         |
|           | Str2Str (PF)           | 0.174                 | 0.326 / 0.307                         | 0.731 / 0.728                         | 0.161             | 0.246 / 0.233                         | 0.615 / 0.644                         |
|           | Str2Str (SDE)          | 0.148                 | 0.349 / 0.340                         | 0.659 / 0.681                         | 0.111             | 0.224 / 0.220                         | 0.521 / 0.545                         |
|           | ESMFlow                | 0.416                 | 0.496 / 0.522                         | 0.856 / 0.893                         | 0.269             | 0.345 / 0.329                         | 0.700 / 0.755                         |
| SLM       | S-T5                   | 0.097                 | 0.144 / 0.166                         | 0.726 / 0.787                         | 0.313             | 0.135 / 0.099                         | 0.437 / 0.392                         |
|           | S-GPT                  | 0.112                 | 0.134 / 0.112                         | 0.571 / 0.562                         | 0.207             | 0.075 / 0.078                         | 0.349 / 0.300                         |
|           | ESM3 (zero shot)       | 0.312                 | 0.473 / 0.466                         | 0.839 / 0.876                         | 0.388             | 0.323 / 0.320                         | 0.627 / 0.717                         |
|           | ESMDiff (ID)           | <b>0.424</b>          | <b>0.502 / 0.517</b>                  | <b>0.851 / 0.883</b>                  | 0.391             | 0.328 / 0.346                         | <b>0.660 / 0.720</b>                  |
|           | ESMDiff (DDPM)         | 0.420                 | 0.489 / 0.515                         | 0.838 / 0.877                         | <b>0.402</b>      | <b>0.341 / 0.288</b>                  | 0.626 / 0.685                         |

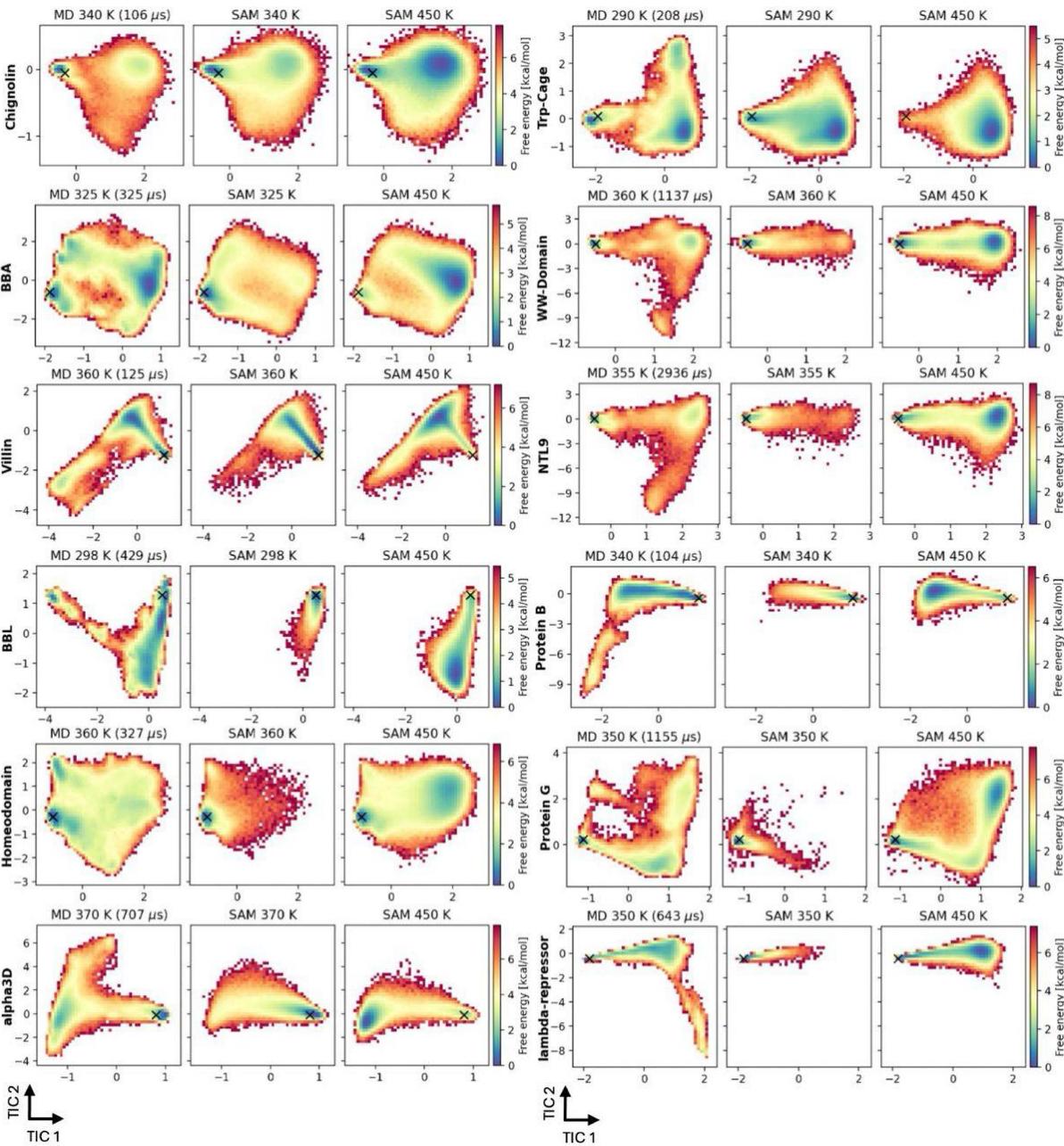
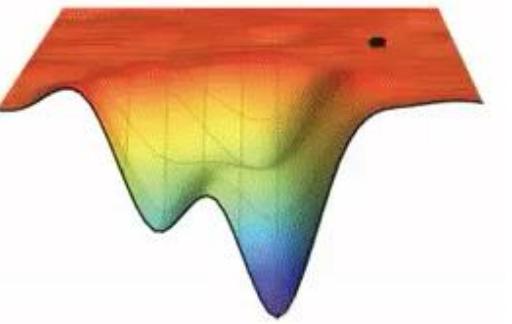
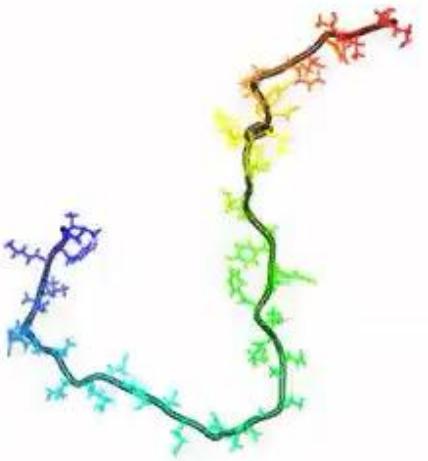
# aSAMt

- Also trained on bulk MD simulations
- Has a tunable “temperature” parameter that predicts unfolded state when raised
- Some evidence that it generalizes dG, and captures differences between closely related proteins
- However, not always to sample distinct, dissimilar conformations



# aSAMt (continued)

- Partial generalization to slow dynamics in fast-folding proteins

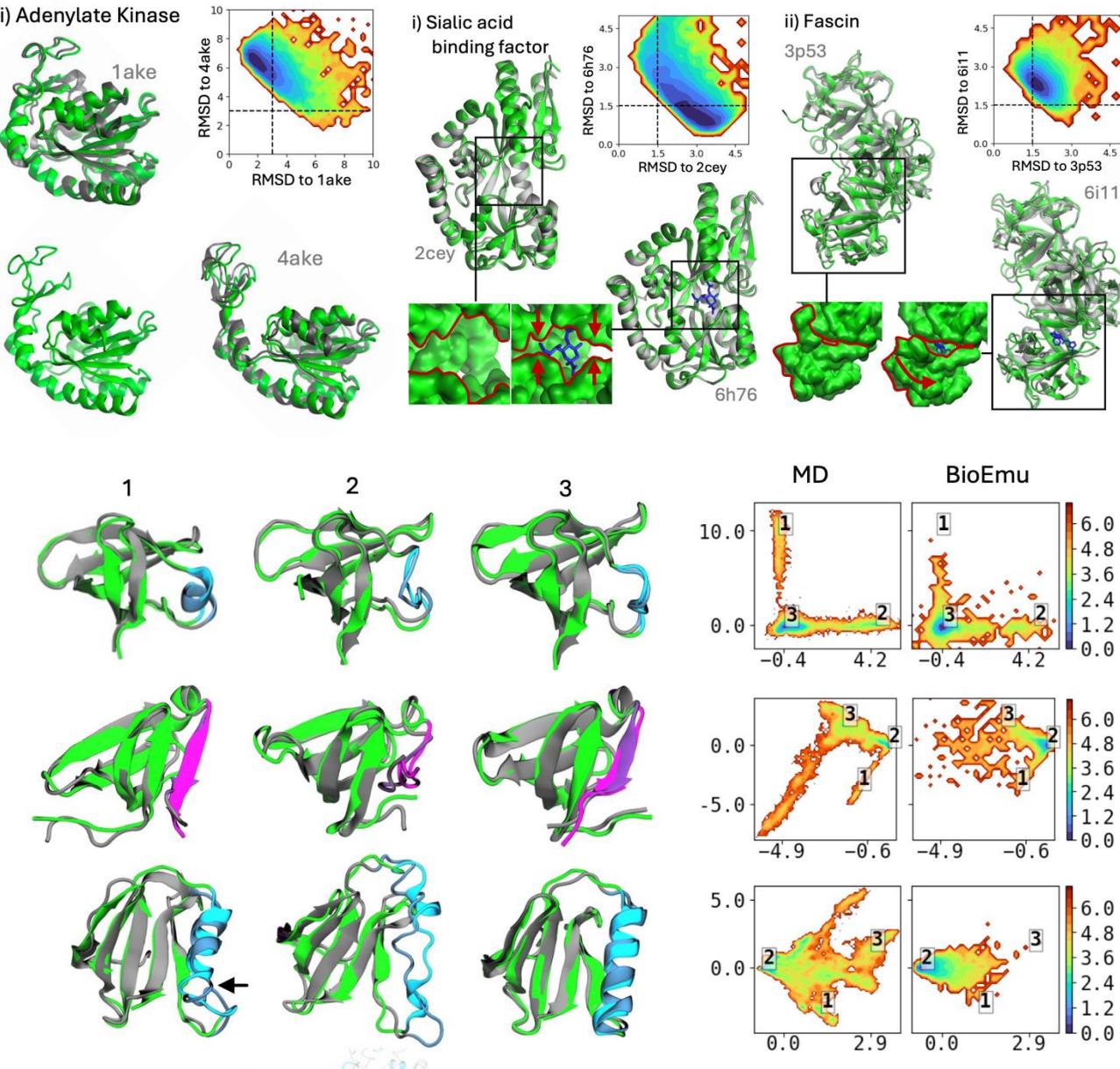


GIF source unknown

Janson et al "Deep generative modeling of temperature-dependent structural ensembles of proteins" bioRxiv 2025

# BioEmu

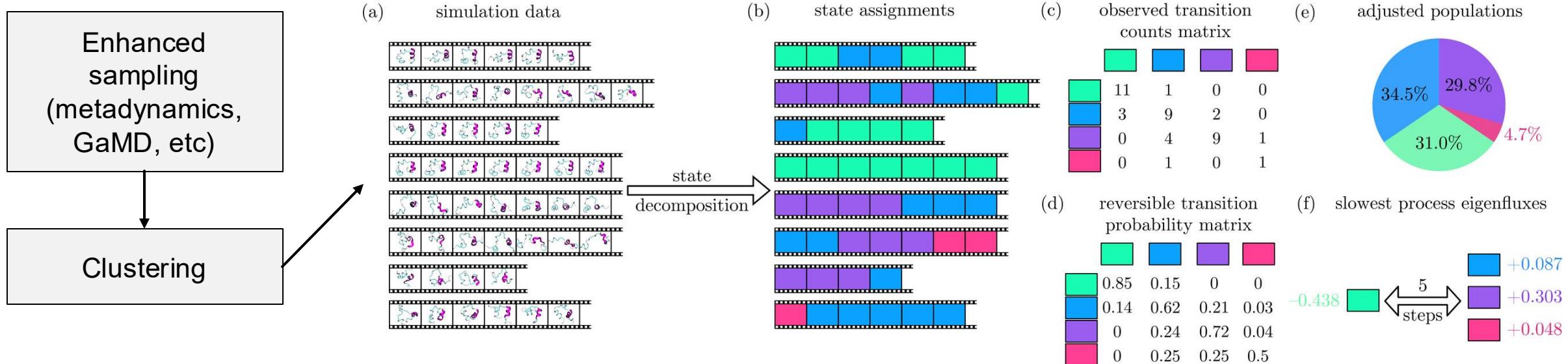
- Among the most promising of the bespoke NNs for conformational modeling
- Independent outputs can sample distinct conformations; but not tested on large proteins
- Is able to model energy landscapes of simple proteins to ~1 kcal/mol @ 300K
- How? Bespoke fine-tuning method trained on 200 milliseconds of MD + 750,000 stability datapoints
- Caveats:
  - Folding/unfolding data used for training happen at different timescales than many conformational interconversion processes



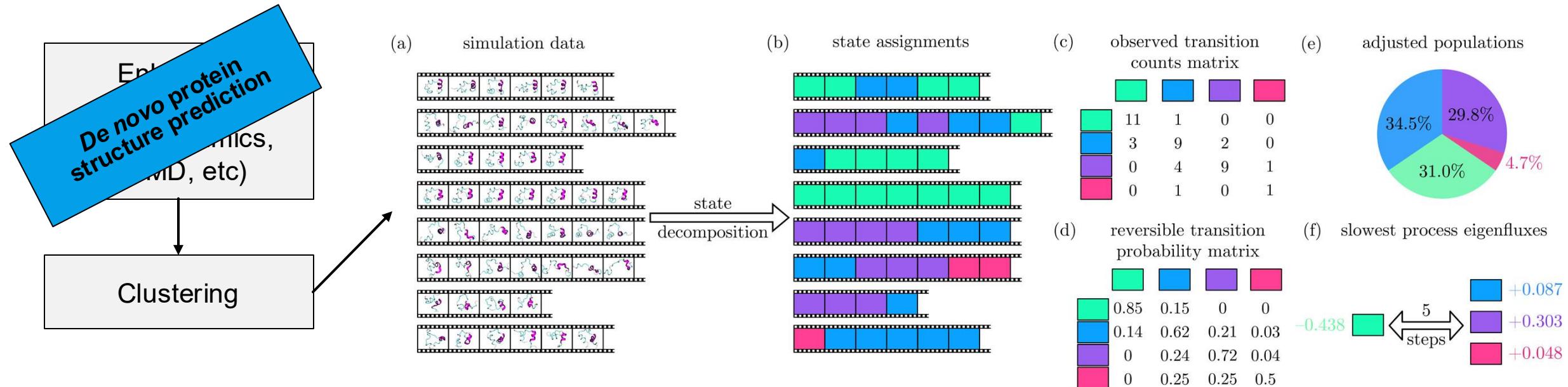
# Why don't protein folding neural networks predict dynamics?

- The problem is difficult:
  - Prediction of protein dynamics is fundamentally different from prediction of protein structure
  - Structure prediction training objective is clear and well-suited to learning from high-quality PDB data (**Bronstein 2024**)
  - Dynamics are less conserved than structure (**Tokuriki 2009**)
- The training data are ill-suited:
  - Dynamics data are ambiguous, sensitive to temperature and (for MD) starting conditions
  - Experimental dynamics data are ambiguous, low-resolution, coarse (SAXS, NMR, FRET)
  - MD simulations are sparse, incomplete, potentially inaccurate
  - Combinations data from different sources introduces noise

# An example MD workflow for free energy calculations

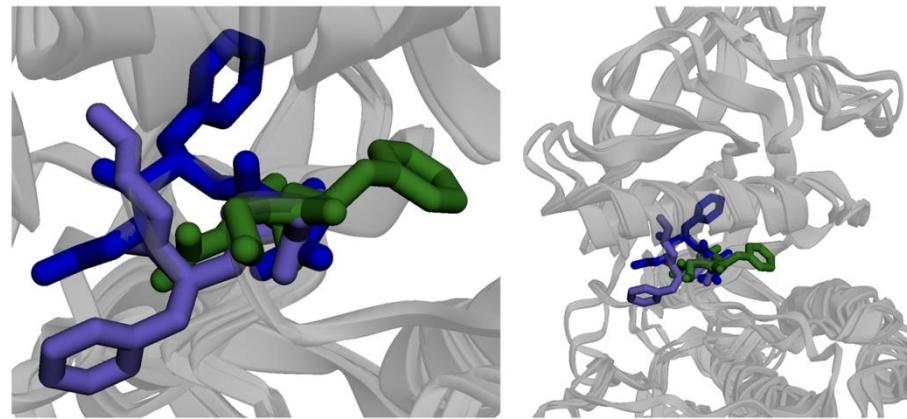


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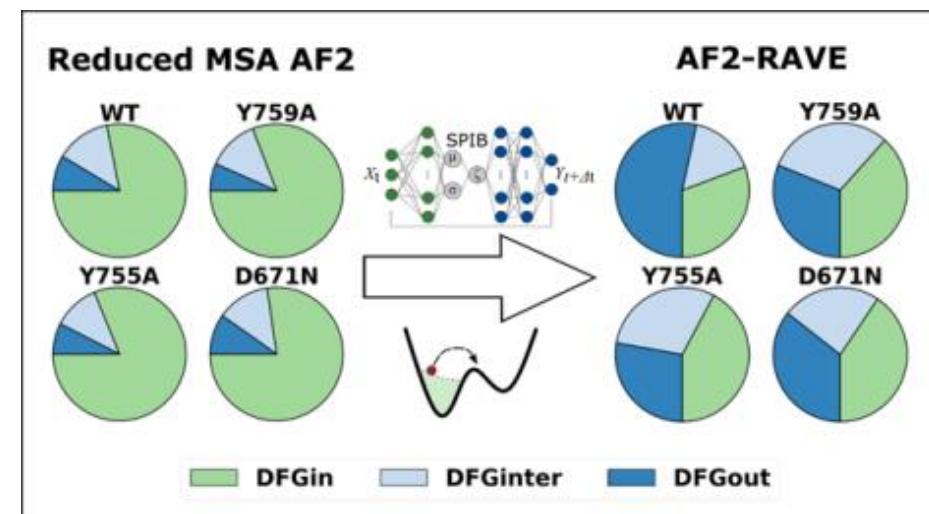


# Replacing enhanced sampling with ensemble prediction

- Several examples involving AlphaFold2 + RAVE ([Vani 2023](#), [Vani 2023a](#))
  - RAVE is a protocol for converting MD trajectories and snapshots into Boltzmann-weighted state assignments
  - Kinases with/without ligand ([Gu 2024](#)), GPCRs ([Vani 2023](#)), SARS-CoV-2 RBD ([Teng 2025](#))
- Collective variables from short MD trajectories of AF2 models match those of much longer simulations (>100  $\mu$ s; [Teng 2025 & Vats 2025](#))
- Can lead to the observation of rare events, such as cryptic pocket opening ([Meller 2023](#), [Vats 2025](#)), conformational interconversion ([Bhakat 2025](#))



DFG-IN DFG-INTER DFG-OUT



Bhakat et al, "Generalizable Protein Dynamics in Serine-Threonine Kinases: Physics is the key" bioRxiv 2025

Gu et al, "Empowering AlphaFold2 for protein conformation selective drug discovery with AlphaFold2-RAVE" eLife 2024

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Teng et al, "AlphaFold2-RAVE: Protein Ensemble Generation with Physics-Based Sampling" ChemRxiv 2025

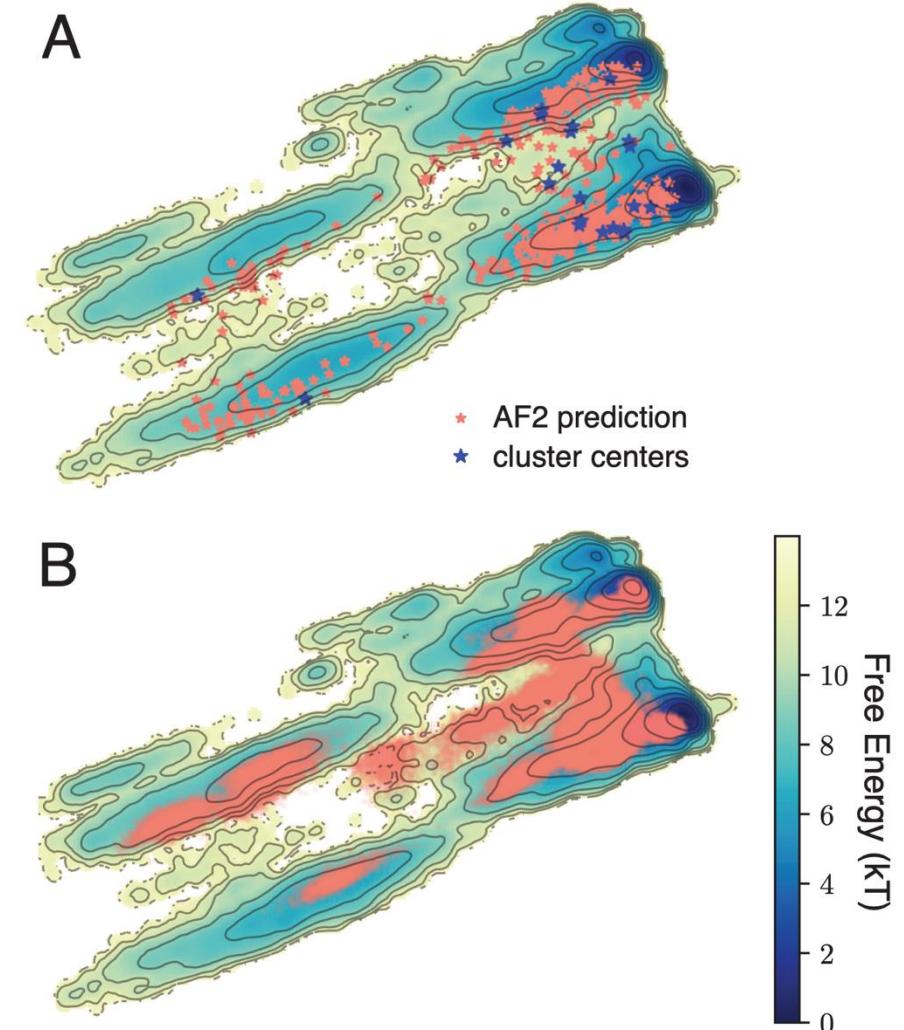
Vani et al, "Exploring kinase Asp-Phe-Gly (DFG) loop conformational stability with AlphaFold2-RAVE" JCIM 2023

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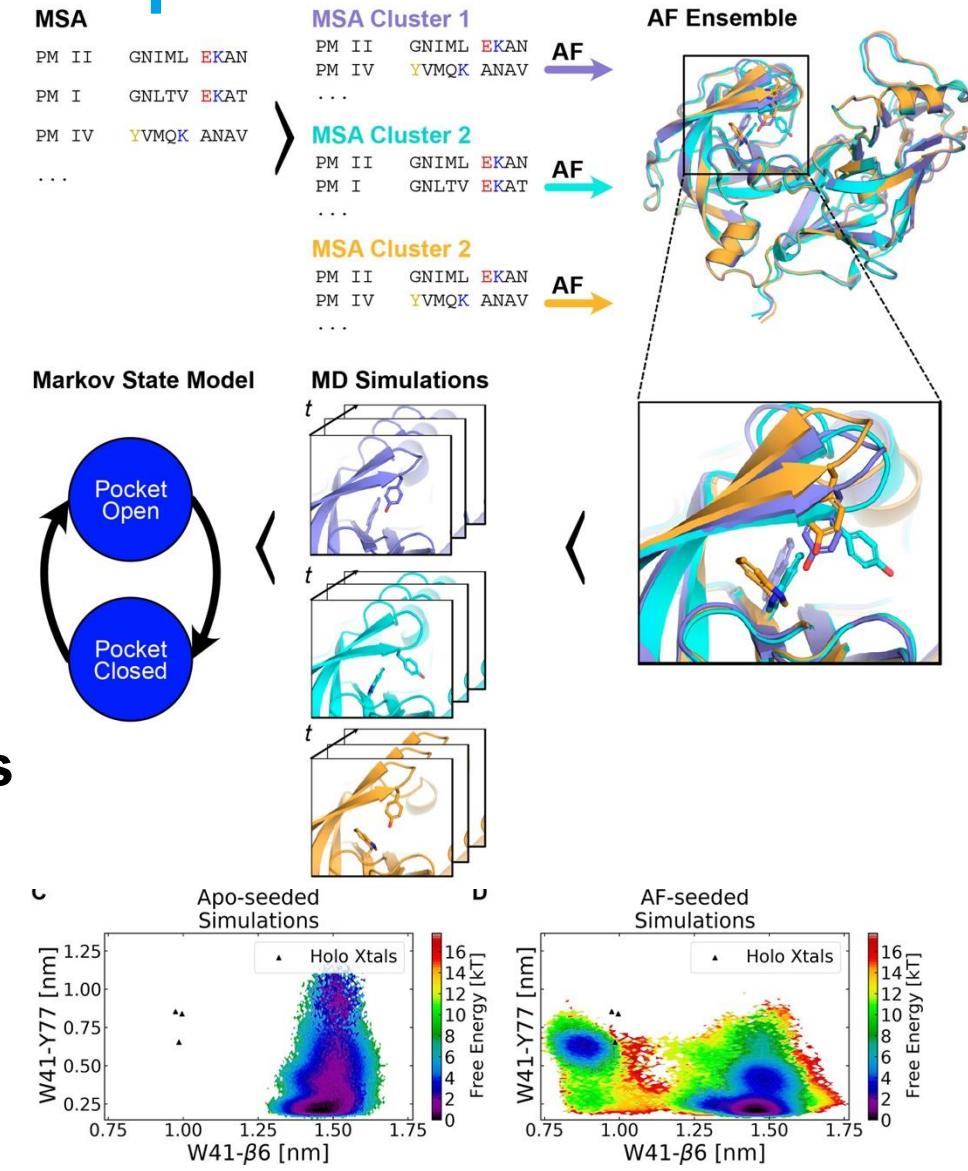
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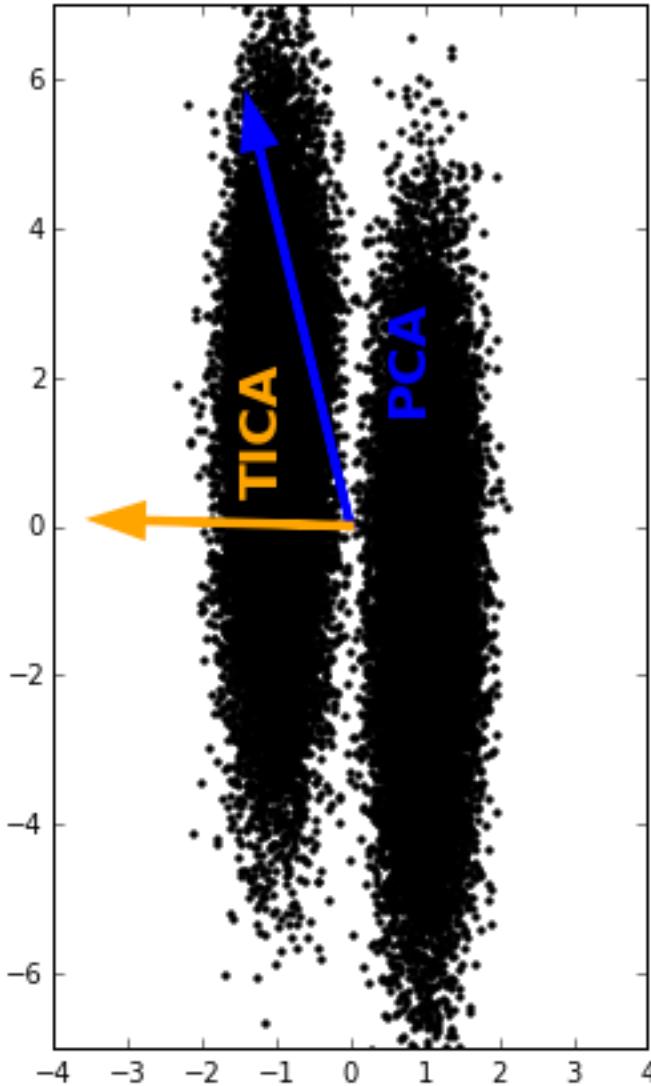
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# Conclusions

- Protein folding neural networks can be re-used for some dynamics prediction problems
- Other problems cannot be addressed by today's suite of tools
- New benchmarks and test cases being released all the time to get a sense of which tools have which strengths
- A separate branch of research is focusing on accelerating MD simulations to take huge time steps (not discussed here)
- **Availability**
  - Most MSA- and template-based approaches available on google colab via [ColabFold](#)
  - Most methods available on GitHub: [AlphaFlow](#), [Rosetta-VAE](#), [AFSample2](#), [Cfold](#), [aSAMt](#), [BioEmu](#); [ESMDiff](#) available on GitHub for non-commercial use
- **Additional resources**
  - “Modeling Boltzmann-weighted structural ensembles of proteins using artificial intelligence–based methods” by Aranganathan et al 2025
  - “Prediction of structural variation” by Kalakoti et al 2025

# SUPPLEMENTAL SLIDES

# What is TICA?



From Frank Noé's blog - [http://docs.markovmodel.org/lecture\\_tica.html](http://docs.markovmodel.org/lecture_tica.html)

