Final Project - EEB590C

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Homework 3

This assignment is due prior to the last day class. You are to self-select and work in groups: 2-3 in a group. For the assignment below submit one R-script. Annotations via comments are highly encouraged. The script should run!

Assignment:

1: Obtain a dataset. This may be one of your own, a dataset from DRYAD, or some other source. Identify hypotheses for this dataset and analyze patterns in the data. You may use any methods learned during the semester, but at least one analysis must come from material learned in weeks 11-13.

USE COMMENTS IN THE R CODE to describe what the patterns you find represent.

```
#load appropriate libraries
library(knitr)
library(RRPP)
library(geomorph)
library(tidyverse)
library(readxl)
library(ade4)
library(vegan)
library(mice)
library(ggplot2)
```

Intro

Epidermal micromorphology in Neotropical bamboo foliage leaves has proven to be a useful tool for differentiating between different species. Certain unusual features have been noted in the epidermal micromorphology of one species of savanna bamboo, Guadua paniculata, including papillae on both leaf surfaces and saddle-shaped silica cells. However, it remains unknown whether the few other species of Guadua primarily found in forests, including G. virgata, have similar micromorphology. Additionally, there are several general trends in Guadua macromorphology, including the G. angustifolia type, which are tall, primarily forest-adapted species, the G. glomerata type, which are scandent species primarily found along rivers, the G. sarcocarpa type, which are similar to G. angustifolia but notable for their tree-killing ability, and species of both savanna and forest bamboos that closely resemble G. paniculata. Here, we analyzed whether micromorphological features such as shape of silica bodies and presence, placement, and shape of stomata and papillae on epidermal cells of foliage leaves were associated with different patterns in macromorphology, habitat type, and country of origin of specimens belonging to a selection of species of Guadua.

TODO : STATE HYPOTHESES

Preprocessing

Read in data

```
#READ IN DATA AS DATAFRAME
mydata<-read_excel(path="data/TransformGuaduaSet.xlsx", col_names = TRUE, na="x")
```

Remove Column V and Y

Remove column V and Y, because there is missing data and we cannot impute the data because it just doesn't make sense to attempt to predict these columns.

V. Adaxial: Frequency of stomates if present on the adaxial surface of foliage leaf blades: 0 = common; 1 = infrequent. Y. Adaxial: Papillae on long cells of the intercostal zone adjacent to the stomates: 0 = not overarching the stomates; 1 = overarching the stomates.

```
to_drop<-c("Adaxial: Papillae on long cells of the intercostal zone adjacent to the stomates:  0 = not
           "Adaxial: Frequency of stomates if present on the adaxial surface of foliage leaf blades:  0

df=data.frame(mydata[,!( names(mydata) %in% to_drop)],
              stringsAsFactors = TRUE)
```

Set columns to factors

Sets all columns to factors and reads them in as a dataframe.

```
df<-data.frame(lapply(df,as.factor))
```

Imputation of missing data

In the last three columns, in some species the papillae of the cells adjacent to the stomata obscured theshape of the subsidiary cells. Therefore, the shapes of these subsidiary cells resulted in missing data that we imputed using the MICE package's logistic regression method.

```
init = mice(df,maxit=0)
meth = init$method
predM=init$predictorMatrix
meth[c(colnames(df[,7:length(df)]))]<-"logreg"
set.seed(183)
imputed<-mice(df,method = meth,
              predictorMatrix = predM,
              m=5,
              rintFlag = FALSE)
imputed<-complete(imputed)
```

To quick visualize this and make sure that there are no NA left we can use sum(is.na()) in sapply().

```
#double check to make sure there are no more NA
check_is_boolean<-sapply(imputed,function(x) sum(is.na(x)))
```

Analysis

Correlation

```
Y<-imputed[,7:ncol(imputed)] # reads in the binary data only
Y<-data.frame(lapply(Y,function(x) as.numeric(levels(x))[x]))
correlation_y<-data.frame(cor(Y)) #calculated the correlation of the Y values
```

PCOA

We get the distance matrix for our binary data using simple matching coefficient. Then we are able to calculate the PCoA on the distance matrix.

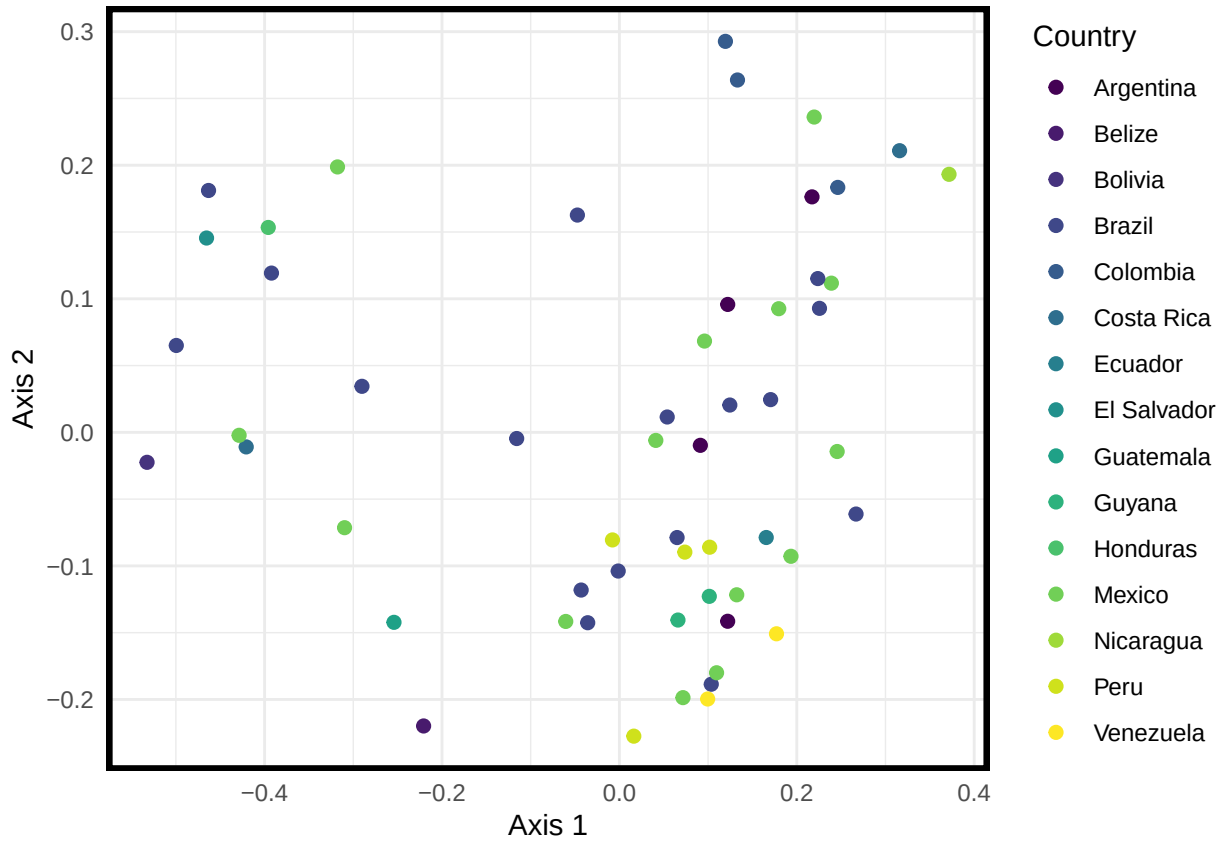
```
#Distance matrix for binary is simple matching coefficient
Y.dist<-dist.binary(Y, method=2, diag = FALSE, upper = FALSE)
Y.dist.matrix<-as.matrix(Y.dist)
PCoA<-cmdscale(Y.dist.matrix, eig= TRUE, x.ret=TRUE, list. = TRUE)    #from vegan
```

PCA ??

```
pca.bamboo<-prcomp(Y.dist.matrix)
```

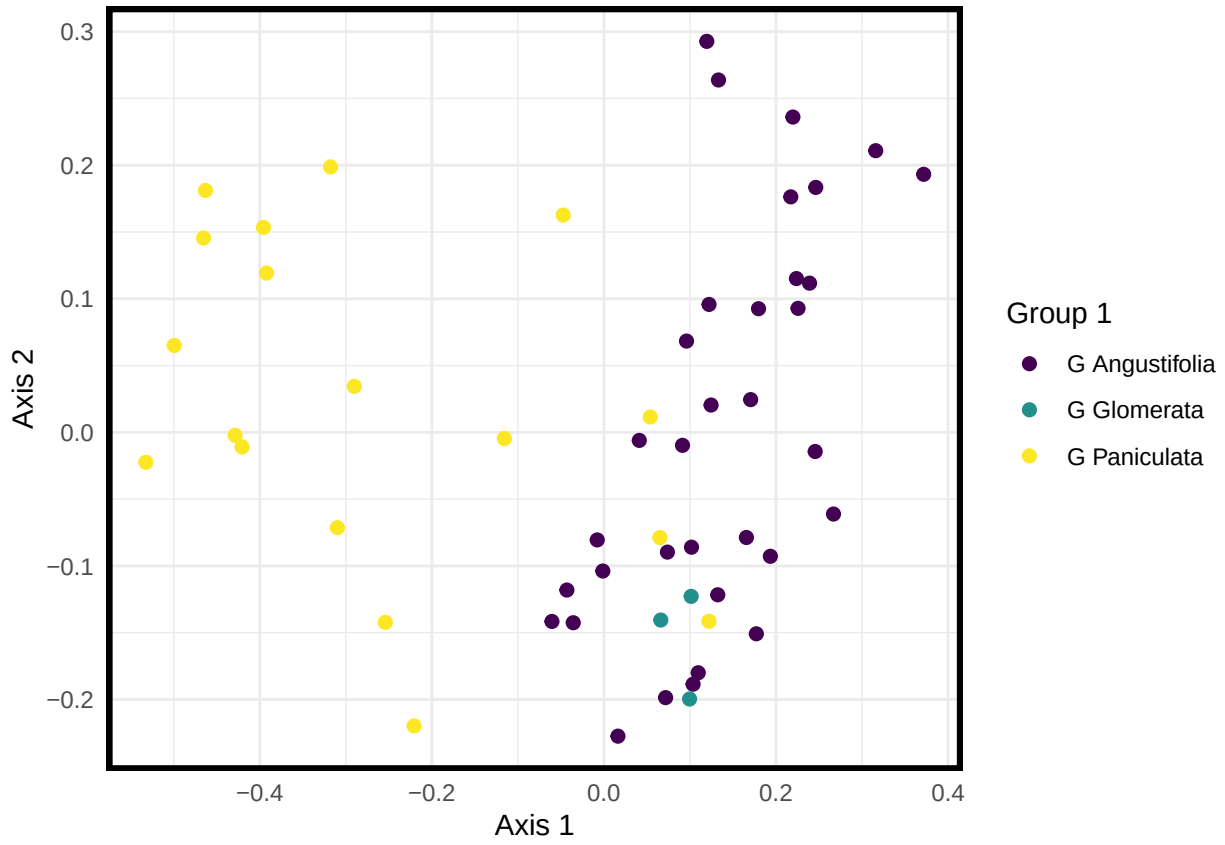
PCoA grouped by Country

```
nice_df <- PCoA$points %>%
  data.frame
nice_df$country <- df$Country
nice_df %>%
  ggplot(aes(X1, X2)) +
  geom_point(aes(color = country), size = 2) +
  scale_color_viridis_d() +
  theme_minimal() +
  theme(
    panel.border = element_rect(size = 2, color = "black", fill = NA)
  ) +
  labs(
    color = "Country",
    x = "Axis 1",
    y = "Axis 2"
  )
```



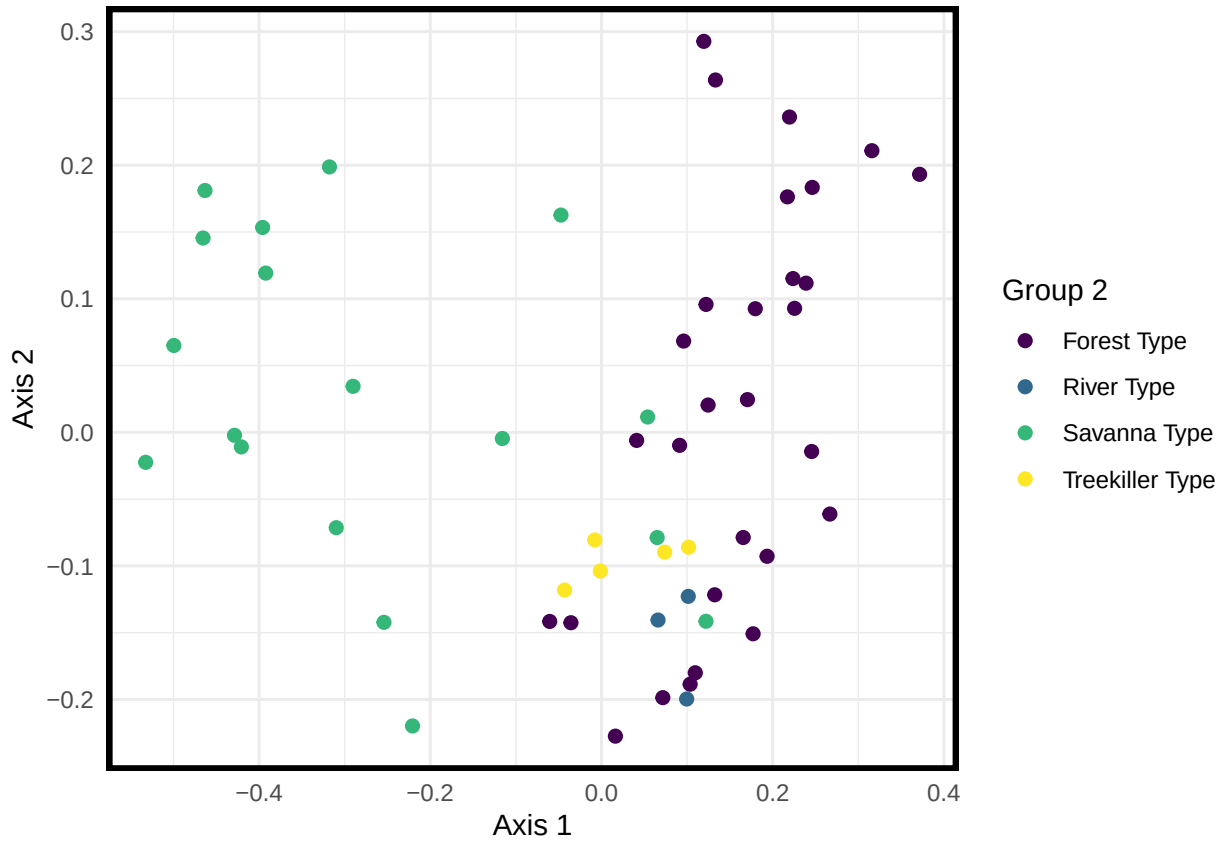
PCoA grouped by Group1

```
nice_df$Group_1<- df$Group_1
nice_df %>%
  mutate(
    Group_1 = str_to_title(str_replace_all(Group_1, "_", " "))
  ) %>%
  ggplot(aes(X1, X2)) +
  geom_point(aes(color = Group_1), size = 2) +
  scale_color_viridis_d() +
  theme_minimal() +
  theme(
    panel.border = element_rect(size = 2, color = "black", fill = NA)
  ) +
  labs(
    color = "Group 1",
    x = "Axis 1",
    y = "Axis 2"
  )
```



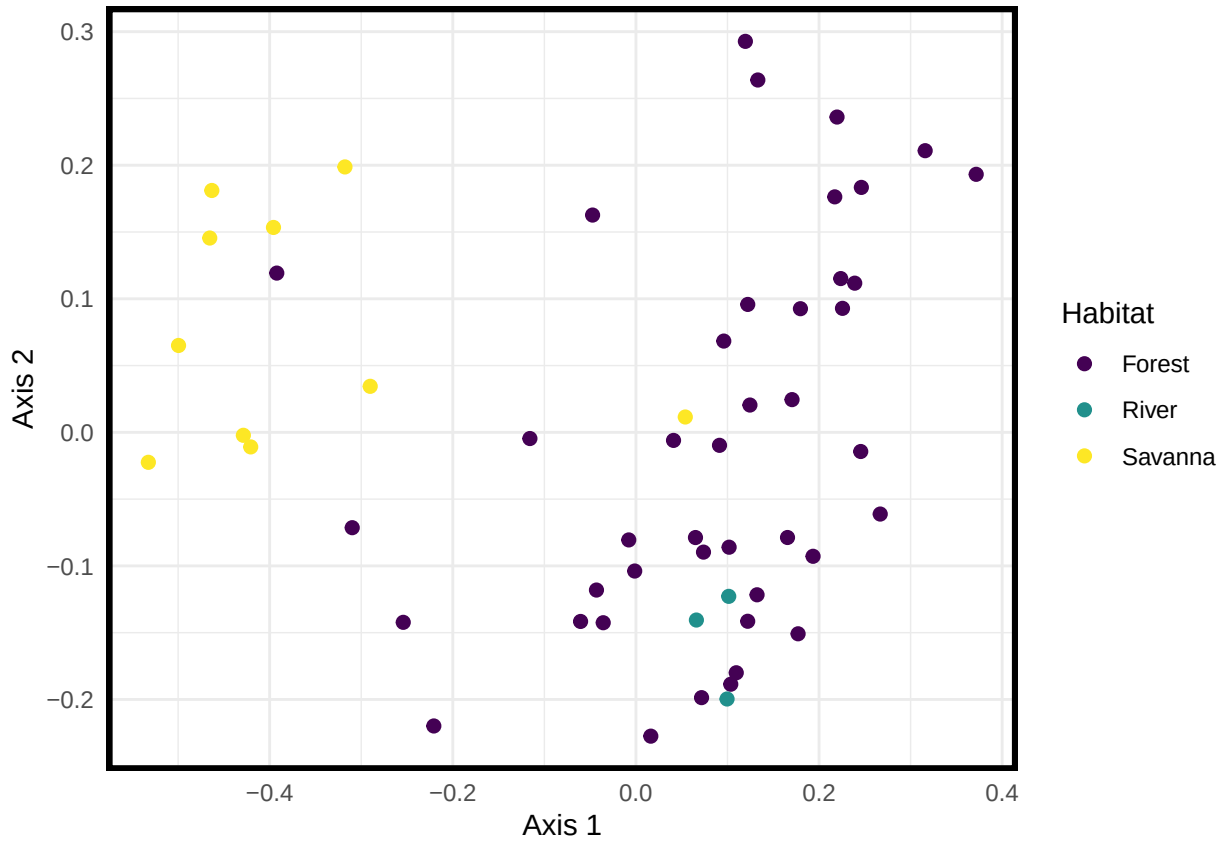
PCoA grouped by Group 2

```
nice_df$Group_2<- df$Group_2
nice_df %>%
  mutate(
    Group_2 = str_to_title(str_replace_all(Group_2, "_", " "))
  ) %>%
  ggplot(aes(X1, X2)) +
  geom_point(aes(color = Group_2), size = 2) +
  scale_color_viridis_d() +
  theme_minimal() +
  theme(
    panel.border = element_rect(size = 2, color = "black", fill = NA)
  ) +
  labs(
    color = "Group 2",
    x = "Axis 1",
    y = "Axis 2"
  )
```



PCoA grouped by Habitat

```
nice_df$Habitat<- df$Habitat
nice_df %>%
  mutate(
    Habitat = str_to_title(str_replace_all(Habitat, "_", " "))
  ) %>%
  ggplot(aes(X1, X2)) +
  geom_point(aes(color = Habitat), size = 2) +
  scale_color_viridis_d() +
  theme_minimal() +
  theme(
    panel.border = element_rect(size = 2, color = "black", fill = NA)
  ) +
  labs(
    color = "Habitat",
    x = "Axis 1",
    y = "Axis 2"
  )
```



Broken Stick Plot

To look into the api of the methods.

cmdscale() : a matrix with k columns whose rows give the coordinates of the points chosen to represent the dissimilarities. aka principal coordinates analysis

```
?cmdscale()
```

```
## starting httpd help server ... done
```

prcomp(): Performs a principal components analysis on the given data matrix and returns the results as an object of class prcomp.

```
?prcomp
```

screeplot(): plots the variances against the number of the principal component

```
?screeplot()
```

```
#TODO

#screeplot(PCoA$points, type = "barplot")

#ERROR
#Not reading in as a principal component analysis
#which principal component analysis to use?
```

To look at what compromises the PC1 variance we look at the rotation. The values that are farther away form 0 are more important for the PC.

```
#pca.bamboo$rotation[,1] #PC1
```

CLUSTERING METHODS?

```
#TODO
```

FACTORIAL MANOVA

```
#Factorial MANOVA via RRPP

# Y<-imputed[,7:ncol(imputed)]
# Y<-data.frame(lapply(Y,function(x) as.numeric(levels(x))[x]))

mydat<-rrpp.data.frame("Y"=Y,
                       "Group_2"= as.factor(imputed$Group_2),
                       "Habitat"=as.factor(imputed$Habitat),
                       "Country" = as.factor(imputed$Country))

model2.rrpp <- lm.rrpp(Y.dist.matrix ~ mydat$Group_2 * mydat$Habitat * mydat$Country,
                       print.progress = FALSE)
```

```
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 180
## Final X columns (rank): 24
## Check coefficients or degrees of freedom in ANOVA to see changes.
```

```
anova(model2.rrpp)
```

```
##
## Analysis of Variance, using Residual Randomization
## Permutation procedure: Randomization of null model residuals
## Number of permutations: 1000
## Estimation method: Ordinary Least Squares
## Sums of Squares and Cross-products: Type I
## Effect sizes (Z) based on F distributions
##
##                               Df      SS      MS     Rsq      F       Z Pr(>F)
## mydat$Group_2                  3  2.9888 0.99626 0.28615 7.2974  6.2057  0.001 **
## mydat$Habitat                  1  0.5926 0.59259 0.05674 4.3406  4.5377  0.001 **
## mydat$Country                 14  1.9162 0.13687 0.18346 1.0026  0.0009  0.495
## mydat$Group_2:mydat$Country    4  0.6422 0.16054 0.06148 1.1760  0.7106  0.247
## mydat$Habitat:mydat$Country    1  0.0729 0.07292 0.00698 0.5341 -0.8921  0.797
## Residuals                     31  4.2322 0.13652 0.40519
## Total                         54 10.4448
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Call: lm.rrpp(f1 = Y.dist.matrix ~ mydat$Group_2 * mydat$Habitat *
##     mydat$Country, print.progress = FALSE)
```

POSTHOC ANALYSIS

Which groups as significantly different from one another?

```
# TODO

# RRPP does have a pairwise posthoc anaylsis to look at
#pairwise comparisons via RRPP
data("Pupfish")
dim(Pupfish$coords)
```

```
## [1]  54 112
```

```
Pupfish$logSize <- log(Pupfish$CS)
Pupfish$Y <- prcomp(Pupfish$coords)$x[, 1:3]

#?pairwise()
#bamboo.groups<-interaction(mydat$Group_2, mydat$Habitat)
#bamboo.groups
#posthoc <- pairwise(fit = model2.rrpp , groups = bamboo.groups, print.progress = FALSE)
#summary(posthoc)

#Dimensionality issues to fix
```

MODEL COMPARISON USING LIKELIHOOD RATIO TEST (LTR)

Setup

```
Y.group2<-lm.rrpp(Y.dist.matrix ~ mydat$Group_2,
                  print.progress=FALSE)
Y.habitat<-lm.rrpp(Y.dist.matrix ~ mydat$Habitat,
                   print.progress=FALSE)
Y.country<-lm.rrpp(Y.dist.matrix ~ mydat$Country,
                   print.progress=FALSE)
Y.group2.habitat<-lm.rrpp(Y.dist.matrix ~ mydat$Group_2 + mydat$Habitat,
                          print.progress=FALSE)
```

```
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 6
## Final X columns (rank): 5
## Check coefficients or degrees of freedom in ANOVA to see changes.
```

```
Y.group2.by.country<-lm.rrpp(Y.dist.matrix ~ mydat$Group_2 + mydat$Country,
                             print.progress=FALSE)
Y.habitat.country<-lm.rrpp(Y.dist.matrix ~ mydat$Habitat + mydat$Country,
                           print.progress=FALSE)
Y.habitat.by.country<-lm.rrpp(Y.dist.matrix ~ mydat$Group_2 * mydat$Habitat,
                              print.progress=FALSE)
```

```
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 12
## Final X columns (rank): 5
## Check coefficients or degrees of freedom in ANOVA to see changes.
```

```
Y.mancova<-lm.rrpp(Y.dist.matrix ~mydat$Group_2 + mydat$Habitat*mydat$Country,
                   print.progress=FALSE)
```

```
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 48
## Final X columns (rank): 21
## Check coefficients or degrees of freedom in ANOVA to see changes.
```

```
Y.mancova_2<-lm.rrpp(Y.dist.matrix ~mydat$Habitat + mydat$Group_2*mydat$Country,
                     print.progress=FALSE)
```

```
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 62
## Final X columns (rank): 23
## Check coefficients or degrees of freedom in ANOVA to see changes.
```

```
Y.full<-lm.rrpp(Y.dist.matrix ~ mydat$Group_2 * mydat$Habitat * mydat$Country,
                print.progress=FALSE)
```

```
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 180
## Final X columns (rank): 24
## Check coefficients or degrees of freedom in ANOVA to see changes.
```

RRPP MODEL COMPARISON

```
#?RRPP::model.comparison()
modelComp1<-model.comparison(Y.full,
                             Y.mancova,
                             Y.mancova_2,
                             Y.habitat.by.country,
                             Y.habitat.country, Y.habitat,
                             Y.group2.by.country,
                             Y.group2.habitat,
                             Y.group2,
                             Y.country,
                             type = "logLik", tol=0.01)
```

```
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 180
## Final X columns (rank): 24
## Check coefficients or degrees of freedom in ANOVA to see changes.
##
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 48
## Final X columns (rank): 21
```

```
## Check coefficients or degrees of freedom in ANOVA to see changes.
##
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 62
## Final X columns (rank): 23
## Check coefficients or degrees of freedom in ANOVA to see changes.
##
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 12
## Final X columns (rank): 5
## Check coefficients or degrees of freedom in ANOVA to see changes.
##
##
## Warning: Because variables in the linear model are redundant,
## the linear model design has been truncated (via QR decomposition).
## Original X columns: 6
## Final X columns (rank): 5
## Check coefficients or degrees of freedom in ANOVA to see changes.
```

```
modelComp1.summ<-as.data.frame(summary(modelComp1))
```

```
##
##
##  Summary statistics for model log-likelihoods
##
## 10 Models compared.
##
##
## mydat$Group_2 + mydat$Habitat + mydat$Country + mydat$Group_2:mydat$Habitat + mydat$Group_2:mydat$Cou
## mydat$Group_2 + mydat$Habitat + mydat$Country + mydat$Habitat:mydat$Country
## mydat$Habitat + mydat$Group_2 + mydat$Country + mydat$Group_2:mydat$Country
## mydat$Group_2 + mydat$Habitat + mydat$Group_2:mydat$Habitat
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##
## mydat$Group_2 + mydat$Habitat + mydat$Country + mydat$Group_2:mydat$Habitat + mydat$Group_2:mydat$Cou
```

```
## mydat$Group_2 + mydat$Habitat + mydat$Country + mydat$Habitat:mydat$Country
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##
## mydat$Group_2 + mydat$Habitat + mydat$Country + mydat$Group_2:mydat$Habitat + mydat$Group_2:mydat$Cou
## mydat$Group_2 + mydat$Habitat + mydat$Country + mydat$Habitat:mydat$Country
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## mydat$Habitat
## mydat$Group_2 + mydat$Country
## mydat$Group_2 + mydat$Habitat
## mydat$Group_2
## mydat$Country
```