

Applied Computational Genomics (PHC 7736)

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Lecture 13: Study Protein from Its Sequence

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Overview

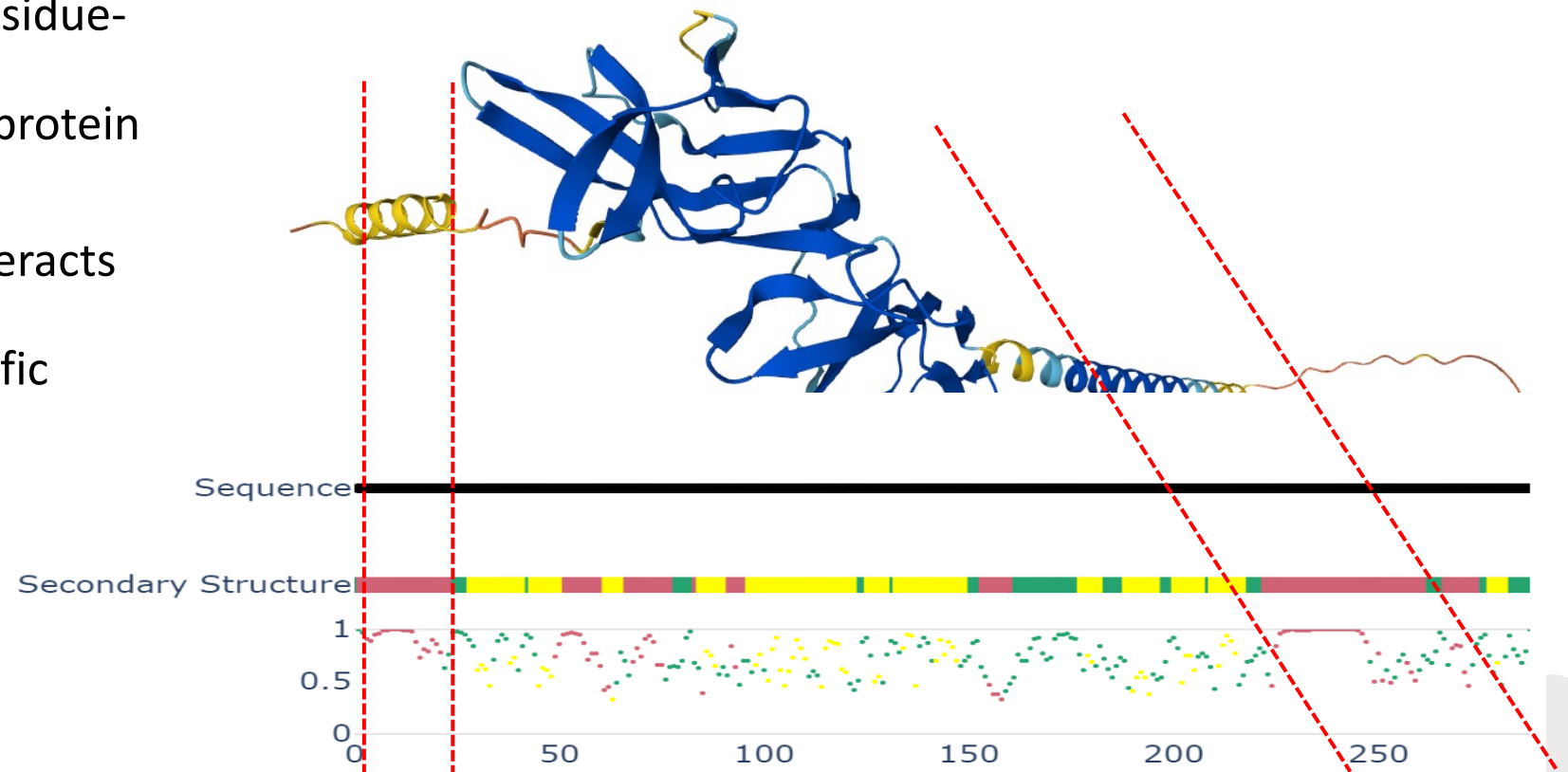
- Introduction to protein solvent accessibility and conservation
- Part 1: Obtain predictive results from ASAquick and MMseqs2
 - In-class work
 - Installation of tools
 - Get the predictive results
 - Short break (~10 mins)
 - Description of the Result format
- Part 2: Analyze the outputs from ASAquick and MMseqs2 to putative annotate protein residue-level solvent accessibility and conservation score
 - In-class work
 - Get the formatted putative annotations
 - Results interpretation

What can be learned from protein sequence

- **Structure**

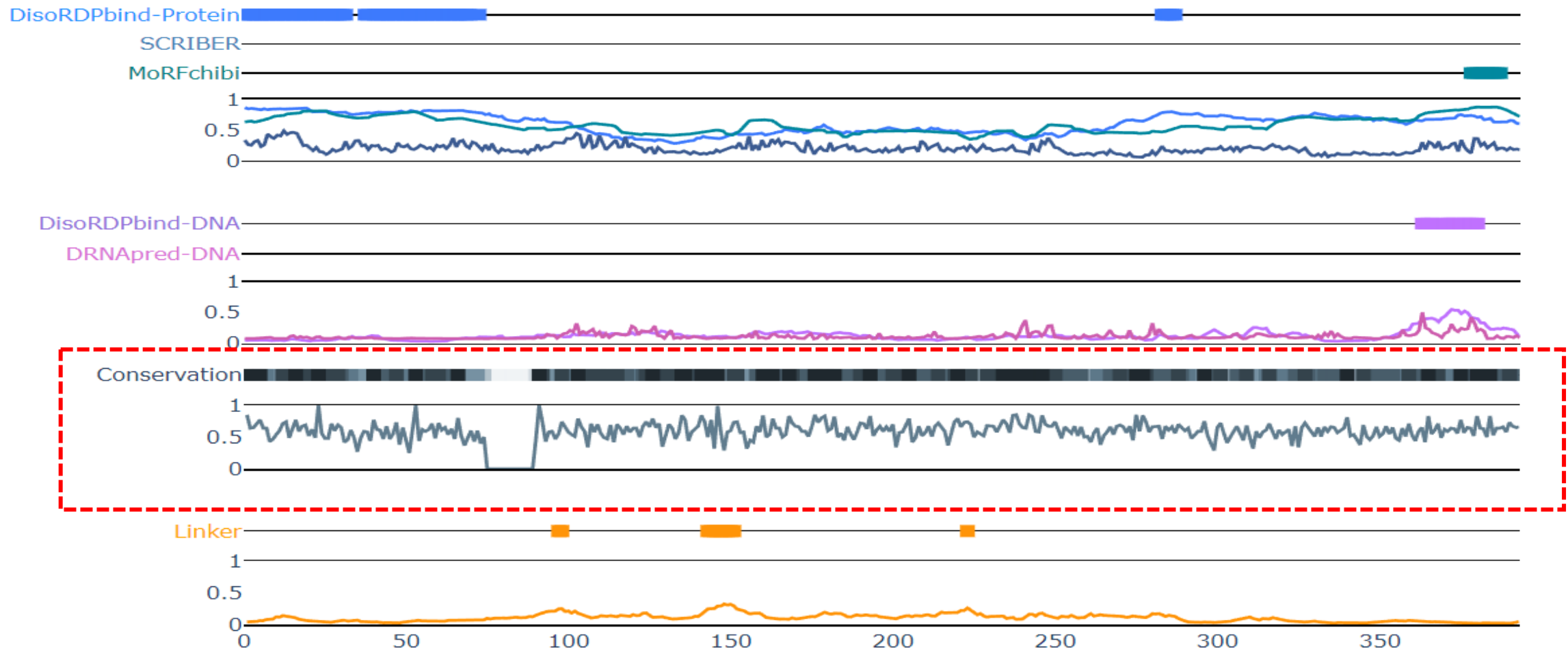
- What can be predicted?
 - 3D structure (AlphaFold, Atom-level)
 - Secondary structure (alpha helix, beta sheet, etc., PSIPRED, residue-level)
 - View the overall fold of the protein
- How can this be used?
 - Understand how protein interacts with other
 - Design drugs targeting specific regions of a protein

- P24071
- Immunoglobulin alpha Fc receptor
- FCAR



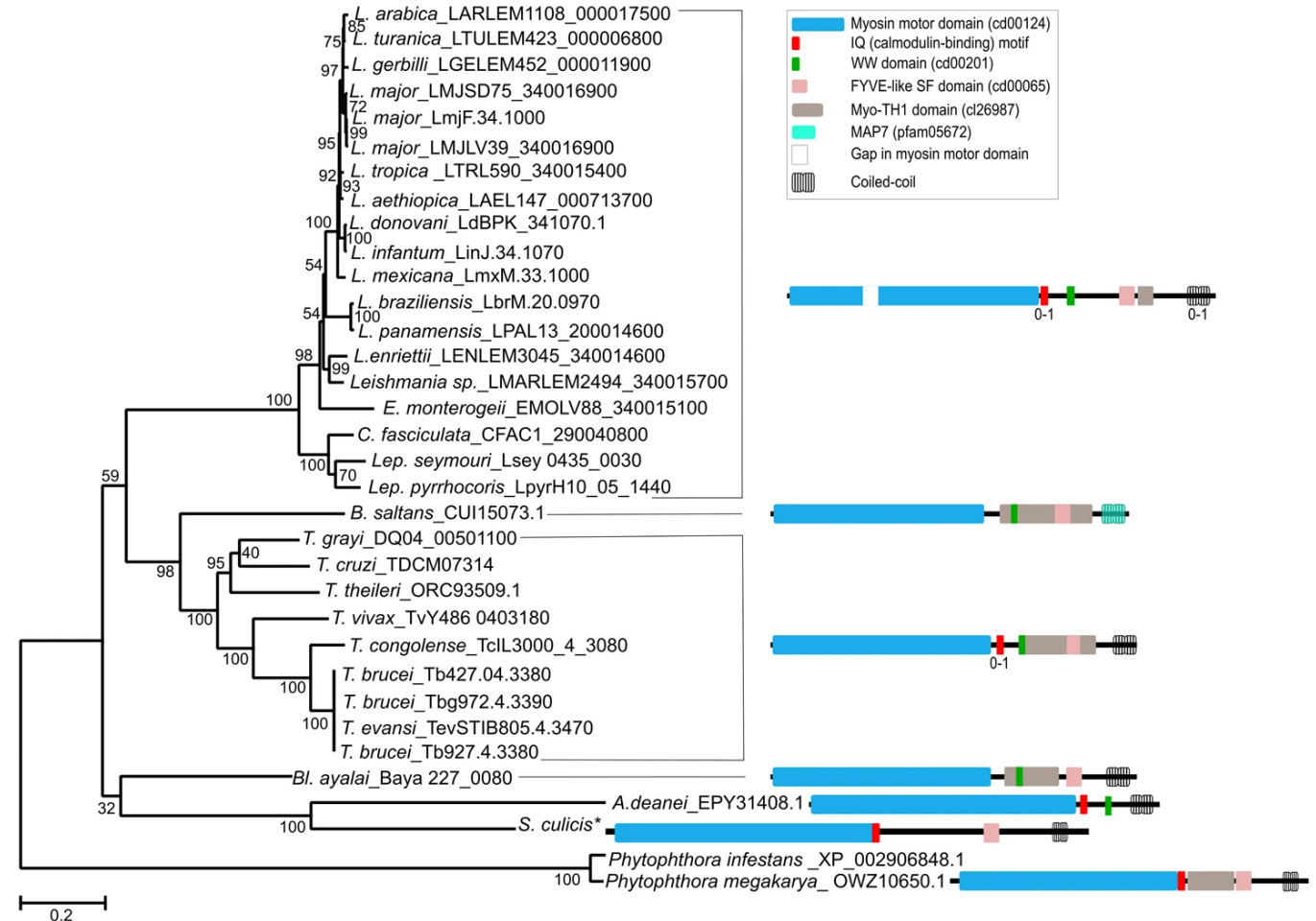
What can be learned from protein sequence (Cont'd)

- Function (P04637 Cellular_tumor_antigen_p53)
 - The highly conserved regions across different species.
 - Identify binding regions. E.g. DNA-binding regions for gene expression regulation.



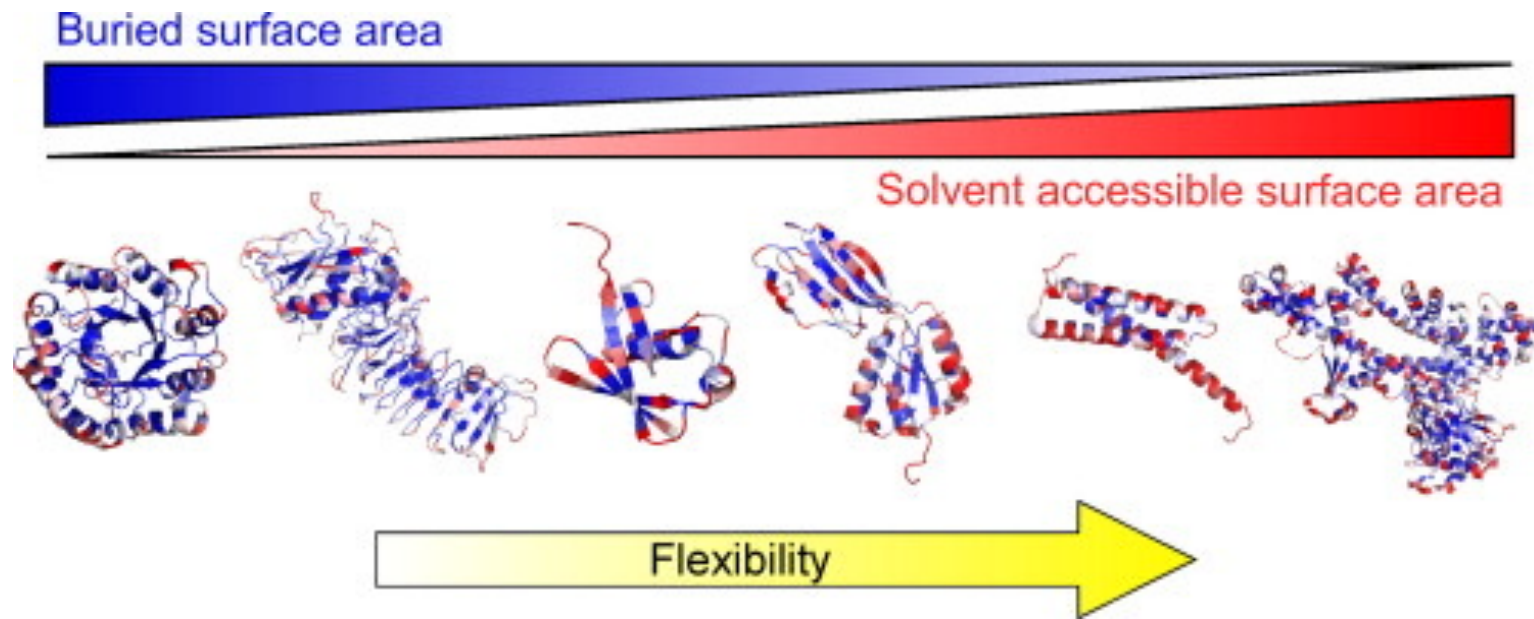
What can be learned from protein sequence (Cont'd)

- Evolution
 - Reveal relationships
 - Similar sequences have a common ancestor
 - Different species comparing may indicate the evolutionary history of those species



Solvent Accessibility

- The propensity of an amino acid that is exposed to the surrounding solvent or buried within the protein core.
- Solvent accessibility can be used to predict the functional and structural properties of a protein.
- ASAquick that are used to annotate protein solvent accessibility
 - Physicochemical properties, 20 parameters related to residue mutation probabilities from BLOSUM62 matrix, the residue length of the protein, its one-residue composition, etc
 - Fast and relatively accurate
 - Outputs include raw solvent accessibility score, relative solvent accessibility, secondary structure, etc.



Protein sequence conservation

- Multiple sequence alignment
- Conservation score
 - Calculated to quantify the degree of sequence conservation at each amino acid residue
- MMseqs2
 - Many to many sequence searching
 - Optimized algorithm that is fast and accurate
 - Outputs: cluster of sequence, sequence-level consensus score, Position-specific scoring matrix

In-class work

- Download installation tutorial
- Download the reference protein sequences and uploaded to the folder where installing MMseqs2

```
$ cd ~/mmseqs2
```

```
$ mkdir dataset
```

```
$ scp ref_protein_seq.fasta.gz bullrocky@sc.rc.usf.edu:/home/b/bullrochky/mmseq2/dataset
```

```
$ gzip -d ref_protein_seq.fasta.gz
```

- Download code run_asaquick.sh, create_mmseqsDB.sh and run_mmseqs.sh upload to each folder
- Run each script

Results

ASAquick

```
[bizhao@sclogin2 asaquick]$ cd GENN+ASAquick/
[bizhao@sclogin2 GENN+ASAquick]$ ls
16VPA.dsspget      asaq.16VPA.dsspget  asaquick  bin    gennstld  install  P01160.fasta  run_asaquick.sh  run.out
AF-P01160-F1-model_v4.pdb  asaq.P01160.fasta  ASAquick  GENN   getpred.sh  LICENSE  README        run.err

[bizhao@sclogin2 GENN+ASAquick]$ cd asaq.P01160.fasta/
[bizhao@sclogin2 asaq.P01160.fasta]$ ls
asa2minmax  asaq.pred  blosnorm  dsspget  genn.gin  physpar  rasaq.pred
```

mmseqs2

```
[bizhao@sclogin2 mmseqs2]$ ls
create_mmseqsDB.sh  dataset  P01160  P01160.fasta  P01160.pssm  profileDB  resultDB  run.err  run_mmseqs.sh  run.out  tmp

[bizhao@sclogin2 mmseqs2]$
```

create the pred_result folder under prot_analysis directory

\$ cd ~/prot_study

\$ mkdir pred_result

\$ cp ./mmseqs2/P01160.pssm ./pred_result

\$ cp ./asaquick/GENN+ASAquick/asaq.P01160.fasta/asaq.pred ./pred_result

Format ASAquick

- The score represents the how much of the surrounding of a residue are occupied by other parts of the chain and how much is accessible to the solvent or to other interactions external to chain.
- To normalize the information, the experimental/theoretical score for each amino acid type are applied. These scores are from ref. Tien MZ. PLOS ONE 8(11):e80635.

```
max_value = {"A":129,"R":274,"N":195,"D":193,"C":167,  
             "E":223,"Q":225,"G":104,"H":224,"I":197,"L":201,"K":197,  
             "M":224,"F":240,"P":159,"S":155,"T":172,"W":285,"Y":263,  
             "V":174}
```

Score = residue["X"]/max_value["X"]

```
14 L 20.4183 2.364  
15 L 14.9352 4.816  
16 A 3.6006 1.2166  
17 F 22.06875 5.468  
18 Q 64.2642 7.556  
19 L 29.106 7.22  
20 L 31.2375 7.274  
21 G 27.378 6.65  
22 Q 77.3682 7.228  
23 T 45.1114 10.62  
24 R 115.3214 8.568  
25 A 41.82 6.664  
26 N 53.41365 10.492  
27 P 56.318 6.342  
28 M 50.7722 6.252
```

Query profile of sequence 0

Pos	Cns	A	C	D	E	F	G	H	I	K	L	M	N	P	Q	R	S	T	V	W	Y
0	M	-1	-1	-1	-1	-3	-1	-1	-1	-1	-3	11	-1	-1	-1	-1	-3	-2	-1	-1	-1
1	S	-1	-1	-1	-1	-3	4	-1	-1	-1	-3	-1	-1	-1	-1	-1	5	-2	-1	-1	-1
2	S	-1	-1	-1	-1	-3	-2	-1	-1	-1	-3	-1	-1	-1	-2	-1	7	-2	-1	-1	-1
3	F	-1	-1	-1	-1	8	-2	-1	-1	-1	-3	-1	-1	-1	-2	-1	-2	-2	-1	-1	-1
4	S	-1	-1	-1	-1	-3	-2	-1	-1	-1	-2	-1	-1	-1	-2	-1	7	-2	-1	-1	-1
5	T	-2	-1	-1	-1	-2	-2	-1	-1	-1	-2	-1	-1	-1	-2	-1	-2	8	-1	-1	-1
6	T	-2	-1	-1	-1	-2	-2	-1	6	-1	-2	-1	-1	-1	-2	-1	-2	6	-1	-1	-1
7	T	-2	-1	-1	-1	-2	-2	-1	-1	-1	-2	-1	-1	-2	-2	-1	-2	8	-1	-1	-1
8	K	-2	-1	-1	-1	-2	-2	-1	-1	6	-2	-2	-1	-2	-2	-1	-2	-2	6	-1	-1
9	S	-2	-1	-1	-1	-2	3	-1	-1	-1	-2	-2	-1	-2	-2	-1	6	-2	-2	-1	-1
10	F	-1	-1	-1	-1	8	-2	-1	-1	-1	-2	-2	-2	-2	-1	-1	-2	-2	-2	-1	-1
11	L	-2	-1	-1	-1	5	-2	-1	-1	-1	4	-2	-2	-1	-1	-1	-2	-2	-2	-1	-1
12	L	-2	-1	-1	-1	-2	-2	-1	-1	-1	6	-2	-2	-1	-1	-1	-2	-2	-2	-1	-1
13	F	-2	-1	-1	-1	7	-2	-1	-1	-1	2	-2	-2	-1	-1	-1	-2	-2	-2	-1	-1
14	L	-2	-1	-1	-1	-2	-2	-1	-1	-1	6	-2	-2	-1	-1	-1	-2	-2	-2	-1	-1
15	A	7	-1	-1	-1	-2	-2	-1	-1	-1	-2	-2	-2	-1	-1	-1	-2	-2	-2	-1	-1
16	F	-2	-1	-1	-1	7	-2	-1	-1	-1	-2	-2	-2	-1	-1	-1	-2	-2	3	-1	-1

Format
MMseqs2

- Position-Specific Scoring Matrix (PSSM)
- Reflect the probability of observing each amino acid at each position
- The conservation score for each position is calculated by comparing with background frequency of that amino acid in protein database

How the conservation score calculated?

```
#the aa_index used to calculate PSSM and obtain the position specific conservation
```

```
blosum62_bg = { 'A' : 7.4,  
                'R' : 5.2,  
                'N' : 4.5,  
                'D' : 5.3,  
                'C' : 2.5,  
                'Q' : 3.4,  
                'E' : 5.4,  
                'G' : 7.4,  
                'H' : 2.6,  
                'I' : 6.8,  
                'L' : 9.9,  
                'K' : 5.8,  
                'M' : 2.5,  
                'F' : 4.7,  
                'P' : 3.9,  
                'S' : 5.7,  
                'T' : 5.1,  
                'W' : 1.3,  
                'Y' : 3.2,  
                'V' : 7.3 }
```

```
bbg_AAs = list(blosum62_bg.keys())
```

```
tot = sum(blosum62_bg.values())
```

```
blosum62_bg = np.array([ blosum62_bg[k] / tot for k in blosum62_bg])
```

```
#function to calculate PSSM
```

```
def calc_pssm_freq_df(pssm_df):
```

```
    df = pssm_df[bbg_AAs]
```

```
    freq = np.exp(df) * blosum62_bg
```

```
    #freq_sum = np.sum(freq,axis=1)
```

```
    return np.array(freq)/np.sum(freq, axis=1)[:,None]
```

```
def calc_relative_entropy(pssm_df):
```

```
    df = calc_pssm_freq_df(pssm_df)
```

```
    cons = np.sum(df*np.log(df/blosum62_bg), axis=1)
```

```
    return cons
```

```
def format_mmseq(filename):
```

```
    pssm_df = pd.read_csv(filename, sep=" ", skiprows=1)
```

```
    print (len(pssm_df))
```

```
    res = calc_relative_entropy(pssm_df)
```

In-class work

- Download python script and shell file, uploaded to sc cluster
- Run the shell file and obtain the result.
- Download and visualize the results.

Results

