

## CONTRIBUTION

We present a modeling framework aimed at studying the contributions of genes and the environment to phenotypic variation in high dimensions. Specifically, we are interested in leveraging high-dimensional phenotypes to study the combined effects of multiple factors on traits like yield in crops, fitness in natural populations, or health in patients. Our approach combines two well established principles:

- Biological systems tend to be modular in organization.
- Complex experimental or breeding designs and observational studies of natural populations with structure require flexible hierarchical models.

Building on our earlier genetic sparse factor model [3], we extend the specification to allow a full linear mixed effect model to capture complex experimental designs. Using an efficient Bayesian MCMC algorithm, the model scales well both computationally and numerically to dimensions relevant to modern phenotype-centric datasets and generates descriptions of modules that are interpretable with respect to the environmental or genetic factors.

## MOTIVATION

High-dimensional phenotypes (ex RNAseq, metabolomics, hyperspectral imaging) promise to provide an unprecedented window into the complex functions of biological organisms. Settings we have in mind include:

- A set of genetically related individuals assigned to 2+ treatments to assess gene-environment interactions.
- Multi-factor experiments with split-plot or repeated measures designs that are common in agriculture settings.
- Case-control studies with individuals of varying ancestry.

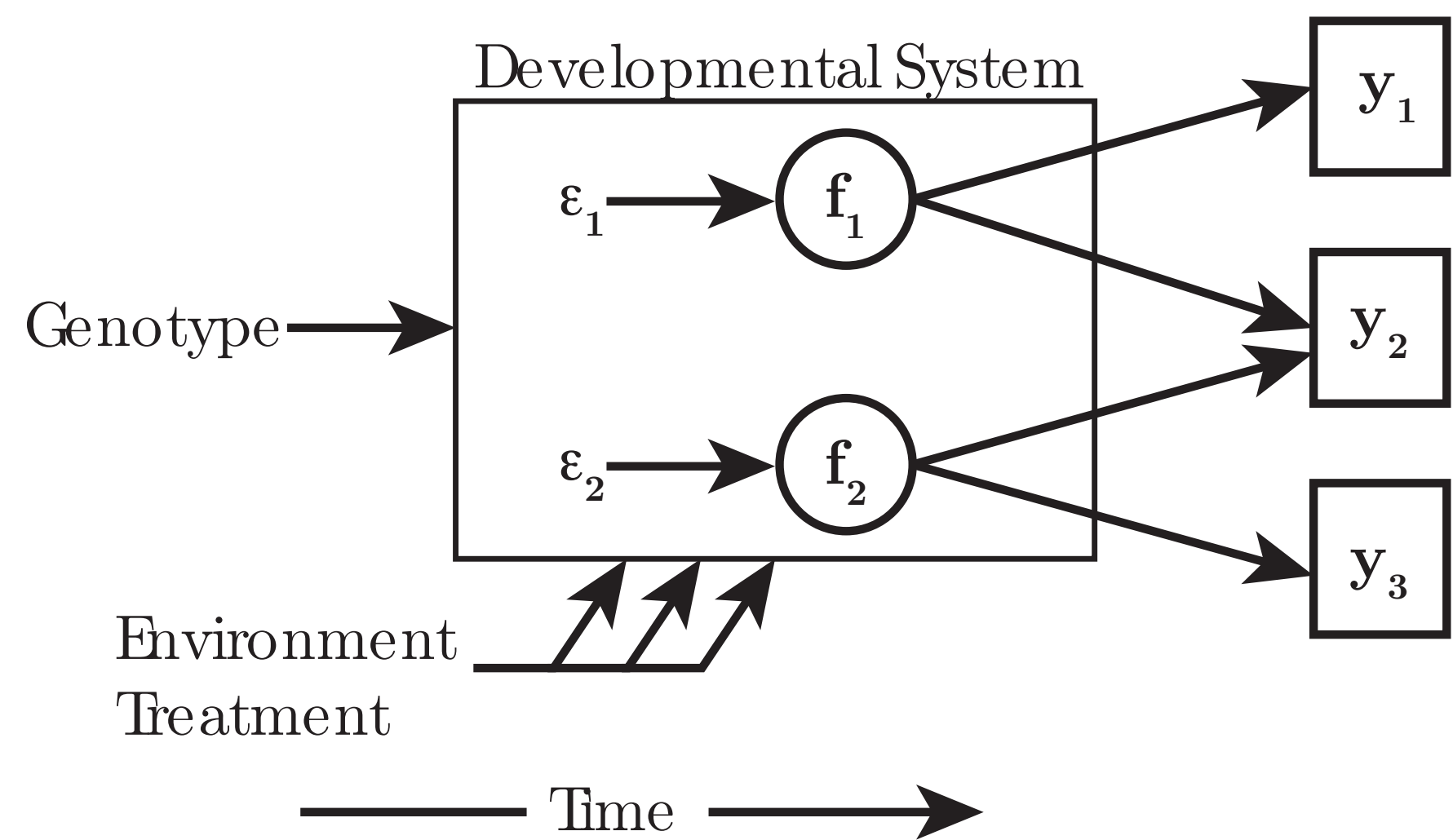
In each case the experimental goals are to identify sets of traits associated with each experimental factor and to understand the biological bases of these effects.

## APPROACH

We view the high-dimensional phenotype as a readout of a modular and time-ordered developmental process (Figure 1). This model has the following implications:

- Experimental factors (genotypes, treatments) cause perturbations to the modules of the developmental system.
- The traits we observe are independent conditional on the state of the underlying developmental system.
- The same module might explain responses to multiple experimental factors (Figure 2).

We therefore focus on identifying the most important modules, which we assume will account for the majority of the effect of the experimental factors on the downstream high-dimensional phenotype.

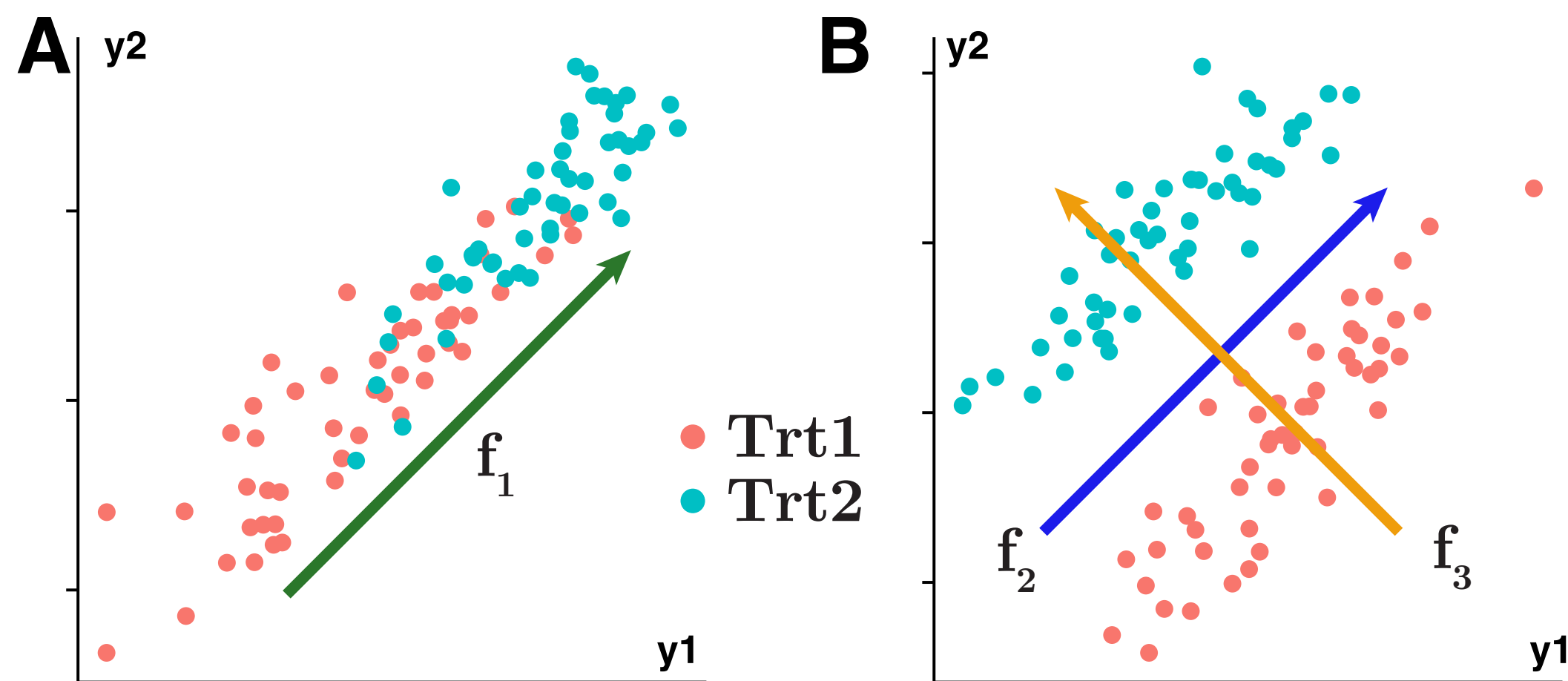


**Figure 1.** We assume that the influence of genotype and treatment on the observed traits  $y_j$  is mediated through developmental modules  $f_i$ .

## ANALYTICAL CHALLENGES

In these examples, the analytical challenges are a combination of those that affect univariate quantitative genetic analyses and those that affect the study of high dimensional data:

- Individuals within (and across) treatments are correlated due to shared genetic history.
- Interactions (ex. Genotype x Treatment) greatly increase the number of parameters for each trait.
- Traits are correlated and may be functionally related
- The number of traits (and between-trait covariances) may be larger than the number of individuals ( $n \ll p$ )



**Figure 2.** When treatment and residual correlations align (as in **A** but not **B**) the same factor can explain variation in both traits.

## MODEL SPECIFICATION

### Sparse factor model

Motivated by the conceptual model in Figure 1, we model the  $n \times p$  phenotype matrix  $\mathbf{Y}$  with a sparse factor model, with  $k$  factors representing latent developmental modules:

$$\mathbf{Y} = \mathbf{f}_1 \boldsymbol{\lambda}'_1 + \mathbf{f}_k \boldsymbol{\lambda}'_k + \dots \mathbf{f}_k \boldsymbol{\lambda}'_k + \mathbf{E} \quad (1)$$

Conditional on the factors, the residuals (rows of  $\mathbf{E}$ ) are independent and assigned Gaussian priors. Sparsity in the factor loadings  $\boldsymbol{\lambda}_l$  is induced with an “infinite factor”[1] prior structure, using local/global shrinkage with increasing shrinkage on higher-indexed factors. This induces a ranking of the factors and reduces the burden of pre-selecting the number of factors.

$$\begin{aligned} \lambda_{jl} &\sim N(0, \tau_k^{-1} \psi_{jl}^{-1}) & \psi_{jl} &\sim \text{Ga}(\nu/2, \nu/2) \\ \delta_1 &\sim \text{Ga}(\alpha_0, \beta_0) & \delta_l &\sim \text{Ga}(\alpha_1, 1) & \pi_l &= \prod_{i=1}^k \delta_i \end{aligned} \quad (2)$$

### Mixed Effect Model (MEM)

Vectors  $\mathbf{f}_l$  represent trait values across the  $n$  individuals for each of the  $k$  latent modules. We assume that each module is independent and model its variation with a linear mixed effect model:

$$\begin{aligned} \mathbf{f}_l &= \mathbf{X} \boldsymbol{\beta}_l + \mathbf{Z}_1 \mathbf{a}_{1l} + \mathbf{Z}_2 \mathbf{a}_{2l} + \boldsymbol{\epsilon}_l & (3) \\ \boldsymbol{\beta}_l &\sim N_b(\mathbf{0}, \sigma_{b_l}^2 (\mathbf{X}'\mathbf{X})^{-1}) & \text{Treatment effect} \\ \mathbf{a}_{1l} &\sim N_r(\mathbf{0}, \sigma_{a_{1l}}^2 \mathbf{A}), \mathbf{a}_{2l} \sim N_r(\mathbf{0}, \sigma_{a_{2l}}^2 \mathbf{A}) & \text{G and GxT effects} \\ \boldsymbol{\epsilon}_l &\sim N_r(\mathbf{0}, \sigma_{e_l}^2 \mathbf{I}) & \text{Residual variation} \\ \sigma_{b_l}^2 + \sigma_{a_{1l}}^2 + \sigma_{a_{2l}}^2 + \sigma_{e_l}^2 &= 1 & (4) \end{aligned}$$

## Summary

The key features of this specification are:

- Specifying the MEM for  $\mathbf{f}_l$  instead of  $\mathbf{y}_j$  is a massive reduction of complexity because the number of traits is small  $k \ll p$  and the latent traits are assumed uncorrelated.
- While factors with shared genetic, treatment and residual components are preferred (Figure 2), purely residual factors are allowed.
- Given a small  $k$ , not all variation may be accounted for by the latent factors. We model each trait’s residual vector (row of  $\mathbf{E}$ ) with a parallel MEM, and place an additional prior  $\pi_j$  on the proportion of variation in that trait accounted for by the  $k$  factors.

## Implementation

We use a Gibbs sampler implemented in R/Rcpp to estimate the posterior distribution of all model parameters and also coded the model in Stan[2].

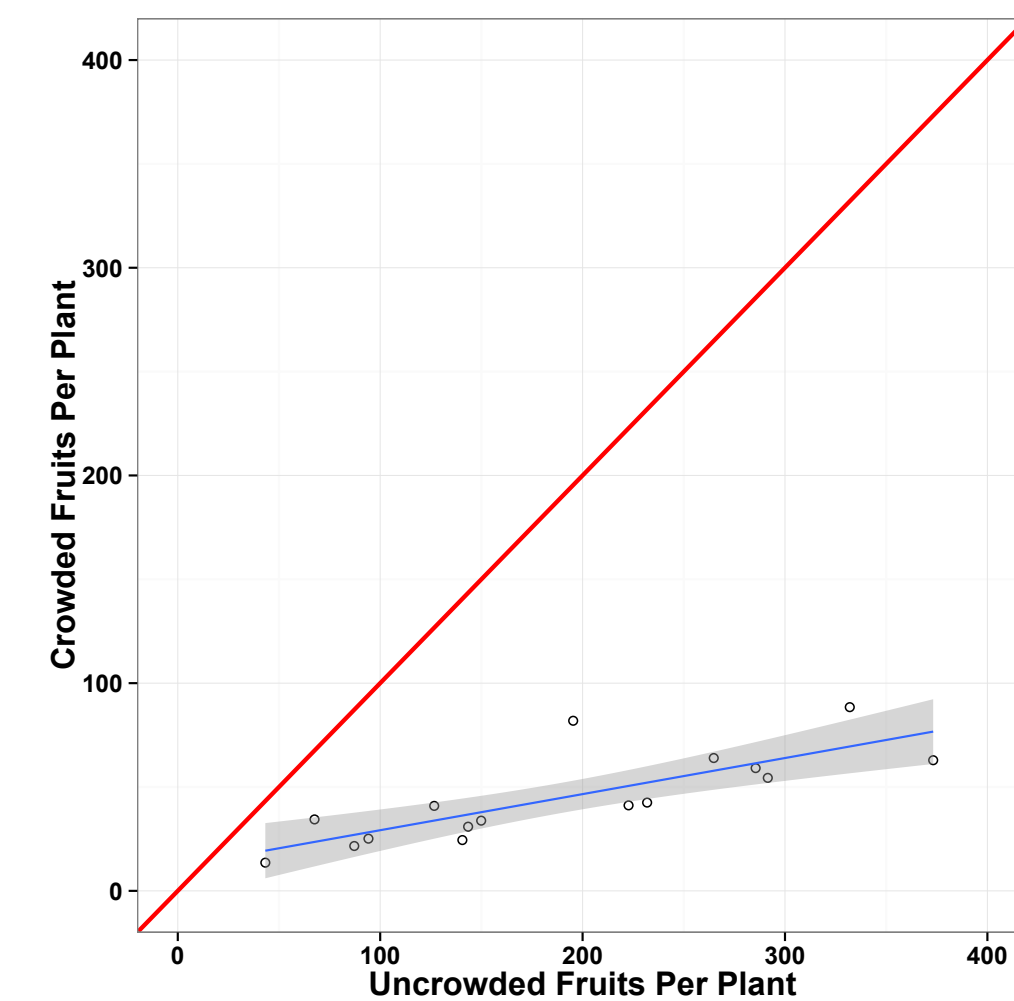
## ACKNOWLEDGEMENTS

We would like to thank Upendra K Devisetty and Julin Maloof for sharing the gene expression data, NSF grants DMS-1418261 and IIS 1546331 to SM and IOS-1402495 to RJCM and the UC Davis Department of Plant Sciences for funding.

## DATA EXAMPLE

### Background

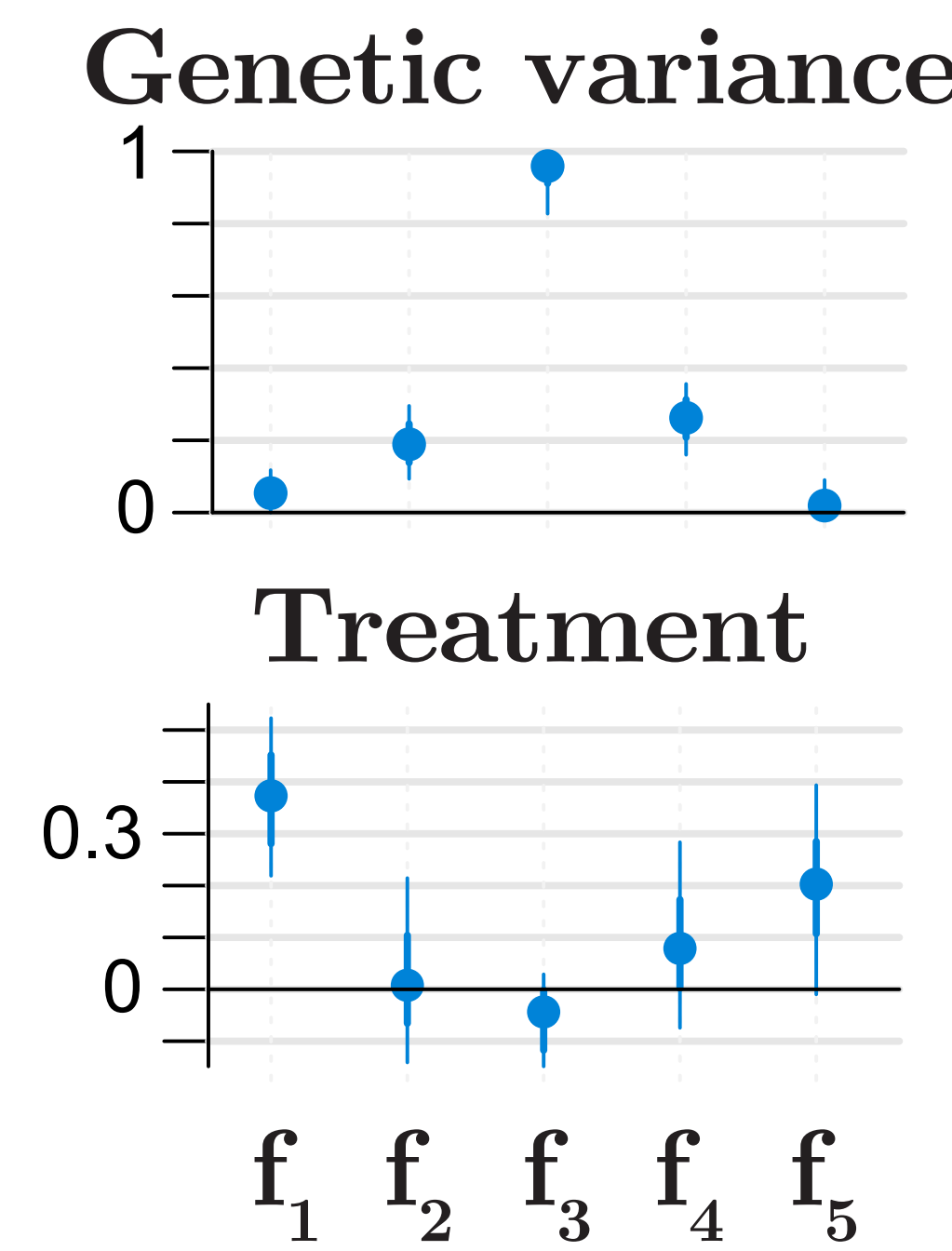
When plants grow in close proximity to one another they alter their development and physiology to compete for light at the top of the canopy having negative consequences on final yield. This response is regulated through the shade avoidance pathway involving a large gene network of transcription factors, hormone biosynthesis genes and metabolic genes. We have subset a large genetical genomics dataset collected on a recombinant inbred line (RIL) population of *Brassica rapa* grown in either crowded or uncrowded conditions in the field to examine known shade avoidance genes [Kazu reference].



**Figure 4.** Comparison of per plant fruit number on a subset of genotypes from the *Brassica rapa* RIL population grown in crowded or uncrowded field conditions.

## Results

Our model identified five large factors that accounted for the majority of the covariance in the data. Interestingly, the two genes that have the greatest positive factor loadings for factor three are PHYA (PHYTOCHROME A) a photoreceptor gene involved in shade detection, and a gene that regulates expression of PHYA, SPA (SUPPRESSOR OF PHYA). The factor loadings for factor one include a top positive loading gene, TAA1 (TRYPTOPHAN AMINOTRANSFERASE OF ARABIDOPSIS 1), an auxin biosynthesis gene, and a top negative loading gene, PIN3 (PIN-FORMED 3), is an auxin transport gene. These modeling results are promising that the loadings explaining either large genetic or treatment effects are selecting known subnetworks involved in the shade avoidance pathway.



**Figure 5.** Among-line genetic variance and response to crowded conditions for each of the five major factors.

## REFERENCES

- [1] A Bhattacharya and D B Dunson. Sparse Bayesian infinite factor models. *Biometrika*, 98(2):291–306, May 2011.
- [2] Bob Carpenter, Andrew Gelman, Matt Hoffman, Daniel Lee, Ben Goodrich, Michael betancourt, marcus A Brubaker, Jiqiang Guo, Peter Li, and Allen Riddell. Stan: A Probabilistic Programming Language. *J Stat Softw*, 2015.