

FirstLast>__A03_DataExploration.Rmd

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Fall 2023

OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Rename this file <FirstLast>_A03_DataExploration.Rmd (replacing <FirstLast> with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

TIP: If your code extends past the page when knit, tidy your code by manually inserting line breaks.

TIP: If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse, lubridate), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the subcommand to read strings in as factors.

```
getwd() #checking WD
```

```
## [1] "/home/guest/EDE_Fall2023"
```

```
knitr::opts_chunk$set(echo = TRUE) #knitting, lubridate, and tidyverse, here, ggplot2 download  
library(lubridate)  
library(tidyverse)  
library(here)  
library(ggplot2)
```

```

setwd(here())

#uploading two datasets
NeonicsFile <- here('Data','Raw','ECOTOX_Neonicotinoids_Insects_raw.csv')
print(NeonicsFile)

## [1] "/home/guest/EDE_Fall2023/Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv"

Neonics <- read.csv(
  file = here('Data','Raw','ECOTOX_Neonicotinoids_Insects_raw.csv'),
  stringsAsFactors = T)

LitterFile <- here('Data','Raw','NEON_NIWO_Litter_massdata_2018-08_raw.csv')
print(LitterFile)

## [1] "/home/guest/EDE_Fall2023/Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv"

Litter <- read.csv(
  file = here('Data','Raw','NEON_NIWO_Litter_massdata_2018-08_raw.csv'),
  stringsAsFactors = T)

```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: It is likely that neonicotinoids can harm or kill unintended insects (not just pests for agriculture but other species needed for other ecosystems)

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: It can help explain how carbon moves through an ecosystem.

4. How is litter and woody debris sampled as part of the NEON network? Read the NEON_Litterfall_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. Debris is dried individually in each category and the mass is recorded. 2. Size must be a $D < 2$ cm and a $L < 50$ cm. 3. Traps were placed to get the debris.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
dim(Neonics) #get dimensions
```

```
## [1] 4623 30
```

```
#4623 by 30
```

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
summary(Neonics$Effect)
```

```
##      Accumulation      Avoidance      Behavior      Biochemistry
##             12             102             360             11
##      Cell(s)      Development      Enzyme(s)      Feeding behavior
##             9             136             62             255
##      Genetics      Growth      Histology      Hormone(s)
##            82             38             5             1
##      Immunological      Intoxication      Morphology      Mortality
##             16             12             22             1493
##      Physiology      Population      Reproduction
##             7             1803             197
```

```
#Top 5: Population (1803), Mortality (1493), Behavior (360), Feeding behavior (255), Reproduction (197)
```

```
#ANSWER: If you see a change in population, death, behavior, or reproduction, it could signal that the
```

Answer: Top 5: Population (1803), Mortality (1493), Behavior (360), Feeding behavior (255), Reproduction (197) If you see a change in population, death, behavior, or reproduction, it could signal that the insecticide has had a negative impact on a species.

7. Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed. [TIP: The `sort()` command can sort the output of the summary command...]

```
summary(Neonics$Species.Common.Name)
```

```
##      Honey Bee      Parasitic Wasp
##             667             285
##      Buff Tailed Bumblebee      Carniolan Honey Bee
##             183             152
##      Bumble Bee      Italian Honeybee
##            140             113
##      Japanese Beetle      Asian Lady Beetle
##             94             76
##      Euonymus Scale      Wireworm
```

##	75	69
##	European Dark Bee	Minute Pirate Bug
##	66	62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
##	45	39
##	Aphid Family	Cabbage Looper
##	38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
##	Ladybird Beetle	Mason Bee
##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order

##		17		17
##		Egg Parasitoid		Insect Class
##		17		17
##	Moth And Butterfly Order		Oystershell Scale Parasitoid	
##		17		17
##	Hemlock Woolly Adelgid Lady Beetle		Hemlock Woolly Adelgid	
##		16		16
##		Mite		Onion Thrip
##		16		16
##	Western Flower Thrips		Corn Earworm	
##		15		14
##	Green Peach Aphid		House Fly	
##		14		14
##	Ox Beetle		Red Scale Parasite	
##		14		14
##	Spined Soldier Bug		Armoured Scale Family	
##		14		13
##	Diamondback Moth		Eulophid Wasp	
##		13		13
##	Monarch Butterfly		Predatory Bug	
##		13		13
##	Yellow Fever Mosquito		Braconid Parasitoid	
##		13		12
##	Common Thrip		Eastern Subterranean Termite	
##		12		12
##	Jassid		Mite Order	
##		12		12
##	Pea Aphid		Pond Wolf Spider	
##		12		12
##	Spotless Ladybird Beetle		Glasshouse Potato Wasp	
##		11		10
##	Lacewing		Southern House Mosquito	
##		10		10
##	Two Spotted Lady Beetle		Ant Family	
##		10		9
##	Apple Maggot		(Other)	
##		9		670

*#They are all bees/wasps! Other (670), Honey Bee (667), Parasitic Wasp (285),
#Buff Tailed Bumblebee (183), Carniolan Honey Bee (152), Bumble Bee (140),
#Italian Honeybee (113)*

*#They are all uncategorized or some sort of Bee/Wasp. These may be of interest
#because they are less likely to be the main targets of the insecticides--
#perhaps studies are checking to see if there are unintended consequences to these species.*

```
insect <- sort(summary(Neonics$Species.Common.Name), decreasing = TRUE)
insect
```

##	(Other)		Honey Bee
##		670	667
##	Parasitic Wasp		Buff Tailed Bumblebee
##		285	183

##	Carniolan Honey Bee	Bumble Bee
##	152	140
##	Italian Honeybee	Japanese Beetle
##	113	94
##	Asian Lady Beetle	Euonymus Scale
##	76	75
##	Wireworm	European Dark Bee
##	69	66
##	Minute Pirate Bug	Asian Citrus Psyllid
##	62	60
##	Parastic Wasp	Colorado Potato Beetle
##	58	57
##	Parasitoid Wasp	Erythrina Gall Wasp
##	51	49
##	Beetle Order	Snout Beetle Family, Weevil
##	47	47
##	Sevenspotted Lady Beetle	True Bug Order
##	46	45
##	Buff-tailed Bumblebee	Aphid Family
##	39	38
##	Cabbage Looper	Sweetpotato Whitefly
##	38	37
##	Braconid Wasp	Cotton Aphid
##	33	33
##	Predatory Mite	Ladybird Beetle Family
##	33	30
##	Parasitoid	Scarab Beetle
##	30	29
##	Spring Tiphia	Thrip Order
##	29	29
##	Ground Beetle Family	Rove Beetle Family
##	27	27
##	Tobacco Aphid	Chalcid Wasp
##	27	25
##	Convergent Lady Beetle	Stingless Bee
##	25	25
##	Spider/Mite Class	Tobacco Flea Beetle
##	24	24
##	Citrus Leafminer	Ladybird Beetle
##	23	23
##	Mason Bee	Mosquito
##	22	22
##	Argentine Ant	Beetle
##	21	21
##	Flatheaded Appletree Borer	Horned Oak Gall Wasp
##	20	20
##	Leaf Beetle Family	Potato Leafhopper
##	20	20
##	Tooth-necked Fungus Beetle	Codling Moth
##	20	19
##	Black-spotted Lady Beetle	Calico Scale
##	18	18
##	Fairyfly Parasitoid	Lady Beetle
##	18	18

##	Minute Parasitic Wasps	Mirid Bug
##	18	18
##	Mulberry Pyralid	Silkworm
##	18	18
##	Vedalia Beetle	Araneoid Spider Order
##	18	17
##	Bee Order	Egg Parasitoid
##	17	17
##	Insect Class	Moth And Butterfly Order
##	17	17
##	Oystershell Scale Parasitoid	Hemlock Woolly Adelgid Lady Beetle
##	17	16
##	Hemlock Woolly Adelgid	Mite
##	16	16
##	Onion Thrip	Western Flower Thrips
##	16	15
##	Corn Earworm	Green Peach Aphid
##	14	14
##	House Fly	Ox Beetle
##	14	14
##	Red Scale Parasite	Spined Soldier Bug
##	14	14
##	Armoured Scale Family	Diamondback Moth
##	13	13
##	Eulophid Wasp	Monarch Butterfly
##	13	13
##	Predatory Bug	Yellow Fever Mosquito
##	13	13
##	Braconid Parasitoid	Common Thrip
##	12	12
##	Eastern Subterranean Termite	Jassid
##	12	12
##	Mite Order	Pea Aphid
##	12	12
##	Pond Wolf Spider	Spotless Ladybird Beetle
##	12	11
##	Glasshouse Potato Wasp	Lacewing
##	10	10
##	Southern House Mosquito	Two Spotted Lady Beetle
##	10	10
##	Ant Family	Apple Maggot
##	9	9

Answer: They are all bees/wasps! Other (670), Honey Bee (667), Parasitic Wasp (285), Buff Tailed Bumblebee (183), Carniolan Honey Bee (152), Bumble Bee (140), Italian Honeybee (113)

#They are all uncategorized or some sort of Bee/Wasp. These may be of interest because they are less likely to be the main targets of the insecticides– perhaps studies are checking to see if there are unintended consequences to these species.

- Concentrations are always a numeric value. What is the class of `Conc.1..Author.` column in the dataset, and why is it not numeric?

```
class(Neonics$Conc.1..Author)
```

```
## [1] "factor"
```

```
#It is a factor because some of the numbers have slashes at the end/ other  
#various symbols, so they can't  
#be classified specifically as a number or not
```

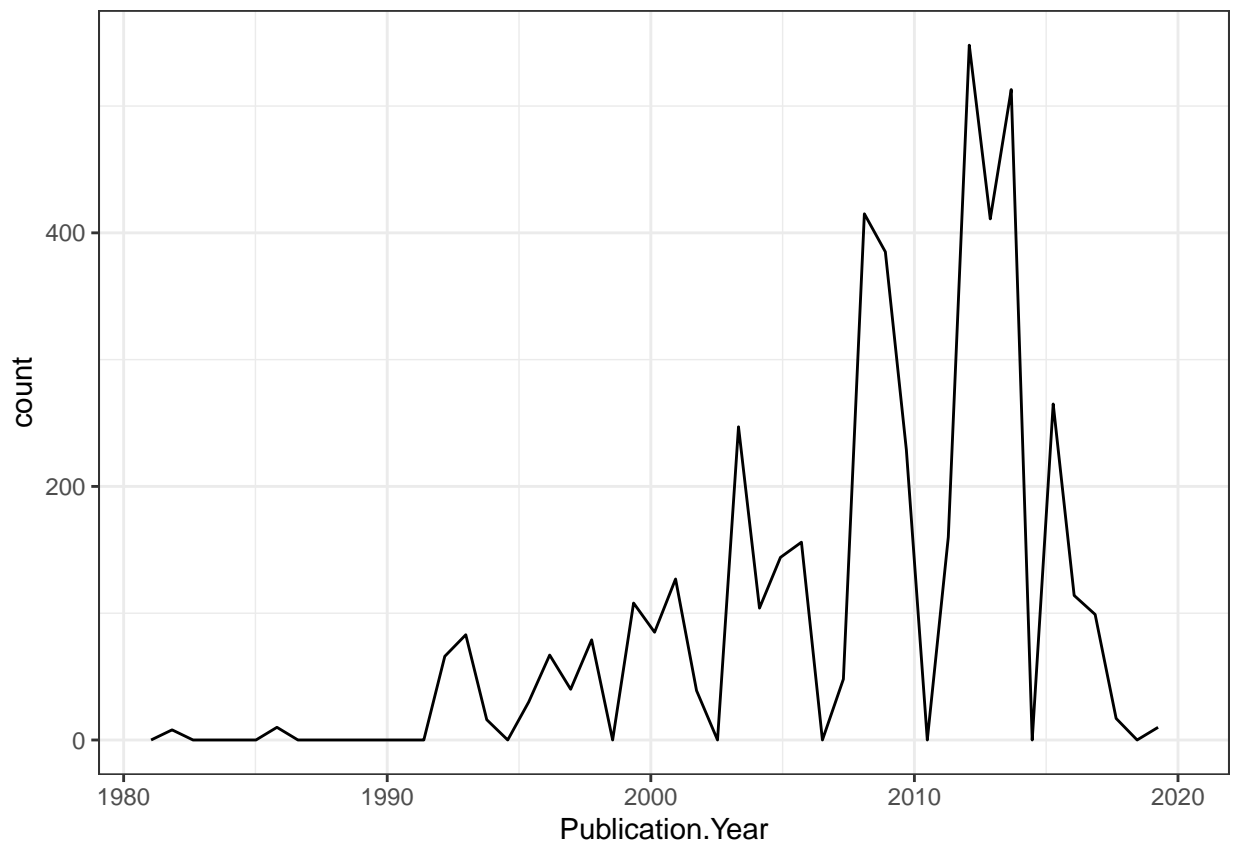
Answer: It is a factor because some of the numbers have slashes at the end/ other various symbols, so they can't be classified specifically as a number or not

Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics,  
  aes(x = Publication.Year))+  
  geom_freqpoly(bins = 50)+  
  scale_x_continuous(limits = c(1981,2020))+ #changing x axis  
  theme_bw()
```

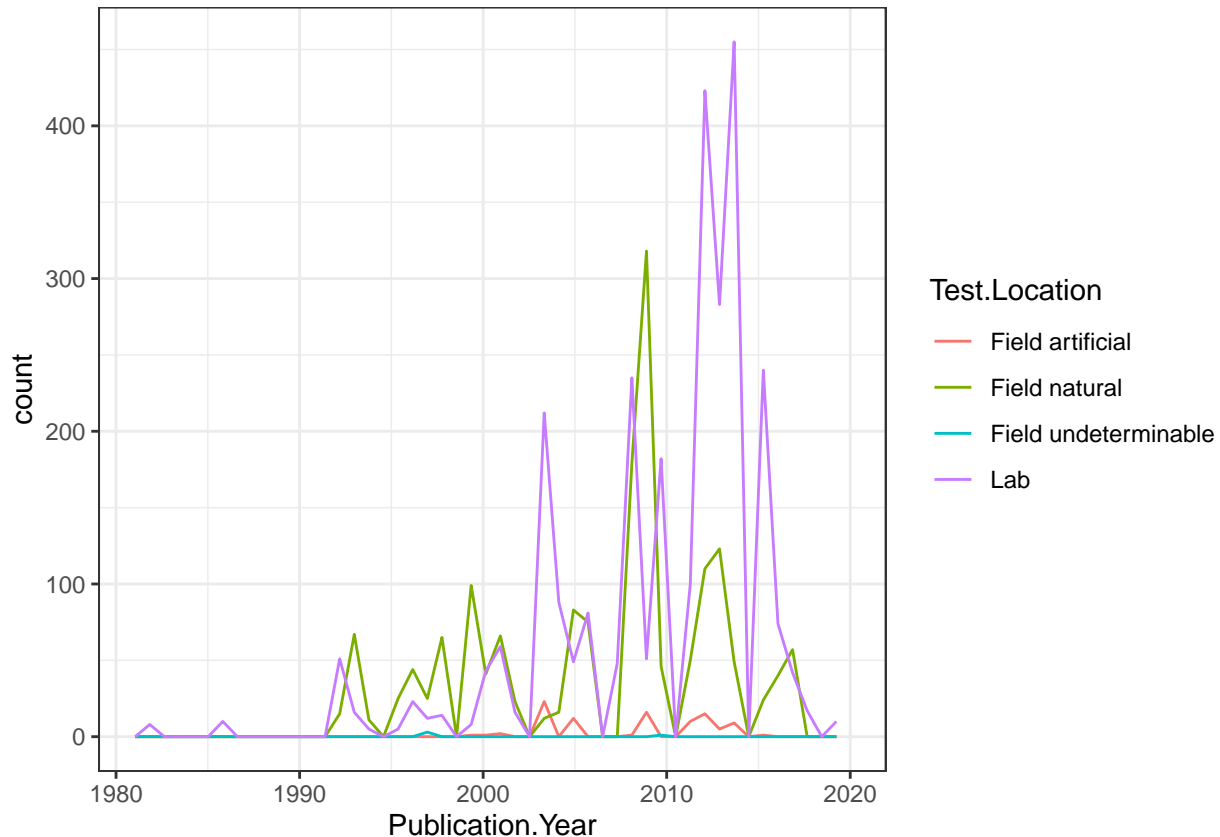
```
## Warning: Removed 3 rows containing missing values ('geom_path()').
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
ggplot(Neonics,  
  aes(x = Publication.Year, color = Test.Location))+ #change color by test location  
  geom_freqpoly(bins = 50)+  
  scale_x_continuous(limits = c(1981,2020))+  
  theme_bw()
```

Warning: Removed 12 rows containing missing values ('geom_path()').



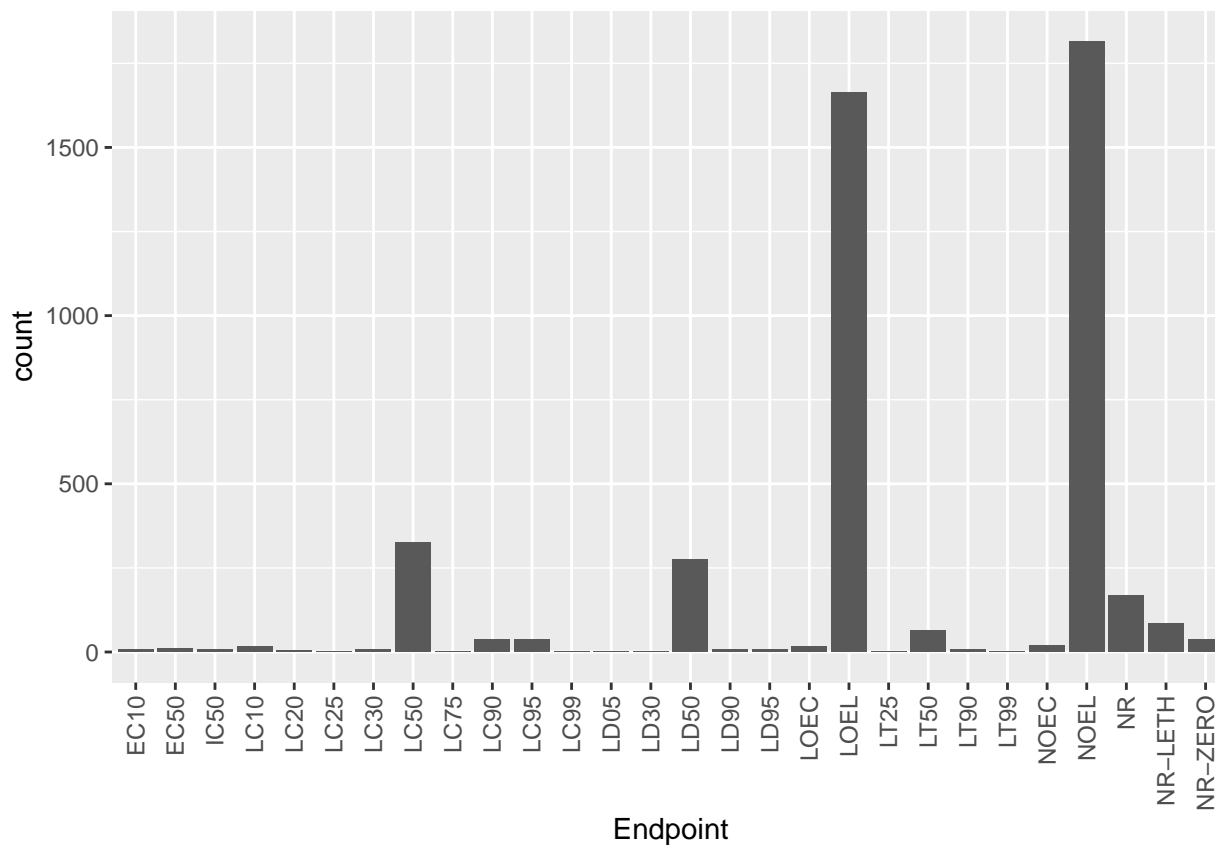
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common is the lab and Field natural, and they do seem to change over time. Lab peaks around 2014, and Field natural peaks around 2009.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

[TIP: Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics,  
  aes(x = Endpoint))+  
  geom_bar()+ #creates endpoints graph  
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))
```



Answer: The two most common endpoints are LOEL and NOEL, which according to the appendix are defined as LOEL (Lowest-observable-effect-level) and NOEL (No-observable-effect-level).

Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
#Collection Date is a factor, not a date.
```

```
class(Litter$collectDate)
```

```
## [1] "factor"
```

```
#it is a factor, not a date.
```

```
Round2 <- unique(Litter$collectDate)
Round2
```

```
## [1] 2018-08-02 2018-08-30
## Levels: 2018-08-02 2018-08-30
```

```
#August 2 and Aug 30 are sampling dates
```

```
#year month day conversion below
```

```
Litter$collectDate <- as.Date(Litter$collectDate, format = "%Y-%m-%d")  
class(Litter$collectDate)
```

```
## [1] "Date"
```

13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

```
summary(Litter$plotID)
```

```
## NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 NIWO_061  
##      20      19      18      15      14       8      16      17  
## NIWO_062 NIWO_063 NIWO_064 NIWO_067  
##      14      14      16      17
```

```
Special <- unique(Litter$plotID)  
Special
```

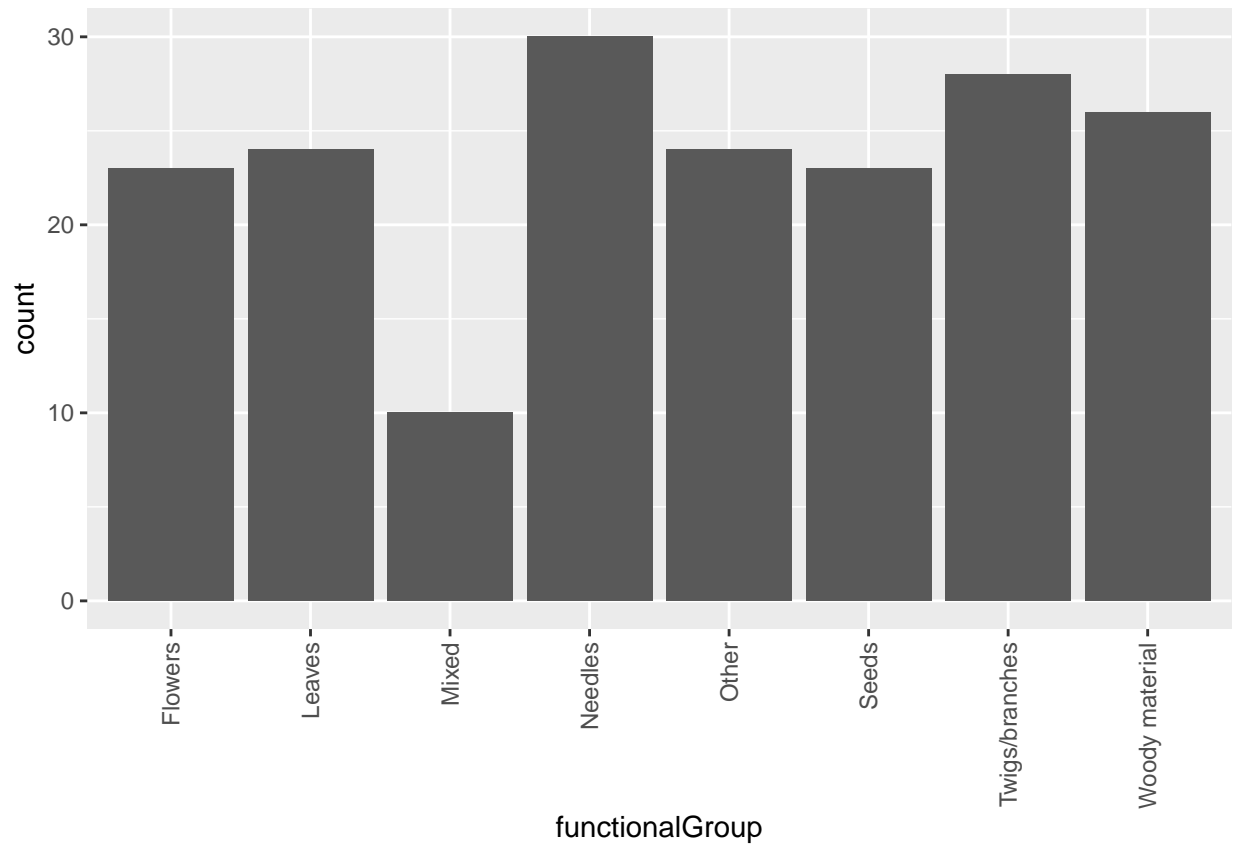
```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051  
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057  
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

```
#Summary lists 12 plots sampled + their frequency. Unique tells me the different types of samples and
```

Answer: Summary lists 12 plots sampled + their frequency. Unique tells me the different types of samples and the total number of different groups, but not their frequency

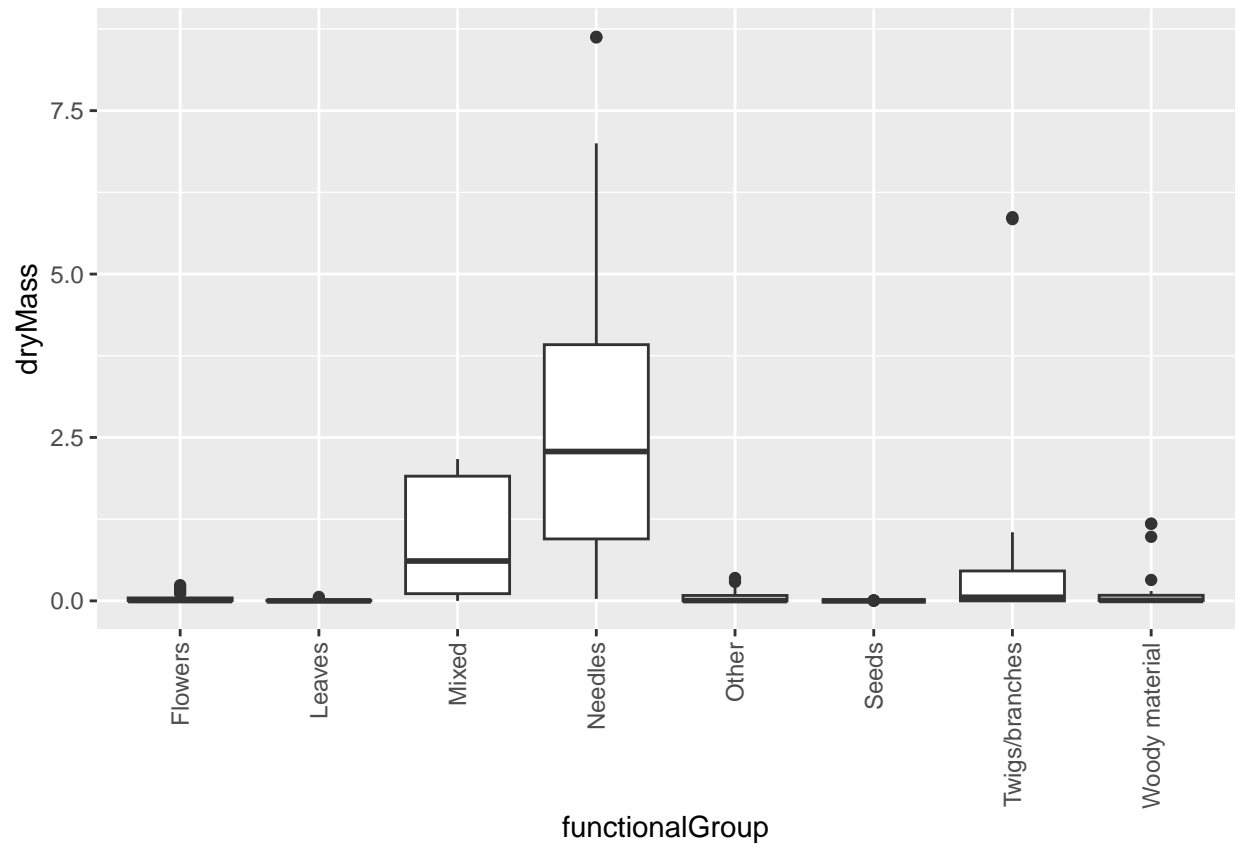
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter,  
       aes(x = functionalGroup))+  
  geom_bar()+ #bar graph  
theme(axis.text.x =element_text(angle = 90, vjust = 0.5, hjust=1))
```

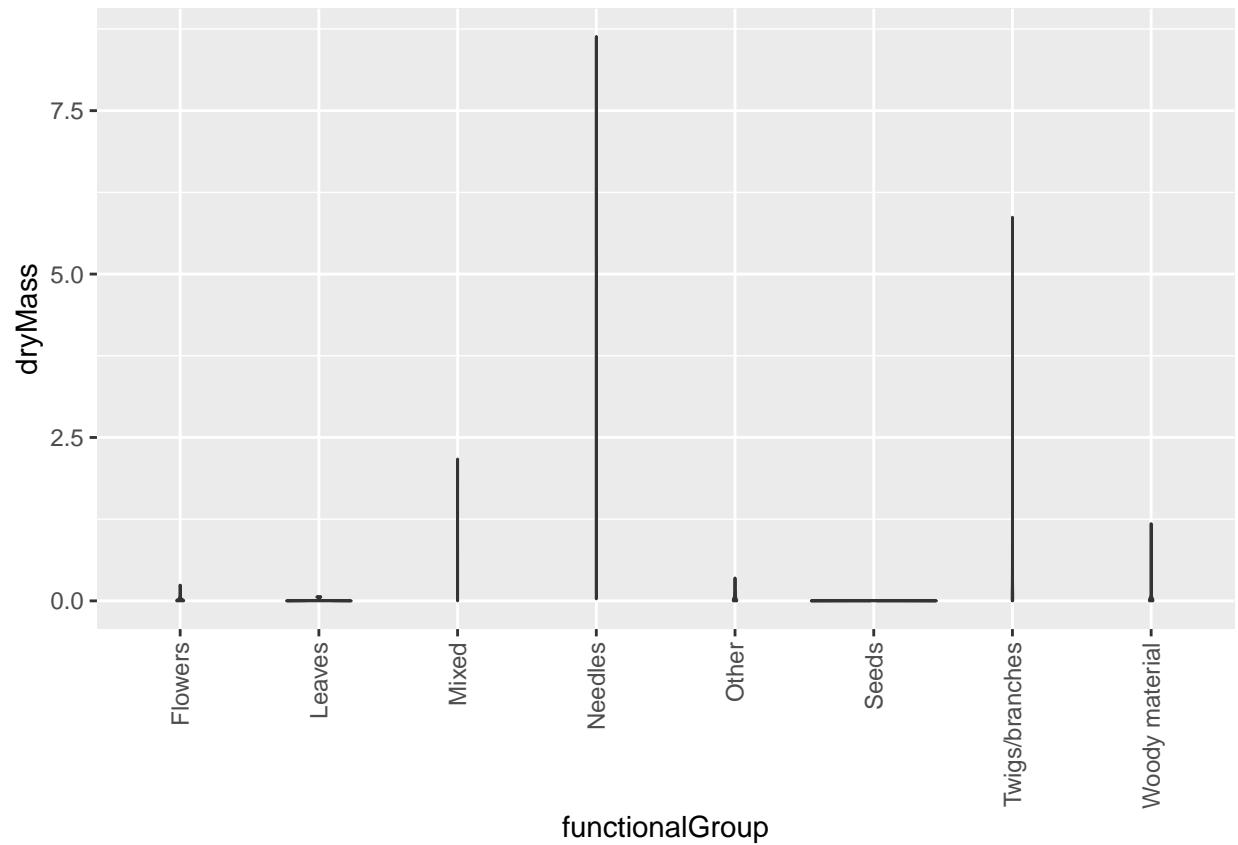


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
ggplot(Litter,
  aes(y= dryMass, x = functionalGroup))+
  geom_boxplot()+ #boxplot
  theme(axis.text.x =element_text(angle = 90, vjust = 0.5, hjust=1))
```



```
ggplot(Litter,
  aes(y= dryMass, x = functionalGroup))+
  geom_violin()+ #violin
  theme(axis.text.x =element_text(angle = 90, vjust = 0.5, hjust=1))
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: It shows the spread and outliers better. The violin does not have enough width to show a clear image.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles, Mixed, and Twig branches have the highest mean dryMass.