

Collective Emotions and Social Resilience in the Digital Traces After a Terrorist Attack

David García and Bernard Rimé

August 31st, 2018 (refactored on October 10th, 2022)

1. Data description

```
library(sfsmisc)
library(ggplot2)
library(zoo)
library(dplyr)
library(magrittr)
library(arm)
library(texreg)
Sys.setlocale("LC_ALL", 'en_US.UTF-8')
source("Scripts/AuxFunctions.R")
```

```
load("Data/Tweets.RData")
Tweets %>% mutate(day=as.Date(date)) -> Tweets
Tweets$user <- Tweets$user_id
Tweets %>% group_by(date) -> Tweets
print(paste("N tweets:", nrow(Tweets))) # 7666170
```

```
## [1] "N tweets: 17899591"
```

```
Tweets %>% filter(date>="2015-04-01" & date < "2015-10-01") -> TweetsBL
TweetsBL %>% mutate(w=weekdays(as.Date(date))) -> TweetsBL
print(paste("N tweets in baseline:", nrow(TweetsBL))) # 7666170
```

```
## [1] "N tweets in baseline: 7666170"
```

```
TweetsBL %>% group_by(date) %>% summarise(nt=length(n)) %>% summarise(mntw=mean(nt)) -> ntweetsBL
Tweets %>% summarise(ntweets=length(n)) -> dts
dts$bl <- rep(ntweetsBL$mntw, length(dts$ntweets))
```

```
dts %>% filter(date >= "2015-10-16" & date < "2015-12-12") -> dts2
```

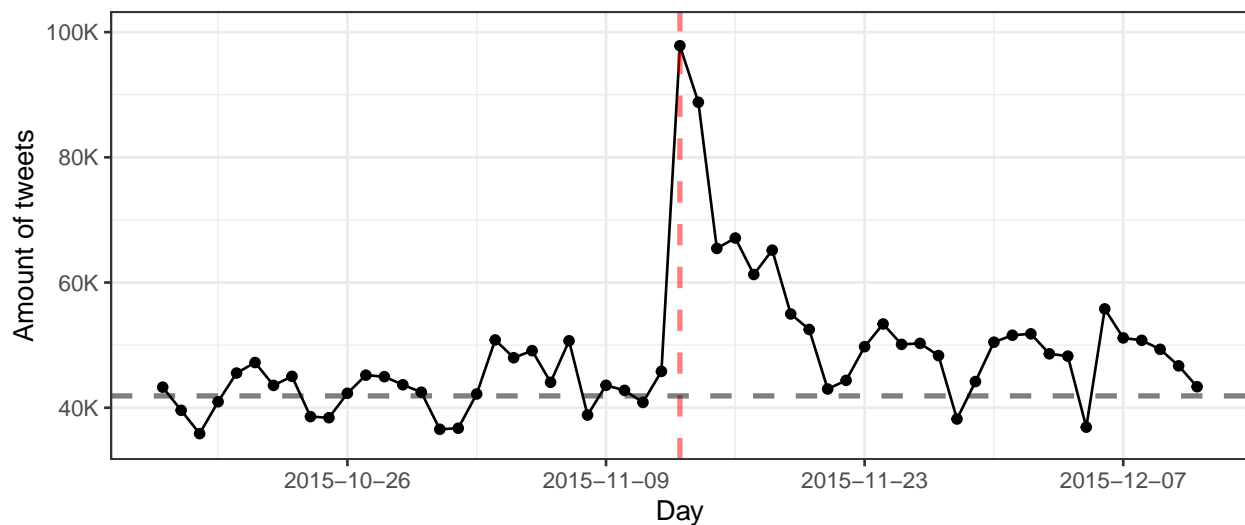
```
print(paste("N tweets in window:", sum(dts2$ntweets))) # 2766054
```

```
## [1] "N tweets in window: 2766054"
```

```

plt <- ggplot()
plt <- plt + geom_line(data = dts2, mapping=aes(x=as.Date(date), y=ntweets))
plt <- plt + geom_point(data = dts2, mapping=aes(x=as.Date(date), y=ntweets))
plt <- plt + scale_x_date("Day", date_breaks = "2 weeks") +
  scale_y_continuous(name="Amount of tweets", breaks=seq(20000,100000,20000),
    labels=c("20K","40K","60K","80K","100K")) +
  expand_limits(y=c(35000,100000))
plt <- plt + theme_bw() + geom_vline(xintercept=as.numeric(as.Date("2015-11-13")),
  col=rgb(1,0,0,0.5), lwd=1, lty=2)
plt <- plt + geom_hline(yintercept = dts$bl[1], col=rgb(0,0,0,0.5), lwd=1,lty=2)
plt

```



2. Affective Reactions

```

TweetsBL %>% group_by(w) %>% summarise(bl=mean(posemo/n)) -> posBL
TweetsBL %>% group_by(w) %>% summarise(bl=mean(negemo/n)) -> negBL

Tweets %>% filter(as.Date(date) >= "2015-10-16" & as.Date(date) < "2015-12-12") -> Tsel
posts <- ciTS(Tsel, "posemo", R=10000)
posts$w <- weekdays(as.Date(posts$date))
posts <- inner_join(posts, posBL)
save(posts, file="temp/posemoCITS.RData")

negts <- ciTS(Tsel, "negemo", R=10000)
negts$w <- weekdays(as.Date(negts$date))
negts <- inner_join(negts, negBL)
save(negts, file="temp/negemoCITS.RData")

load("temp/posemoCITS.RData")
load("temp/negemoCITS.RData")

d1 <- as.Date("2015-11-13")
dtbreaks <- c(d1 + seq(-27,0,by=3), d1 + seq(3,27,by=3))

```

```

plt <- plotts(ts=posts, ylab="Affect Terms", col="darkblue", bgcolor = rgb(0,0,1,0.25))
plt <- plotts(ts=negts, col="darkred", bgcolor = rgb(1,0,0,0.25), plt=plt)
plt <- plt + scale_x_date("Day", breaks = dtbreaks, date_labels = "%b %d")
plt <- plt + theme(axis.text.x = element_text(angle = 30, hjust = 1, colour = "black", size = 10))

Zday <- which(posts$date == "2015-11-13")[1]
negFit <- TSmodel1(log(negts$mid/negts$bl), Zday)
negDF <- data.frame(x=negts$date[2:length(negts$date)],y=negFit$fitted.values)
plt <- plt + geom_line(data=negDF, aes(x=as.Date(x),y=y), col= rgb(1,0,0), lty=2)

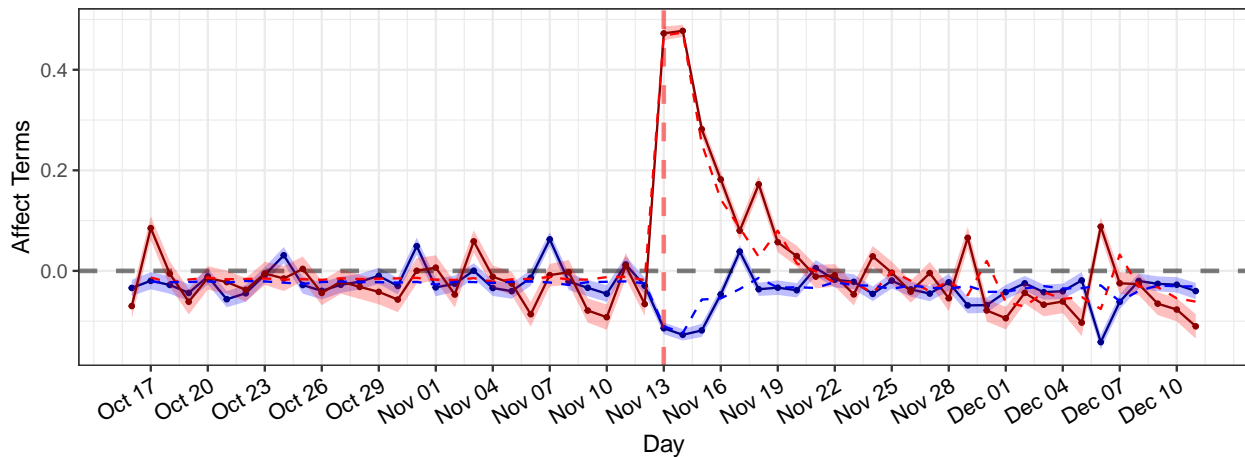
posFit <- TSmodel1(log(posts$mid/posts$bl), Zday)
posDF <- data.frame(x=posts$date[2:length(posts$date)],y=posFit$fitted.values)
plt <- plt + geom_line(data=posDF, aes(x=as.Date(x),y=y), col= rgb(0,0,1), lty=2)

plt

```

```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

```
## Warning: Use of 'ts$bl' is discouraged. Use 'bl' instead.
```



```

texreg(list(posFit, negFit), custom.model.names = c("PA", "NA"),
       digits=4, bold=0.05)

```

```

simulates <- coef(sim(posFit, n.sims=20000))
print("Intercept:")

```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```

##          2.5%          50%          97.5%
## -0.03358442 -0.02376761 -0.01377786

```

	PA	NA
(Intercept)	− 0.0239 *** (0.0051)	− 0.0175 * (0.0075)
ypre:postFALSE	−0.0652 (0.1276)	−0.0644 (0.2073)
ypre:postTRUE	0.2604 * (0.1124)	0.5669 *** (0.0772)
zTRUE	− 0.0882 ** (0.0310)	0.4812 *** (0.0532)
z2TRUE	− 0.0701 * (0.0323)	0.2240 *** (0.0626)
AIC	−222.5376	−165.9640
BIC	−210.3855	−153.8119
Log Likelihood	117.2688	88.9820
Deviance	0.0498	0.1366
Num. obs.	56	56

*** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$

Table 1: Statistical models

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## -0.31431033 -0.06360629  0.18612096
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## 0.04565718 0.26113473 0.47802588
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## -0.14837677 -0.08828218 -0.02859040
```

```
print("z2TRUE")
```

```
## [1] "z2TRUE"
```

```
quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%  
## -0.132710877 -0.069666619 -0.007595419
```

```
library(tseries)  
confint.default(posFit)
```

```
##          2.5 %          97.5 %  
## (Intercept)  -0.03385149 -0.013877238  
## ypre:postFALSE -0.31531176  0.184887999  
## ypre:postTRUE  0.04013304  0.480760085  
## zTRUE         -0.14885563 -0.027450230  
## z2TRUE        -0.13336449 -0.006749594
```

```
summ <- summary(posFit)  
print(paste("Positive terms model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "Positive terms model R2: 0.320636290180013"
```

```
shapiro.test(posFit$residuals)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data:  posFit$residuals  
## W = 0.88288, p-value = 5.772e-05
```

```
kpss.test(posFit$residuals)
```

```
## Warning in kpss.test(posFit$residuals): p-value greater than printed p-value
```

```
##  
## KPSS Test for Level Stationarity  
##  
## data:  posFit$residuals  
## KPSS Level = 0.24758, Truncation lag parameter = 3, p-value = 0.1
```

```
cor.test(sqrt(abs(posFit$residuals)), posFit$fitted.values)
```

```
##  
## Pearson's product-moment correlation  
##  
## data:  sqrt(abs(posFit$residuals)) and posFit$fitted.values
```

```
## t = 1.1988, df = 54, p-value = 0.2359
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## -0.1064014 0.4066892
## sample estimates:
##      cor
## 0.1610029
```

```
simulates <- coef(sim(negFit, n.sims=20000))
print("Intercept:")
```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## -0.032487129 -0.017661267 -0.002873199
```

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## -0.4703191 -0.0694061 0.3457771
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## 0.4181270 0.5660027 0.7128982
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## 0.3775782 0.4821834 0.5861031
```

```
print("z2TRUE")
```

```
## [1] "z2TRUE"
```

```
quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.1046636 0.2242888 0.3477586
```

```
confint.default(negFit)
```

```
##              2.5 %      97.5 %  
## (Intercept) -0.03223176 -0.002839171  
## ypre:postFALSE -0.47070331  0.341947757  
## ypre:postTRUE  0.41563237  0.718138851  
## zTRUE         0.37693212  0.585414794  
## z2TRUE        0.10136199  0.346683142
```

```
summ <- summary(negFit)  
print(paste("Negative terms model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "Negative terms model R2: 0.814094076168924"
```

```
shapiro.test(negFit$residuals)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data:  negFit$residuals  
## W = 0.91346, p-value = 0.0006731
```

```
kpss.test(negFit$residuals)
```

```
## Warning in kpss.test(negFit$residuals): p-value greater than printed p-value
```

```
##  
## KPSS Test for Level Stationarity  
##  
## data:  negFit$residuals  
## KPSS Level = 0.070959, Truncation lag parameter = 3, p-value = 0.1
```

```
cor.test(sqrt(abs(negFit$residuals)), negFit$fitted.values)
```

```
##  
## Pearson's product-moment correlation  
##  
## data:  sqrt(abs(negFit$residuals)) and negFit$fitted.values  
## t = -1.7877, df = 54, p-value = 0.07944
```

```
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## -0.47006823 0.02827807
## sample estimates:
##      cor
## -0.2363798
```

```
TweetsBL %>% group_by(w) %>% summarise(bl=mean(sad/n)) -> sadBL
TweetsBL %>% group_by(w) %>% summarise(bl=mean(ang/n)) -> angBL
TweetsBL %>% group_by(w) %>% summarise(bl=mean(anx/n)) -> anxBL
```

```
angts <- ciTS(Tsel, "ang", R=10000)
angts$w <- weekdays(as.Date(angts$date))
angts <- inner_join(angts, angBL)
save(angts, file="temp/angCITS.RData")
```

```
anxts <- ciTS(Tsel, "anx", R=10000)
anxts$w <- weekdays(as.Date(anxts$date))
anxts <- inner_join(anxts, anxBL)
save(anxts, file="temp/anxCITS.RData")
```

```
sadts <- ciTS(Tsel, "sad", R=10000)
sadts$w <- weekdays(as.Date(sadts$date))
sadts <- inner_join(sadts, sadBL)
save(sadts, file="temp/sadCITS.RData")
```

```
load("temp/anxCITS.RData")
load("temp/angCITS.RData")
load("temp/sadCITS.RData")
```

```
plt <- plotts(ts=sadts, ylab="Negative Affect Terms",
              col="darkblue", bgcolor = rgb(0,0,1,0.25))
plt <- plotts(ts=angts, col="darkred", bgcolor = rgb(1,0,0,0.25), plt=plt)
plt <- plotts(ts=anxts, col="darkorange", bgcolor = rgb(1,140/255,0,0.25), plt=plt)
```

```
Zday <- which(posts$date == "2015-11-13")[1]
sadFit <- TSmodel1(log(sadts$mid/sadts$bl), Zday)
DF <- data.frame(x=sadts$date[2:length(sadts$date)], y=sadFit$fitted.values)
plt <- plt + geom_line(data=DF, aes(x=as.Date(x), y=y), col= rgb(0,0,1), lty=2)
```

```
angFit <- TSmodel1(log(angts$mid/angts$bl), Zday)
DF <- data.frame(x=angts$date[2:length(angts$date)], y=angFit$fitted.values)
plt <- plt + geom_line(data=DF, aes(x=as.Date(x), y=y), col= rgb(1,0,0), lty=2)
```

```
anxFit <- TSmodel1(log(anxts$mid/anxts$bl), Zday)
DF <- data.frame(x=anxts$date[2:length(anxts$date)], y=anxFit$fitted.values)
plt <- plt + geom_line(data=DF, aes(x=as.Date(x), y=y), col= rgb(1,140/255,0), lty=2)
```

```
plt
```

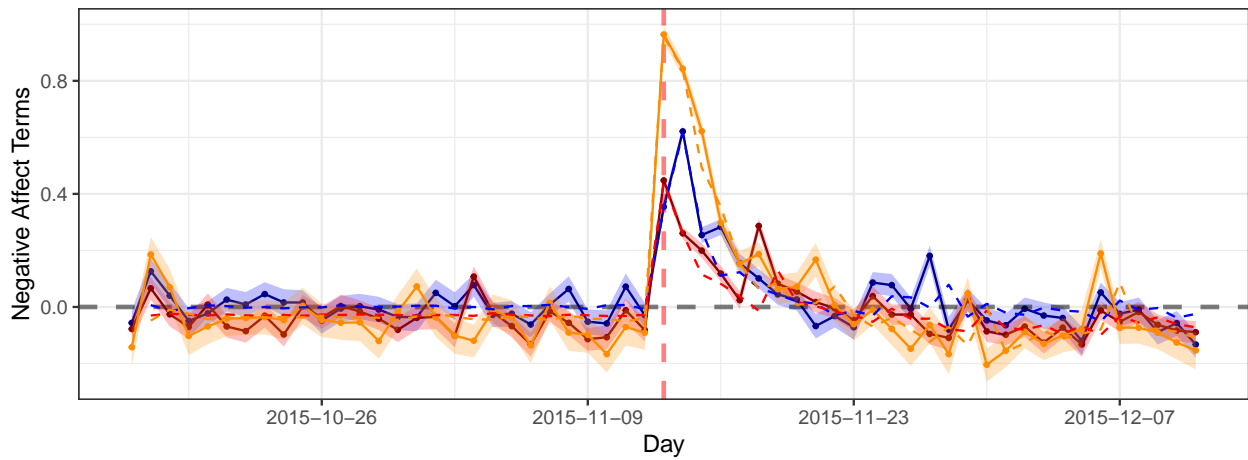
```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

```
## Warning: Use of 'ts$bl' is discouraged. Use 'bl' instead.
```


	Sadness	Anger	Anxiety
(Intercept)	0.0003 (0.0093)	-0.0268* (0.0101)	-0.0314* (0.0133)
ypre:postFALSE	-0.1116 (0.2353)	0.0452 (0.1903)	0.1161 (0.2072)
ypre:postTRUE	0.4337*** (0.0824)	0.5444*** (0.1170)	0.6220*** (0.0728)
zTRUE	0.3446*** (0.0674)	0.4700*** (0.0689)	0.9988*** (0.0914)
z2TRUE	0.4604*** (0.0723)	0.0422 (0.0847)	0.2713* (0.1128)
AIC	-137.0705	-135.4260	-104.2293
BIC	-124.9184	-123.2739	-92.0772
Log Likelihood	74.5352	73.7130	58.1146
Deviance	0.2289	0.2357	0.4115
Num. obs.	56	56	56

*** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$

Table 2: Statistical models



```
texreg(list(sadFit, angFit, anxFit),
        custom.model.names = c("Sadness", "Anger", "Anxiety"),
        digits=4, bold=0.05)
```

```
simulates <- coef(sim(sadFit, n.sims=20000))
print("Intercept:")
```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## -0.0179329047  0.0003464633  0.0189051349
```

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## -0.5713856 -0.1105952  0.3386838
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.2743307 0.4331621 0.5966726
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.2120646 0.3434533 0.4759827
```

```
print("z2TRUE")
```

```
## [1] "z2TRUE"
```

```
quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.3226935 0.4609608 0.6009262
```

```
summ <- summary(sadFit)  
confint.default(sadFit)
```

```
##              2.5 %      97.5 %  
## (Intercept) -0.01789236 0.01852496  
## ypre:postFALSE -0.57273097 0.34961751  
## ypre:postTRUE  0.27225981 0.59508548  
## zTRUE          0.21248197 0.47668488  
## z2TRUE         0.31867101 0.60218966
```

```
print(paste("Sadness terms model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "Sadness terms model R2: 0.72509524065696"
```

```
shapiro.test(sadFit$residuals)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data: sadFit$residuals  
## W = 0.9678, p-value = 0.1393
```

```
kpss.test(sadFit$residuals)
```

```
## Warning in kpss.test(sadFit$residuals): p-value greater than printed p-value
```

```
##  
## KPSS Test for Level Stationarity  
##  
## data: sadFit$residuals  
## KPSS Level = 0.27604, Truncation lag parameter = 3, p-value = 0.1
```

```
cor.test(sqrt(abs(sadFit$residuals)), sadFit$fitted.values)
```

```
##  
## Pearson's product-moment correlation  
##  
## data: sqrt(abs(sadFit$residuals)) and sadFit$fitted.values  
## t = -1.4816, df = 54, p-value = 0.1443  
## alternative hypothesis: true correlation is not equal to 0  
## 95 percent confidence interval:  
## -0.43779302 0.06883722  
## sample estimates:  
## cor  
## -0.1976401
```

```
simulates <- coef(sim(angFit, n.sims=20000))  
print("Intercept:")
```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%  
## -0.046736751 -0.026681029 -0.007115979
```

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%  
## -0.32302353  0.04639009  0.41504681
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%  
## 0.3196843 0.5453978 0.7783533
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%  
## 0.3373233 0.4693421 0.6034956
```

```
print("z2TRUE")
```

```
## [1] "z2TRUE"
```

```
quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%  
## -0.12710741  0.04195016  0.20669433
```

```
summ <- summary(angFit)  
confint.default(angFit)
```

```
##          2.5 %          97.5 %  
## (Intercept) -0.04655919 -0.007003452  
## ypre:postFALSE -0.32780784  0.418184553  
## ypre:postTRUE  0.31499807  0.773720618  
## zTRUE          0.33490204  0.605124221  
## z2TRUE         -0.12376716  0.208086024
```

```
print(paste("Anger terms model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "Anger terms model R2: 0.630283227850662"
```

```
shapiro.test(angFit$residuals)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data: angFit$residuals  
## W = 0.85791, p-value = 9.754e-06
```

```
kpss.test(angFit$residuals)
```

```
## Warning in kpss.test(angFit$residuals): p-value greater than printed p-value
```

```
##  
## KPSS Test for Level Stationarity  
##  
## data: angFit$residuals  
## KPSS Level = 0.1018, Truncation lag parameter = 3, p-value = 0.1
```

```
cor.test(sqrt(abs(angFit$residuals)), angFit$fitted.values)
```

```
##  
## Pearson's product-moment correlation  
##  
## data: sqrt(abs(angFit$residuals)) and angFit$fitted.values  
## t = -1.4684, df = 54, p-value = 0.1478  
## alternative hypothesis: true correlation is not equal to 0  
## 95 percent confidence interval:  
## -0.43637155 0.07058583  
## sample estimates:  
## cor  
## -0.195951
```

```
simulates <- coef(sim(anxFit, n.sims=20000))  
print("Intercept:")
```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%  
## -0.057190760 -0.031669709 -0.005954387
```

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## -0.2856313  0.1185355  0.5253398
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.4790579 0.6215181 0.7641173
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.8234559 0.9986836 1.1784208
```

```
print("z2TRUE")
```

```
## [1] "z2TRUE"
```

```
quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.0539678 0.2708852 0.4918722
```

```
summ <- summary(anxFit)  
confint.default(anxFit)
```

```
##              2.5 %      97.5 %  
## (Intercept) -0.05746755 -0.00540306  
## ypre:postFALSE -0.29007127  0.52218201  
## ypre:postTRUE  0.47935823  0.76457837  
## zTRUE         0.81974455  1.17793407  
## z2TRUE        0.05025417  0.49241980
```

```
print(paste("Anxiety terms model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "Anxiety terms model R2: 0.84568694624521"
```

```
shapiro.test(anxFit$residuals)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data: anxFit$residuals  
## W = 0.94105, p-value = 0.008617
```

```
kpss.test(anxFit$residuals)
```

```
## Warning in kpss.test(anxFit$residuals): p-value greater than printed p-value
```

```
##  
## KPSS Test for Level Stationarity  
##  
## data: anxFit$residuals  
## KPSS Level = 0.082316, Truncation lag parameter = 3, p-value = 0.1
```

```
cor.test(sqrt(abs(anxFit$residuals)), anxFit$fitted.values)
```

```
##  
## Pearson's product-moment correlation  
##  
## data: sqrt(abs(anxFit$residuals)) and anxFit$fitted.values  
## t = -0.93662, df = 54, p-value = 0.3531  
## alternative hypothesis: true correlation is not equal to 0  
## 95 percent confidence interval:  
## -0.3768102 0.1411579  
## sample estimates:  
## cor  
## -0.1264347
```

3. Social processes

```
TweetsBL %>% group_by(w) %>% summarise(bl=mean(soc/n)) -> SocBL  
TweetsBL %>% group_by(w) %>% summarise(bl=mean(prosoc/n)) -> ProSocBL  
TweetsBL %>% group_by(w) %>% summarise(bl=mean(frenchValues/n)) -> FVBL  
  
Socts <- ciTS(Tsel, "soc", R=10000)  
Socts$w <- weekdays(as.Date(Socts$date))  
Socts <- inner_join(Socts, SocBL)  
save(Socts, file="temp/SocCITS.RData")
```

```
ProSocts <- ciTS(Tsel, "prosoc", R=10000)
ProSocts$w <- weekdays(as.Date(ProSocts$date))
ProSocts <- inner_join(ProSocts, ProSocBL)
save(ProSocts, file="temp/ProSocCITS.RData")
```

```
FVts <- ciTS(Tsel, "frenchValues", R=10000)
FVts$w <- weekdays(as.Date(FVts$date))
FVts <- inner_join(FVts, FVBL)
save(FVts, file="temp/frenchValuesCITS.RData")
```

```
load("temp/SocCITS.RData")
```

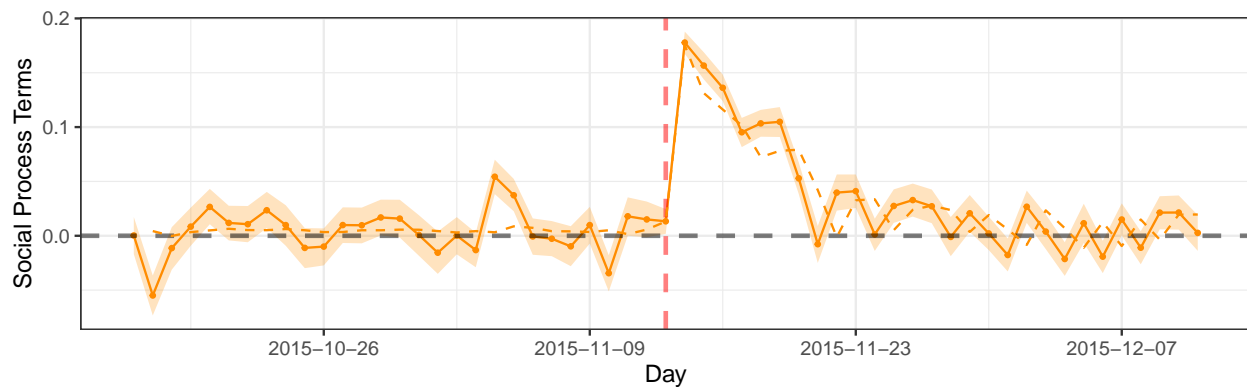
```
plt <- plotts(ts=Socts, ylab="Social Process Terms",
             col="darkorange", bgcolor = rgb(1,144/255,0,0.25))
```

```
Zday <- which(Socts$date == "2015-11-13")[1]
SocFit <- TSmodel1(log(Socts$mid/Socts$bl), Zday)
DF <- data.frame(x=Socts$date[2:length(Socts$date)], y=SocFit$fitted.values)
plt <- plt + geom_line(data=DF, aes(x=as.Date(x), y=y), col= rgb(1,144/255,0), lty=2)

plt
```

```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

```
## Warning: Use of 'ts$bl' is discouraged. Use 'bl' instead.
```



```
texreg(SocFit, custom.model.names = c("Social"), digits=4, bold=0.05)
```

```
simulates <- coef(sim(SocFit, n.sims=20000))
print("Intercept:")
```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## -0.002678329  0.004308643  0.011114040
```


	Social
(Intercept)	0.0043 (0.0036)
ypre:postFALSE	0.0810 (0.1453)
ypre:postTRUE	0.7146 *** (0.0754)
zTRUE	0.0076 (0.0238)
z2TRUE	0.1616 *** (0.0238)
AIC	-253.0178
BIC	-240.8657
Log Likelihood	132.5089
Deviance	0.0289
Num. obs.	56

*** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$

Table 3: Statistical models

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## -0.20710348  0.08179196  0.36700532
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## 0.5676856  0.7144005  0.8606857
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## -0.038667499  0.008093485  0.055198961
```

```

print("z2TRUE")

## [1] "z2TRUE"

quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))

##      2.5%      50%      97.5%
## 0.1161985 0.1614211 0.2080157

summ <- summary(SocFit)
confint.default(SocFit)

##              2.5 %      97.5 %
## (Intercept) -0.002708557 0.01125288
## ypre:postFALSE -0.203714229 0.36576467
## ypre:postTRUE  0.566851413 0.86239467
## zTRUE         -0.039102850 0.05437763
## z2TRUE         0.114842490 0.20827864

print(paste("Social process terms model R2:", (1-summ$deviance/summ$null.deviance)))

## [1] "Social process terms model R2: 0.727505894368885"

shapiro.test(SocFit$residuals)

##
##  Shapiro-Wilk normality test
##
## data:  SocFit$residuals
## W = 0.99025, p-value = 0.9307

kpss.test(SocFit$residuals)

## Warning in kpss.test(SocFit$residuals): p-value greater than printed p-value

##
##  KPSS Test for Level Stationarity
##
## data:  SocFit$residuals
## KPSS Level = 0.079223, Truncation lag parameter = 3, p-value = 0.1

cor.test(sqrt(abs(SocFit$residuals)), SocFit$fitted.values)

##
##  Pearson's product-moment correlation
##
## data:  sqrt(abs(SocFit$residuals)) and SocFit$fitted.values
## t = -0.28917, df = 54, p-value = 0.7736

```

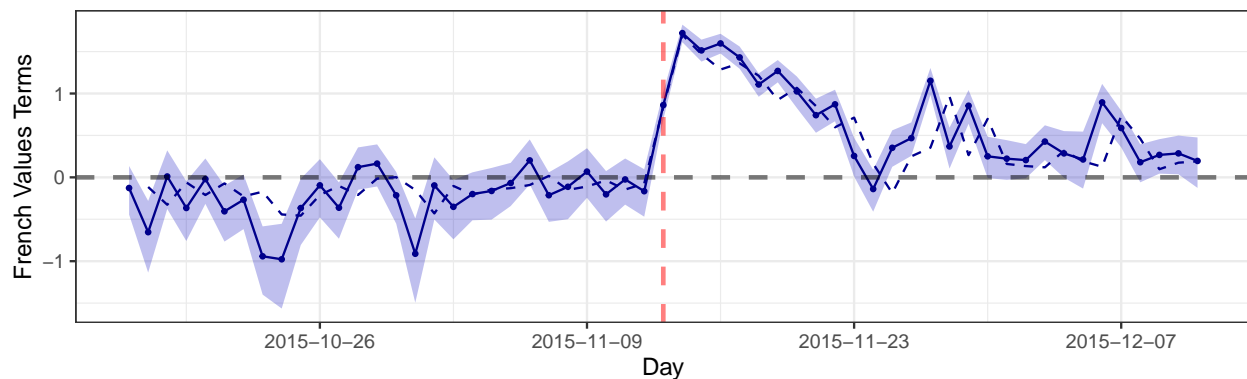
```
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## -0.2991287 0.2259155
## sample estimates:
## cor
## -0.03932035
```

```
load("temp/frenchValuesCITS.RData")
plt <- plotts(ts=FVts, ylab="French Values Terms",
             col="darkblue", bgcolor = rgb(0,0,0.75,0.25))

Zday <- which(FVts$date == "2015-11-13")[1]
FVFit <- TSmodel1(log(FVts$mid/FVts$bl), Zday)
DF <- data.frame(x=FVts$date[2:length(FVts$date)], y=FVFit$fitted.values)
plt <- plt + geom_line(data=DF, aes(x=as.Date(x), y=y), col= rgb(0,0,144/255), lty=2)
plt
```

```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

```
## Warning: Use of 'ts$bl' is discouraged. Use 'bl' instead.
```



```
texreg(FVFit, custom.model.names = c("French Values Terms"), digits=4, bold=0.05)
```

```
simulates <- coef(sim(FVFit, n.sims=20000))
print("Intercept:")
```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## -0.19816807 -0.06442813  0.06549143
```

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

	French Values Terms
(Intercept)	-0.0639 (0.0674)
ypre:postFALSE	0.4023* (0.1987)
ypre:postTRUE	0.8916*** (0.1011)
zTRUE	0.9704** (0.3472)
z2TRUE	0.9944** (0.3492)
AIC	46.9645
BIC	59.1166
Log Likelihood	-17.4822
Deviance	6.1217
Num. obs.	56

*** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$

Table 4: Statistical models

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## 0.005039597 0.401113710 0.789266876
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## 0.7009989 0.8922652 1.0917788
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##          2.5%          50%          97.5%
## 0.2870458 0.9689702 1.6594691
```

```
print("z2TRUE")
```

```
## [1] "z2TRUE"
```

```
quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%  
## 0.3097633 0.9916495 1.6798260
```

```
summ <- summary(FVFit)  
confint.default(FVFit)
```

```
##              2.5 %      97.5 %  
## (Intercept) -0.19614136 0.06825486  
## ypre:postFALSE 0.01295194 0.79171428  
## ypre:postTRUE 0.69345962 1.08968676  
## zTRUE        0.28983941 1.65101132  
## z2TRUE       0.30993792 1.67893619
```

```
print(paste("French Values terms model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "French Values terms model R2: 0.715291698184095"
```

```
shapiro.test(FVFit$residuals)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data:  FVFit$residuals  
## W = 0.98072, p-value = 0.5071
```

```
kpss.test(FVFit$residuals)
```

```
##  
## KPSS Test for Level Stationarity  
##  
## data:  FVFit$residuals  
## KPSS Level = 0.5964, Truncation lag parameter = 3, p-value = 0.02296
```

```
cor.test(sqrt(abs(FVFit$residuals)), FVFit$fitted.values)
```

```
##  
## Pearson's product-moment correlation  
##  
## data:  sqrt(abs(FVFit$residuals)) and FVFit$fitted.values  
## t = -1.2658, df = 54, p-value = 0.211  
## alternative hypothesis: true correlation is not equal to 0  
## 95 percent confidence interval:  
## -0.41417327 0.09749468  
## sample estimates:  
##      cor  
## -0.169757
```

	Prosocial Terms
(Intercept)	0.0408 *** (0.0081)
ypre:postFALSE	0.2317 (0.1509)
ypre:postTRUE	0.6544 *** (0.0449)
zTRUE	0.1405 *** (0.0326)
z2TRUE	0.4072 *** (0.0328)
AIC	-218.8325
BIC	-206.6804
Log Likelihood	115.4162
Deviance	0.0532
Num. obs.	56

*** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$

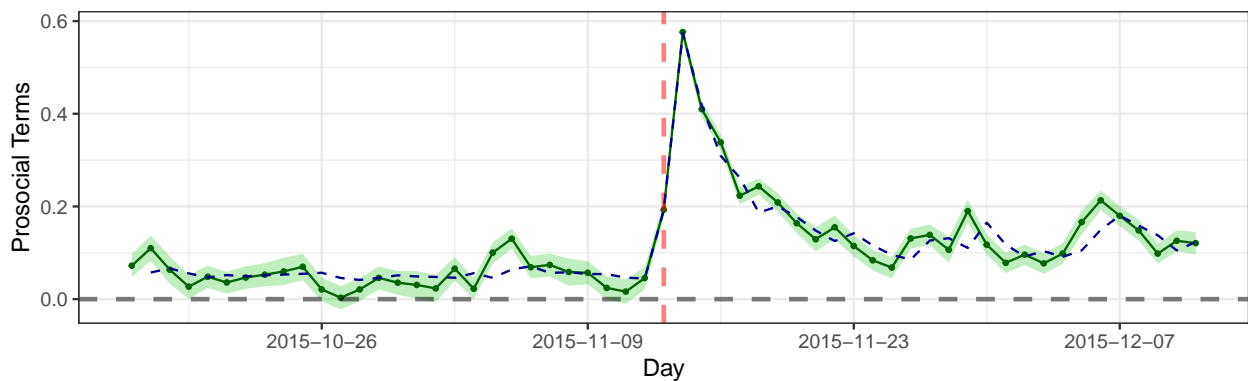
Table 5: Statistical models

```
load("temp/ProSocCITS.RData")
plt <- plotts(ts=ProSocTs, ylab="Prosocial Terms",
             col="darkgreen", bgcolor = rgb(0,0.75,0,0.25))

Zday <- which(ProSocTs$date == "2015-11-13")[1]
ProSocFit <- TSmodel1(log(ProSocTs$mid/ProSocTs$bl), Zday)
DF <- data.frame(x=ProSocTs$date[2:length(ProSocTs$date)], y=ProSocFit$fitted.values)
plt <- plt + geom_line(data=DF, aes(x=as.Date(x), y=y), col= rgb(0,0,144/255), lty=2)
plt
```

Warning: Use of 'ts\$mid' is discouraged. Use 'mid' instead.

Warning: Use of 'ts\$bl' is discouraged. Use 'bl' instead.



```
texreg(ProSocFit, custom.model.names = c("Prosocial Terms"), digits=4, bold=0.05)
```

```
simulates <- coef(sim(ProSocFit, n.sims=20000))
print("Intercept:")
```

```
## [1] "Intercept:"
```

```
quantile(simulates[10001:20000,1], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## 0.02514143 0.04062941 0.05656255
```

```
print("ypre:postFALSE")
```

```
## [1] "ypre:postFALSE"
```

```
quantile(simulates[10001:20000,2], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## -0.06218908 0.23234455 0.52765040
```

```
print("ypre:postTRUE")
```

```
## [1] "ypre:postTRUE"
```

```
quantile(simulates[10001:20000,3], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## 0.5647845 0.6545555 0.7400445
```

```
print("zTRUE")
```

```
## [1] "zTRUE"
```

```
quantile(simulates[10001:20000,4], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## 0.07613559 0.14053265 0.20437299
```

```
print("z2TRUE")
```

```
## [1] "z2TRUE"
```

```
quantile(simulates[10001:20000,5], probs = c(0.025, 0.5, 0.975))
```

```
##      2.5%      50%      97.5%
## 0.3430520 0.4074965 0.4724248
```

```
summ <- summary(ProSocFit)
confint.default(ProSocFit)
```

```
##              2.5 %      97.5 %
## (Intercept)  0.02487821 0.05667417
## ypre:postFALSE -0.06403942 0.52742572
## ypre:postTRUE  0.56647622 0.74229704
## zTRUE         0.07656915 0.20447626
## z2TRUE        0.34297309 0.47141470
```

```
print(paste("prosocial terms model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "prosocial terms model R2: 0.905744466411463"
```

```
shapiro.test(ProSocFit$residuals)
```

```
##
##  Shapiro-Wilk normality test
##
## data:  ProSocFit$residuals
## W = 0.93085, p-value = 0.003221
```

```
kpss.test(ProSocFit$residuals)
```

```
## Warning in kpss.test(ProSocFit$residuals): p-value greater than printed p-value
```

```
##
##  KPSS Test for Level Stationarity
##
## data:  ProSocFit$residuals
## KPSS Level = 0.072436, Truncation lag parameter = 3, p-value = 0.1
```

```
cor.test(sqrt(abs(ProSocFit$residuals)), ProSocFit$fitted.values)
```

```
##
##  Pearson's product-moment correlation
##
## data:  sqrt(abs(ProSocFit$residuals)) and ProSocFit$fitted.values
## t = -0.48618, df = 54, p-value = 0.6288
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
##  -0.3233061  0.2003612
## sample estimates:
##           cor
## -0.06601682
```

4. Emotional Synchronization Effect


```

Tweets$emo <- Tweets$posemo + Tweets$negemo

Tweets %>% filter(date>="2015-11-13" & date <= "2015-11-27")-> TweetsDFAttack
TweetsDFAttack %>% group_by(userid) -> TweetsDFAttack
TweetsDFAttack %>% summarise(avgEmo=mean((posemo+negemo)/n), nT=n()/15, # 15 days after the attacks
                             avgSoc=mean(soc/n), avgProsoc=mean(prosoc/n),
                             avgFV=mean(frenchValues/n), avgPA=mean(posemo/n), avgNA=mean(negemo/n),
                             avgAng=mean(ang/n), avgAnx = mean(anx/n), avgSad=mean(sad/n)) -> userDF

Tweets %>% filter(date>="2015-08-13" & date <= "2015-11-12")-> TweetsDFBL
TweetsDFBL %>% group_by(userid) -> TweetsDFBL
TweetsDFBL %>% summarise(nTBL = length(n)/92, # number of days in baseline period
                          avgEmoBL=mean((posemo+negemo)/n), socBL=mean(soc/n), prosocBL=mean(prosoc/n),
                          FVBL=mean(frenchValues/n),
                          PABL=mean(posemo/n), NABL=mean(negemo/n),
                          anxBL=mean(anx/n), sadBL=mean(sad/n), angerBL=mean(ang/n), iBL=mean(i/n)) -> userDFBL

userDF <- full_join(userDF, userDFBL, by="userid")

Tweets %>% filter(date>="2015-11-28" & date <= "2016-02-27")-> TweetsDFPost
TweetsDFPost %>% group_by(userid) -> TweetsDFPost
TweetsDFPost %>% summarise( nTpost=n(),
                             PAPost = mean(posemo/n), NAPost = mean(negemo/n),
                             anxPost = mean(anx/n), sadPost = mean(sad/n), angPost = mean(ang/n),
                             EmoPost = mean((posemo+negemo)/n),
                             socPost=mean(soc/n), prosocPost =mean(prosoc/n), FVPost = mean(frenchValues/n)) -> userDFPost
userDF <- full_join(userDF, userDFPost, by="userid")

save(userDF, file="temp/userDF.RData")

sel <- data.frame(userid=userDF$userid, sel=userDF$avgEmo>userDF$avgEmoBL)
sel$user <- sel$userid

pTS <- PairTSW(Tweets, sel, "soc", R=1000,w=30)
save(pTS, file="temp/SynchSoc.RData")

pTS <- PairTSDifW(Tweets, sel, "soc", R=1000,w=30)
save(pTS, file="temp/SynchDifSoc.RData")

pTS <- PairTSW(Tweets, sel, "prosoc", R=1000,w=30)
save(pTS, file="temp/SynchProSoc.RData")

pTS <- PairTSDifW(Tweets, sel, "prosoc", R=1000,w=30)
save(pTS, file="temp/SynchDifProSoc.RData")

pTS <- PairTSW(Tweets, sel, "frenchValues", R=1000,w=30)
save(pTS, file="temp/SynchFV.RData")

pTS <- PairTSDifW(Tweets, sel, "frenchValues", R=1000,w=30)
save(pTS, file="temp/SynchDifFV.RData")

```

	Model 1
(Intercept)	0.0365 *** (0.0005)
PABL	0.3093 *** (0.0078)
NABL	0.3345 *** (0.0127)
socBL	-0.0096 (0.0104)
iBL	0.2411 *** (0.0076)
log(nTBL)	-0.0015 *** (0.0001)
AIC	-170257.2162
BIC	-170195.6190
Log Likelihood	85135.6081
Deviance	88.8427
Num. obs.	49001

*** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$

Table 6: Self-selection model based on personality correlates

```
load("temp/userDF.RData")
attach(userDF)

sum(!is.na(nT) & nT>0)
sum(!is.na(nTBL) & nTBL>0)
sum(!is.na(nTpost) & nTpost>0)
sum((!is.na(nT) & nT>0) & (!is.na(nTBL) & nTBL>0) & (!is.na(nTpost) & nTpost>0))

periodUserDF <- subset(userDF, (!is.na(nT) & nT>0) & (!is.na(nTBL) & nTBL>0) & (!is.na(nTpost) & nTpost>0))
save(periodUserDF, file="temp/periodUserDF.RData")
```

4.0.1 Self-selection

```
load("temp/periodUserDF.RData")
periodUserDF %>% dplyr::select(avgEmo, PABL, NABL, socBL, iBL, nTBL) -> cDF

lin <- bayesglm(avgEmo ~ PABL+NABL+socBL+iBL+log(nTBL), data=cDF)

texreg(lin, digits=4, bold=0.05, caption="Self-selection model based on personality correlates")
```

```
confint(lin)
```

```
##                2.5 %        97.5 %
## (Intercept)  0.035532334  0.037525639
## PABL        0.294195301  0.324772563
## NABL        0.310028013  0.360011595
## socBL       -0.030275168  0.010757290
```

```
## iBL          0.226220989  0.256190957
## log(nTBL)    -0.001774636 -0.001225168
```

```
summ <- summary(lin)
print(paste("Self-selection model R2:", (1-summ$deviance/summ$null.deviance)))
```

```
## [1] "Self-selection model R2: 0.0928057185364379"
```

4.1 Difference time series visualizations

```
load("temp/SynchSoc.RData")

d1 <- as.Date("2015-11-13")
dtbreaks <- c(d1 + seq(-4*7*8,0,by=7*4), d1 + seq(7*8,4*7*8,by=7*4))

Tts <- data.frame(date=pTS$date, mid=pTS$Tmid*100, low=pTS$Tlow*100, hi=pTS$Thi*100)
Fts <- data.frame(date=pTS$date, mid=pTS$Fmid*100, low=pTS$Flow*100, hi=pTS$Fhi*100)
plt <- plotts(Tts, nolog=TRUE,
              zero=FALSE, dtbreaks= "4 weeks", col="red", bgcolor = rgb(1,0,0,0.25))
plt <- plotts(Fts, col="blue", nolog=TRUE, add=TRUE, plt=plt,
              bgcolor= rgb(0,0,1,0.25), zero=FALSE)
plt <- plt + geom_vline(xintercept=as.numeric(as.Date("2015-11-27")))
plt <- plt + scale_x_date("Day", breaks = dtbreaks, date_labels = "%b %d", limits=c(as.Date("2015-06-30"),
plt
```

```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

```
## Warning: Removed 61 row(s) containing missing values (geom_path).
```

```
## Warning: Removed 61 rows containing missing values (geom_point).
```

```
## Warning: Removed 61 row(s) containing missing values (geom_path).
```

```
## Warning: Removed 61 rows containing missing values (geom_point).
```

```
load("temp/SynchSoc.RData")
refTS <- pTS
load("temp/SynchDifSoc.RData")
pTS$low <- pTS$low/refTS$Flow*100
pTS$mid <- pTS$mid/refTS$Fmid*100
pTS$hi <- pTS$hi/refTS$Fhi*100

plt <- plotts(pTS, nolog=TRUE, bgcolor= rgb(0.95,0.75,0,0.25), col="darkorange",
              zero=TRUE)
plt <- plt + geom_vline(xintercept=as.numeric(as.Date("2015-11-27")))
plt <- plt + scale_x_date("Day", breaks = dtbreaks, date_labels = "%b %d", limits=c(as.Date("2015-06-30"),
plt
```

```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

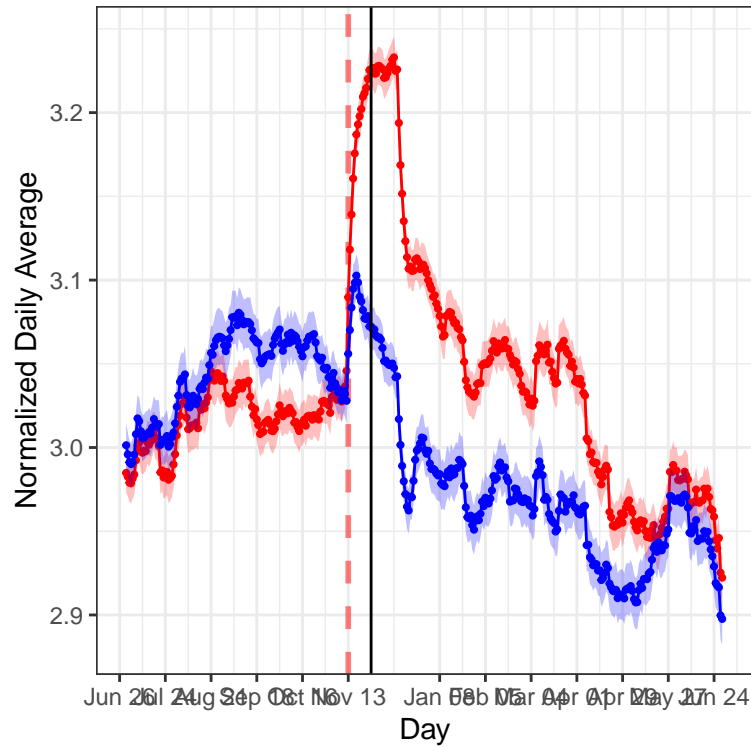


Figure 1: Social Process Terms Synchronization

```
## Warning: Removed 61 row(s) containing missing values (geom_path).
```

```
## Warning: Removed 61 rows containing missing values (geom_point).
```

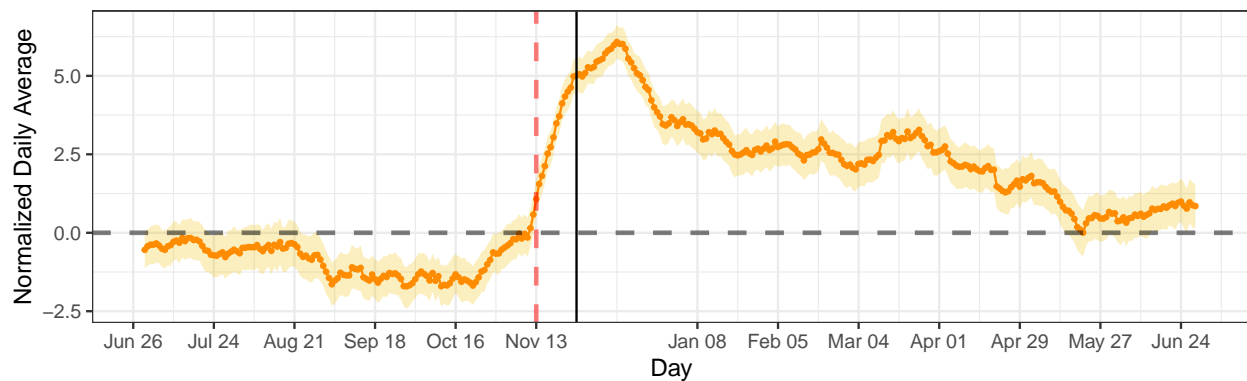


Figure 2: Social Process Terms Synchronization

```
load("temp/SynchProSoc.RData")
d1 <- as.Date("2015-11-13")
dtbreaks <- c(d1 + seq(-4*7*8,0,by=7*8), d1 + seq(7*8,4*7*8,by=7*8))

Tts <- data.frame(date=pTS$date, mid=pTS$Tmid, low=pTS$Tlow, hi=pTS$Thi)
Fts <- data.frame(date=pTS$date, mid=pTS$Fmid, low=pTS$Flow, hi=pTS$Fhi)
plt <- plotts(Tts, nolog=TRUE,
```

```

      zero=FALSE, dtbreaks= "8 weeks", col="red", bgcolor = rgb(1,0,0,0.25))
plt <- plotts(Fts, col="blue", nolog=TRUE, add=TRUE, plt=plt,
      bgcolor= rgb(0,0,1,0.25), zero=FALSE)
plt <- plt + geom_vline(xintercept=as.numeric(as.Date("2015-11-27")))
plt <- plt + scale_x_date("Day", breaks = dtbreaks, date_labels = "%b %d")
plt

```

Warning: Use of 'ts\$mid' is discouraged. Use 'mid' instead.

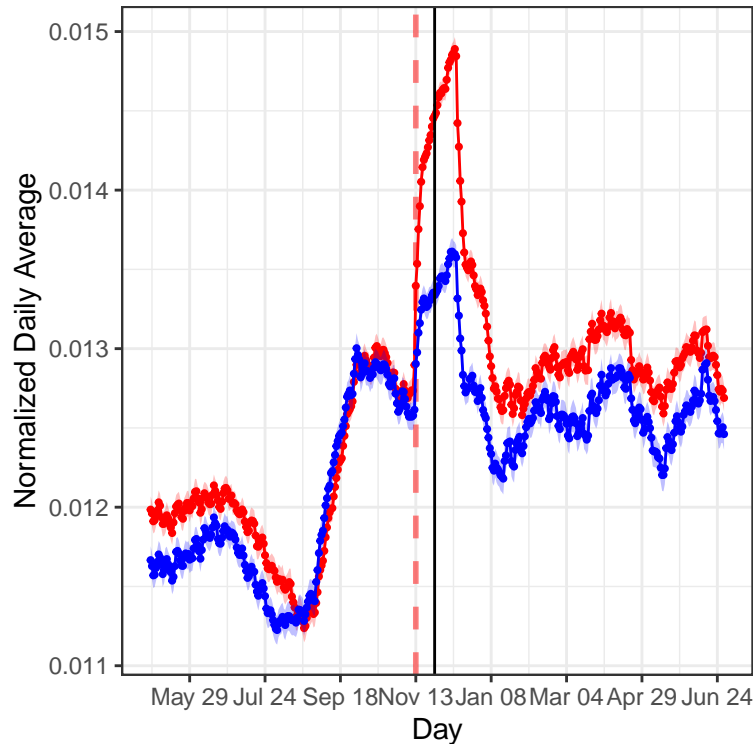


Figure 3: Prosocial Terms Synchronization

```

load("temp/SynchProSoc.RData")
refTS <- pTS
load("temp/SynchDifProSoc.RData")
pTS$low <- pTS$low/refTS$Flow*100
pTS$mid <- pTS$mid/refTS$Fmid*100
pTS$hi <- pTS$hi/refTS$Fhi*100

plt <- plotts(pTS, nolog=TRUE, bgcolor= rgb(0,0.5,0,0.25), col="darkgreen",
      zero=TRUE)
plt <- plt + geom_vline(xintercept=as.numeric(as.Date("2015-11-27")))
plt <- plt + scale_x_date("Day", breaks = dtbreaks, date_labels = "%b %d", limits=c(as.Date("2015-06-30"),
      as.Date("2015-12-31")))
plt

```

Warning: Use of 'ts\$mid' is discouraged. Use 'mid' instead.

Warning: Removed 61 row(s) containing missing values (geom_path).

```
## Warning: Removed 61 rows containing missing values (geom_point).
```

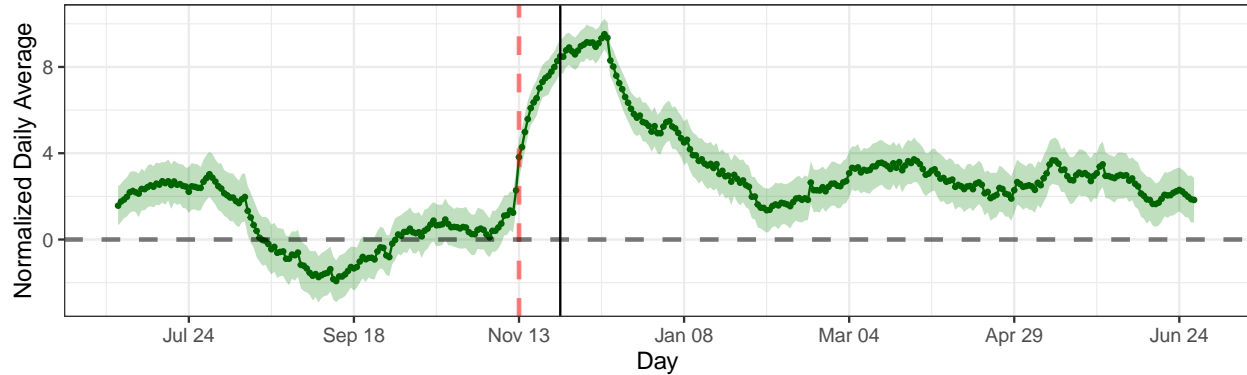


Figure 4: Prosocial Terms Synchronization

```
load("temp/SynchFV.RData")
d1 <- as.Date("2015-11-13")
dtbreaks <- c(d1 + seq(-4*7*8,0,by=7*8), d1 + seq(7*8,4*7*8,by=7*8))

Tts <- data.frame(date=pTS$date, mid=pTS$Tmid, low=pTS$Tlow, hi=pTS$Thi)
Fts <- data.frame(date=pTS$date, mid=pTS$Fmid, low=pTS$Flow, hi=pTS$Fhi)
plt <- plotts(Tts, nolog=TRUE,
              zero=FALSE, dtbreaks= "8 weeks", col="red", bgcolor = rgb(1,0,0,0.25))
plt <- plotts(Fts, col="blue", nolog=TRUE, add=TRUE, plt=plt,
              bgcolor= rgb(0,0,1,0.25), zero=FALSE)

plt <- plt + geom_vline(xintercept=as.numeric(as.Date("2015-11-27")))
plt <- plt + scale_x_date("Day", breaks = dtbreaks, date_labels = "%b %d")
plt
```

```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

```
load("temp/SynchFV.RData")
refTS <- pTS
load("temp/SynchDifFV.RData")
pTS$low <- pTS$low/refTS$Flow*100
pTS$mid <- pTS$mid/refTS$Fmid*100
pTS$hi <- pTS$hi/refTS$Fhi*100

plt <- plotts(pTS, nolog=TRUE, bgcolor= rgb(0,0,0.5,0.25), col="darkblue",
              zero=TRUE)
plt <- plt + geom_vline(xintercept=as.numeric(as.Date("2015-11-27")))
plt <- plt + scale_x_date("Day", breaks = dtbreaks, date_labels = "%b %d", limits=c(as.Date("2015-06-30"),
as.Date("2016-06-30")))
plt
```

```
## Warning: Use of 'ts$mid' is discouraged. Use 'mid' instead.
```

```
## Warning: Removed 61 row(s) containing missing values (geom_path).
```

```
## Warning: Removed 61 rows containing missing values (geom_point).
```

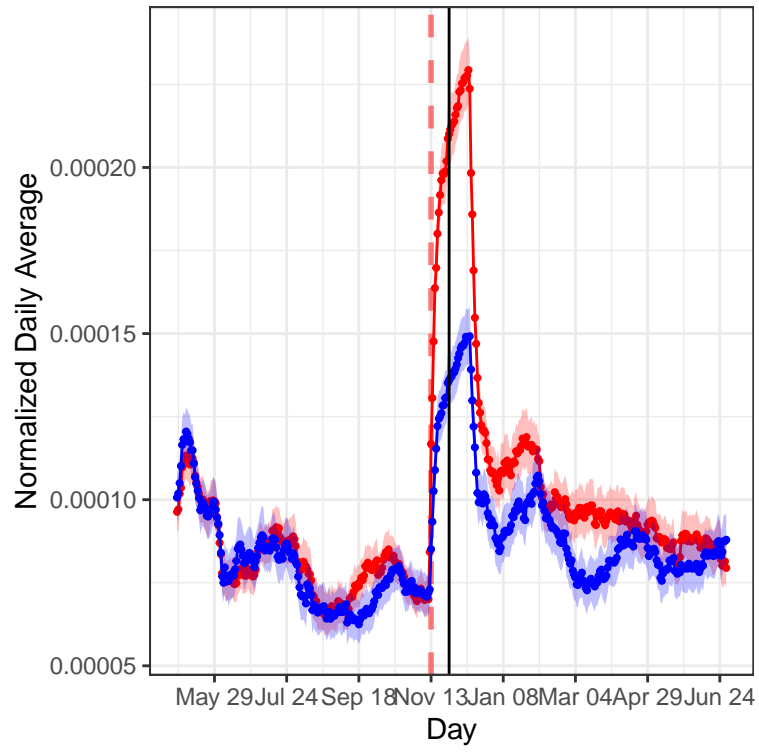


Figure 5: French Value Terms Synchronization

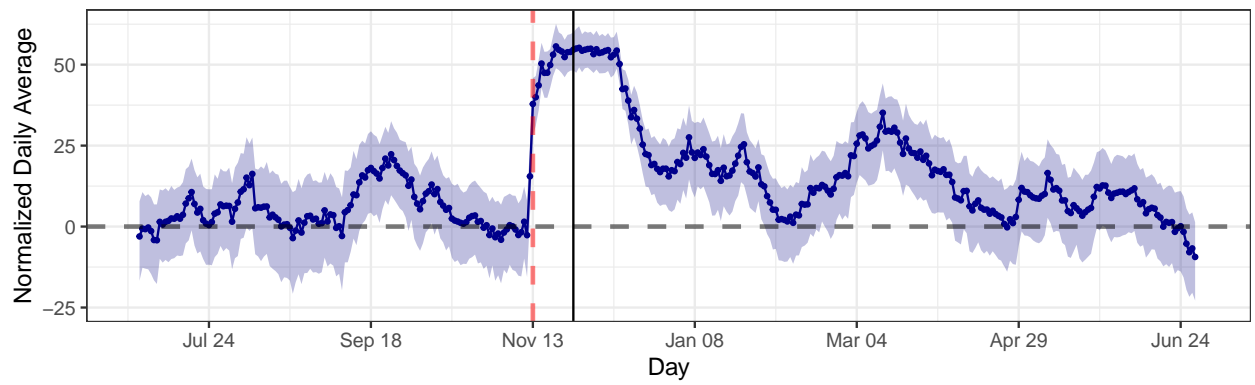


Figure 6: French Value Synchronization

4.2 Mediation analysis

```
library(mediation)
Nrep <- 10000

load("periodUserDF.RData")
periodUserDF %>% dplyr::select(avgEmo, PABL, NABL, socBL, iBL, nTBL) -> cDF
prelin <- glm(avgEmo ~ PABL+NABL+socBL+iBL+log(nTBL), data=cDF)

userDF <- periodUserDF
userDF$emo <- as.numeric(scale(userDF$avgEmo - predict(prelin, newdata=userDF)))
userDF$socBL <- as.numeric(scale(userDF$socBL))
userDF$prosocBL <- as.numeric(scale(userDF$prosocBL))
userDF$FVBL <- as.numeric(scale(userDF$FVBL))
userDF$PABL <- as.numeric(scale(userDF$PABL))
userDF$NABL <- as.numeric(scale(userDF$NABL))
userDF$iBL <- as.numeric(scale(userDF$iBL))
userDF$lnTBL <- as.numeric(scale(log(userDF$nTBL)))

userDF$socPost <- as.numeric(scale(userDF$socPost))
userDF$avgSoc <- as.numeric(scale(userDF$avgSoc))
med.fit <- glm(avgSoc ~ emo + socBL + PABL + NABL + iBL + lnTBL, data = userDF)
out.fit <- glm(socPost ~ avgSoc + emo + socBL + PABL + NABL + iBL + lnTBL, data = userDF)

med.soc <- mediate(med.fit, out.fit, treat = "emo", mediator = "avgSoc", control.value=quantile(userDF$emo, 0.5))
save(med.soc, file="temp/med.soc.RData")

userDF$prosocPost <- as.numeric(scale(userDF$prosocPost))
userDF$avgProsoc <- as.numeric(scale(userDF$avgProsoc))
med.fit <- glm(avgProsoc ~ emo + prosocBL + PABL + NABL + iBL + lnTBL, data = userDF)
out.fit <- glm(prosocPost ~ avgProsoc + emo + prosocBL + PABL + NABL + iBL + lnTBL, data = userDF)

med.prosoc <- mediate(med.fit, out.fit, treat = "emo", mediator = "avgProsoc", control.value=quantile(userDF$emo, 0.5))
save(med.prosoc, file="temp/med.prosoc.RData")

userDF$FVPost <- as.numeric(scale(userDF$FVPost))
userDF$avgFV <- as.numeric(scale(userDF$avgFV))
med.fit <- glm(avgFV ~ emo + FVBL + PABL + NABL + iBL + lnTBL, data = userDF)
out.fit <- glm(FVPost ~ avgFV + emo + FVBL + PABL + NABL + iBL + lnTBL, data = userDF)

med.FV <- mediate(med.fit, out.fit, treat = "emo", mediator = "avgFV", control.value=quantile(userDF$emo, 0.5))
save(med.FV, file="temp/med.FV.RData")

userDF$PAPost <- as.numeric(scale(userDF$PAPost))
userDF$avgPA <- as.numeric(scale(userDF$avgPA))
med.fit <- glm(avgPA ~ emo + socBL + PABL + NABL + iBL + lnTBL, data = userDF)
out.fit <- glm(PAPost ~ avgPA + emo + socBL + PABL + NABL + iBL + lnTBL, data = userDF)

med.PA <- mediate(med.fit, out.fit, treat = "emo", mediator = "avgPA", control.value=quantile(userDF$emo, 0.5))
save(med.PA, file="temp/med.PA.RData")

userDF$NAPost <- as.numeric(scale(userDF$NAPost))
userDF$avgNA <- as.numeric(scale(userDF$avgNA))
```



```

med.fit <- glm(avgNA~ emo + socBL + PABL + NABL + iBL + lnTBL, data = userDF)
out.fit <- glm(NAPost ~ avgNA + emo + socBL + PABL + NABL + iBL + lnTBL, data = userDF)

med.NA <- mediate(med.fit, out.fit, treat = "emo", mediator = "avgNA", control.value=quantile(userDF$emo,
save(med.NA, file="temp/med.NA.RData")

```

```

library(mediation)
load("temp/med.soc.RData")
summary(med.soc)

```

```

##
## Causal Mediation Analysis
##
## Nonparametric Bootstrap Confidence Intervals with the Percentile Method
##
## (Inference Conditional on the Covariate Values Specified in 'covariates')
##
##           Estimate 95% CI Lower 95% CI Upper p-value
## ACME           0.0160      0.0131      0.02 <2e-16 ***
## ADE            0.0224      0.0105      0.03  2e-04 ***
## Total Effect    0.0384      0.0267      0.05 <2e-16 ***
## Prop. Mediated   0.4172      0.2960      0.62 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Sample Size Used: 49001
##
##
## Simulations: 10000

```

```

load("temp/med.prosoc.RData")
summary(med.prosoc)

```

```

##
## Causal Mediation Analysis
##
## Nonparametric Bootstrap Confidence Intervals with the Percentile Method
##
## (Inference Conditional on the Covariate Values Specified in 'covariates')
##
##           Estimate 95% CI Lower 95% CI Upper p-value
## ACME           0.00922      0.00722      0.01 <2e-16 ***
## ADE            0.01571      0.00442      0.03  0.0076 **
## Total Effect    0.02492      0.01375      0.04 <2e-16 ***
## Prop. Mediated   0.36975      0.23530      0.69 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Sample Size Used: 49001
##
##
## Simulations: 10000

```

```
load("temp/med.FV.RData")
summary(med.FV)
```

```
##
## Causal Mediation Analysis
##
## Nonparametric Bootstrap Confidence Intervals with the Percentile Method
##
## (Inference Conditional on the Covariate Values Specified in 'covariates')
##
##           Estimate 95% CI Lower 95% CI Upper p-value
## ACME           3.44e-03    4.45e-04      0.01 <2e-16 ***
## ADE            -5.29e-03   -1.56e-02      0.00  0.27
## Total Effect   -1.85e-03   -1.07e-02      0.01  0.67
## Prop. Mediated -1.85e+00   -1.20e+01     11.50  0.67
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Sample Size Used: 49001
##
##
## Simulations: 10000
```

```
load("temp/med.PA.RData")
summary(med.PA)
```

```
##
## Causal Mediation Analysis
##
## Nonparametric Bootstrap Confidence Intervals with the Percentile Method
##
## (Inference Conditional on the Covariate Values Specified in 'covariates')
##
##           Estimate 95% CI Lower 95% CI Upper p-value
## ACME           0.0567    0.0422      0.07 <2e-16 ***
## ADE            0.0678    0.0498      0.09 <2e-16 ***
## Total Effect    0.1245    0.1119      0.14 <2e-16 ***
## Prop. Mediated  0.4556    0.3353      0.58 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Sample Size Used: 49001
##
##
## Simulations: 10000
```

```
load("temp/med.NA.RData")
summary(med.NA)
```

```
##
## Causal Mediation Analysis
```

```

##
## Nonparametric Bootstrap Confidence Intervals with the Percentile Method
##
## (Inference Conditional on the Covariate Values Specified in 'covariates')
##
##           Estimate 95% CI Lower 95% CI Upper p-value
## ACME           0.05632      0.04770      0.07 <2e-16 ***
## ADE            0.00522     -0.00718      0.02    0.4
## Total Effect   0.06154      0.05065      0.07 <2e-16 ***
## Prop. Mediated 0.91516      0.74132      1.13 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Sample Size Used: 49001
##
##
## Simulations: 10000

```