# Bioinformatics at the Institute for Animal Health

Michael Watson

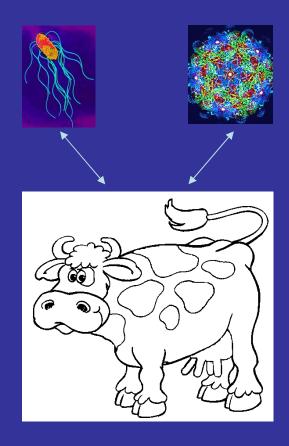
#### Research at the IAH

- Infectious diseases of farm animals
  - Chicken
    - Salmonella
    - Eimeria (coccidiosis)
    - Marek's disease
  - Cow
    - Foot and Mouth
    - Pestivirus
    - Streptococcal infection (mastitis)

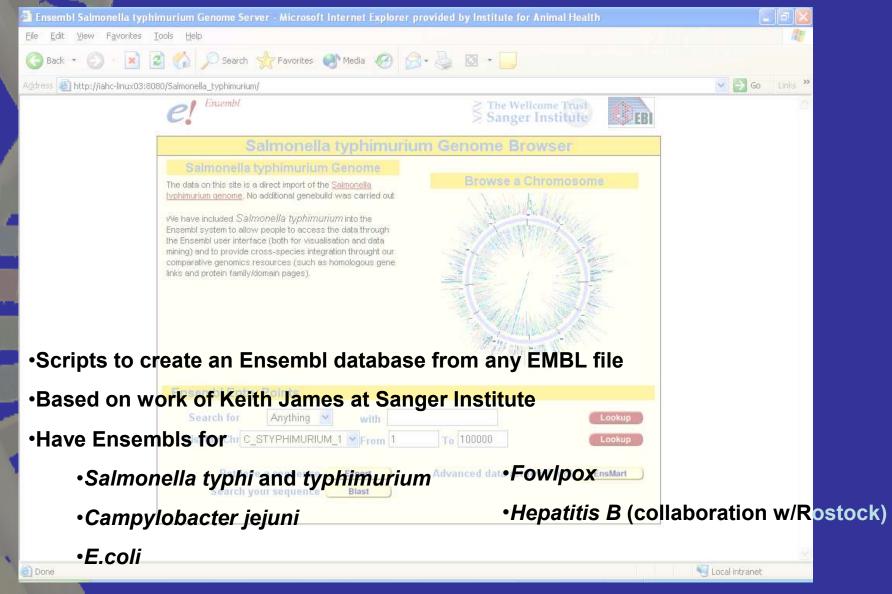
#### Pig

- African Swine Fever
- Foot and Mouth
- Sheep
  - TSEs (scrapie)
  - Orbivirus

#### **Host-Pathogen Interactions**

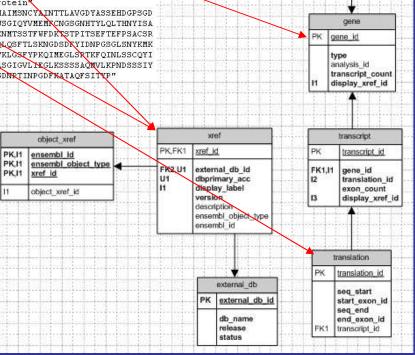


## Ensembl



# **Basic Functionality**

complement (210264..211439) gene FT /locus tag="STY0201" /gene="staG" FT FT /note="synonym: yadC" FT complement (210264..211439) /locus tag="STY0201" FT FT /protein id="NP 454792.1" FT /gene="staG" FT /transl table=11 /note-"Similar to Serratia marcescens fimbria a protein FT precursor smfA SW:YMA SERMA (P13421) (174 aa) fasta FT scores: E(): 0.96, 24.6% id in 142 aa, and to Salmonella FT typhimurium long polar fimbria protein a precursor lpfA FT FT SW:LPFA SALTY (P43660) 178 aa) fasta scores: E(): 2.3, FT 26.3% id in 156 aa Orthologue of E. coli yadC (YADC ECOLI); Fasta hit to NADC ECOLI (412 aa), 34% FT identity in 407 aa overlap" FT /db xref="GI:16759175" FT FT /codon start=1 /product="putative fimbrial protein" FT /translation="MLLKNTTWFAAFFLMMAIMSNCYAINTTLAVGDYASSEHDGPSGD FT FT SVFTDNSHNFGQTIALHKETALRQITVFNUSGIQYVMEMCNGSGNHTYLQLTHNYISA GKSYNGHPLYKTSIPGFYFTIEMTFDOPAENMTSSTFWFDKYSTPITSEFTEFPSACSR TNVYSNLGKLMYGLKIYAYVDBOFAPTEAGLOSFTLSKNGDSDRYIDNPGSGLSNYKMK FT FNLAATGLKAVUPTCSASTISGTNVSGSTVKLGSFYPKQIMEGLSRTKFQINLSSCQYI FT FT NNIEVKLASNNVGTKNTSLLTNNSTSNTKASGIGVLNEGLKSSSSAONVLKPNDSSSIY FT KDTTNNTGDGSPVGSATKSLYFQATLKPDGDNRTINPGDFKATAQFSITYP"



gene description

gene id

description

## Microarrays - BASE

- Open Source
- Free and Easy to set up
- Handles
  - Array Production (LIMS)
  - Experimental Annotation
  - Experimental Results
  - Experimental Analysis
  - MIAME



### **MAGE-ML**

```
<?xml version="1.0" encoding="ISO-8859-1" ?>
 <!DOCTYPE MAGE-ML (View Source for full doctype...)>
- <MAGE-ML identifier="BASE:dev:MAGE-ML">
 - <BioMaterial_package>
   - <Compound_assnlist>
       <Compound identifier="COMP:IAH-C-2" isSolvent="false" name="2" />
       <Compound identifier="COMP:IAH-C-1" isSolvent="false" name="1" />
     </Compound assnlist>
   - <BioMaterial_assnlist>
     - <LabeledExtract identifier="S:IAH-C-labeledextract-2" name="in vivo exp1 Cy3.e1.l1">
      - <MaterialType_assn>
          <OntologyEntry category="MaterialType" value="unknown" />
         </MaterialType_assn>
       - < Treatments_assnlist>
         - <Treatment identifier="T:IAH-C-le2" order="0">
          + < Protocol Applications_assnlist>
          - <Action assn>
              <OntologyEntry category="unknown" value="Labeling" />
            </Action assn>
          + <SourceBioMaterialMeasurements_assnlist>
          </Treatment>
         </Treatments_assnlist>
      + <Labels assnreflist>
       </LabeledExtract>
     - <BioSample identifier="S:IAH-C-biosample-2" name="in vivo exp1 Cy3.e1">
       - <MaterialType_assn>
          <OntologyEntry category="MaterialType" value="unknown" />
         </MaterialType_assn>
      - < Treatments assnlist>
         - <Treatment identifier="T:IAH-C-ex2" order="0">
          A < Protocol Applications_assnlist>
          - <Action_assn>
              <OntologyEntry category="unknown" value="Extraction" />
```

# ArrayPipe: Normalisation Pipeline

- Based on R and Bioconductor
- Setup:
  - Apache

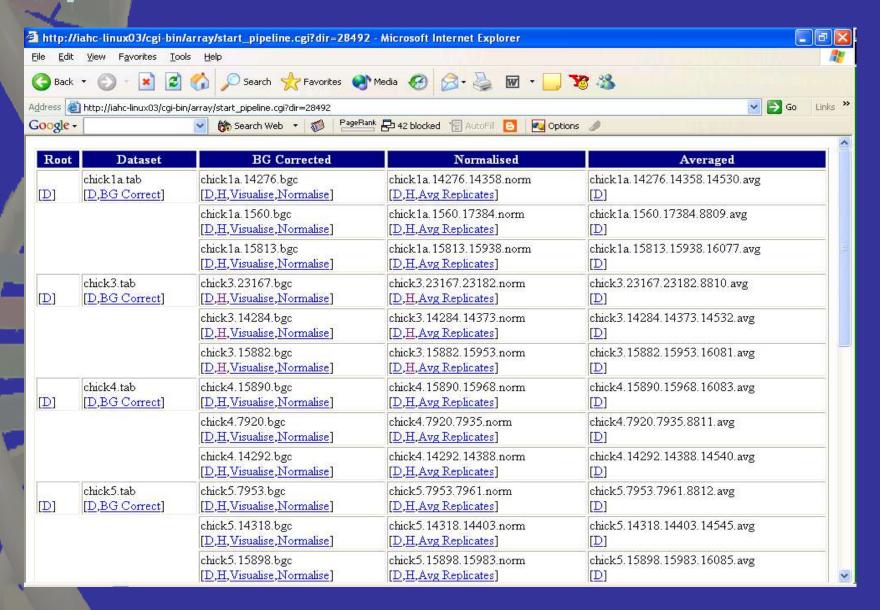


- Perl

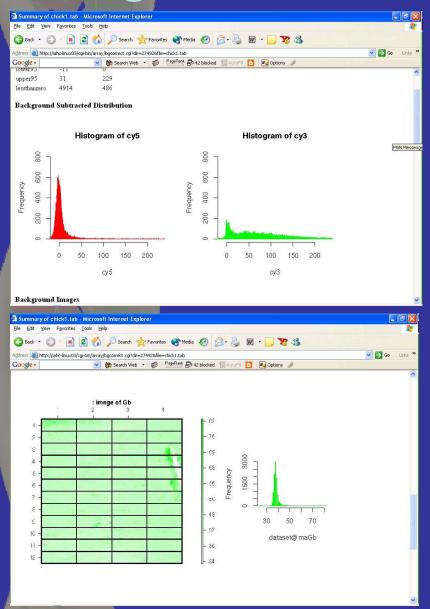


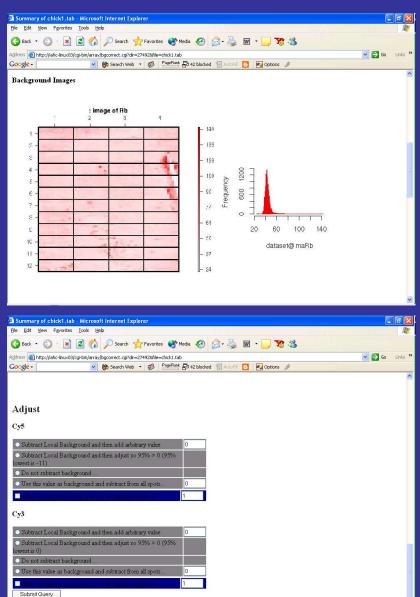
Implements a Flexible user-driven pipeline

#### **Pipeline**



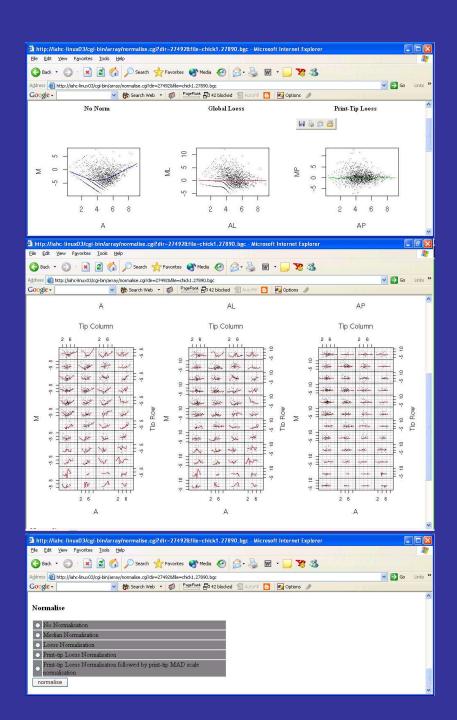
### **Background Subtraction**





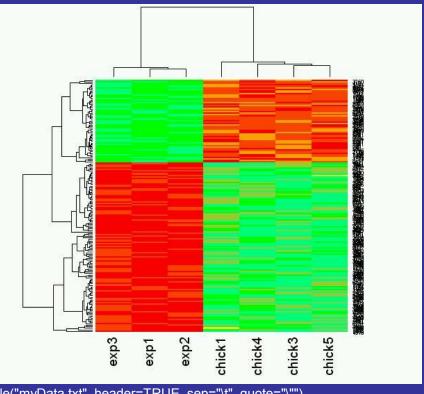
#### Normalisation

- •M v A Plots
  - •Global
  - Print-tip
- Normalisation
  - None
  - Median
  - ·Loess
  - Print-tip Loess
    - Print-tip Loess + MAD



# Microarray Analysis

- Use R/Bioconductor
- Limma
  - Linear models
  - Bayesian Statistics
- Cluster Analysis
- PCA



```
d <- read.table("myData.txt", header=TRUE, sep="\t", quote="\"")
smalld <- d[d$p<0.001,]
dd <- dist(smalld[,3:9], method="euclidean")
h <- hclust(dd, method="complete")
td <- t(smalld[,3:9])
tdd <- dist(td, method="euclidean")
th <- hclust(tdd, method="complete")
col = c("green", "springgreen", "yellowgreen", "yellow", "orange", "orangered", "red")
heatmap(as.matrix(smalld[,3:9]), as.dendrogram(h), as.dendrogram(th), col = mycol)</pre>
```



???