

Notes

David Helekal

September 3, 2020

Contents

1	Introduction	2
2	Methods	4
2.1	Coalescent Preliminaries	4
2.2	Inhomogenous Coalescent	4
2.2.1	Exponential Growth	4
2.3	Coalescent with Local Population Structure	4
3	Results	5
3.1	Implementation Notes	5
3.2	Exponential Growth	5
3.2.1	Phylogeny Simulation	5
3.2.2	MCMC inference	5
3.3	Coalescent with Local Population Structure	5
3.3.1	Phylogeny Simulation	5
3.3.2	MCMC inference	5
4	Discussion	6
5	Bibliography	7

Chapter 1

Introduction

In epidemiology, it is often desired to be able to reconstruct the history of a pathogen population and its structure. The problem of reconstructing the history of a pathogen population can be tackled using phylodynamics. Phylodynamics utilises genomic data to assemble phylogenies, which are then used to infer the population size history. This is possible by viewing a phylogeny as a realisation of a coalescent process, with appropriately rescaled time. This claim can be justified by viewing the coalescent as a Moran model, run backwards in time with the time rate equal to the population size [1].

Within this report we will first introduce the coalescent process for phylodynamic inference, review its inhomogeneous generalisation, and finally introduce the main result of this work, a new model capable of doing local phylodynamic inference, i.e. on a subset of the whole population capable of detecting and modelling clonal expansions.

Clonal expansions are a process in which a particular sub-set of a given bacterial strain undergoes explosive population growth that can be traced back to a particular individual [2]. The presence of clonal expansions in bacterial populations have been of long-standing interest and is implicated in epidemic processes, where an outbreak can be traced to a single ancestor [2, 3, 4, 5]. This often happens when a particular strain or individual obtains a variant of a particular gene that confers evolutionary advantage, for example, antibiotic resistance [6, 7, 5].

The presence of clonal expansions leaves an imprint in the overall population structure of a given bacterial strain, the particular topology associated with this often being referred to as star-like [2, 3]. The problem of detecting hidden population structure corresponding to clonal expansions has become a problem of interest in epidemiology and outbreak surveillance [8].

While methods to detect inhomogeneities in the population structure and size have been of interest since the early days of genetic sequencing [2, 3], the interest in the problem increased with whole genome sequencing becoming more accessible and affordable [6, 9, 10].

Despite the problems of inferring population size from a genealogy and detecting heterogeneities in the population size of the entire population being intrinsically tied, all but one method [8], to our knowledge, rely either on manual detection or indirect detection. We aim to propose a simulation for the formation of clonal expansions in genealogy using the structured coalescent process, and devise a fully bayesian method for joint estimation and detection of relative population size and clonal expansions.

Chapter 2

Methods

2.1 Coalescent Preliminaries

2.2 Inhomogenous Coalescent

2.2.1 Exponential Growth

2.3 Coalescent with Local Population Structure

Chapter 3

Results

3.1 Implementation Notes

3.2 Exponential Growth

3.2.1 Phylogeny Simulation

3.2.2 MCMC inference

3.3 Coalescent with Local Population Structure

3.3.1 Phylogeny Simulation

3.3.2 MCMC inference

Chapter 4

Discussion

Chapter 5

Bibliography

Bibliography

- [1] R. C. Griffiths et al. “Sampling theory for neutral alleles in a varying environment”. In: *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences* 344.1310 (June 29, 1994). Publisher: Royal Society, pp. 403–410. DOI: [10.1098/rstb.1994.0079](https://royalsocietypublishing.org/doi/10.1098/rstb.1994.0079). URL: <https://royalsocietypublishing.org/doi/10.1098/rstb.1994.0079> (visited on 08/28/2020).
- [2] J. M. Smith et al. “How clonal are bacteria?” In: *Proceedings of the National Academy of Sciences* 90.10 (May 15, 1993), pp. 4384–4388. ISSN: 0027-8424, 1091-6490. DOI: [10.1073/pnas.90.10.4384](http://www.pnas.org/cgi/doi/10.1073/pnas.90.10.4384). URL: <http://www.pnas.org/cgi/doi/10.1073/pnas.90.10.4384> (visited on 07/29/2020).
- [3] Brian G. Spratt et al. “Displaying the relatedness among isolates of bacterial species – the eBURST approach”. In: *FEMS Microbiology Letters* 241.2 (2004). _eprint: <https://onlinelibrary.wiley.com/doi/pdf/10.1016/j.femsle.2004.11.015>, pp. 129–134. ISSN: 1574-6968. DOI: [10.1016/j.femsle.2004.11.015](https://onlinelibrary.wiley.com/doi/abs/10.1016/j.femsle.2004.11.015). URL: <https://onlinelibrary.wiley.com/doi/abs/10.1016/j.femsle.2004.11.015> (visited on 07/29/2020).
- [4] Christophe Fraser, William P. Hanage, and Brian G. Spratt. “Neutral microepidemic evolution of bacterial pathogens”. In: *Proceedings of the National Academy of Sciences of the United States of America* 102.6 (Feb. 8, 2005), pp. 1968–1973. ISSN: 0027-8424. DOI: [10.1073/pnas.0406993102](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC548543/). URL: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC548543/> (visited on 07/29/2020).
- [5] Alice Ledda et al. “Re-emergence of methicillin susceptibility in a resistant lineage of *Staphylococcus aureus*”. In: *Journal of Antimicrobial Chemotherapy* 72.5 (May 1, 2017). Publisher: Oxford Academic, pp. 1285–1288. ISSN: 0305-7453. DOI: [10.1093/jac/dkw570](https://academic.oup.com/jac/article/72/5/1285/2930201). URL: <https://academic.oup.com/jac/article/72/5/1285/2930201> (visited on 07/29/2020).
- [6] Matthew T. G. Holden et al. “A genomic portrait of the emergence, evolution, and global spread of a methicillin-resistant *Staphylococcus aureus* pandemic”. In: *Genome Research* 23.4 (Apr. 2013), pp. 653–664. ISSN: 1549-5469. DOI: [10.1101/gr.147710.112](https://doi.org/10.1101/gr.147710.112).
- [7] Li-Yang Hsu et al. “Evolutionary dynamics of methicillin-resistant *Staphylococcus aureus* within a healthcare system”. In: *Genome Biology* 16.1

- (Apr. 23, 2015), p. 81. ISSN: 1465-6906. DOI: [10.1186/s13059-015-0643-z](https://doi.org/10.1186/s13059-015-0643-z). URL: <https://doi.org/10.1186/s13059-015-0643-z> (visited on 07/29/2020).
- [8] Erik M. Volz et al. “Identification of Hidden Population Structure in Time-Scaled Phylogenies”. In: *Systematic Biology* (). DOI: [10.1093/sysbio/syaa009](https://academic.oup.com/sysbio/advance-article/doi/10.1093/sysbio/syaa009/5734655). URL: <https://academic.oup.com/sysbio/advance-article/doi/10.1093/sysbio/syaa009/5734655> (visited on 07/01/2020).
 - [9] Bethany L. Dearlove and Simon D. W. Frost. “Measuring Asymmetry in Time-Stamped Phylogenies”. In: *PLOS Computational Biology* 11.7 (July 6, 2015). Publisher: Public Library of Science, e1004312. ISSN: 1553-7358. DOI: [10.1371/journal.pcbi.1004312](https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1004312). URL: <https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1004312> (visited on 07/29/2020).
 - [10] Vegard Eldholm et al. “Four decades of transmission of a multidrug-resistant *Mycobacterium tuberculosis* outbreak strain”. In: *Nature Communications* 6.1 (May 11, 2015). Number: 1 Publisher: Nature Publishing Group, p. 7119. ISSN: 2041-1723. DOI: [10.1038/ncomms8119](https://www.nature.com/articles/ncomms8119). URL: <https://www.nature.com/articles/ncomms8119> (visited on 07/29/2020).