

Reporting

Report Date: Jul 28th, 2025
Receipt Date: Jul 18th, 2025
Collection Date: Jul 17th, 2025
Specimen: Blood
Status: FINAL

Sameer Desai

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Additional Recipient: N/A



Complete Tumor Response Map on page 4

Summary of Genomic & Epigenetic Biomarkers with Associated Treatment Options

✓ Approved in indication
⌚ Approved in other indication
✗ Lack of response

DETECTED ALTERATION(S) / BIOMARKER(S)	ASSOCIATED FDA-APPROVED THERAPIES	CLINICAL TRIALS (SEE PAGE 5)	% CFDNA OR COPY NUMBER
PTEN Loss (Single Copy Deletion)	⌚ Capivasertib	Yes	DETECTED
TP53 Y205C	None	Yes	22.3%
ATM Loss (Single Copy Deletion)	None	Yes	DETECTED
PALB2 Loss (Single Copy Deletion)	None	Yes	DETECTED
RB1 Biallelic Loss (Homozygous Deletion)	None	No	DETECTED

Variants of uncertain clinical significance listed on following pages.

Additional Biomarkers

Tumor Mutational Burden (TMB)	5.69 mut/Mb
MSI-High	NOT DETECTED
Confidence Patient is Biomarker Negative	≥99% - Likelihood that the patient is free of biomarkers associated with FDA-approved therapies.
Tumor Fraction[#]	42.5% - Based on 1099 patient-specific biomarkers

[#] Tumor fraction is defined as the proportion of tumor molecules present in the cfDNA within the submitted specimen and is based on epigenomic signals.

Patient Biomarkers

Pharmacogenomics

CYP2D6 *1/*41 Normal metabolizer

This genotype results in normal CYP2D6 enzyme activity. Activity score: 1.25

TPMT *1/*3C Intermediate metabolizer

This genotype results in an intermediate metabolizer phenotype and may require dose reduction when prescribed azathioprine, 6-mercaptopurine (6-MP), and thioguanine. See guidelines (pharmgkb.org) for additional information.

UGT1A1 *1/*28 Intermediate metabolizer

This genotype results in intermediate UGT1A1 enzyme activity. Irinotecan is expected to be metabolized at a level between normal and poor metabolizers leading to increased toxicity, such as neutropenia or hyperbilirubinemia. See guidelines (pharmgkb.org) for additional information.

DPYD, HLA-B No abnormal variant detected in reportable alleles (see Interpretations)

Human Leukocyte Antigen (HLA) Genotyping

HLA-A: A*23:01, A*68:01

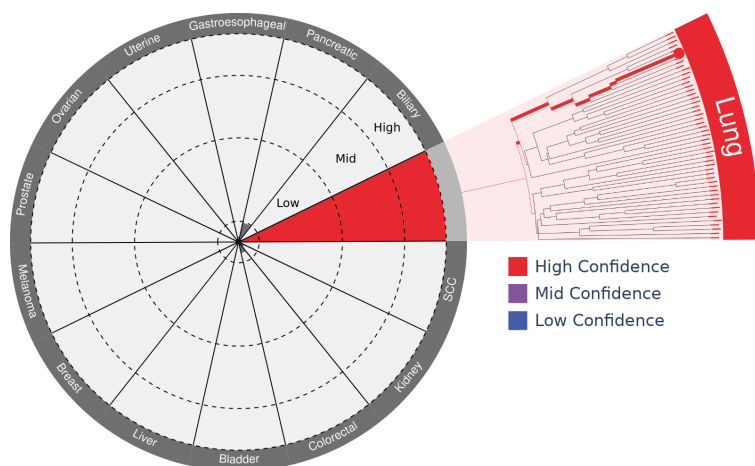
HLA-B: B*18:01, B*37:01

HLA-C: C*06:02, C*07:01

Results reflect evaluable alleles in HLA-A, -B, and -C. HLA Class I results are intended as an aid for oncology therapy selection and/or clinical trial matching and are not validated for transplant allele typing.

Molecular Tumor Type

These findings are not a diagnosis of cancer and should be interpreted in the context of other clinical information, by a qualified medical professional.



Molecular Tumor Typing

Primary Prediction: **Lung, 100%**

Percentage represents confidence level in tumor type prediction

Molecular Lung Subtype

Lung adenocarcinoma: <10%

Lung squamous cell carcinoma: <10%

Small cell lung carcinoma: >90%

Percentage represents proportional contribution of each subtype

A molecular prediction of tumor type and/or subtype based on DNA methylation signatures is provided as a supplement and not a replacement of standard clinical and pathologic evaluation. These results are not considered diagnostic and should be interpreted in the context of other clinical information by a qualified medical professional. Confirmatory microscopic evaluation of the tumor may be considered if clinically indicated.

Comments

Reported by: SYS

Variants of Uncertain Clinical Significance

ATM E2290D (3.1%)	GSK3B G34Afs*4 (2.6%)	LTK L684F (13.4%)	RAF1 K493Q (24.3%)
DNMT3B H677Y (0.3%)	HRAS G151V (0.3%)	NOTCH3 D2010D (0.2%)	
FZD8 E8E (0.3%)	JAK2 Q175P (19.1%)	PTPRD A945A (2.2%)	

Variants of Uncertain Clinical Significance: The functional consequences and/or clinical significance of these alterations is uncertain. Relevance of therapies targeting these alterations is unknown.

Variants of Potential Clonal Hematopoiesis

DNMT3A R771Q (0.5%)	KMT2C Y2218* (0.9%)	SF3B1 K666N (0.5%)
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Variants are categorized as of potential clonal hematopoietic origin using a proprietary algorithm that discriminates variants of non-solid-tumor origin, the majority of which are likely to arise from hematopoietic sources. The specificity of the test for annotating clonal hematopoiesis in cfDNA is >98%.

We evaluated this sample for MSI, TMB, and alterations in 744 genes, including the following guideline-recommended genes for NSCLC

ALK **BRAF** **EGFR** **ERBB2 (HER2)** **KRAS** **MET** **NRG1** **NTRK** **RET** **ROS1**

Guardant360 Tumor Response Map

The Guardant360 Tumor Response Map illustrates the variant allele fraction (% cfDNA) of observed somatic variants at each sample submission. Amplifications are not plotted, and only the first and last five test dates are plotted. Please see the Physician Portal (portal.guardanthealth.com) for the Tumor Response Map with all test dates.

Highest Variant
Allele Fraction

22.3%



JUL-17-2025

DETECTED ALTERATION(S) / BIOMARKER(S)

% CFDNA OR COPY NUMBER

Tumor Fraction

42.5%

TP53 Y205C

22.3%

ATM Loss (Single Copy Deletion)

DETECTED

Deletions not graphed above

Plasma Copy Number 1.7

DNMT3A R771Q

0.5%

Variant of Potential Clonal Hematopoiesis §

KMT2C Y2218*

0.9%

Variant of Potential Clonal Hematopoiesis §

PALB2 Loss (Single Copy Deletion)

DETECTED

Deletions not graphed above

Plasma Copy Number 1.6

PTEN Loss (Single Copy Deletion)

DETECTED

Deletions not graphed above

Plasma Copy Number 1.6

RB1 Biallelic Loss (Homozygous Deletion)

DETECTED

Deletions not graphed above

Plasma Copy Number 1.2

SF3B1 K666N

0.5%

Variant of Potential Clonal Hematopoiesis §

The table above annotates the variant allele fraction (% cfDNA) detected in this sample, listed in descending order.

§ See definitions section for more detail

Available Clinical Trials (within the same state as the ordering physician)

There may be additional trials not listed here. Visit: portal.guardanthealth.com or email clientservices@guardanthealth.com with A1473811 in the subject line of the email, for additional trials.

ALTERATION	TRIAL ID / CONTACT	TITLE	PHASE	SITE(S)
PTEN Loss (Single Copy Deletion)	NCT03994796 Priscilla Brastianos, MD,pbrastianos@partners.org,617-724-1074	Genetic Testing in Guiding Treatment for Patients With Brain Metastases	Phase 2	Neptune, NJ Summit, NJ Pennington, NJ
	NCT05238922 Incyte Corporation Call Center (US), medinfo@incyte.com,1.855.463.3463	Study of INCB123667 in Subjects With Advanced Solid Tumors	Phase 1	Hackensack, NJ
	NCT05932862 Exelixis Clinical Trials,druginfo@exelixis.com,1-888-393-5494)	A Phase 1 Study of XL309 (ISM3091) Alone and in Combination in Patients With Advanced Solid Tumors	Phase 1	New Brunswick, NJ
	Visit portal.guardanthealth.com for trials not within the same state as the physician's office			
ATM Loss (Single Copy Deletion)	NCT05238922 Incyte Corporation Call Center (US), medinfo@incyte.com,1.855.463.3463	Study of INCB123667 in Subjects With Advanced Solid Tumors	Phase 1	Hackensack, NJ
	NCT05566574 Nancy Lee, MD,leen2@mskcc.org,212-639-3341	A Study of RP-3500 in Combination With Standard Radiation Therapy in People With Solid Tumor Cancer	Phase 1 /Phase 2	Basking Ridge, NJ Montvale, NJ Middletown, NJ
	NCT05932862 Exelixis Clinical Trials,druginfo@exelixis.com,1-888-393-5494)	A Phase 1 Study of XL309 (ISM3091) Alone and in Combination in Patients With Advanced Solid Tumors	Phase 1	New Brunswick, NJ
	Visit portal.guardanthealth.com for trials not within the same state as the physician's office			
PALB2 Loss (Single Copy Deletion)	NCT05238922 Incyte Corporation Call Center (US), medinfo@incyte.com,1.855.463.3463	Study of INCB123667 in Subjects With Advanced Solid Tumors	Phase 1	Hackensack, NJ
	NCT05932862 Exelixis Clinical Trials,druginfo@exelixis.com,1-888-393-5494)	A Phase 1 Study of XL309 (ISM3091) Alone and in Combination in Patients With Advanced Solid Tumors	Phase 1	New Brunswick, NJ
	Visit portal.guardanthealth.com for trials not within the same state as the physician's office			
TP53 Y205C	Visit portal.guardanthealth.com for trials not within the same state as the physician's office			
More clinical trial options available at portal.guardanthealth.com				

Interpretation

Somatic Alterations: Guardant360 detects somatic alterations in circulating cell-free DNA isolated from a patient's blood specimen in the genes listed in Table 1. These genomic alterations are cancer-associated somatic variants, some of which have been associated with either increased or reduced clinical response to specific treatments. The percentage of altered cell-free DNA circulating (% cfDNA) in blood is related to the unique tumor biology of each patient. Factors that may affect the % cfDNA of detected somatic alterations include tumor growth, turn over, size, heterogeneity, vascularization, disease progression, and treatment.

Amplification: Guardant360 detects amplifications in the genes listed in Table 1. Gene amplification results in increased copies of the gene present in the cfDNA. The reported absolute copy number value represents the average copy number for the detected gene that was detected in circulating cfDNA. With the exception of sex-linked genes such as *AR*, 2 copies are expected in the absence of amplification. As the absolute number of copies in circulation is dependent on both tumor fraction and the magnitude of the tumor amplification, amplifications are reported on a semi-quantitative scale.

Amplifications are reported in two levels:

Medium (++) : Amplification magnitude is below the 50th percentile of amplifications detected by Guardant360.

High (+++) : Amplification magnitude is above the 50th percentile.

Unlike tissue based gene amplification tests (e.g. IHC or FISH), Guardant360 assesses the total representation of a given gene in all circulating cell-free DNA present in the patient's blood sample including material derived from the tumor and healthy tissue alike. As such, the absolute level of amplification present in the blood depends both on the tumor-derived cfDNA content and on the degree of amplification within that fraction and cannot be inferred from bulk cfDNA interrogation. For example, a positive Guardant360 test could represent a small population of cells with extremely high levels of the detected gene amplification. Alternatively, it could represent a large population of cells with low to medium levels of the detected gene amplifications. The exact correlation between amplification detected by Guardant360 compared to IHC or FISH and how each test differentially guides patient management is an area of active investigation.

Biomarker Negative: Guardant360 assesses the level of certainty (confidence score) that the sample is negative for clonal/driver FDA-approved genomic biomarkers in that tumor type (see <https://www.guardantcomplete.com/hcp/solutions/guardant360> for a list of eligible biomarkers in lung and colorectal cancers). It is intended to support clinical decision making regarding further diagnostic procedures. Therapy annotations and wildtype status are provided only for colorectal samples with confidence scores >95%. The algorithm is based on biomarker prevalence from the literature, modified by the specific test result, based on analytical test characteristics and a large tissue-plasma concordance training set. For lung cancer and colorectal cancer samples that are determined to be FDA-approved biomarker negative on Guardant360 with confidence scores >90%, the concordance with tissue, i.e. the predictive value of the negative result, is 96%. As subclonal alterations may exist at any variant allele fraction, the algorithm result applies only to clonal/driver alterations. Samples with confidence scores <90% have >10% chance for being positive for relevant biomarkers in the tumor; therefore consideration for further testing by tissue is recommended as clinically indicated.

Clonal Hematopoiesis (CH): All SNVs and Indels detected on the Guardant 360 assay will be evaluated for the likelihood that they are derived from hematopoietic sources. The categorization of these variants as potential clonal hematopoietic origin by an algorithm is not intended to be used for the diagnosis of a hematolymphoid malignancy. The clinical significance of these variants requires evaluation of other clinical and laboratory parameters, such as peripheral blood counts, patient age, and treatment history. Non-tumor derived variants may arise from hematolymphoid tissues, such as those associated with clonal hematopoiesis, and/or include variants from clonal processes in other organ sites.

Deletion (Del): Guardant360 detects short deletions in exons of certain genes (see Table 1), including potential splice site-disrupting events.

Fusion: Fusion events are gene rearrangements that fuse two previously distinct genes into a single genetic unit. Guardant360 detects fusions in the genes listed in Table 1.

Genomic Instability Status (GIS): Homologous recombination deficiency (HRD) is characterized by inability to accurately repair double-stranded breaks in DNA and sensitivity to DNA-damaging agents such as platinum-based chemotherapy and poly-ADP ribose polymerase (PARP) inhibitors. HRD status can be determined by testing for either deleterious mutations in homologous recombination repair (HRR) genes or genomic instability (PMID: 37769224). Guardant360 reports a qualitative genomic instability status (GIS) in prostate, breast, ovarian, and pancreatic cancers based on a probabilistic prediction measured by a combination of genomic instability biomarkers, including loss of heterozygosity, large-scale state transitions, telomere allelic imbalance, and specific SNV signatures. The positive agreement of GIS with HRD status as assessed by mutations in HRR genes is 79.5%, and the negative agreement is 97.6% . A "Not Detected" result indicates that signal in the plasma sample did not exceed a predefined threshold, and does preclude genomic instability in the tumor. A "Not evaluable" result is returned for samples with large discordances in observed and expected copy numbers or germline variant allele fraction distributions, indicating low confidence of background signal assessment and potentially inconclusive results.

Homozygous Deletion: Homozygous deletion results when both copies of a gene are lost.

Human Leukocyte Antigen: All samples are evaluated for germline MHC Class I genes in the *HLA-A*, *HLA-B*, and *HLA-C* loci at two-field high resolution (IPD-IMGT/HLA release 3.44.0) from cfDNA. The accuracy of Guardant360 for HLA detection compared to orthogonal methods is 97.3%. The assay cannot differentiate between loss of heterozygosity and homozygosity when only one allele is detected in samples with very high tumor fractions where the majority of cfDNA is tumor derived. A Not Evaluable result indicates ambiguous support for multiple alleles. HLA alleotyping results are intended as a supplement to standard of care in therapy selection and/or an aid in clinical trial eligibility assessment, not a replacement for standard methodologies in immunohistochemistry testing (PMID 34551229 & 37328642).

Indel: Guardant360 detects short insertions or deletions in the exons of certain genes (see Table 1). This type of variant represents frameshift, in-frame and complex alterations comprising both an insertion and a deletion. These variants can result in the substitution of one or more amino acids with the reading frame preserved (in-frame), or disrupt the reading frame (frameshift) leading to a premature stop codon.

Insertion (Ins): Guardant360 detects short insertions in exons of certain genes (see Table 1), including potential splice site-disrupting events.

MET exon 14 skipping: This type of genomic alteration is predicted to result in the skipping of *MET* exon 14 during RNA splicing, resulting in oncogenic activation of the *MET* protein.

Molecular Breast Subtype: Guardant360 evaluates the methylation profile of the sample across target panel regions to infer tumor-associated DNA methylation changes. In breast cancer patients, distinct cfDNA methylation signatures characterize hormone receptor (HR), HER2-positive, and triple negative breast cancer (TNBC) subtypes. Guardant360 reports the level of certainty (confidence score) for the presence of these subtypes, when the tumor fraction in blood is >0.5%. The accuracy for detecting HR, HER2, and TNBC status compared to standard of care methodologies such as immunohistochemistry (IHC) and in-situ hybridization (ISH) is 85.7%, 86.3%, and 91.9%, respectively. Detection of the HER2-positive subtype was developed and validated in cases with high HER2 expression (IHC 3+, IHC 2+, or ISH+ in paired tissue); the test has not been validated for detection of HER2-low or HER2-ultralow disease. When the confidence score for any subtype is reported as <10%, the signal in plasma is below the detection limit of the assay; this does not preclude the presence of the subtype in the patient's tumor. The molecular breast tumor subtyping provided by Guardant360 is intended to be a supplement rather than replacement for standard of care methodologies. If clinically indicated, confirmatory testing with a validated standard assay is recommended.

Molecular Lung Subtype: Guardant360 evaluates the methylation profile of a sample across target panel regions to infer tumor-associated DNA methylation changes. In lung cancer patients, histologic subtypes (adenocarcinoma (LUAD), squamous cell carcinoma (LUSC), and small cell carcinoma (SCLC)) are associated with distinct cfDNA methylation signatures (PMID: 30317601, PMID: 35941262). Guardant360 assesses the relative proportional contribution of each lung subtype to the unique methylation signature to the tumor, when the tumor fraction in blood is >0.1%. The accuracy for detecting the predominant subtype for LUAD, LUSC and SCLC is 93.44%, 83.51%, 81.08%, respectively. The estimated proportions for each subtype are provided as an annotation and not intended to be a quantitative output. When the estimated proportion for any subtype is reported as <10%, the signal in plasma is below the detection limit of the assay; this does not preclude the presence of that subtype in the tumor. This prediction is intended to supplement histologic characterization and confirmatory tissue evaluation may be warranted if clinically indicated.

Molecular Tumor Type: Molecular Tumor Type (MTT) is a prediction of tumor type based on DNA methylation signatures. Primary and secondary MTT predictions, along with an associated confidence score, are provided as an annotation to supplement standard clinical and pathologic diagnostic evaluation, particularly for adjudication of carcinoma of unknown primary (CUP). The associated confidence scores represent the level of certainty of the MTT prediction: 0.1-0.5 (Low), 0.5-0.8 (Mid), 0.8-1.0 (High). The MTT algorithm has demonstrated 88.5%-100% agreement between MTT predictions and clinicopathologic diagnosis depending on tumor type. MTT predictions are not provided if the confidence score is below predefined thresholds or the patient's tumor type is not supported by the algorithm (see <https://www.guardantcomplete.com/hcp/solutions/guardant360>). The presence of two CSO predictions does not necessarily indicate that multiple primary cancers are present.

Nonsense Mutation: A point mutation that results in a premature stop gain.

Pharmacogenomics: All samples are evaluated for germline variants in Tier 1 and select Tier 2 alleles that modulate toxicity/sensitivity to chemotherapeutic agents in *CYP2D6*, *DPYD*, *TPMT*, *UGT1A1*, and the HLA allele *HLA-B*57:01* (*for more information including reportable alleles and therapy indications see <https://www.guardantcomplete.com/hcp/solutions/guardant360>). Germline allelotype is inferred based on cfDNA sequencing data and metabolizer status is a prediction based on the diplotype. Interpretations are limited to oncologic therapies. Activity scores are calculated using CPIC guidelines. The accuracy of Guardant360 for Pharmacogenomics allelotyping compared to PCR-based methods is 96.3%. A Not Evaluable result indicates that the copy number of the gene cannot be determined and/or there is ambiguous support for multiple alleles. As the test does not detect all variants, *1 represents the absence of a detectable variant. *CYP2D6* hybrid and duplicate alleles will be represented with 3 values (*1/*2/*3) and complex alleles are not reported. Only the HLA allele *HLA-B*57:01* associated with increased risk of drug hypersensitivity will be reported if detected. Drug response can be influenced by other clinical factors and therapy decisions should take these variables into consideration. Functional confirmation is recommended when conflicting evidence exists in the literature.

Promoter Methylation: Guardant360 detects promoter methylation in the genes listed in Table 1. Hypermethylation of normally unmethylated CpG islands located in gene promoters correlates with transcriptional repression that can serve as an alternative to coding region mutations for inactivation of tumor suppressor genes (Baylin et al., 1997). DNA methylation of key CG dinucleotides within the promoter blocks accessibility of transcription factors to their binding motifs, thereby silencing gene expression. DNA hypermethylation occurs early in tumorigenesis and typically increases with tumor progression (Ehrlich 2019).

Rearrangement: Guardant360 detects rearrangements in the genes listed in Table 1. Rearrangement refers to large genomic events that result in single or multi-exon deletions (>3Kb in size) or gene fusions.

Reversion: Reversions are secondary mutations, often small indels, that convert an initial frameshift mutation into an in-frame internal deletion that still produces a partly functional protein product.

Single Copy Deletion + Co-Occurring Loss of Function (LOF) Variant: Occurs when a single copy of a gene is lost due to whole gene deletion in the presence of a loss-of-function variant in the same gene.

Single Copy Deletion: Occurs when a single copy of a gene is lost due to a whole gene deletion.

Somatic Alterations Not Detected (ND): In some cases, Guardant360 does not detect any somatic alterations. Somatic alterations may be present that are below the limit of detection of this test. Certain sample or variant characteristics may result in reduced analytic sensitivity. The absence of detectable somatic alterations in circulating cell-free DNA does not preclude the presence of somatic alterations in the tumor.

Splice Site: Splice site variants disrupt the donor and/or acceptor splice site(s), leading to abnormal mRNA splicing and altered protein levels and/or function.

Synonymous Alteration: This sequence change does not alter the amino acid at this position and is unlikely to be a therapeutic target. Clinical correlation is advised.

Tumor Mutational Burden (TMB): Checkpoint inhibition has been FDA-approved for patients who have no satisfactory alternative treatment option with a tissue TMB score of 10 mut/Mb or higher. In a large clinical study of blood TMB in patients with newly diagnosed non-small cell lung cancer (NSCLC) using Guardant360, Guardant360 TMB score of 16 mut/Mb, 76th percentile, correlates with a tissue TMB score of 10 mut/Mb (Rizvi, N.A., et al. JAMA Oncology, 2020). The distribution of tissue TMB scores has been shown to differ across tumor types (Samstein, R.M, et al. Nature Genetics, 2019). Similarly, in a landscape analysis of Guardant360 TMB, 80th percentile corresponds to the following TMB scores: NSCLC - 20.2 mut/Mb; colorectal cancer - 20.1 mut/Mb; breast cancer- 15.3 mut/Mb; prostate cancer - 13.4 mut/Mb; pancreatic cancer - 11.4 mut/Mb; head and neck squamous cell cancer (HNSCC)- 17.4 mut/Mb; cholangiocarcinoma - 10.5 mut/Mb.

Variants of Uncertain Clinical Significance: The functional consequences and/or clinical significance of alterations are unknown. Relevance of therapies targeting these alterations is uncertain.

Viral Status: Guardant360 targets viral genome sequences from *EBV* and 14 strains of high-risk HPV (HPV16, HPV18, HPV31, HPV33, HPV35, HPV39, HPV45, HPV51, HPV52, HPV56, HPV58, HPV59, HPV66, and HPV68). The reportable range for *EBV* is ≥10.64 normalized viral genome copies, and the reportable range for HPV is ≥3.6 normalized viral genome copies. The intended use population for *EBV* viral genome detection are patients with advanced-stage gastric and nasopharyngeal carcinoma, and for HPV is patients with advanced-stage HPV-mediated malignancies in the head and neck and lower anogenital tract. Both viruses are reported in carcinoma of unknown primary. The sensitivity of Guardant360 for viral detection compared to standard of care methods is 93.6%, and the specificity is 96.6%. The test cannot distinguish between infectious and neoplastic sources of viral DNA and is not a substitute for diagnostic tissue-based testing for newly-diagnosed cancers.

About the Test

The Guardant360 assay was developed and its performance characteristics were determined by Guardant Health, Inc. This test has not been cleared or approved by the U.S. Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary. This test may be used for clinical purposes and should not be regarded as investigational or for research only. Guardant Health's clinical reference laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high complexity clinical laboratory testing. The laboratory report should be interpreted in the context of other clinical information and laboratory, pathology, and imaging studies by a qualified medical professional prior to initiating or changing a patient's treatment plan. The selection of any, all, or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) is entirely at the discretion of the treating medical professional. Drug and trial information are based on the diagnosis written on the submitted test request form; this information is not based on any supplemental information provided by the requesting medical professional, including pathology reports or other molecular studies. Some drugs listed in this report may not be approved or cleared by the FDA for the indicated use. Guardant Health makes no endorsement, expressed or implied, of any product, physician, or procedure contained in this report. This report makes no promises or guarantees that a particular medication will affect (or not affect) the clinical outcome of any patient.

Testing Performed at: Guardant Health

Laboratory Director: Martina Lefterova, MD PhD | CLIA ID: 05D2070300 |
CAP #: 8765297 | 505 Penobscot Drive, Redwood City, CA, 94063, USA

Method and Limitations

Guardant360 sequences 744 cancer-associated genes to identify somatic and germline alterations. Cell-free DNA (cfDNA) is extracted from plasma, partitioned based on methylation status, enriched for targeted regions, and sequenced using the Illumina platform and hg19 as the reference genome. All exons are sequenced in some genes; only clinically significant exons are sequenced in other genes. The types of genomic alterations detected by Guardant360 include single nucleotide variants (SNVs), gene amplifications, fusions/rearrangements, short insertions/deletions (indels, longest detected, 87 base pairs), copy number deletions, promoter methylation and splice site disrupting events (see Table 1). Microsatellite Instability (MSI) is assessed for all cancer types by evaluating somatic changes in the length of repetitive sequences on the Guardant360 panel. A "Not Detected" result in samples where the highest % cfDNA is < 0.2% is an inconclusive result because it does not preclude MSI-High status in tissue. Tumor mutational burden (TMB) score is calculated for all cancer types from somatic SNVs and indels in exons of 497 genes detected in cfDNA, followed by adjusting for tumor shedding levels and the size of the panel. A "Not Evaluable" result is an inconclusive result in samples where the evidence of tumor shedding is insufficient and it does not preclude TMB-High status in tissue. Promoter methylation status is assessed for all cancer types by calculating the % cfDNA methylated molecules overlapping defined promoter regions of 47 genes and is reported as Detected or Not Detected based on the established sensitivity. Promoter methylation may not always be associated with gene inactivation; orthogonal confirmation and clinical correlation are recommended as indicated. Methylation-based tumor fraction is estimated in all cancer types by evaluating multiple patient-specific differentially methylated regions associated with cancer. The quantifiable range for tumor fraction ranges from 0.05% to 90%. Tumor fractions less than 0.05% are reported as <0.05%, those higher than 90% are indicated as >90%. All SNVs and Indels detected on the Guardant360 assay will be evaluated for the likelihood that they are derived from hematopoietic sources by a proprietary algorithm that identifies variants of nonsolid-tumor origin that may originate from hematopoietic or non-hematopoietic clonal processes. The test is not intended to be used for the diagnosis of hematolymphoid malignancy. Certain sample or variant characteristics, such as low cfDNA concentration, may result in reduced analytic sensitivity. Guardant360 cannot discern the source of circulating cfDNA, and for some variants in the range of ~40 to 60% cfDNA, the test cannot easily distinguish germline variants from somatic alterations. Guardant360 is not validated for the detection of germline or de novo variants that are associated with hereditary cancer risk. Tissue genotyping should be considered when plasma genotyping is negative, if clinically appropriate.

Additional information is available

Therapeutic annotations are based on publicly available information. A more detailed Guardant360 report (including Additional Clinical Trials, Detailed Therapy Results, Relevance of Detected Alterations, and References) is available through our online portal at portal.guardanthealth.com. To set up an account, contact Client Services at [855.698.8887](tel:855.698.8887) or clientservices@guardanthealth.com.

If you would like to receive this additional information for this report, or with every Guardant360 report, contact Client Services.

Table 1: Genes on the Guardant360 Panel

Guardant360 reports single nucleotide variants, splice site mutations, and insertion and deletion variants (indels) in the genes listed below and reports other variant types in select genes as indicated.

ABCB1	BRD2	CRTC1	EML4	FGF7	HLA-C ^h §	LGR6	NF1 [^]	PIK3R2	RB1 [^] #	SMARCE1	TP63
ABL1	BRD3	CSF1R	EMSY	FGF8	HNF1A	LIG1	NF2 [^]	PIK3R3	RBBP6	SMC1A	TP73
ABL2	BRD4	CSF3R	EP300	FGF9	HNRNPDL	LIG4	NFE2L2	PIM1	RBM10	SMC3	TPMT ^P
ABRAXAS1	BRIP1 [^]	CTC1	EPCAM	FGFR1 [†] #	HOXB13	LMO1	NFKBIA	PIN1	RBMX	SMO	TRAF2
ACVR1	BSG	CTCF	EPHA3	FGFR2 [†] #	HPV (high-risk) [~] §	LRP1B	NHEJ1	PKM	RECQL	SNCAIP	TRAF3
ACVR1B	BTG1	CTLA4	EPHA5	FGFR3 [†] #	HRAS	LRP2	NKX2-1	PLCG2	RECQL4	SOC1	TRAF7
ACVR2A	BTG2	CTNNA1	EPHA7	FGFR4	HSD3B1	LRP5	NOTCH1	PLEKHS1	RET [†] #	SOC3	TRIM24
ADARB2	BTK	CTNNA1	EPHB1	FH	HSP90AA1	LRP6	NOTCH2	PLRG1	REV3L	SOS1	TRIP13
ADGRA2	BUB1B	CUL3	ERBB2 [†]	FLCN	ICOSLG	LTK	NOTCH3	PMS1	RGS1	SOX10	TSC1
ADGRG4	C9orf78	CUL4A	ERBB3	FLT1	ID3	LYN	NOTCH4	PMS2 [^]	RHEB	SOX17	TSC2 [^]
AFDN	CALR	CUX1	ERBB4	FLT3	IDH1	LZTR1	NOVA1	POLA1	RHOA	SOX2	TSHR
AGGF1	CARD11	CWC22	ERCC1	FLT4	IDH2	MAD2L2	NPM1	POLD1	RHOB	SOX9	TSHZ2
AIP	CASP8	CXCR4	ERCC2	FOXO1	IDO1	MALT1	NPRL2	POLE [^]	RICTOR	SPEN	TYMP
AKT1	CASR	CYLD	ERCC3	FOXO2	IFNG	MAP2K1	NPRL3	POLH	RIF1	SPOP	TYMS
AKT1S1	CAV1	CYP17A1	ERCC4	FOXO1	IFNGR1	MAP2K2	NRAS	POLQ	RILPL1	SRC	TYRO3
AKT2	CBFB	CYP19A1	ERCC5	FOXP1	IFNGR2	MAP2K4	NRG1 [†] #	POT1	RIT1	SRSF2	U2AF1
AKT3	CBL	CYP2C19	ERCC6	FOXP2	IFNW1	MAP3K1	NSD1	POU2F2	RNASEH2B	SRY	UBE2T
ALB	CBLB	CYP2D6 ^P §	ERCC6L2	FUBP1	IGF1	MAP3K13	NSD2	PPARG	RNF43	SS18	UGT1A1 ^P
ALK [†] #	CCAR1	CYP3A4	ERCC8	FUBP3	IGF1R	MAP4K3	NSD3	PPIG	ROBO1	STAG2	UIMC1
ALOX12B	CCN6	DAXX	EREG	FUS	IGF2	MAPK1	NSRP1	PPM1D	ROBO2	STAT1	ULBP1
ALOX15B	CCNA2	DCUN1D1	ERF	FYN	IGF2BP3	MAPK3	NTHL1	PPP2CA	ROS1 [†] #	STAT3	ULBP3
ALOX5	CCNB1	DDIT3	ERG	FZD1	IGF2R	MAPKAP1	NTRK1 [†] #	PPP2R1A	RPA1	STAT4	USP28
AMER1	CCND1 [†]	DDR1	ERRF1	FZD10	IKBKE	MARK2	NTRK2 [†] #	PPP2R2A	RPS27A	STK11 [^] #	USP7
APC [^]	CCND2 [†]	DDR2	ESR1 [†] #	FZD2	IKZF1	MAX	NTRK3 [†] #	PPP3CA	RPS6KA3	STK19	USP9X
APEX1	CCND3	DDX17	ETS1	FZD3	IL1R1	MCL1	NUMA1	PPP6C	RPS6KB1	STK40	VEGFA
APLN	CCNE1 [†]	DDX18	ETV1	FZD4	IL2RA	MDC1	NUMB	PRDM1	RPS6KB2	STN1	VEGFB
AR [†]	CCNE2	DDX27	ETV4	FZD5	IL2RB	MDM2	NUP93	PREX1	RPTOR	SUFU	VHL
ARAF	CD274	DDX3X	ETV5	FZD6	IL2RG	MDM4	NUTM1	PREX2	RRAGC	SYK	VIRMA
ARFRP1	CD276	DDX41	ETV6	FZD7	IL7R	MED12	P2RY8	PRKAR1A	RSP01	SYNCRIP	WBP11
ARHGAP35	CD74	DEPDC5	EWSR1	FZD8	INHBA	MEF2B	PABPC1	PRKCI	RSP02	TACSTD2	WEE1
ARID1A [^]	CD79A	DEPTOR	EXO1	FZD9	INPP4B	MEN1	PAK1	PRKDC	RSP04	TAF1L	WRN
ARID1B [^]	CD79B	DHX15	EZH1	GAS6	INSL6	MERTK	PAK3	PRKN	RUNX1	TAP1	WT1
ARID2	CDC27	DHX16	EZH2	GATA1	IRF1	MET [†] #	PALB2 [^] #	PRMT5	RUNX1T1	TAP2	WWP1
ASXL1	CDC5L	DHX36	FAAP100	GATA2	IRF2	MGA	PARG	PRPF40B	RXRα	TAPBP	XBP1
ATM [^] #	CDC7	DHX9	FAAP20	GATA3	IRF4	MGMT [^] §	PARP1	PRPF4B	RYBP	TBC1D7	XPA
ATMIN	CDC73	DICER1	FAAP24	GATA4	IRS2	MITF	PARP2	PSENN1	SAHMD1	TBX3	XPC
ATR [^]	CDH1 [^]	DIS3L2	FANCA [^]	GATA6	JAK1	MKNK1	PAX3	PSMB10	SDC4	TCERG1	XPO1
ATRX	CDH6	DLL4	FANCB	GEN1	JAK2	MLH1 [^]	PAX5	PSMB8	SDHA [^]	TCF7L2	XRCC1
AURKA	CDK11A	DNAJB1	FANCC	GID4	JAK3	MLH3	PAX7	PSMB9	SDHA2	TEK	XRCC2
AURKB	CDK12 [^] #	DNMT1	FANCD2	GLI1	JUN	MLST8	PAX8	PTCH1	SDHB [^]	TEN1	XRCC3
AURKC	CDK4 [†]	DNMT3A	FANCE [^]	GNA11	KAT6A	MPL	PAXIP1	PTDSS1	SDHC [^]	TENT5C	XRCC4
AXIN1	CDK6 [†]	DNMT3B	FANCF [^]	GNA13	KAT6B	MRAS	PBRM1 [^]	PTEN [^] #	SDHD [^]	TERT [†]	XRCC5
AXIN2	CDK7	DOT1L	FANCG [^]	GNAQ	KDM4A	MRE11	PCBP1	PTPN11	SEM1	TET1	XRCC6
AXL	CDK8	DPYD ^P	FANCI	GNAS	KDM5A	MSH2 [^]	PCBP2	PTPN2	SERPINB3	TET2	YAP1
B2M [^]	CDKN1A	DUSP4	FANCL [^]	GNAS	KDM5B	MSH3	PCDH15	PTPRD	SERPINB4	TFE3	YES1
BABAM1	CDKN1B [^]	DYNLL1	FANCM	GPC3	KDM5C	MSH6 [^]	PDCD1	PTPRS	SES2	TFRC	ZC3H13
BABAM2	CDKN1C	DYRK2	FAS	GREM1	KDM6A	MTAP [^] #	PDCD1LG2	PTPRT	SETD2	TGFBF1	ZC3H18
BAP1	CDKN2A [^] #	E2F3	FAT1	GRIN2A	KDR	MTHFR	PDE7A	QKI	SF3B1	TGFBF2	ZC3H4
BARD1	CDKN2B	EBV [~] §	FBXW7	GSK3B	KEAP1 [^] #	MTOR	PDGFRA [†]	RAB35	SF3B3	THRAP3	ZMYM3
BCL2	CDKN2C	ECT2L	FCGR2A	GSTM1	KIN	MUTYH [^]	PDGFRB	RAC1	SH2D1A	TIA1	ZNF217
BCL2L1	CEBPA	EFTUD2	FCGR3A	GSTP1	KIT [†]	MYB	PDK1	RAD18	SHLD1	TIPARP	ZNF703
BCL2L2	CELF4	EGFR [†] #	FEN1	H3-4	KLF4	MYC [†]	PDPK1	RAD21	SHLD2	TMEM127	ZNRF3
BCL6	CEP295	EIF1AX	FGF1	H3F3A	KLHL6	MYCL	PHF6	RAD50 [^]	SLC34A2	TMPPRSS2	ZRSR2
BCOR	CFAP20	EIF4A1	FGF10	HACD4	KLHL9	MYCN	PHLPP1	RAD51 [^]	SLFN11	TNFAIP3	
BCORL1	CHD4	EIF4A2	FGF12	HDAC2	KMT2A	MYD88	PHLPP2	RAD51B	SLIT2	TNFRSF14	
BCR	CHEK1 [^]	EIF4A3	FGF14	HDAC6	KMT2B	MYOD1	PHOX2B	RAD51C [^] #	SMAD2	TNFRSF1A	
BIRC5	CHEK2 [^] #	EIF4B	FGF19	HELQ	KMT2C	NAB2	PIAS4	RAD51D [^] #	SMAD3	TNK2	
BLM	CIC	EIF4E	FGF2	HES1	KMT2D	NBN	PIK3C2B	RAD52	SMAD4 [^]	TNP01	
BMPR1A	CMTM4	EIF4E2	FGF23	HEY1	KNSTRN	NCOR1	PIK3CA [†]	RAD54L	SMARCA2	TOP1	
BRAF [†] #	CMTM6	ELAVL1	FGF3	HEYL	KRAS [†]	NCR1	PIK3CB	RAET1E	SMARCA4	TOP2A	
BRCA1 [^] #	CNOT3	ELAVL2	FGF4	HGF	LATS1	NCR3	PIK3CD	RAF1 [†]	SMARCAL1	TOPA21	
BRCA2 [^] #	CREBBP	ELF3	FGF5	HLA-A ^h §	LGR4	NEGR1	PIK3CG	RARA	SMARCB1	TP53 [^]	
BRCC3	CRKL	ELOC	FGF6	HLA-B ^P h §	LGR5	NELFE	PIK3R1	RASA1	SMARCD1	TP53BP1	

[†] Guardant360 reports alterations in the promoter region of this gene.

[†] Guardant360 reports amplifications of this gene.

^{*} Guardant360 reports copy number deletions in this gene.

[#] Guardant360 reports rearrangements in this gene.

[§] Guardant360 does not report SNVs or indels from this feature.

[^] Guardant360 reports promoter methylation of this gene.

[~] Guardant360 reports detection of this virus.

^h Guardant360 reports HLA alleles in this gene.

^P Guardant360 reports pharmacogenomics results of this gene.