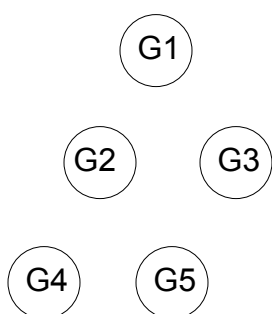


“Change is the end result of all  
true learning.”  
-- Leo Buscaglia

## Independent Events



If  $G1, \dots, G5$  are independent, then  
the joint probability  
 $p(G1, G2, G3, G4, G5)$   
 $= p(G1) p(G2) p(G3) p(G4) p(G5)$

Example:

$D$  = “Dowell gives the lecture today”.

$R$  = “It is raining outside today”

Whether it is rain or shine outside doesn’t  
affect whether Dowell is giving the lecture  
today.

$$P(D, R) = p(D) * p(R)$$

## Conditional Probability Distributions

- Conditional probability distributions:  $p(B|A)$  = the probability of  $B$  given  $A$ .

Example:

$D$  = "Dowell gives the lecture today".

$E$  = "today's lecture contains equations"

$P(E, D)$  = Probability that Dowell gives the lecture today and today's lecture contains equations = 0.05.

$P(D) = 1/10 = 0.1$ .

$P(E|D) = P(E, D) / P(D) = 0.05/0.1 = 0.5$ .

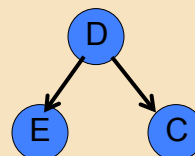
## Conditional Independence

Example:

$D$  = "Dowell gives the lecture today".

$E$  = "today's lecture contains equations"

$C$  = "today's slides are in *Comic Sans* font"



If Dowell is giving the lecture today, then whether today's lecture contains equations doesn't affect whether today's slides are in *Comic Sans*.

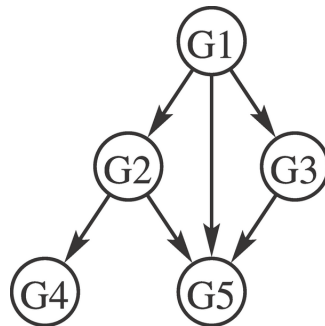
$P(E|D, C) = P(E|D)$

$E$  and  $C$  are conditionally independent given  $D$ .

- In Bayesian networks, each node is independent of its non-descendants, given its parents in the graph.
- Using conditional independence between variables, the joint probability distribution of the models may be represented in a compact manner.

## Can capture biological relationships

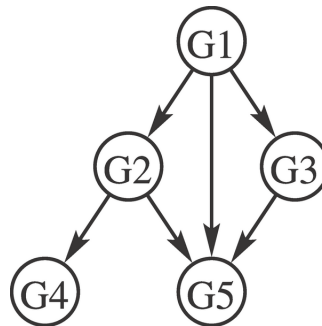
- A **directed acyclic graph (DAG)** such that the nodes represent mRNA expression levels and the edges represent the probability of observing an expression value given the values of the parent nodes.
- The probability distribution for a gene depends only on its regulators (parents) in the network.



**Example:** G4 and G5 share a common regulator G2, i.e., they are conditionally independent given G2.  
 → factorization of the full joint probability distribution into component conditional distributions.

Needham et al. PLOS Comp Bio 2007

## Joint Probability Distribution

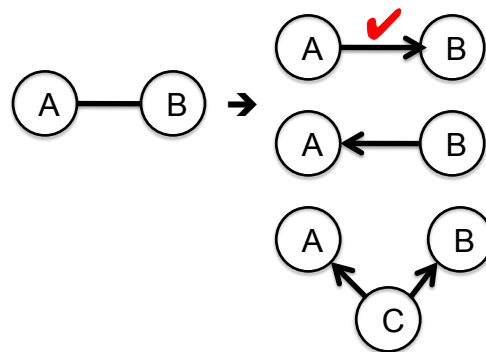


$$\begin{aligned}
 & p(G1, G2, G3, G4, G5) \\
 &= p(G1) p(G2|G1) p(G3|G1) p(G4|G2) p(G5|G1, G2, G3)
 \end{aligned}$$

## What kinds of data contain potential information about gene networks?

### Large expression sets

- Co-expression (correlation of expression levels) implies connectivity
- But correlation  $\neq$  causality



#### Adding causality

- Genetic perturbation: DNA variation at A influences RNA variation at B.
- Time series: A goes up prior to B.
- Prior knowledge

## Bayesian networks

- Advantages:
  - Compact and intuitive representation
  - Integration of prior knowledge
  - Probabilistic framework for data integration
- Limitation: no feedback loop  $\rightarrow$  dynamic Bayesian networks (variables are indexed by time and replicated in the network)
- References:
  - Using Bayesian Network to Analyze Expression Data. Friedman et al. *J. Computational Biology* 7:601-620, 2000.
  - A Primer on Learning in Bayesian Networks for Computational Biology. Needham et al. *PLOS Computational Biology* 2007.

## Using Bayesian Networks

- There are algorithms for inferring Bayesian networks from large collections of data.
- Given a particular network, can determine whether particular datasets are consistent with the inherent probabilistic relationships implied by the graph.
- Ultimately powerful for predicting the impact of a perturbation.

“We have only just hit the tip of the iceberg.  
There’s a whole world of this noncoding RNA. ”

-- Peter Schultz (2005)



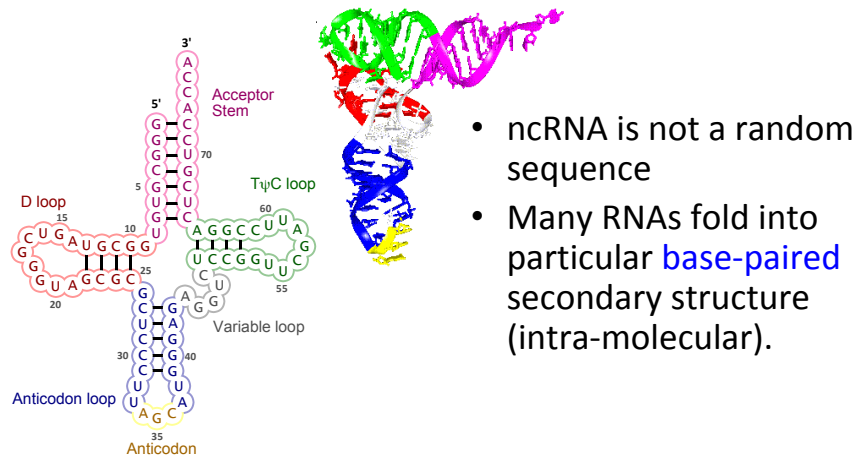
## What do they do?

- [RNA-protein machine](#):
  - Transfer RNA (tRNA).
  - Ribosomal RNA (rRNA).
  - RNAs (snRNAs) in spliceosome.
- [Catalytic RNAs](#) (ribozymes): catalyzing some functions.
  - 1989 Nobel Prize in Chemistry
- [Micro RNAs \(miRNAs\)](#): regulatory roles.
- [Small interfering RNAs \(siRNAs\)](#): RNA silencing
  - The genome's immune system. [Plasterk, *Science* (2002)]
  - 2006 Nobel Prize in Medicine

## What do they do?

- [Riboswitch RNAs](#): a genetic control element, to control gene expression.
  - found in bacteria, archaea, and plants.
- [Small nucleolar RNAs \(snoRNAs\)](#): help the modification of rRNAs.
- [tmRNA \(tRNA like mRNA\)](#): direct abnormal protein degradation.
- [lncRNAs](#): long noncoding RNAs (look like protein coding in terms of exon/intron structure, histone marks, etc)
- [eRNAs](#): enhancer regulation/function?, unstable

## RNA secondary structure

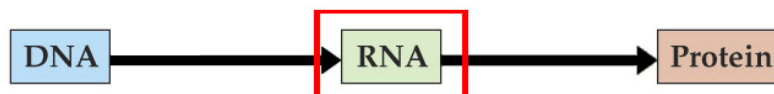
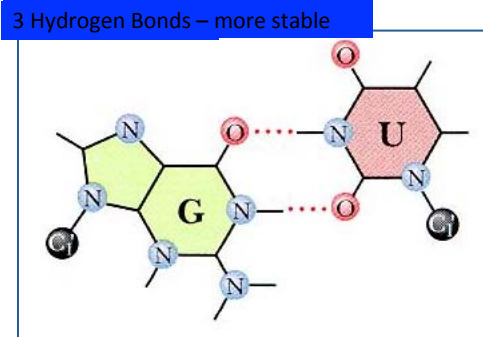


- ncRNA is not a random sequence
- Many RNAs fold into particular **base-paired** secondary structure (intra-molecular).

RNA secondary structure -- a set of non-crossing base pairs

## Many ncRNAs conserve structure

- RNA bases A,C,G,U
- Canonical Base Pairs
  - A-U
  - G-C
  - G-U
- “wobble” pairing
  - Bases can only pair with **one** other base.

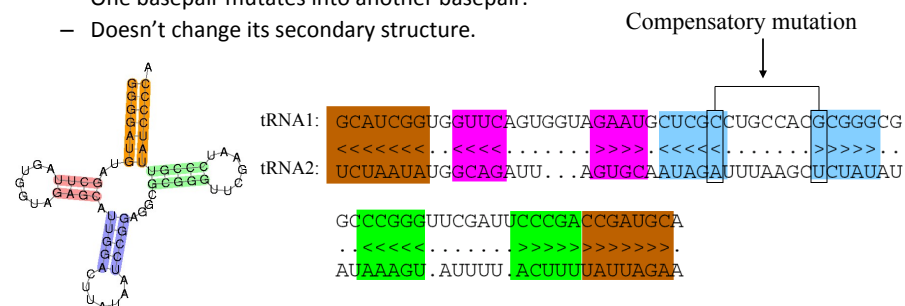


Note: all 16 (including non-canonical) base pairs are actually possible and occasionally observed!!

Image: <http://www.bioalgorithms.info/>

## ncRNA evolution is constrained by its secondary structure

- **Drastic** sequence changes can be tolerated, so long as structure conserved.
- *Compensatory mutations* are very common.
  - One basepair mutates into another basepair.
  - Doesn't change its secondary structure.



<http://www.sanger.ac.uk/Software/Rfam/>

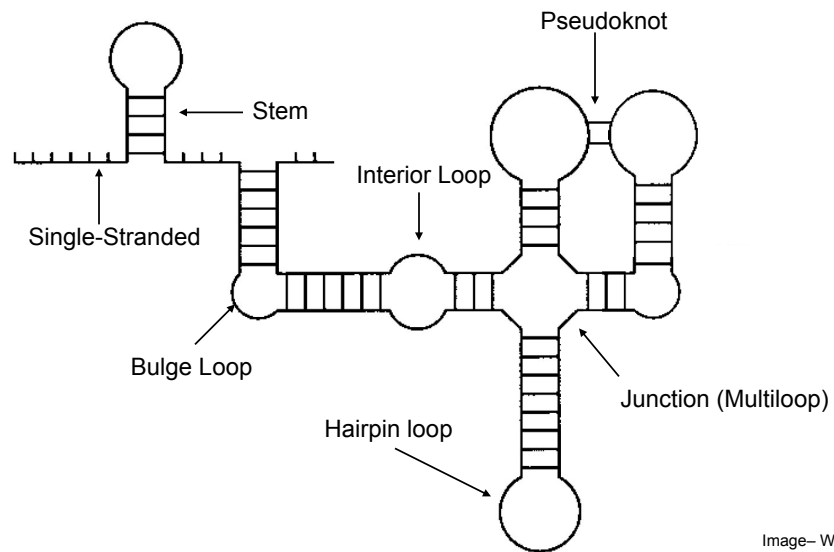
Hence sequence alignment also often “fails” to find ncRNAs!!!

## RNA secondary structure prediction

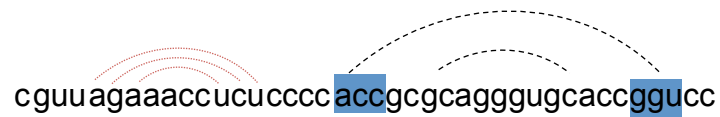
- It is a basic issue in structural ncRNA analysis
- It is important information towards function
- Searching and alignment algorithms are based on these models
- But it is computationally VERY expensive.



## RNA secondary structure elements

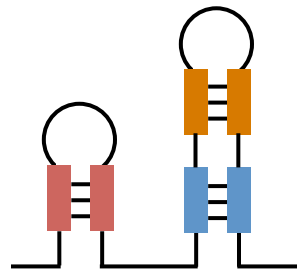


## Stems in nested or parallel pattern



stem (double helix):  
stacked base pairs

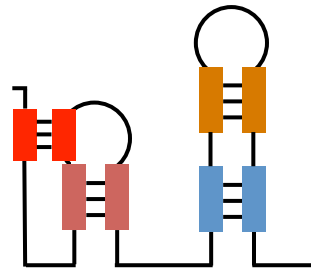
loop: strand of  
unpaired bases



## Stems in crossing patterns are pseudoknots

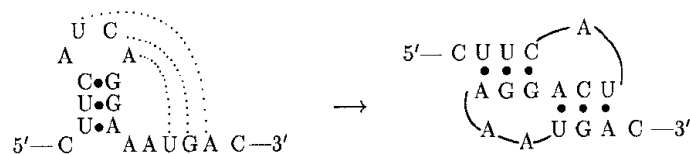
cguu **aga** aacc **ucu** cccc **acc** gc **gca** ggg **ugc** acc **gg** ucc

Pseudoknots: **crossing**  
patterns of stems.



## Pseudoknots

- Pseudoknots are important for certain ncRNAs
- Violate the non-crossing assumption.
- Pseudoknots make most problems **a LOT harder**
- We assume there are no pseudoknots, unless otherwise noted.

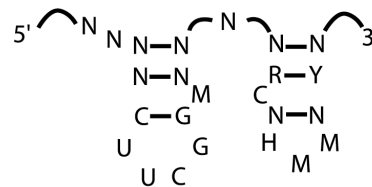
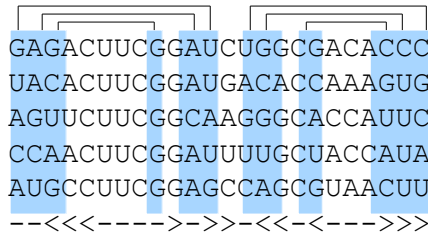
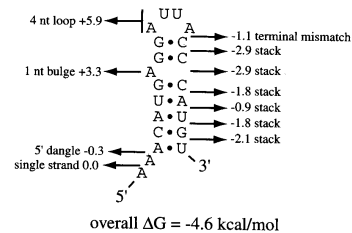


[Rivas and Eddy (1999)]

## RNA secondary structure prediction

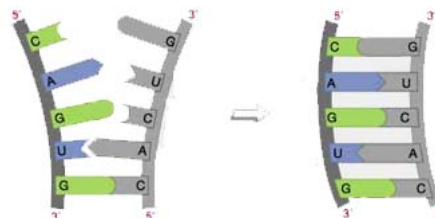
**ab initio structure prediction**  
to predict the structure of a  
single sequence

**Consensus structure prediction**  
to predict the structure shared  
by more than one sequences



## Specialized intra-molecular “sequence alignment” as a method to determine structure

- Bases pair in order to form backbones and determine the secondary structure
- Aligning bases based on their ability to pair with each other gives an algorithmic approach to determining the optimal structure

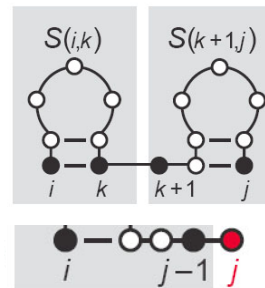


## Base Pair Maximization – Dynamic Programming Algorithm

$S(i,j)$  is the folding of the subsequence of the RNA strand from index  $i$  to index  $j$  which results in the highest number of base pairs

### Maximizing Base Pair

$$S(i,j) = \max \begin{cases} S(i+1, j-1) + 1 & \text{[if } i,j \text{ base pair]} \\ S(i+1, j) \\ S(i, j-1) \\ \max_{i < k < j} S(i, k) + S(k+1, j) \end{cases}$$



Bifurcation

Base pair at  $i$  and  $j$

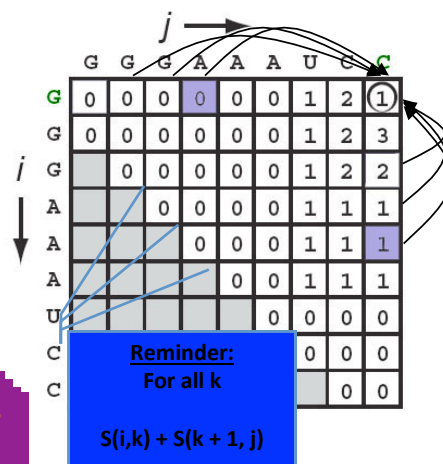
Unmatched at  $i$

Unmatched at  $j$

Images – Sean Eddy

## Base Pair Maximization – Dynamic Programming Algorithm

- Alignment Method
  - Align RNA strand to itself
  - Score increases for feasible base pairs
- Each score independent of overall structure
- Bifurcation adds extra dimension



Images – Sean Eddy

## Dynamic Programming Algorithm

$$S(i,j) = \max \begin{cases} S(i+1, j-1) + M(i,j) \\ S(i+1, j) \\ S(i, j-1) \\ \max_{i < k < j} S(i, k) + S(k+1, j) \end{cases}$$

Note that this is the basic folding algorithm, but we can CHANGE the scoring scheme!

## Base Pair Maximization - Drawbacks

- Base pair maximization will not necessarily lead to the most stable structure
  - May create structure with many interior loops or hairpins which are energetically unfavorable
- Comparable to aligning sequences with scattered matches – not biologically reasonable

Alternative scoring scheme:  
Energy Minimization  
(Thermodynamic Stability)

## Energy Minimization Drawbacks

- Compute only one optimal structure
- Usual drawbacks of purely mathematical approaches
  - Similar difficulties in other algorithms

G-C	$E_{\text{Hairpin loop}} = +4.5$
A A	$E_{\text{Stack}} = -2.4$
G-C	$E_{\text{Interior loop}} = -1.4$
C-G	$E_{\text{Stack}} = -2.1$
G G	
A-U	
C-G	$E_{\text{Total}} = -1.4$

- Protein structure
- Exon finding

Alternative scoring scheme:  
Probabilistic Framework

## Probabilistic Models

- $S(i,j)$  = Score at indices  $i$  and  $j$  in RNA

$$S(i,j) = \max \begin{cases} S(i+1, j-1) + M(i,j) \\ S(i+1, j) \\ S(i, j-1) \\ \max_{i < k < j} S(i, k) + S(k+1, j) \end{cases}$$

$$M_{i,j} = \sum_{x_i, x_j} f_{x_i, x_j} \log_2 \frac{f_{x_i, x_j}}{f_{x_i} f_{x_j}}$$

Frequency of seeing the symbols (A, C, G, U) together in locations  $i$  and  $j$  depending on symbol.

- Frequencies obtained by aligning model to “training data” – consists of sample sequences
  - Reflect values which optimize alignment of sequences to model

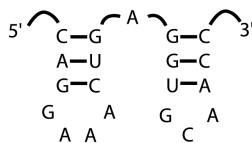
## Stochastic context-free grammars

- Big brother to HMMs
  - Emission and transition probabilities!
- Still folding styled dynamic programming

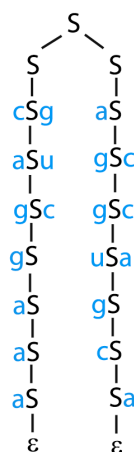
$$S \rightarrow aSa' \mid aS \mid Sa \mid SS \mid \epsilon$$

SCFGs describe structure with “rules” (aka states).

$$S \rightarrow aSa' \mid aS \mid Sa \mid SS \mid \epsilon$$



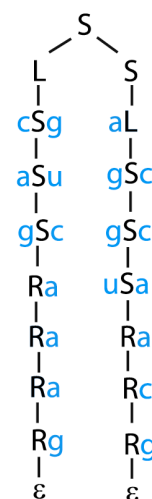
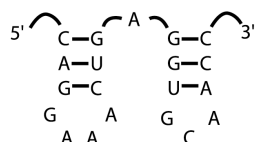
$S \rightarrow SS \rightarrow cSgS \rightarrow caSugS \rightarrow cagScugS \rightarrow$   
 $caggScugS \dots$



## Representational ambiguity

$$S \rightarrow aSa' \mid aL \mid Ra \mid LS$$

$$L \rightarrow aSa' \mid aL$$

$$R \rightarrow Ra \mid \epsilon$$


Grammar designs vary and each capture different “features” of structure

G1:  $S \rightarrow aSa' \mid aS \mid Sa \mid SS \mid \epsilon$

G3:  $S \rightarrow aS \mid T \mid \epsilon$   
 $T \rightarrow Ta \mid aSa' \mid TaSa'$

G5:  $S \rightarrow L \mid LS$   
 $L \rightarrow aFa' \mid a$   
 $F \rightarrow aFa' \mid LS$

G2:  $S \rightarrow aSa' \mid aL \mid Ra \mid LS$   
 $L \rightarrow aSa' \mid aL$   
 $R \rightarrow Ra \mid \epsilon$

G4:  $S \rightarrow aSa'S \mid aS \mid \epsilon$



## Standard Algorithms

(notice parallels to HMMs!)

- **Scoring** (probability of parse tree)

$$P(x, \pi \mid G, \Theta)$$

- **Viterbi / CYK** (highest probability parse tree)

$$\operatorname{argmax}_{\pi} P(x, \pi \mid G, \Theta)$$

- **Inside** (probability of sequence, akin to “Forward”)

$$P(x \mid G, \Theta) = \sum_{\pi} P(x, \pi \mid G, \Theta)$$

## ncRNA gene finding (a computational challenge)

### *de novo* prediction

identify ncRNA regions from  
genomic sequence

### Profile based methods

identify new instances of a  
known family of structural  
ncRNAs

### Classification Methods

use evolutionary signatures to  
distinguish coding, noncoding,  
and other regions