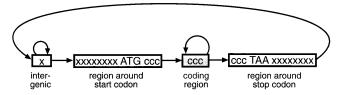


Bacterial Gene finding as an HMM

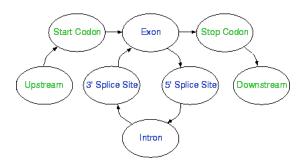
- Nucleotides {A,C,G,T} are the observables
- Different states generates generate nucleotides at different frequencies
 A simple HMM for unspliced genes:



AAAGC ATG CAT TTA ACG AGA GCA CAA GGG CTC TAA TGCCG

 The sequence of states is an annotation of the generated string – each nucleotide is generated in intergenic, start/stop, coding state

What about splicing?



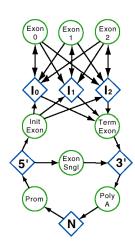
Note that transitions to the same state are left off simply to make the diagram simpler.

Genscan Example

- Developed by Chris Burge 1997
- One of the most accurate *ab initio* programs
- Uses explicit state duration HMM to model gene structure (different length distributions for exons)
- Different model parameters for regions with different GC content

Genscan (Burge and Karlin, 1998)

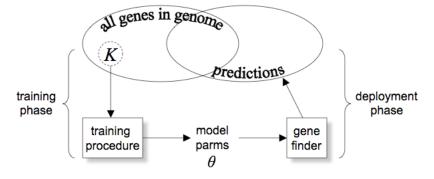
- Dramatic improvement over previous methods
- Generalised HMM
- Different parameter sets for different GC content regions (intron length distribution and exon stats)



Desire a generalizable model.

- · Trained on one dataset
- But performs well on new, never seen before data.
- So what constitutes good performance?

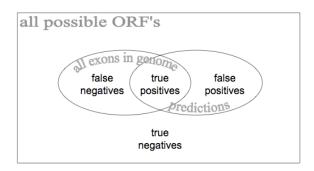
During training of a gene finder, only a subset K of an organism's gene set will be available for training. The gene finder will later be deployed for use in predicting the rest of the organism's genes. Alternatively, the training will use a closely related organism's gene set.



The way in which the *model parameters* are inferred during training can significantly affect the accuracy of the deployed program.

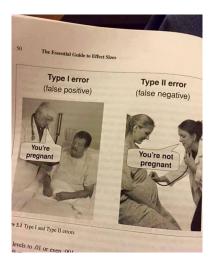
TP, FP, FN, TN

Gene predictions can be evaluated in terms of *true positives* (predicted features that are real), *true negatives* (non-predicted features that are not real), *false positives* (predicted features that are not real), and *false negatives* (real features that were not predicted:

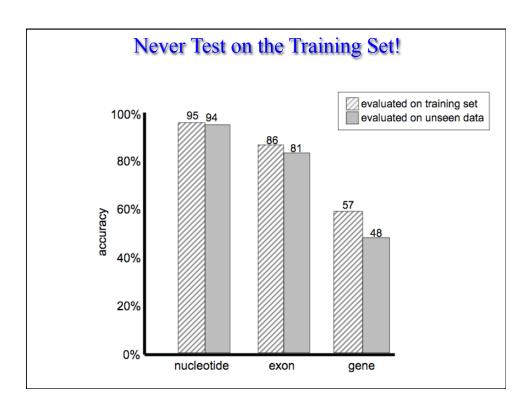


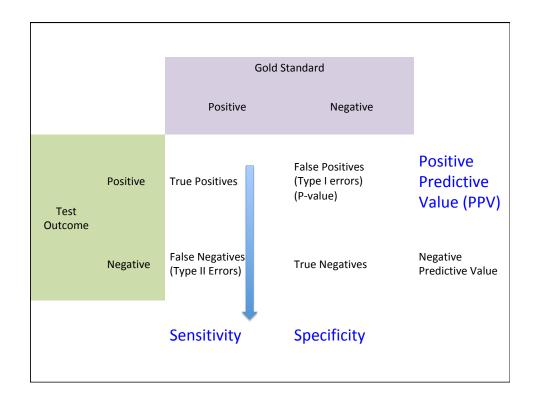
These definitions can be applied at the *whole-gene*, *whole-exon*, or *individual nucleotide* level to arrive at three sets of statistics.

Two distinct types of error ...

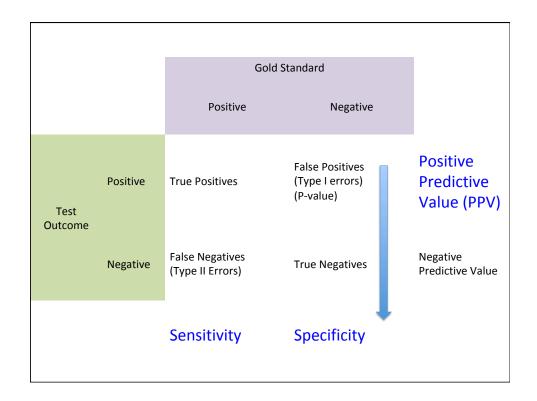


Must first determine what predictions are actually correct.



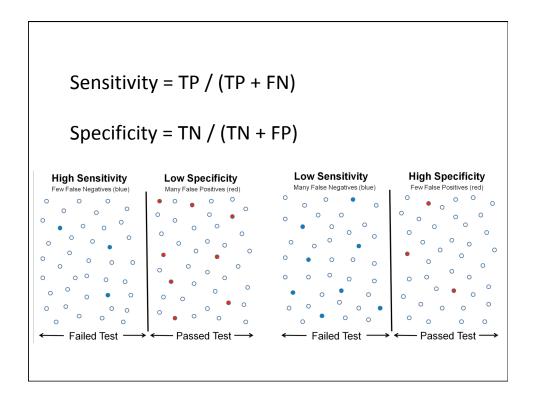


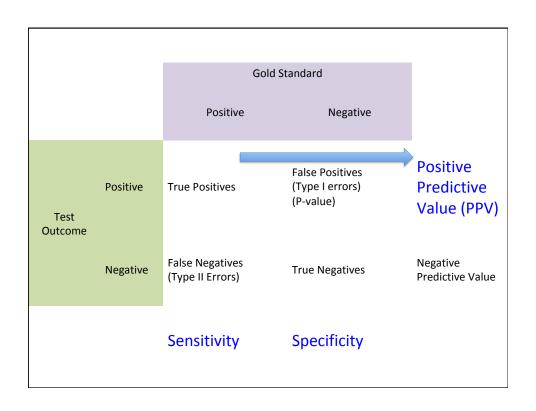
Sensitivity = TP / (TP + FN)



Sensitivity = TP / (TP + FN)

Specificity = TN / (TN + FP)





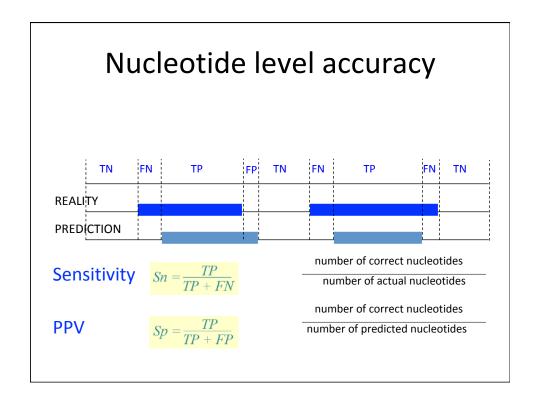
Primary Performance Metrics

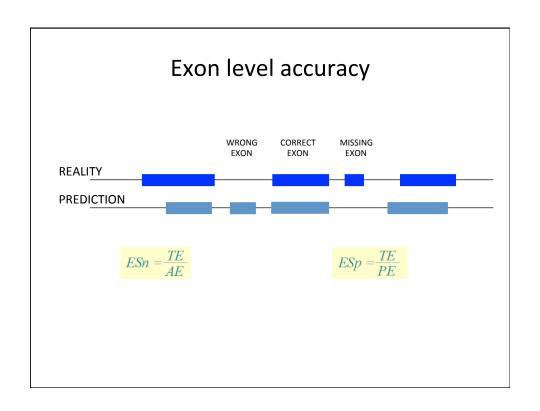
Sensitivity =
$$TP / (TP + FN)$$

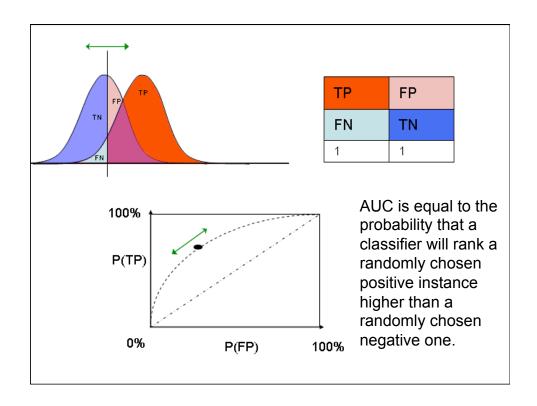
$$PPV = TP / (FP + TP)$$

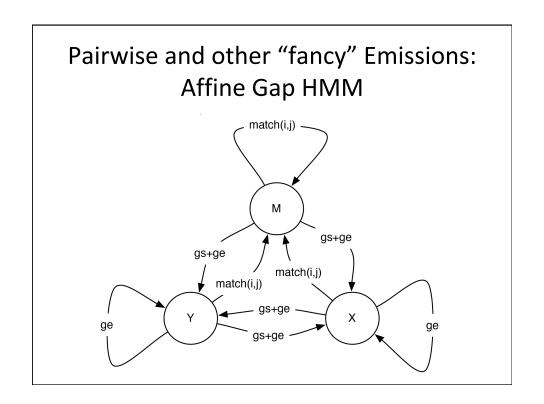
Related Metrics

- False negative rate = FN / (TP + FN)= 1 sensitivity
- False discovery rate = FP / (FP + TP)= 1 PPV
- Accuracy = (TP + TN) / ((TP+FN) + (FP+TN))









Dynamic Programming for the Affine Gap Penalty Case

• to do in $O(n^2)$ time, need 3 matrices instead of 1

M(i, j) best score given that x[i] is aligned to y[j]

 $I_x(i, j)$ best score given that x[i] is aligned to a gap

 $I_{y}(i, j)$ best score given that y[j] is aligned to a gap

Global Alignment DP for the Affine Gap Penalty Case

$$M(i,j) = \max \begin{cases} M(i-1,j-1) + S(x_i,y_j) \\ I_x(i-1,j-1) + S(x_i,y_j) \\ I_y(i-1,j-1) + S(x_i,y_j) \end{cases}$$

$$I_{x}(i,j) = \max \begin{cases} M(i-1,j) + g + s \\ I_{x}(i-1,j) + s \end{cases}$$

$$I_{y}(i,j) = \max \begin{cases} M(i,j-1) + g + s \\ I_{y}(i,j-1) + s \end{cases}$$

The M matrix

$$M(i,j) = \max \begin{cases} M(i-1,j-1) + S(x_i,y_j) \\ I_x(i-1,j-1) + S(x_i,y_j) \\ I_y(i-1,j-1) + S(x_i,y_j) \end{cases}$$

Any kind of alignment is allowed before the match.

——A

The gap matricies $(I_x \text{ and } I_y)$

If previous alignment ends in a match, this must be a new gap.

$$I_x(i,j) = \max \begin{cases} M(i-1,j) + g + s \\ I_x(i-1,j) + s \end{cases}$$

Otherwise we must be extending an existing gap.

