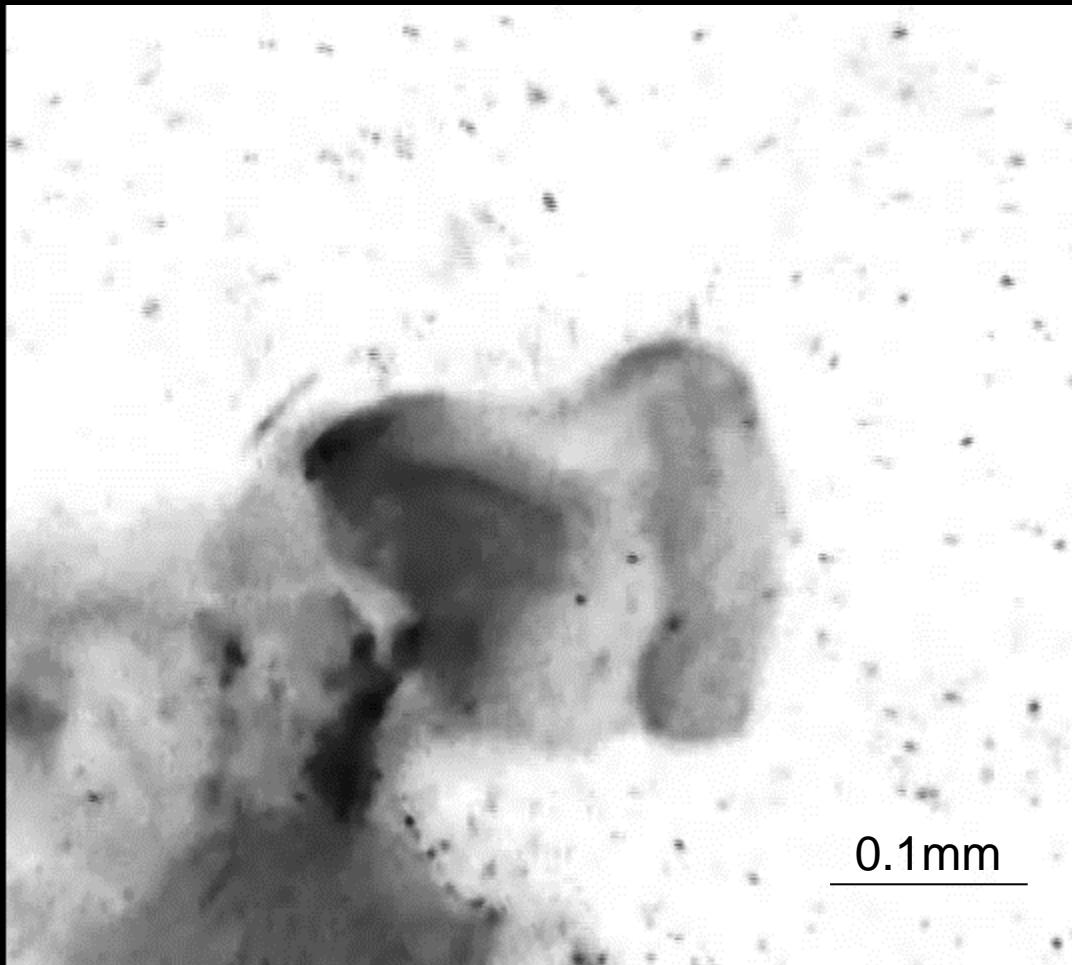


# Cilia-driven flow fields for transport and selective capture of bacteria



Janna Nawroth, Ph.D.

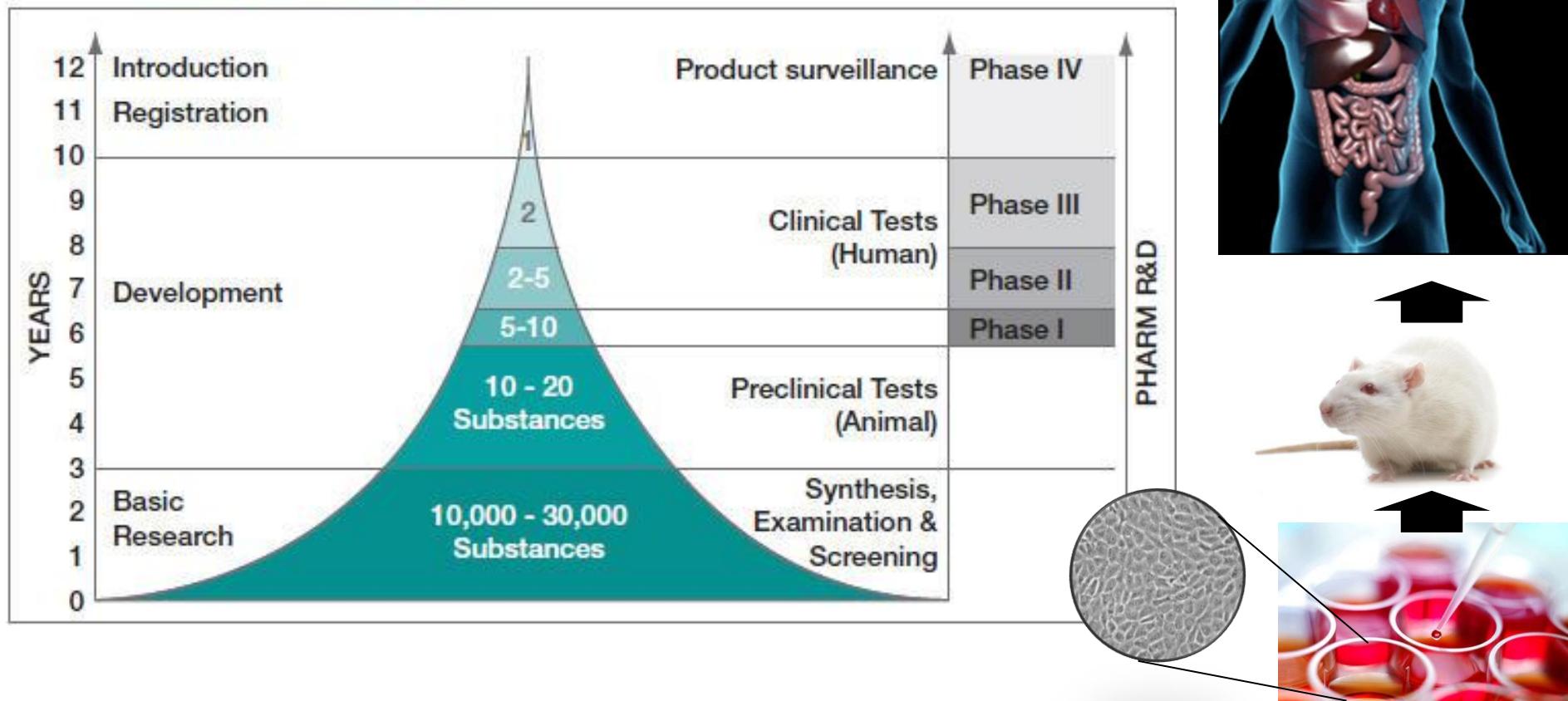
Wyss Institute for Biologically Inspired Engineering, Harvard University  
Active Matter | Kavli Institute for Theoretical Physics | May 15<sup>th</sup>, 2014



# Traditional drug development

Time: 10 years  
Cost: \$1 Billion/drug

Figure 1: The drug development pyramid

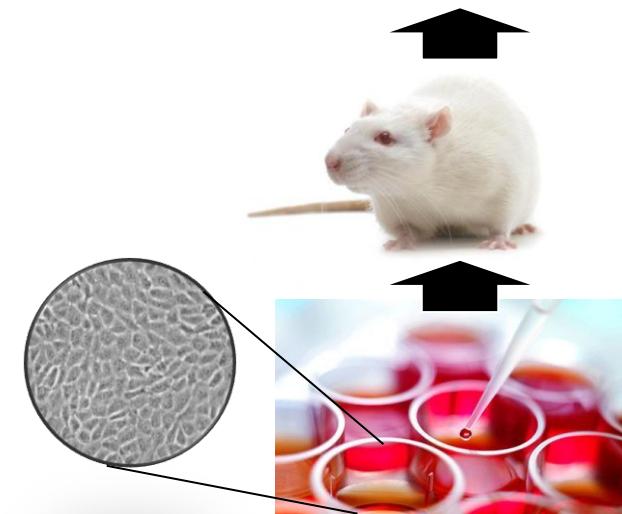
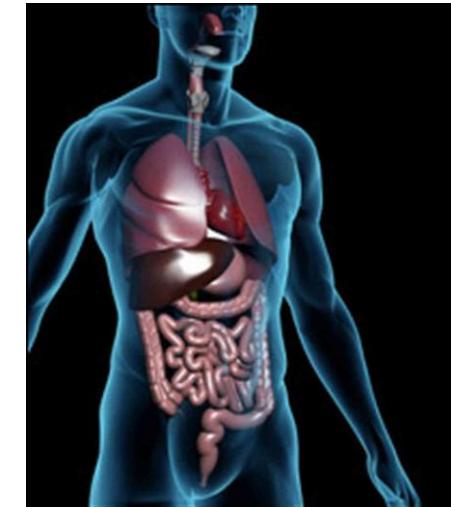
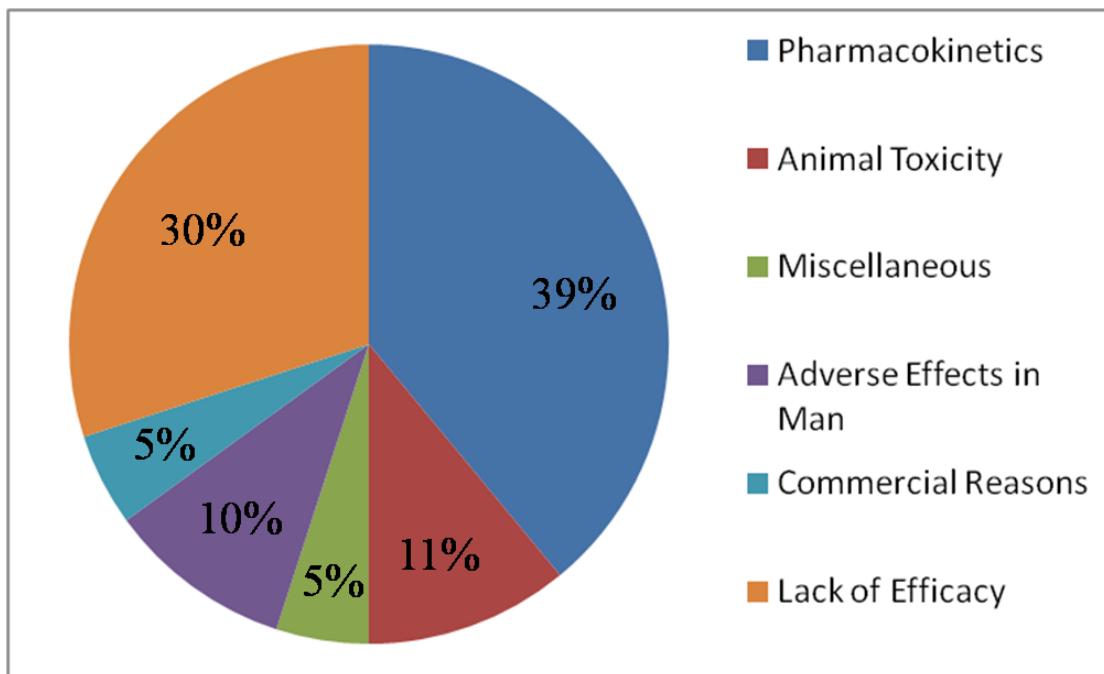




# Traditional drug development

Time: 10 years  
Cost: \$1 Billion/drug

Reasons for drug failure: poor translation  
between test platforms and humans



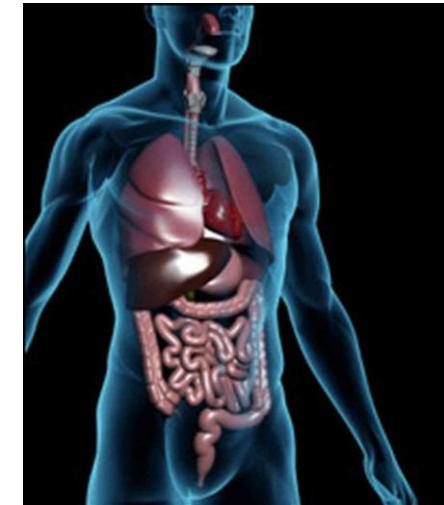


# The quest for a better *in-vitro* model of human disease and drug response

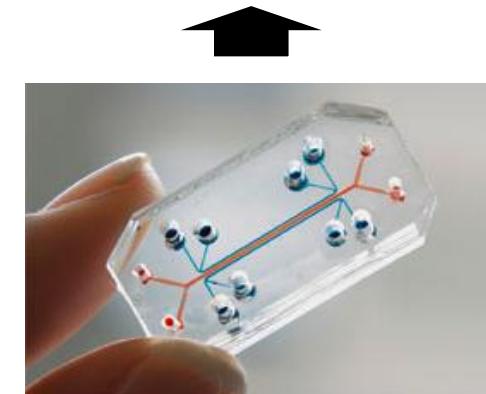
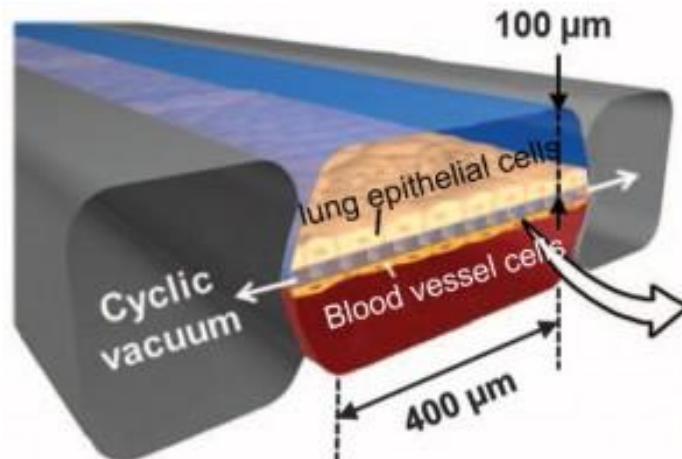
## Organ-on-a-chip

- 3-D microfluidic cell culture chip from human cells
- simulates characteristic mechanics and physiological responses of entire organs

→ “human” platform for drug testing and disease models



Example: lung-blood barrier function





# Organ-on-a-chip design

## Biology

Identify relevant structure-function relationships

## Engineering

Design, build and test structure-function relationships

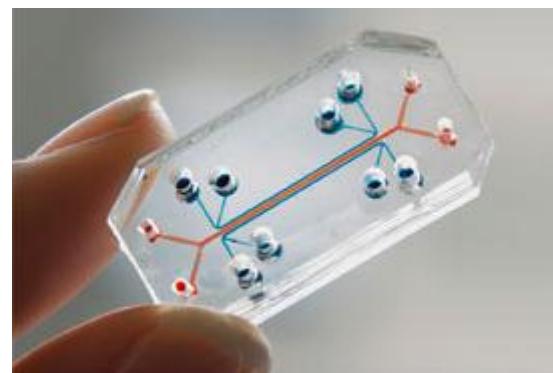
Structure and motion of building blocks (cells, matrix)



Emergent functions



Quantitative metrics of organ fitness





# Organ-on-a-chip design

## Biology

Identify relevant structure-function relationships

Structure and motion of building blocks (cells, matrix)



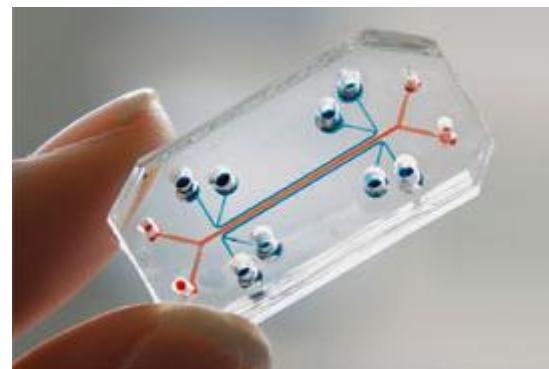
Emergent functions



Quantitative metrics of organ fitness

## Engineering

Design, build and test structure-function relationships



## Example lung chip

Metrics of fitness are

- Structural integrity
- Absorption air → blood

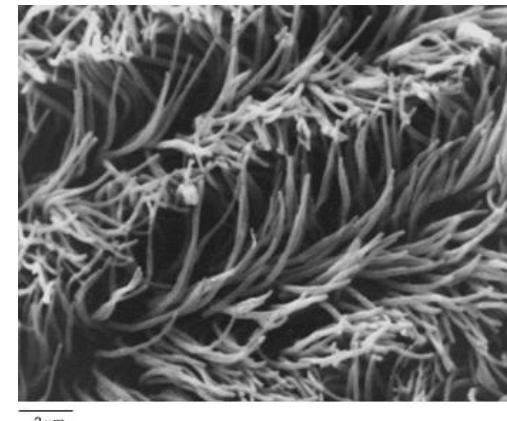


# 2 case studies

**Muscle-powered fluid transport**  
...relevant to cardiovascular system



**Cilia-powered fluid transport**  
... relevant to respiratory organs, brain, Fallopian tube

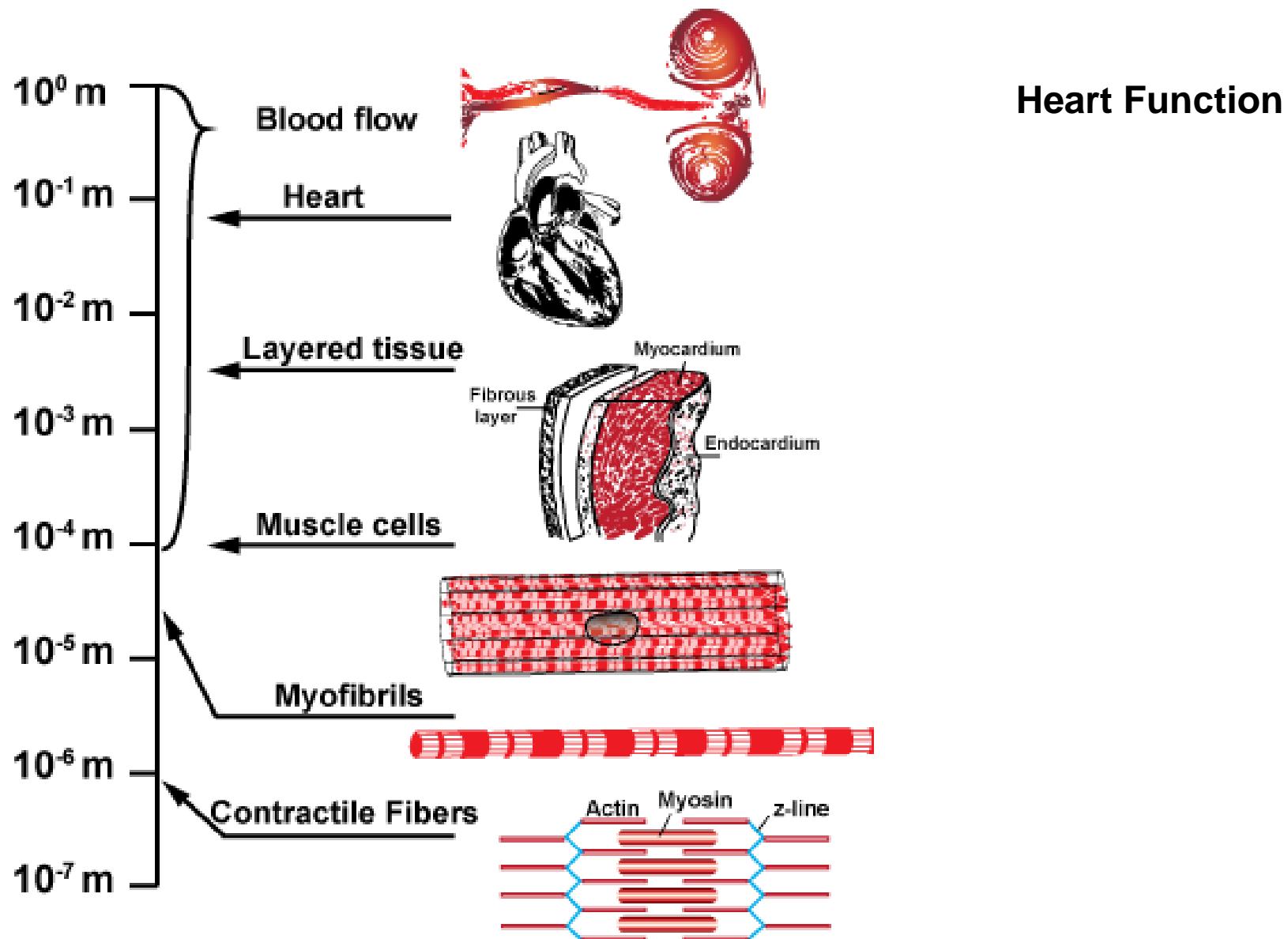


with  
**John Dabiri** (Caltech)  
**Kit Parker** (Harvard)  
**Donald Ingber** (Harvard)

with  
**Eva Kanso** (USC)  
**Margaret McFall-Ngai** (UW Madison)  
**Edward Ruby** (UW Madison)  
**John Dabiri** (Caltech)  
**Scott Fraser** (USC)

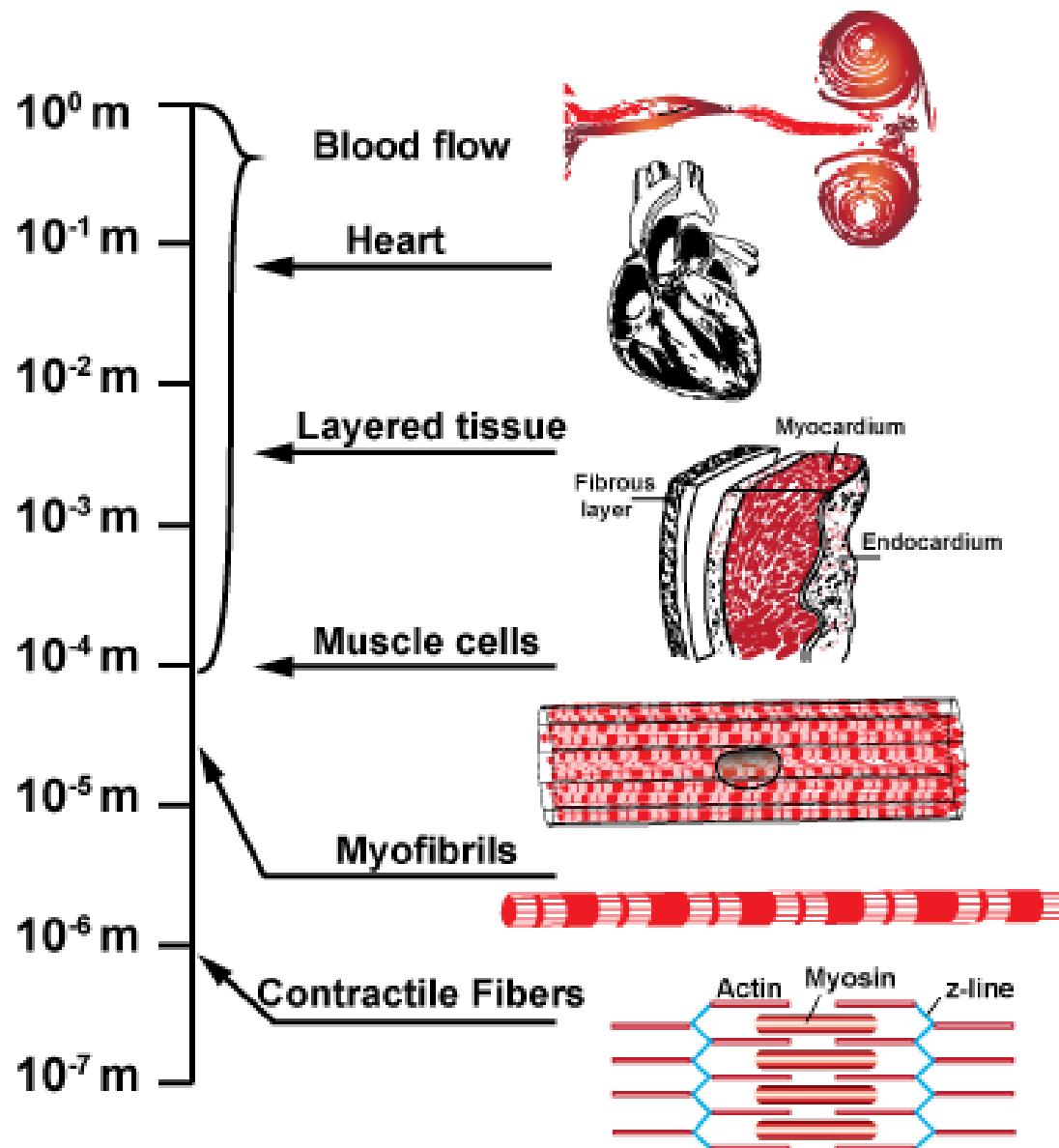


# Case study: human heart





# Case study: human heart



Heart Function



Body-fluid interaction



Tissue geometry

+

Kinematics

+

Microstructure

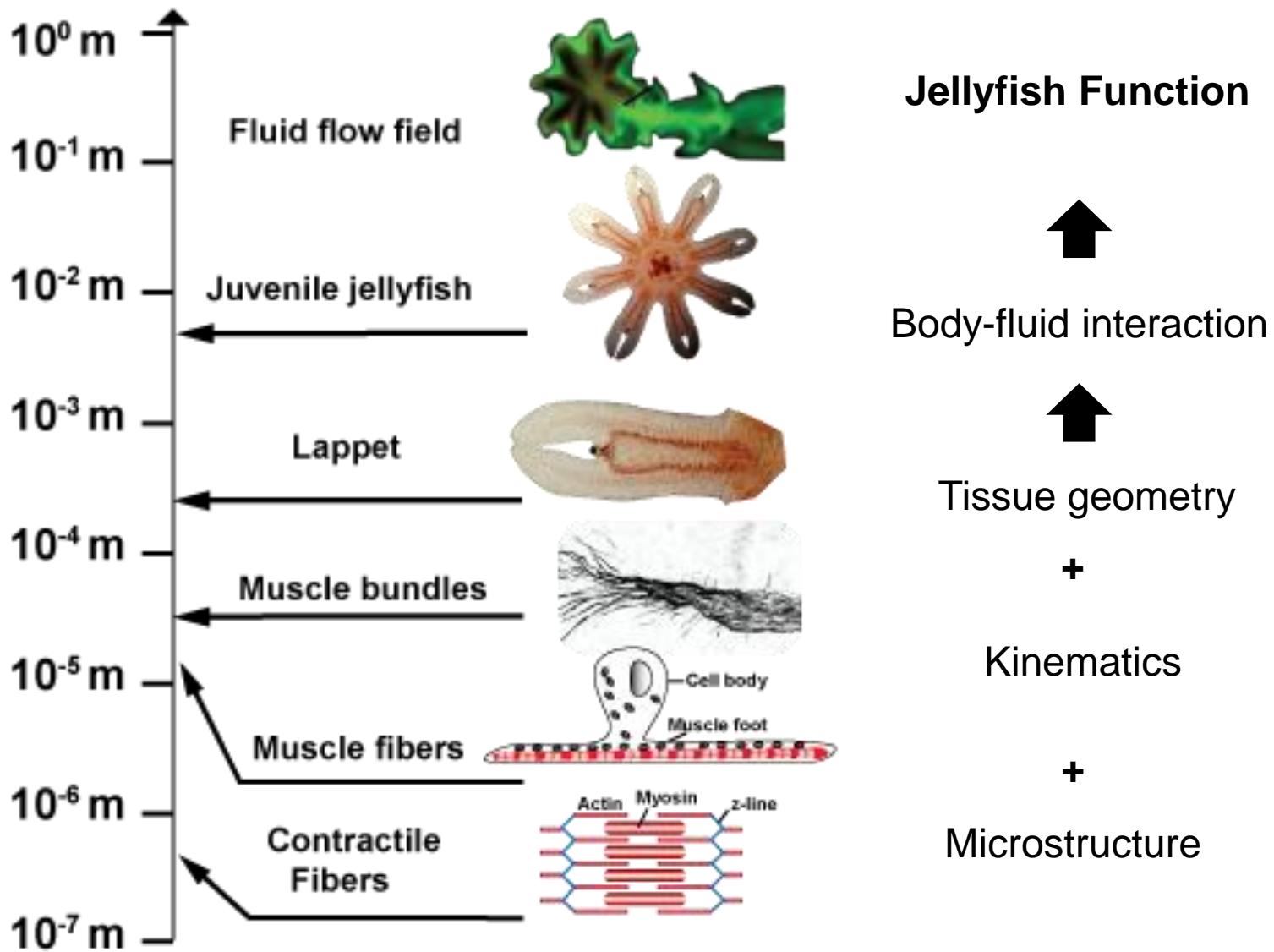


# A simplified heart: jellyfish



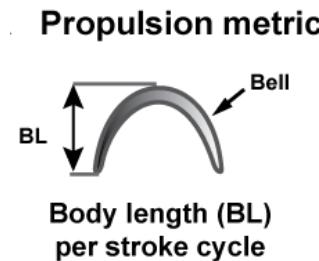
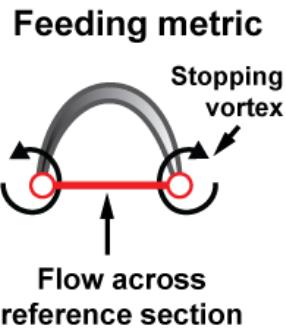


# A simplified heart: jellyfish





# Measuring jellyfish fitness

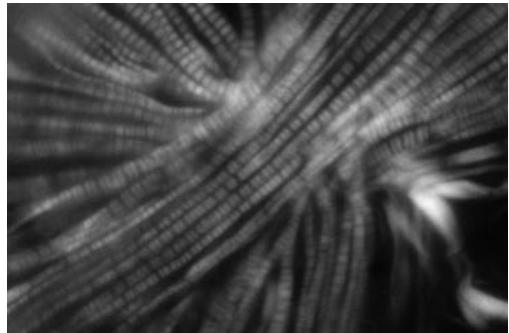


Jellyfish function

- Feeding flux
- Propulsion



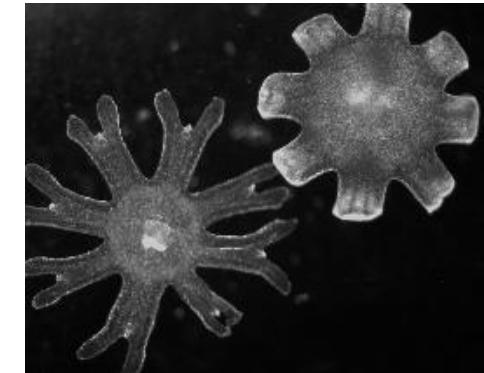
Fluid transport



Muscle fiber  
alignment



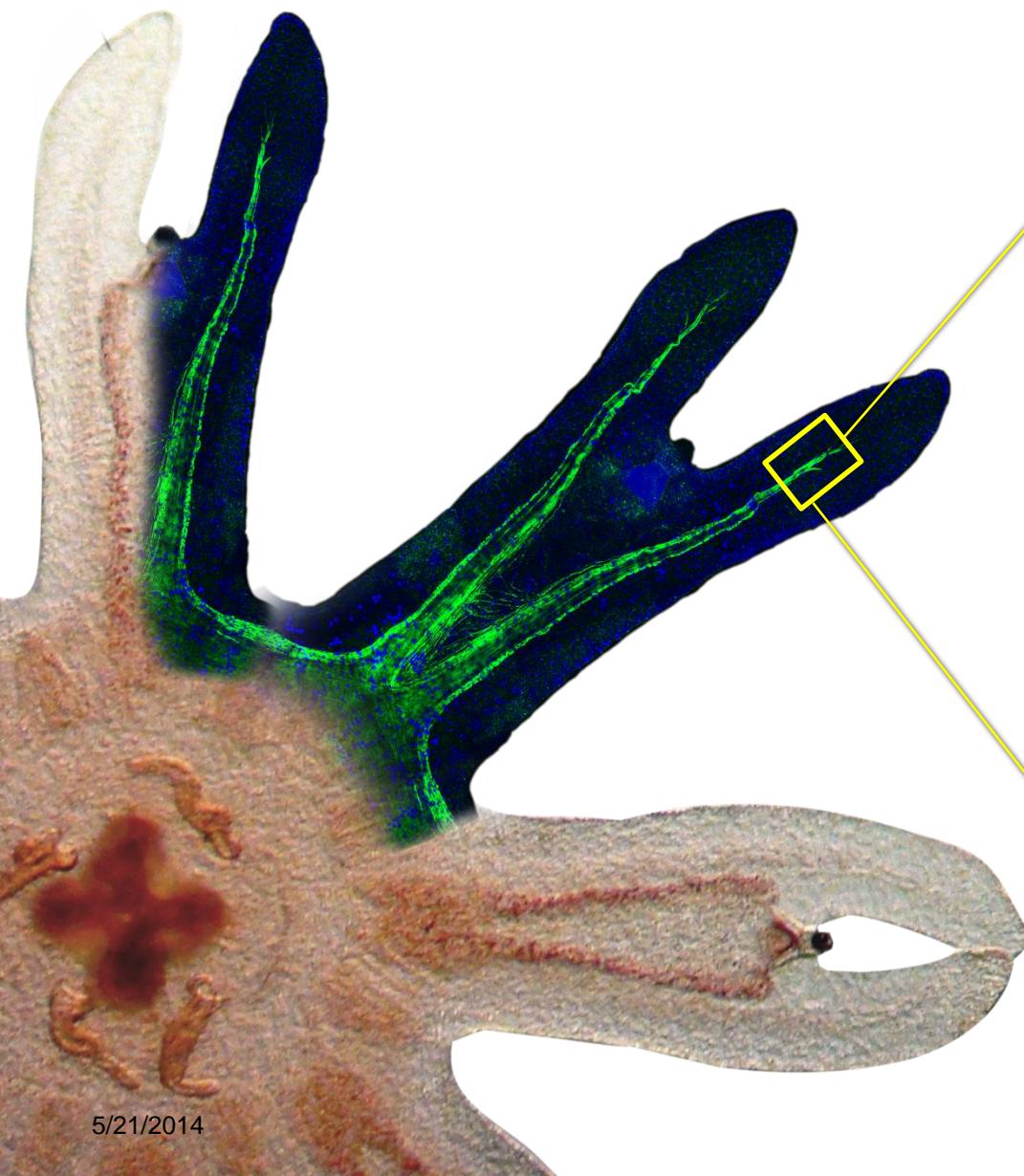
Contraction  
kinematics



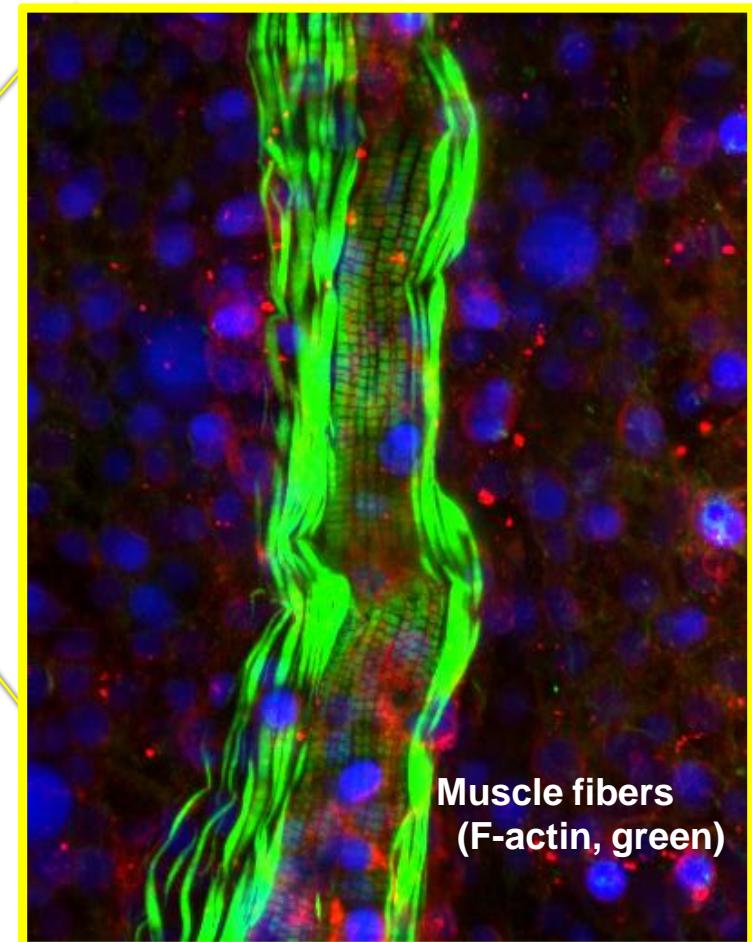
Bell geometry



# Muscle fiber alignment



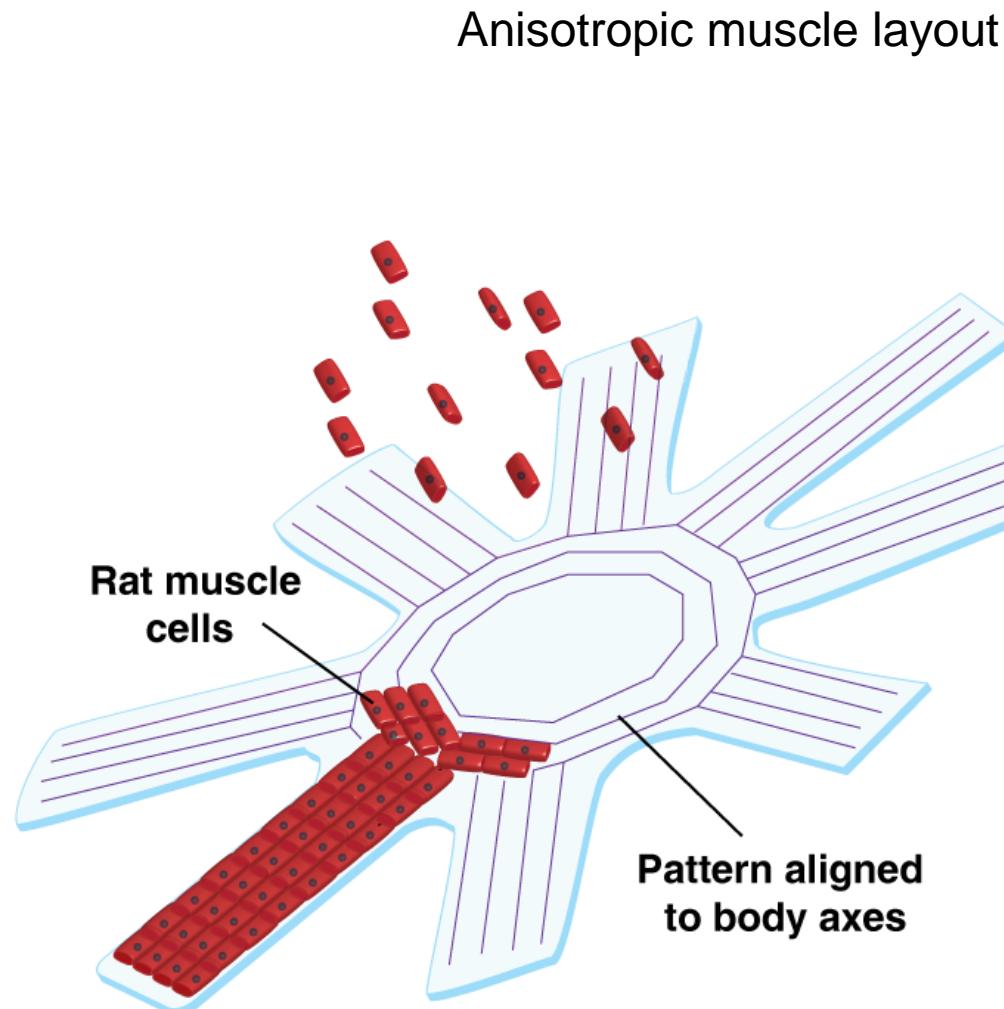
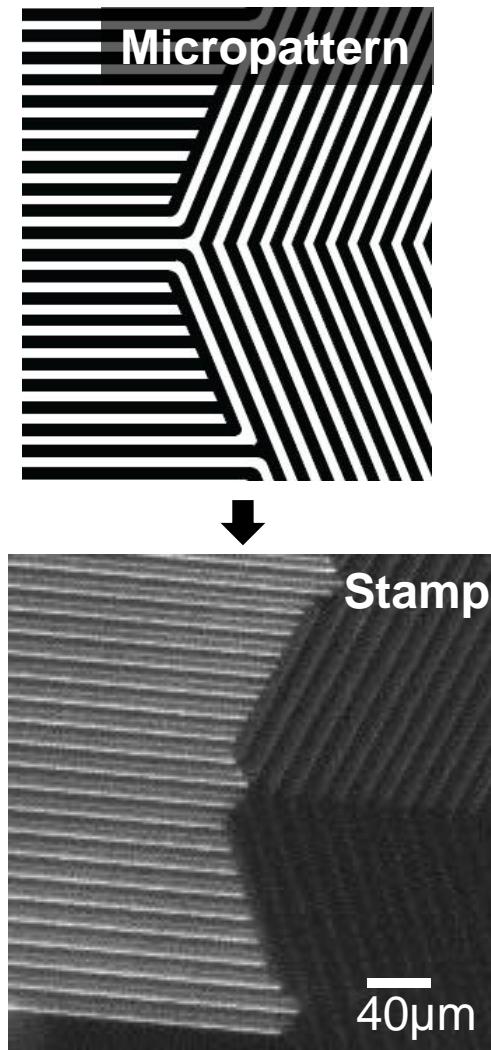
Anisotropic muscle layout



100µm

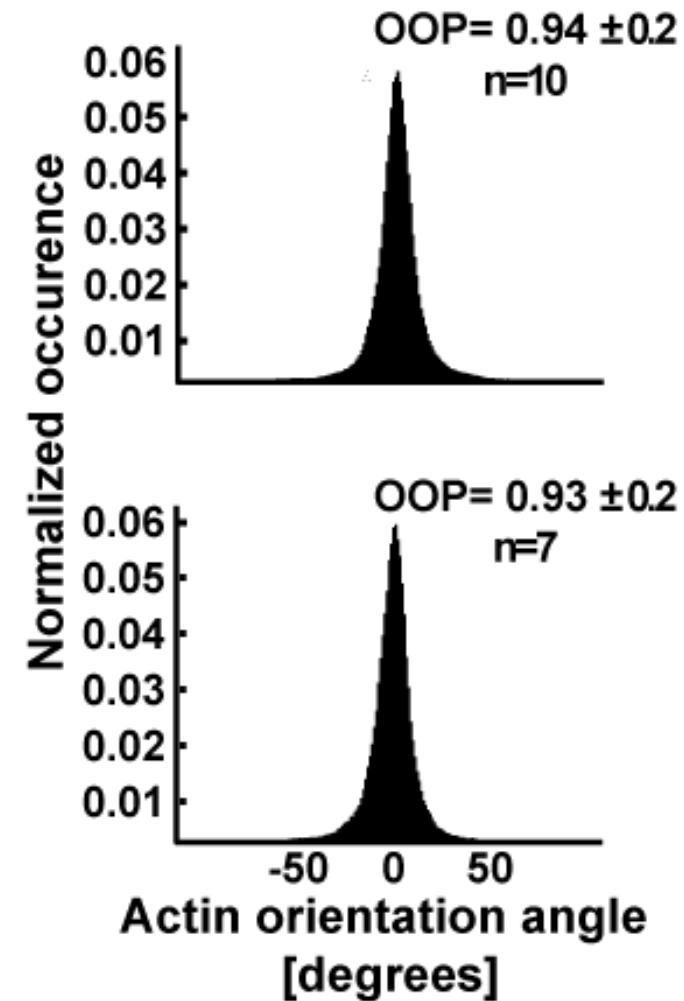
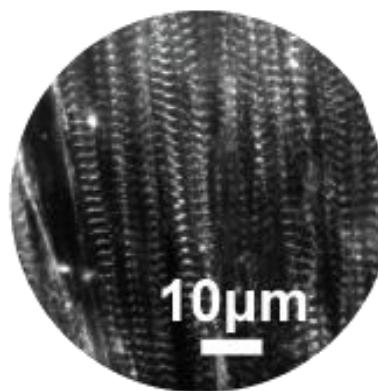
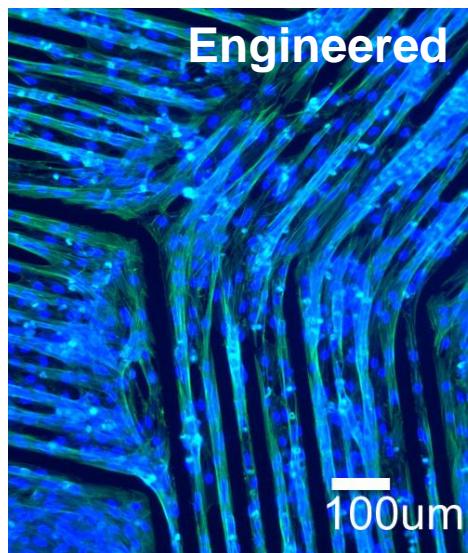
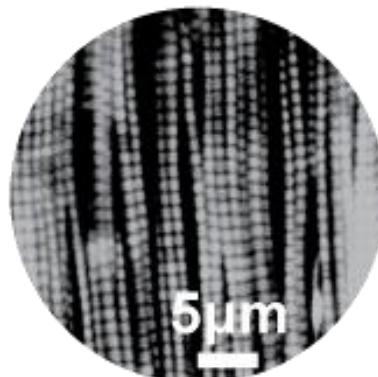
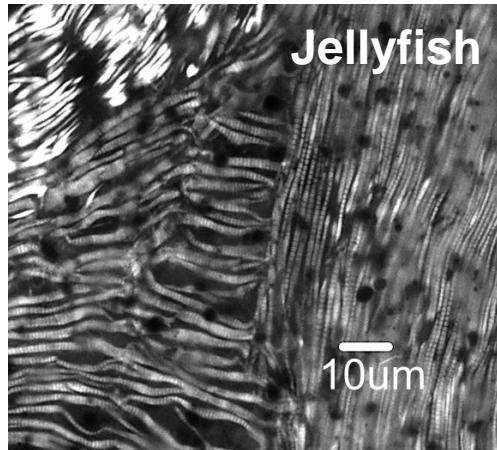


# Muscle fiber alignment

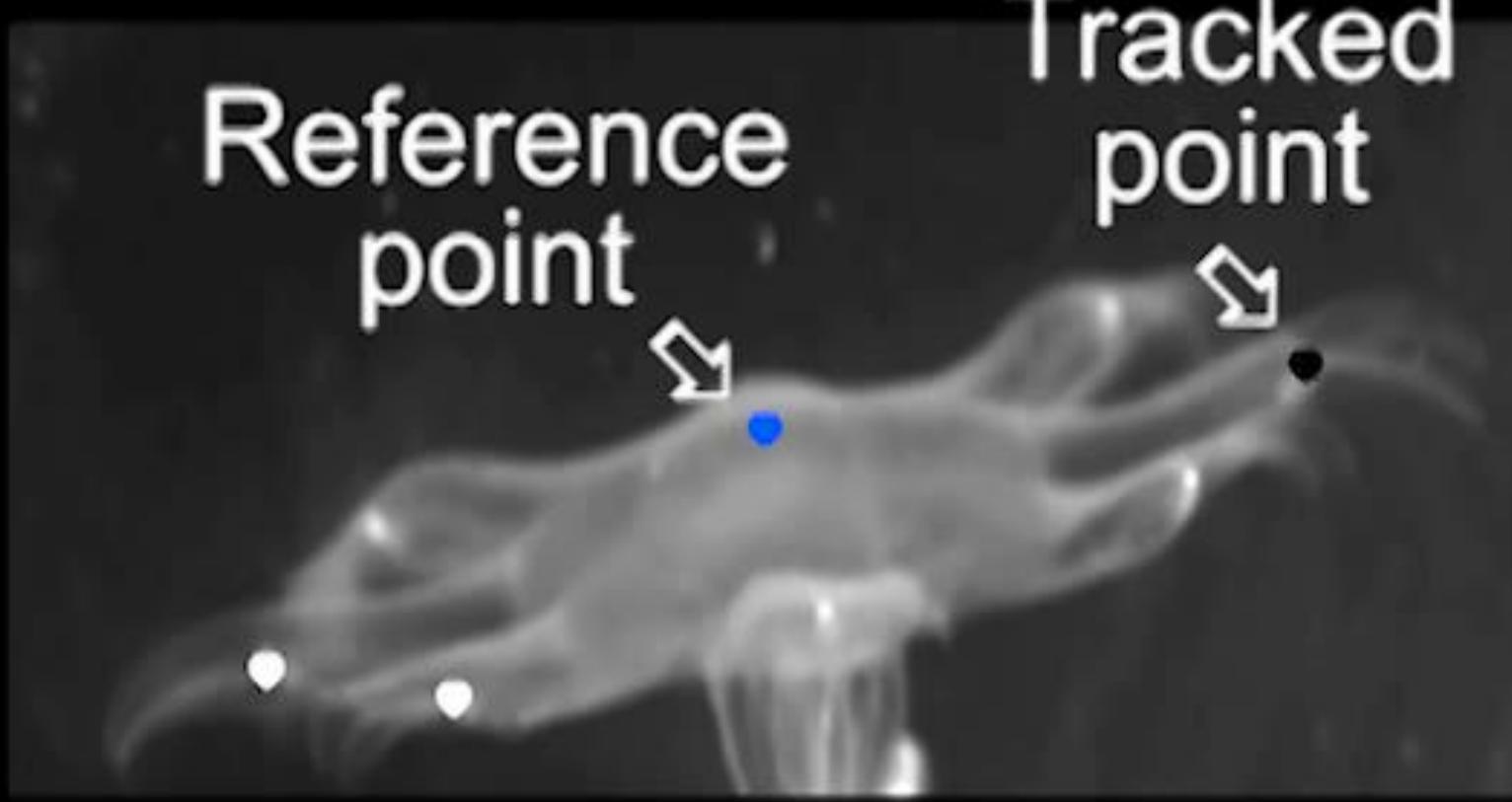




# Muscle fiber alignment



# Bell kinematics

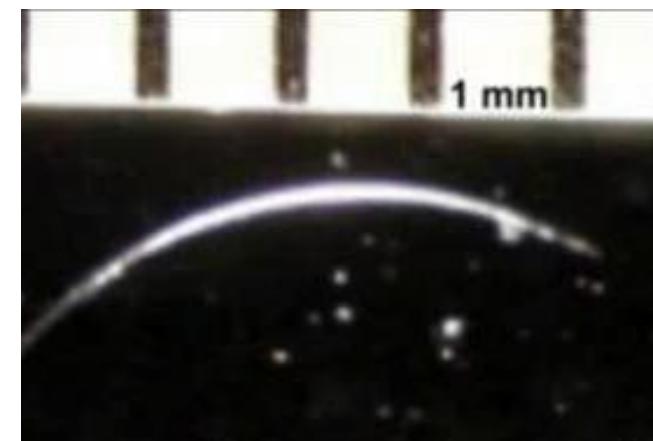
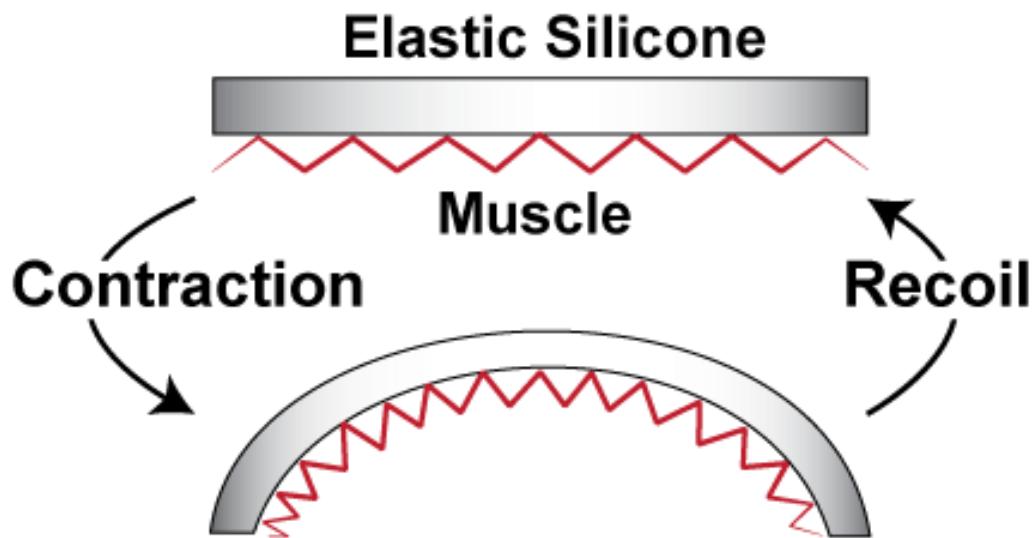


1mm



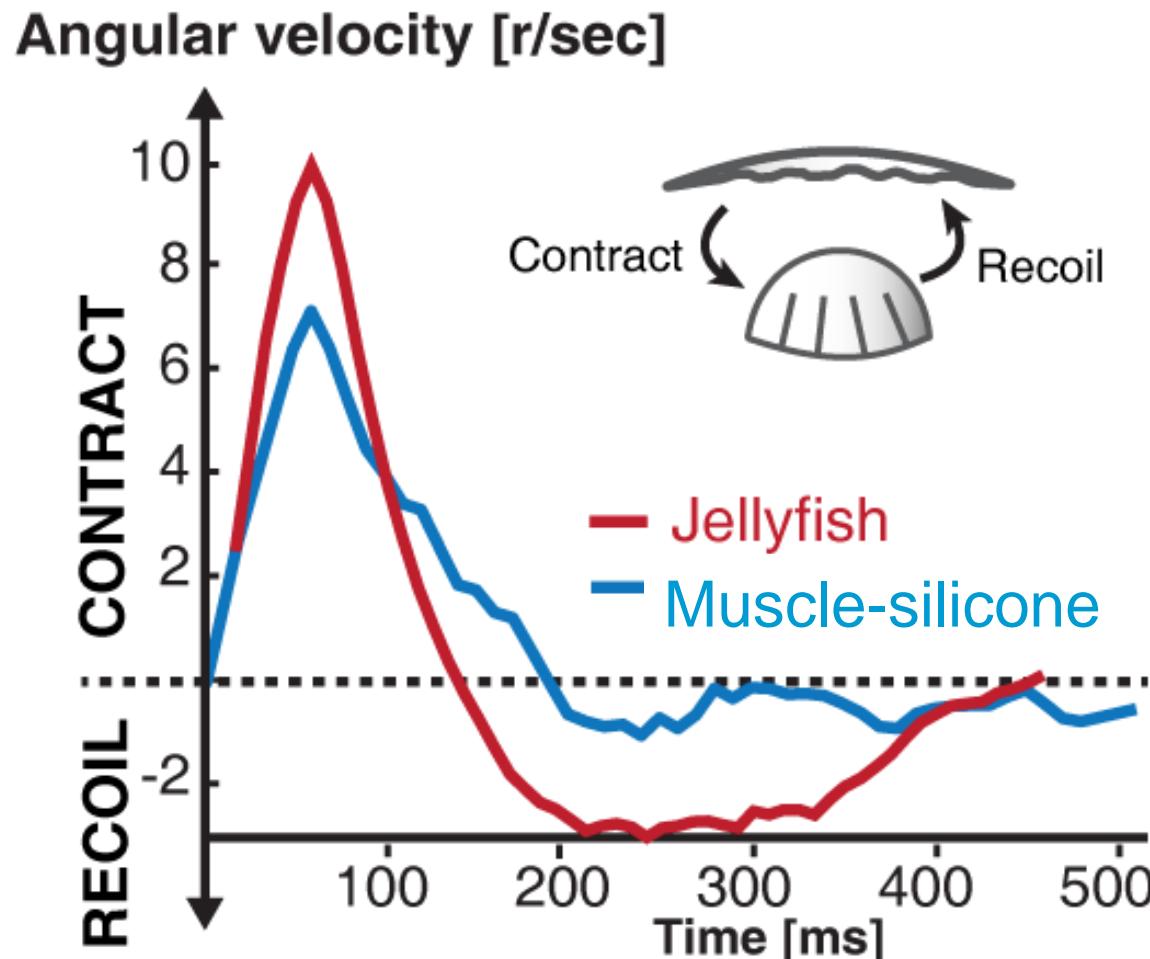
# Bell kinematics

Muscle-elastomer composite



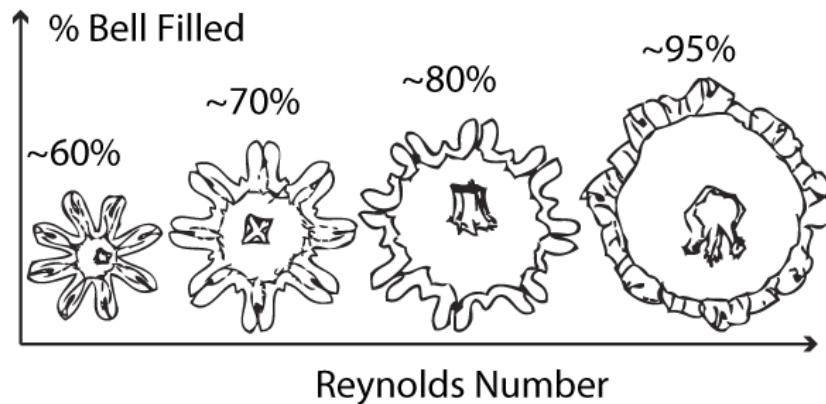


# Bell kinematics





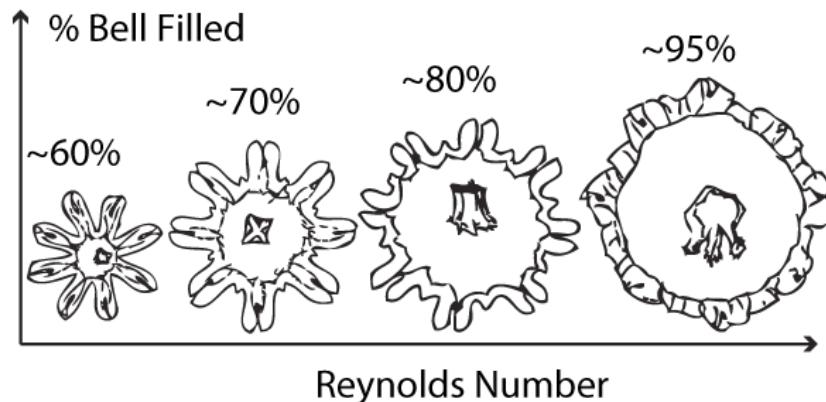
# Body geometry



Feitl, 2009; Nawroth, 2010

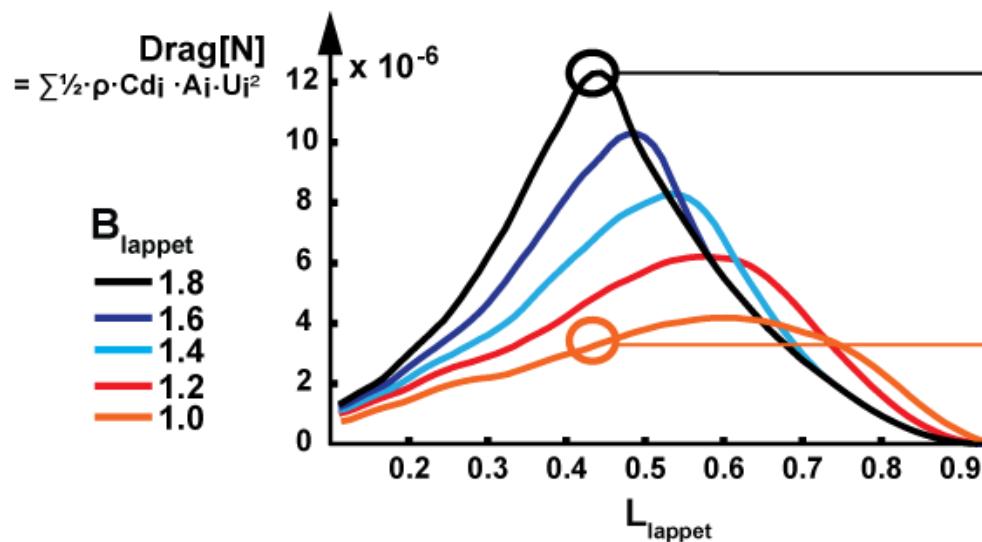


# Body geometry

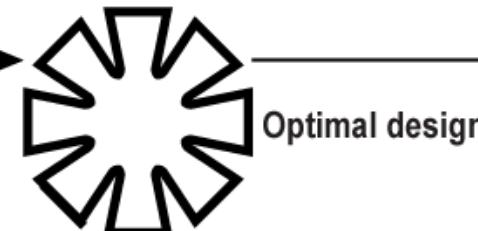


Feitl, 2009; Nawroth, 2010

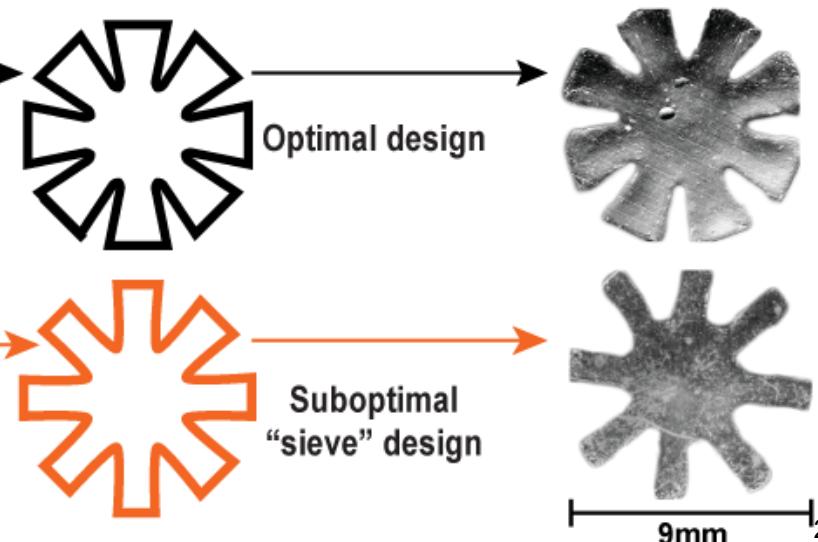
Predicted drag for medusoid geometries



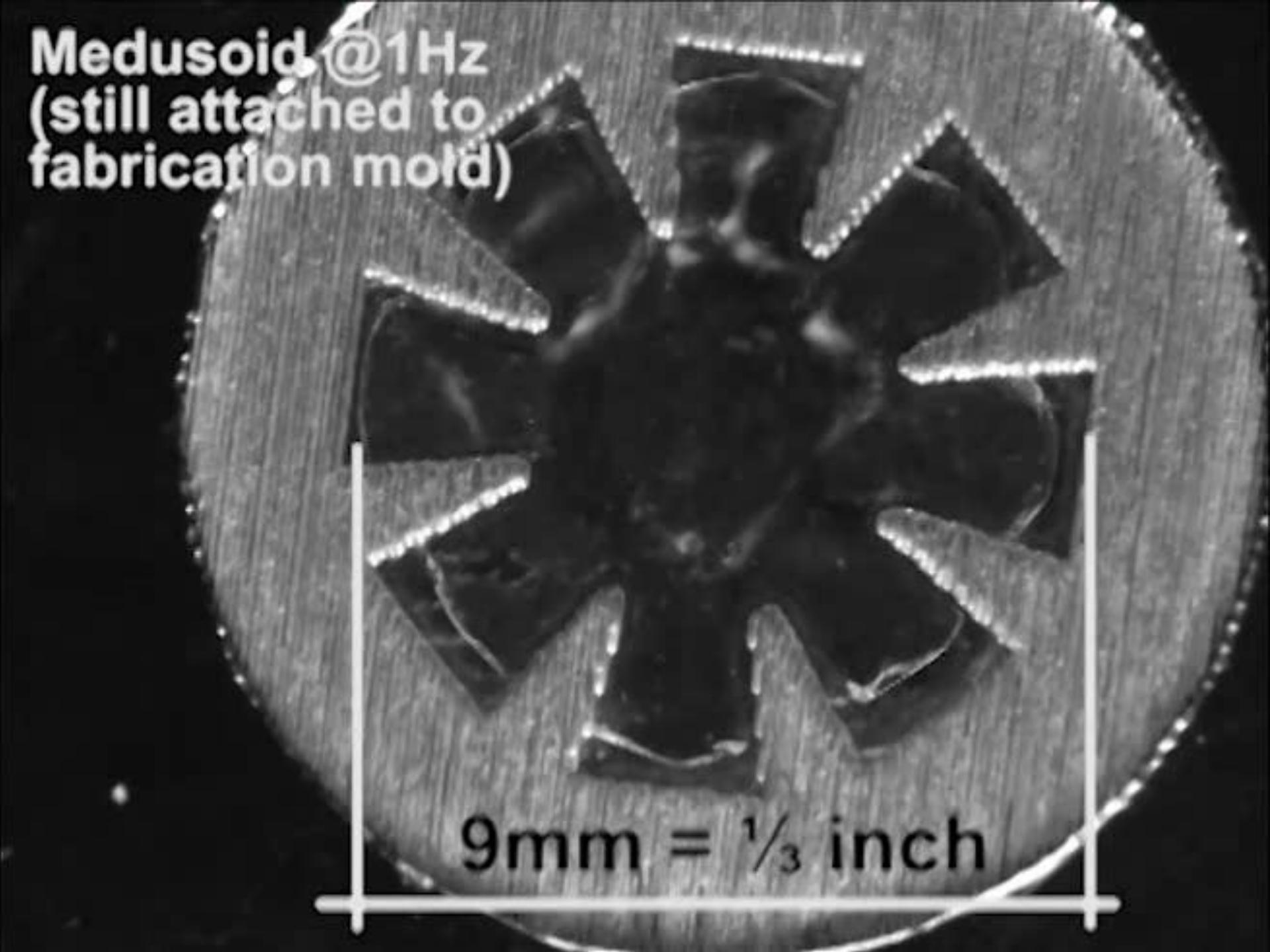
Optimal vs. suboptimal designs



Silicone implementation



Medusoid @1Hz  
(still attached to  
fabrication mold)

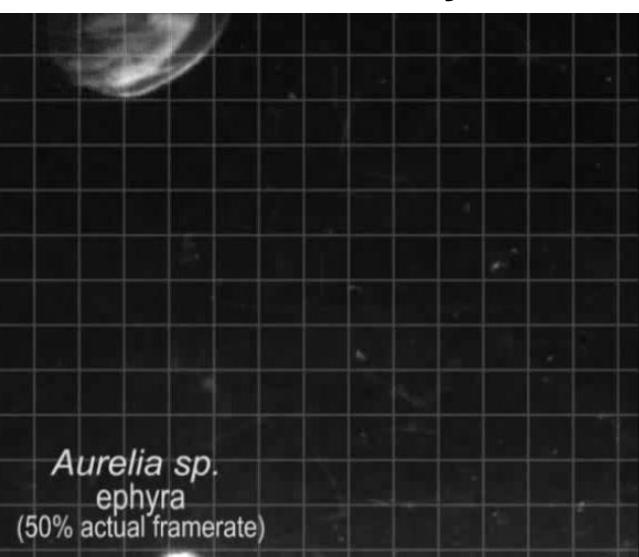


9mm =  $\frac{1}{3}$  inch



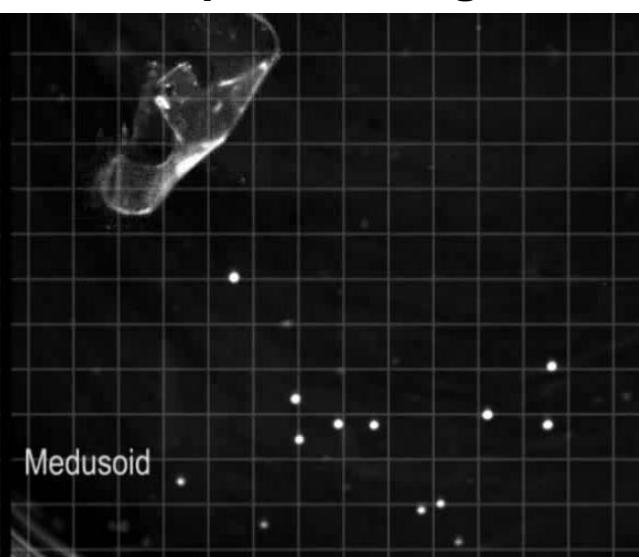
# Real and artificial jellyfish propulsion

Control: Jellyfish



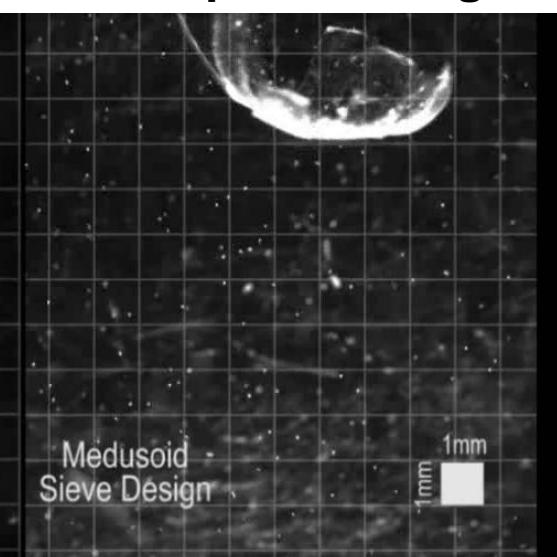
*Aurelia sp.*  
ephyra  
(50% actual framerate)

Optimal design



Medusoid

Suboptimal design



Medusoid -  
Sieve Design

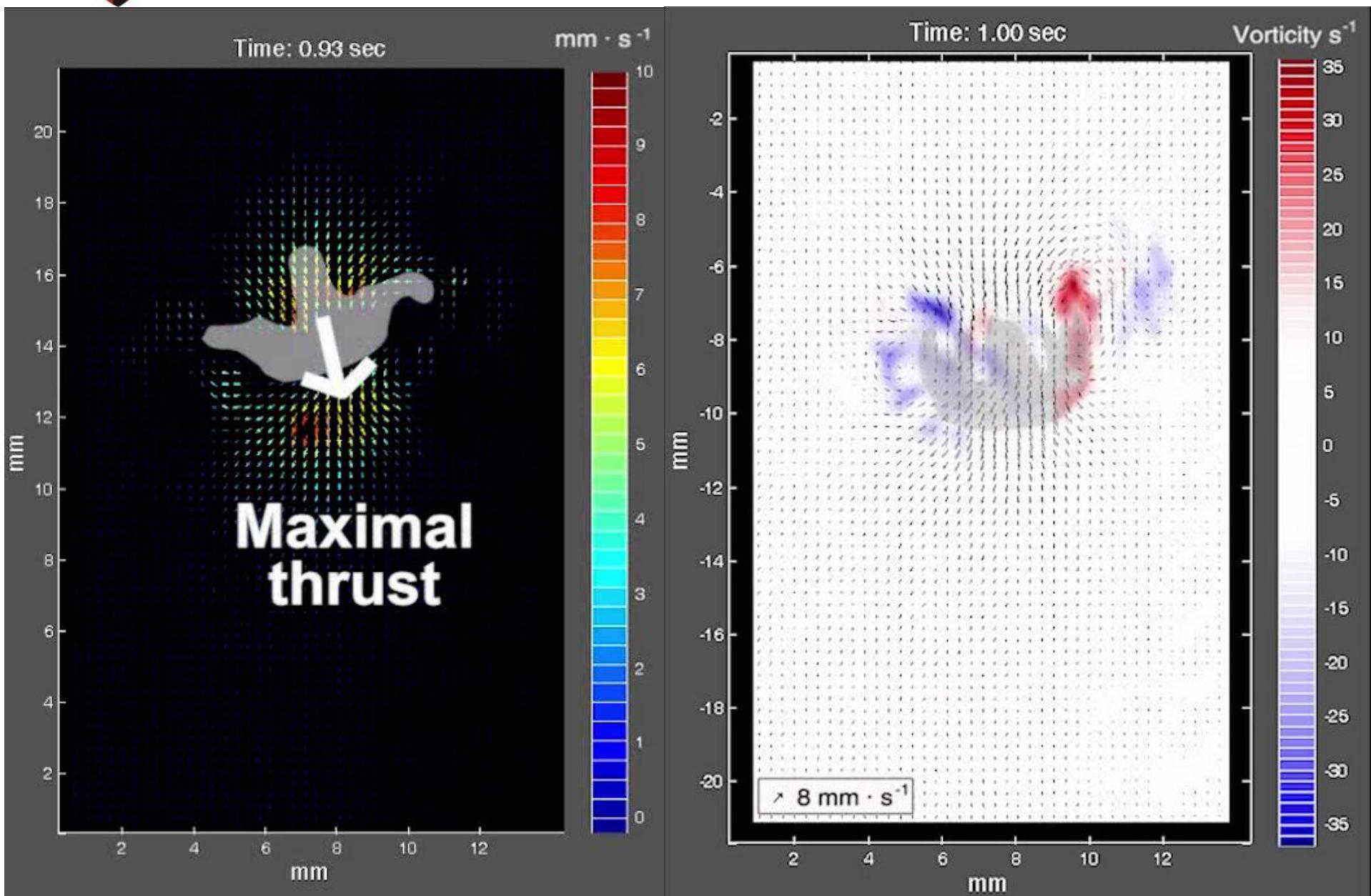
1mm

# Quantifying feeding and propulsion currents



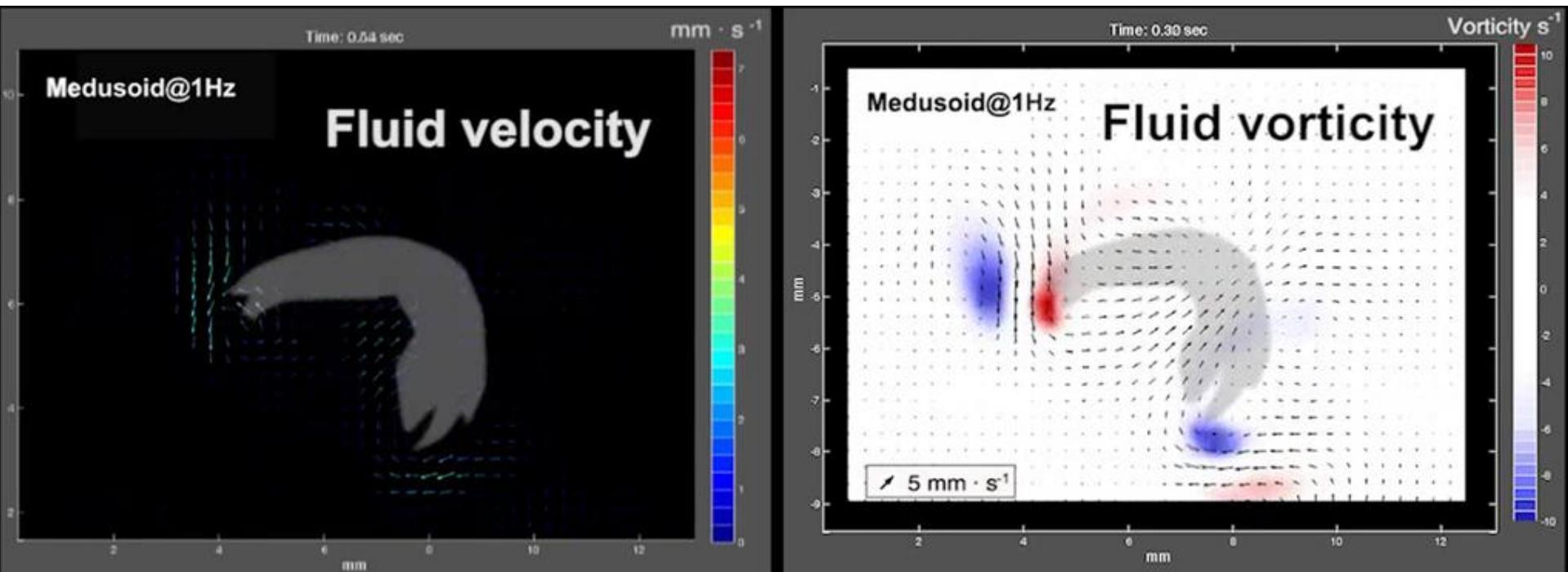


# Control: Flow field in jellyfish





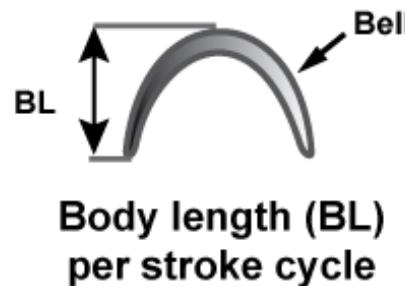
# Flow field in optimal Medusoid



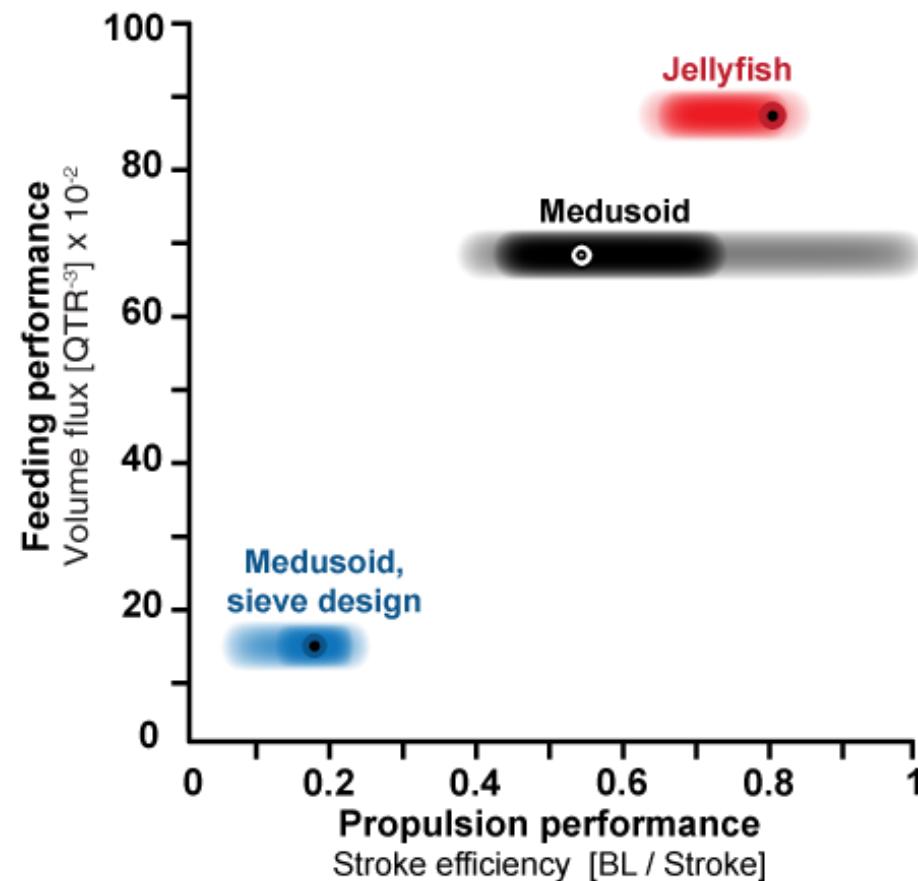


# Fitness metrics

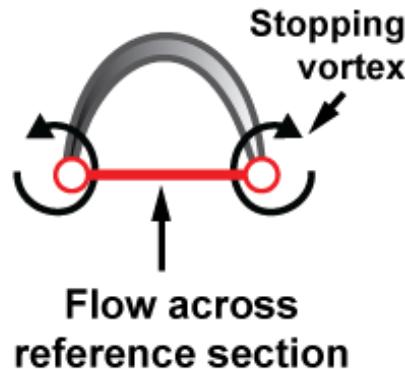
a Propulsion metric



c Performance matrix

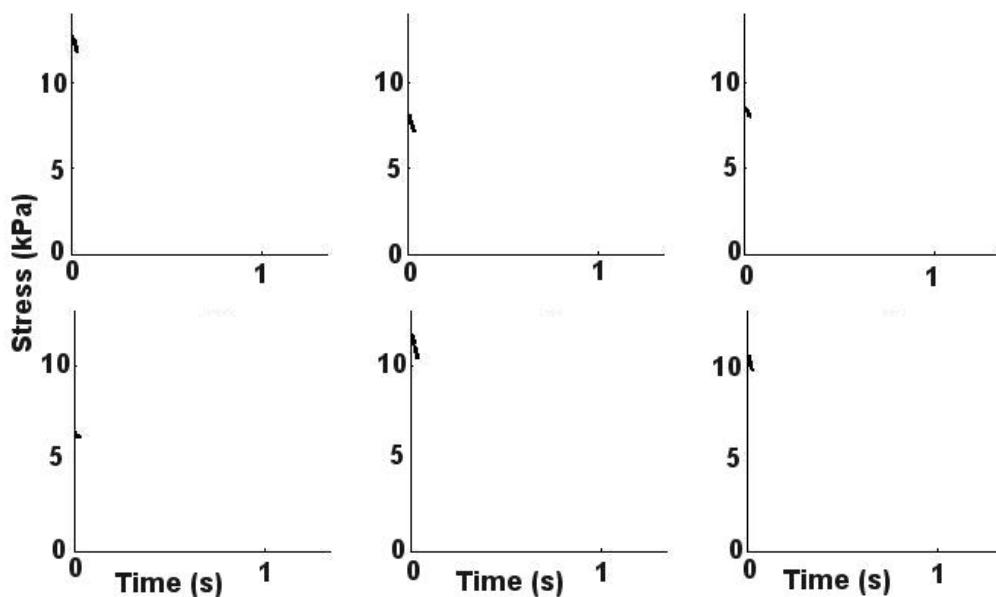
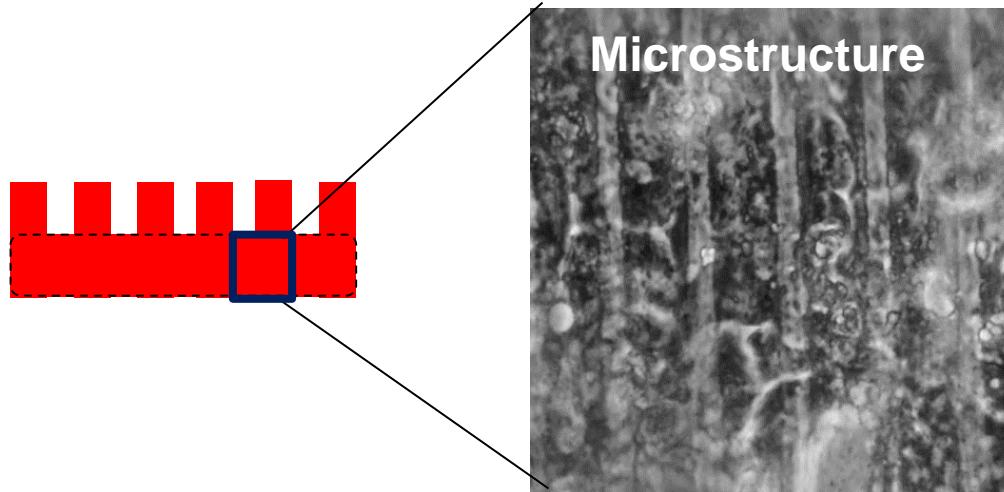


b Feeding metric

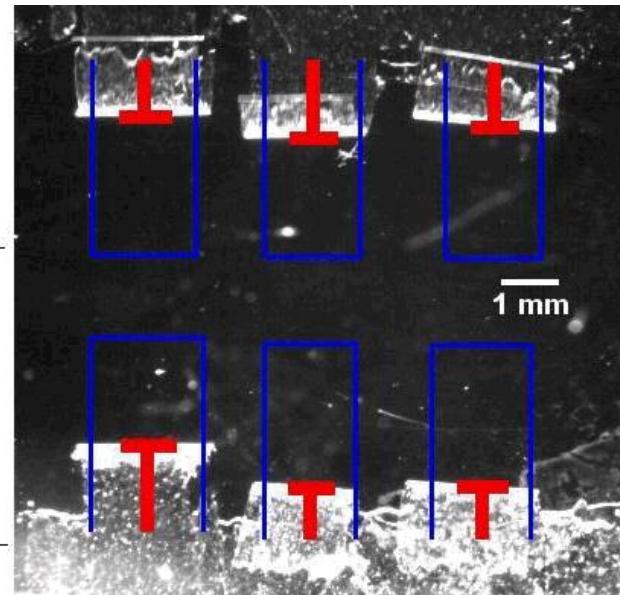




# Heart-on-a-chip: further reduction



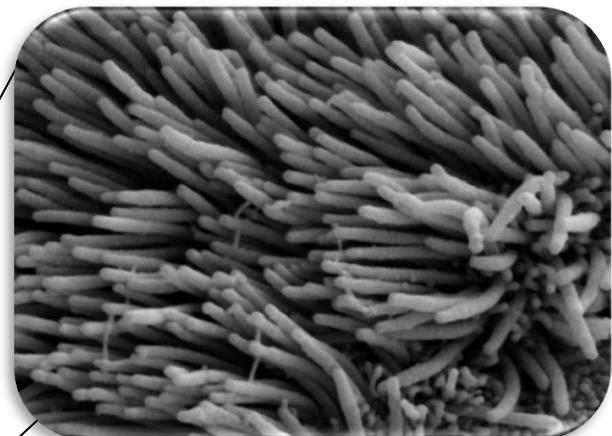
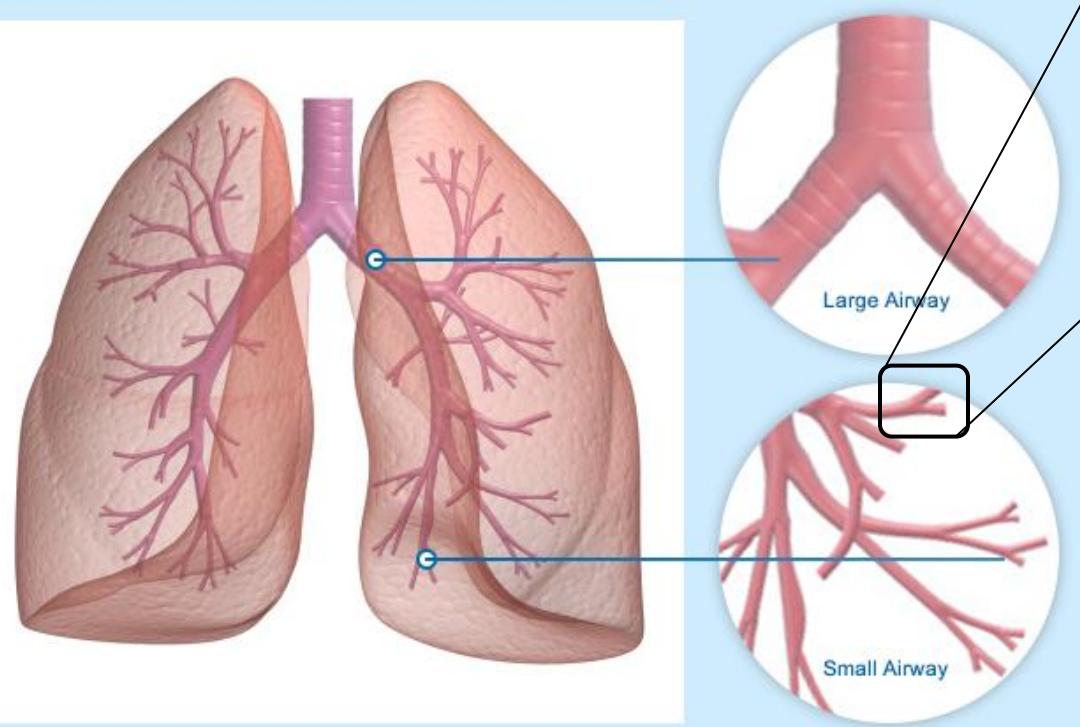
Tissue contractile stress





# Case study: ciliated epithelium

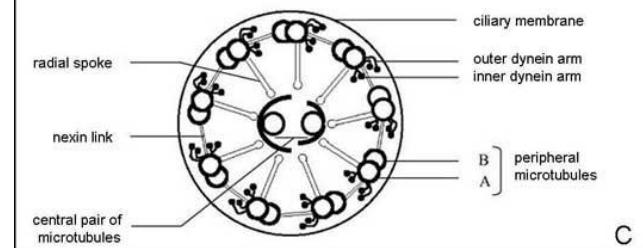
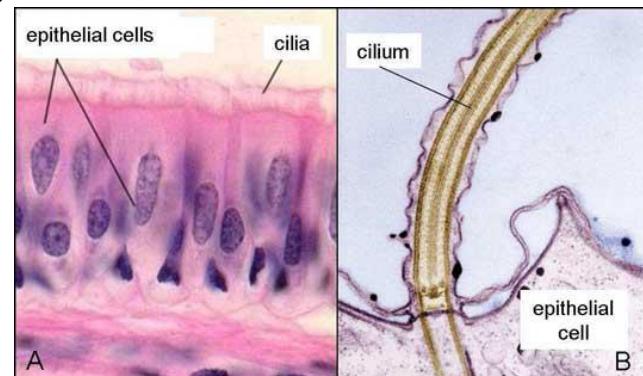
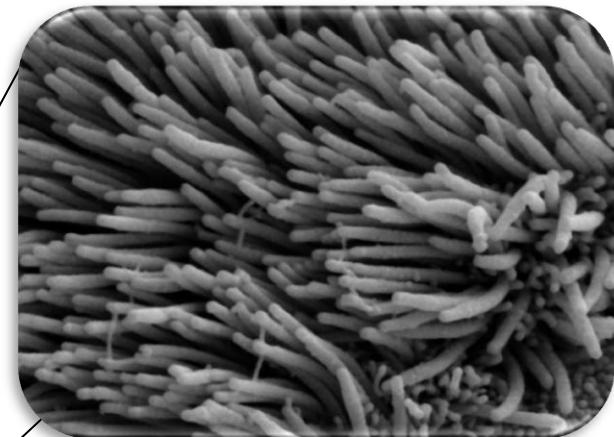
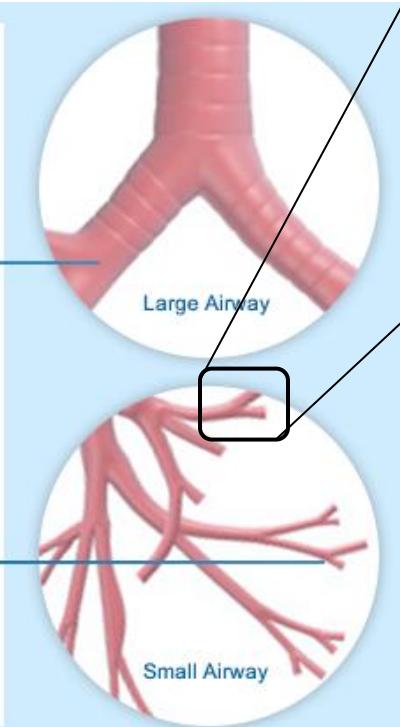
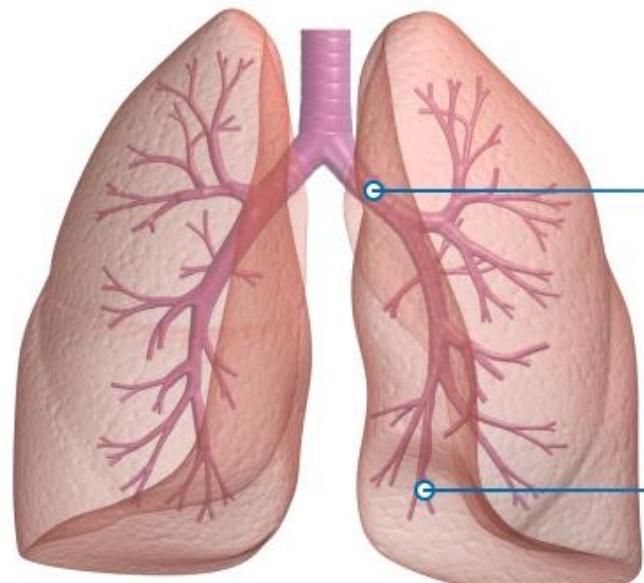
e.g., respiratory epithelium





# Case study: ciliated epithelium

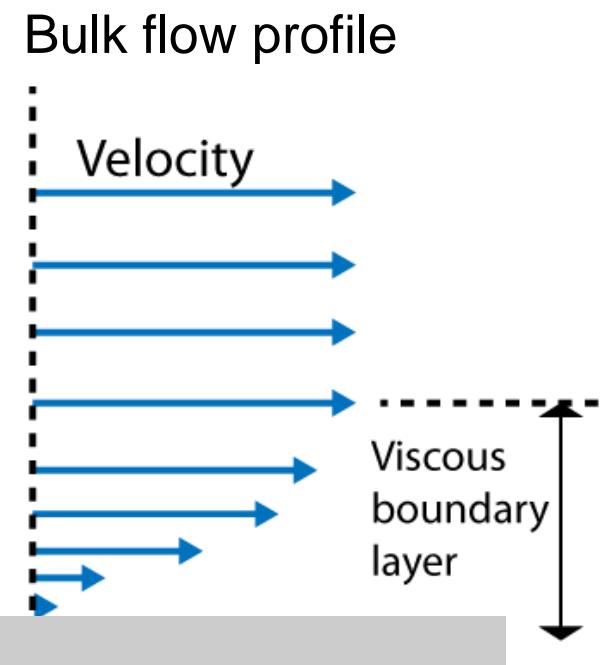
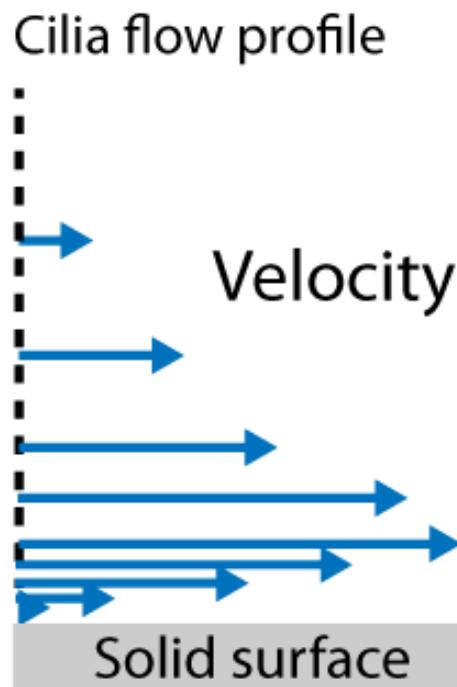
e.g., respiratory epithelium





# Cilia-powered fluid transport

Ciliated epithelium (Paramecium)



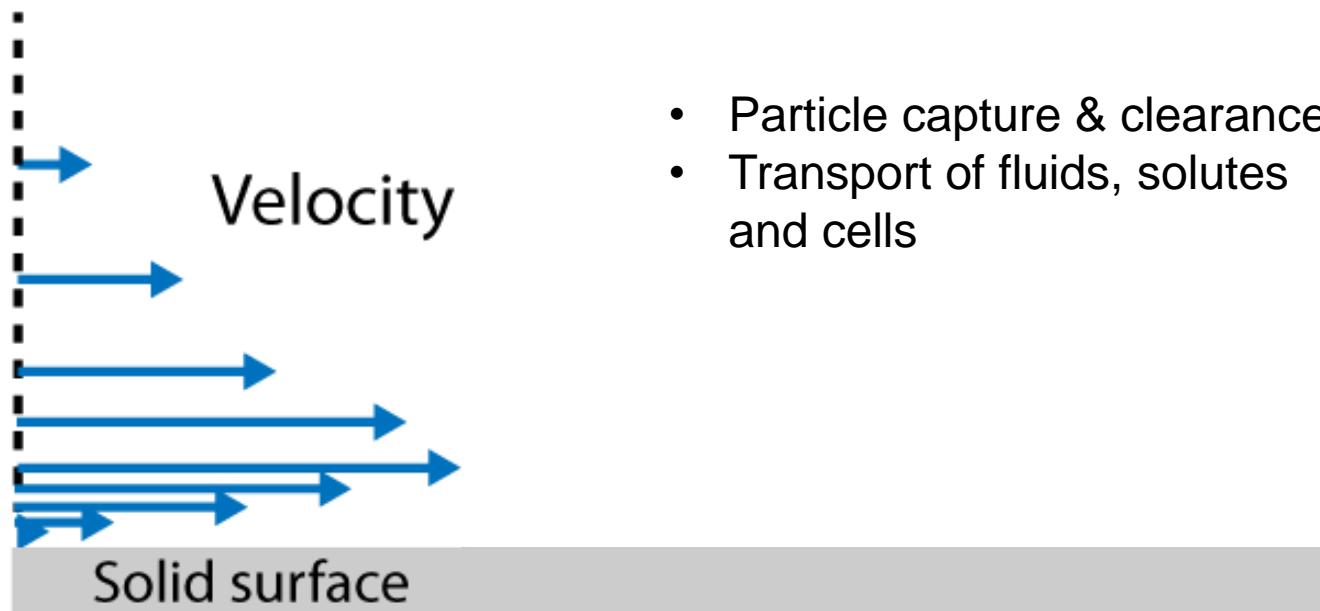


# Cilia-powered fluid transport

Ciliated epithelium (Paramecium)

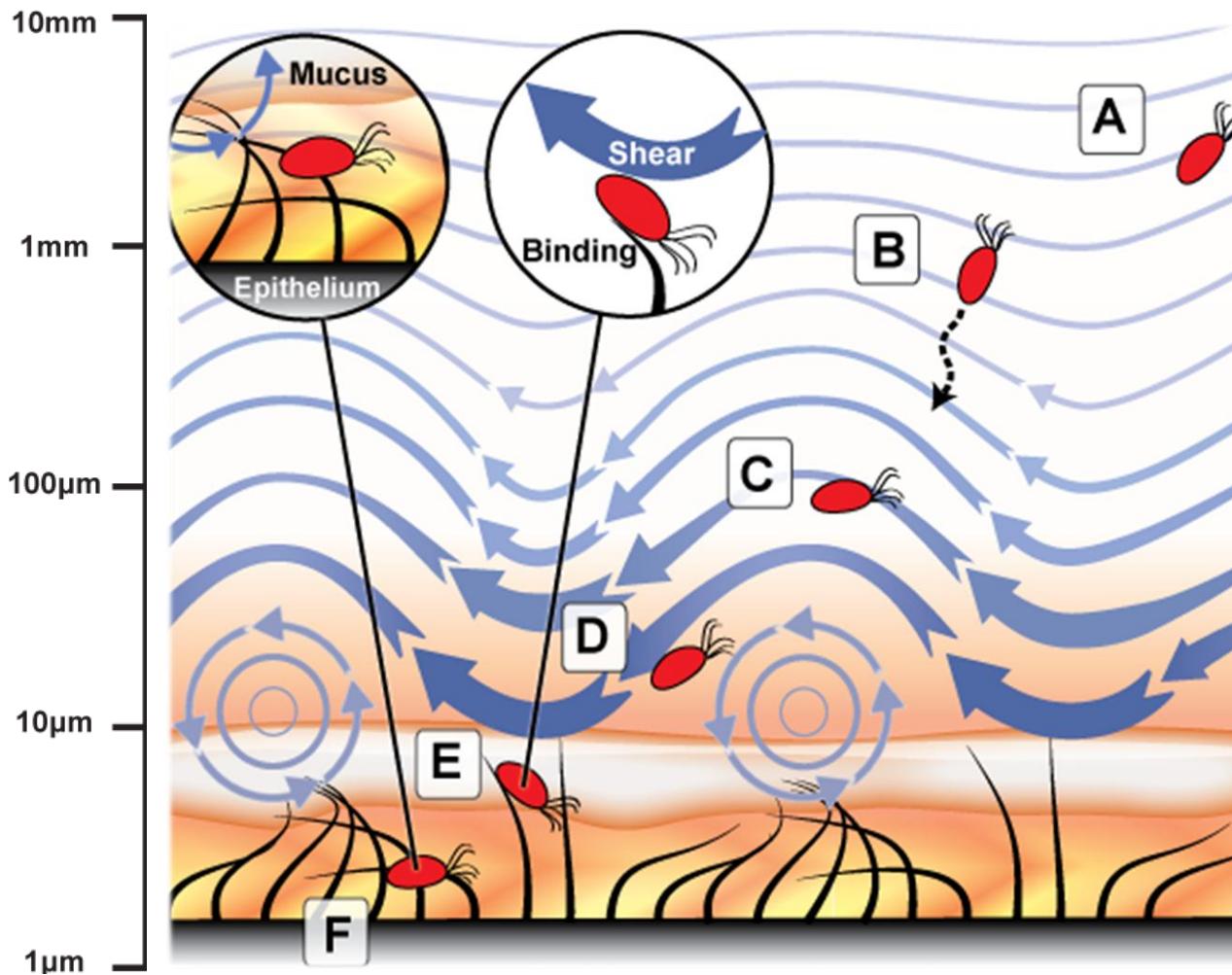


Cilia flow profile





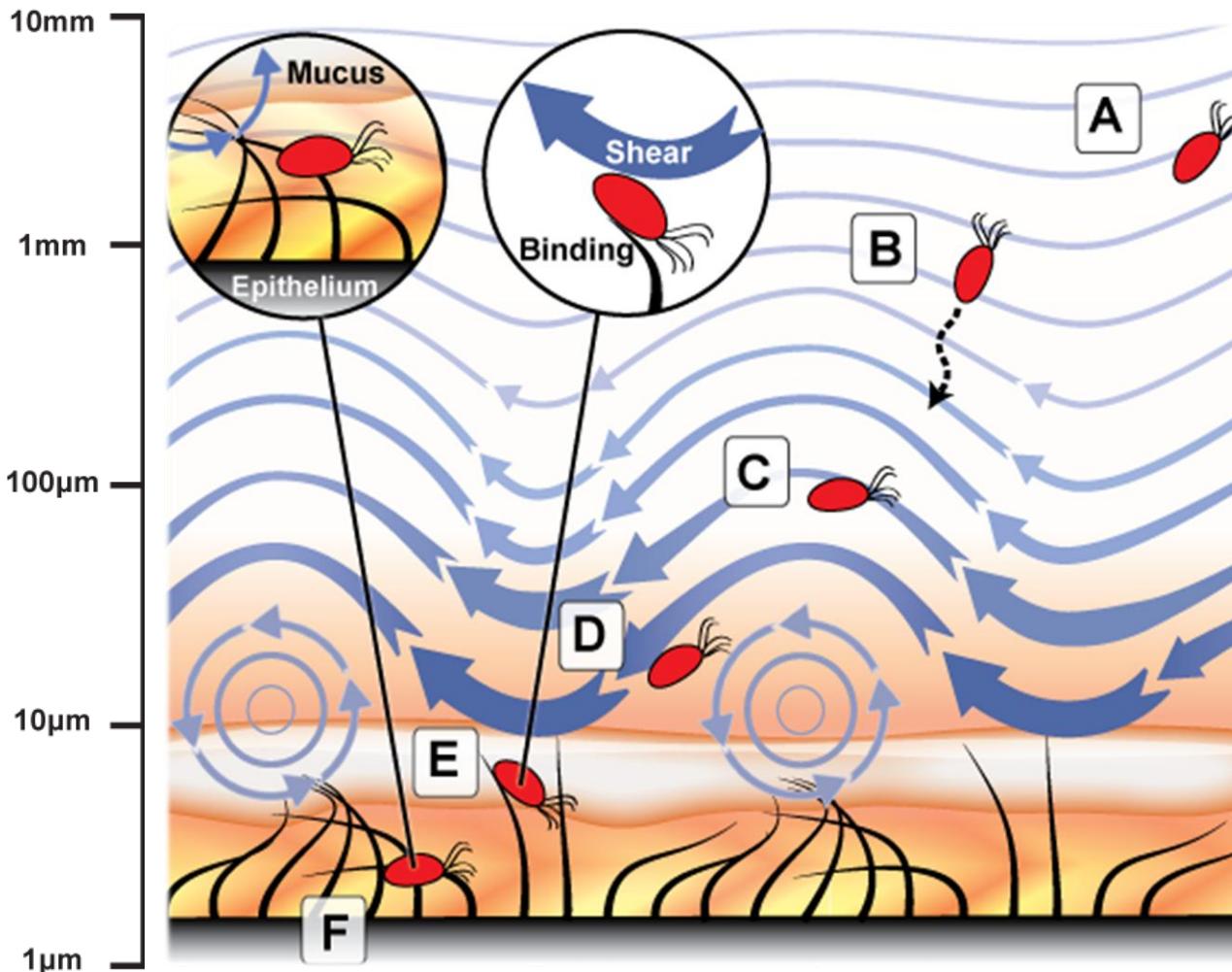
# Structure-function relationships of ciliated surfaces



**Cilia fitness**  
Selective bacterial recruitment



# Structure-function relationships of ciliated surfaces



## Cilia fitness

Selective bacterial recruitment



Fluid transport  
and mixing



Surface geometry

+

Metachronal wave

+

Ciliary structure and  
kinematics

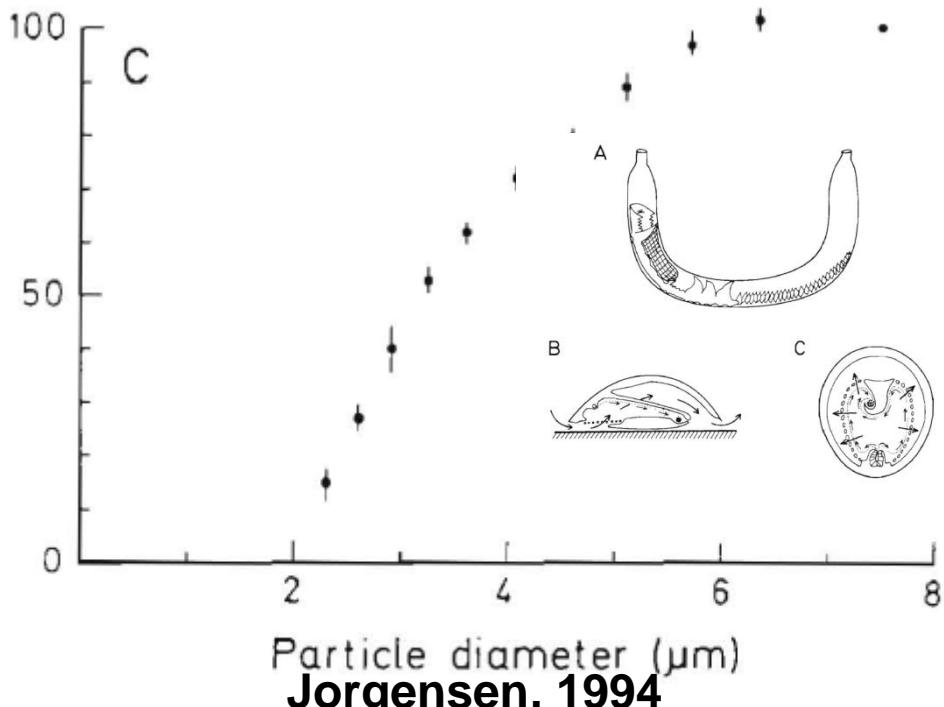


# A master of selective bacterial recruitment: The Hawaiian bobtail squid

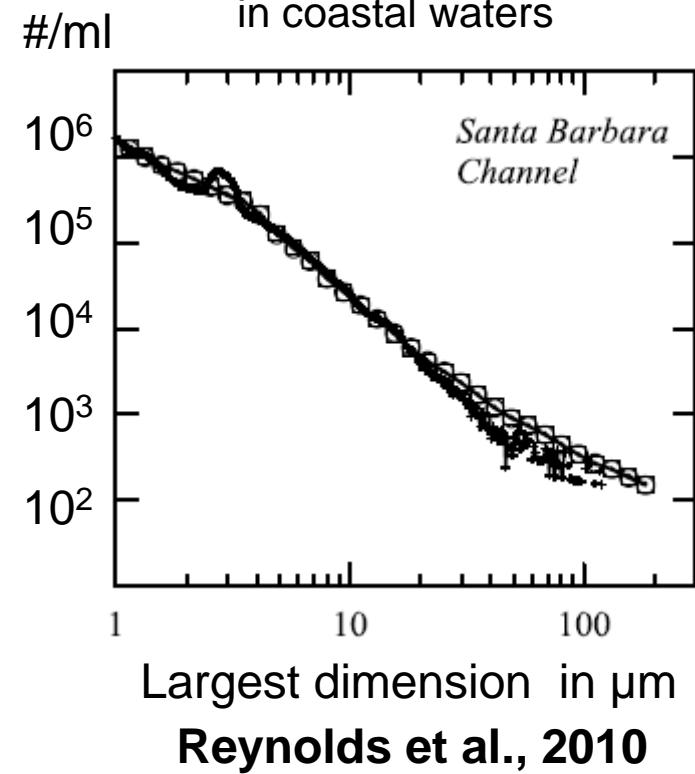


Internal ciliated organ captures 1  $\mu\text{m}$  bacteria species (*Vibrio fischeri*) from huge microbial background (0.5%), and wide range of particle sizes

Retention efficiency in typical ciliary filter feeders

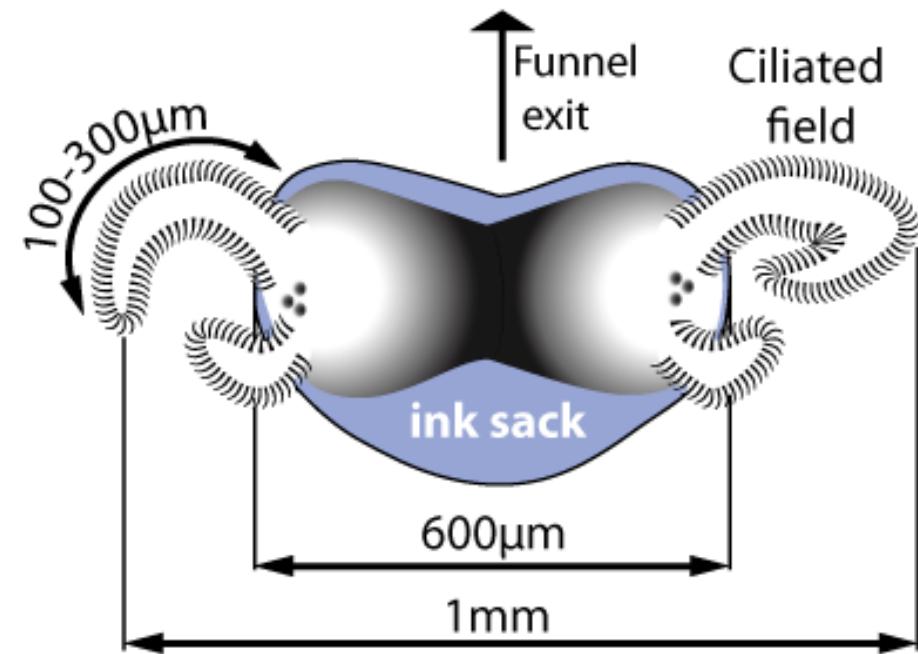
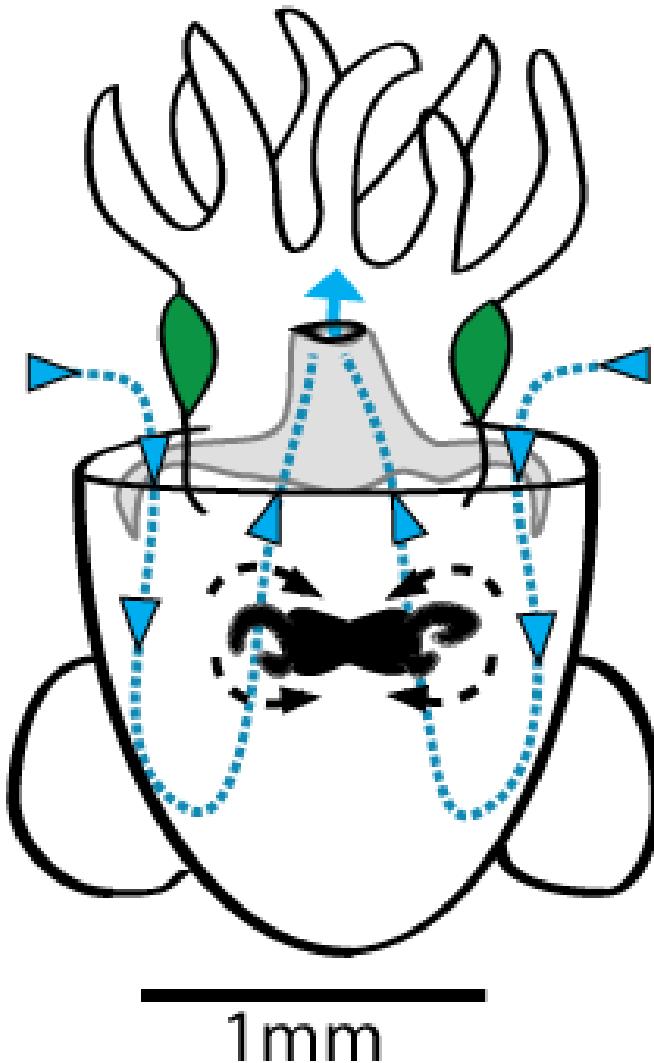


Particle size distribution  
in coastal waters





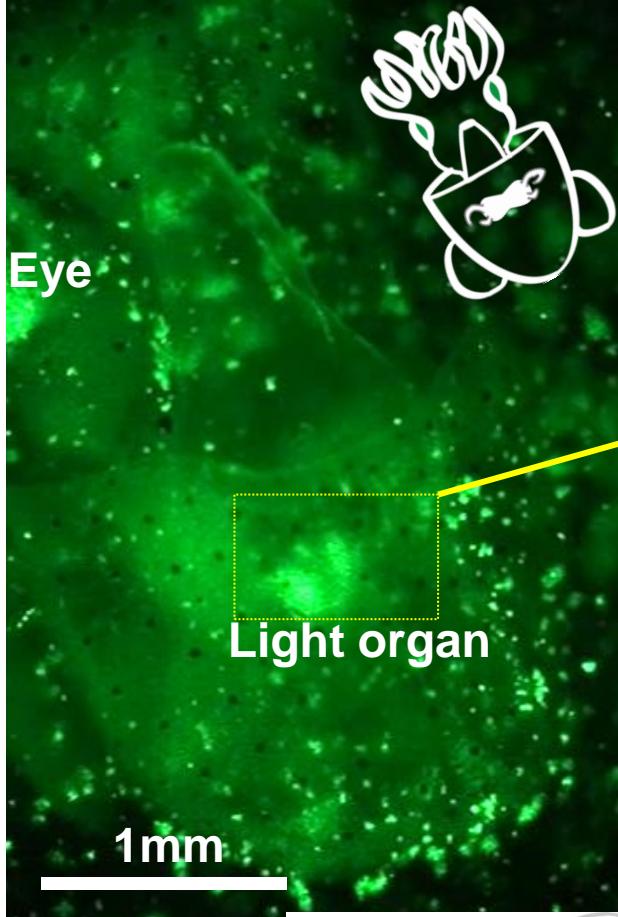
# The squid ciliated organ



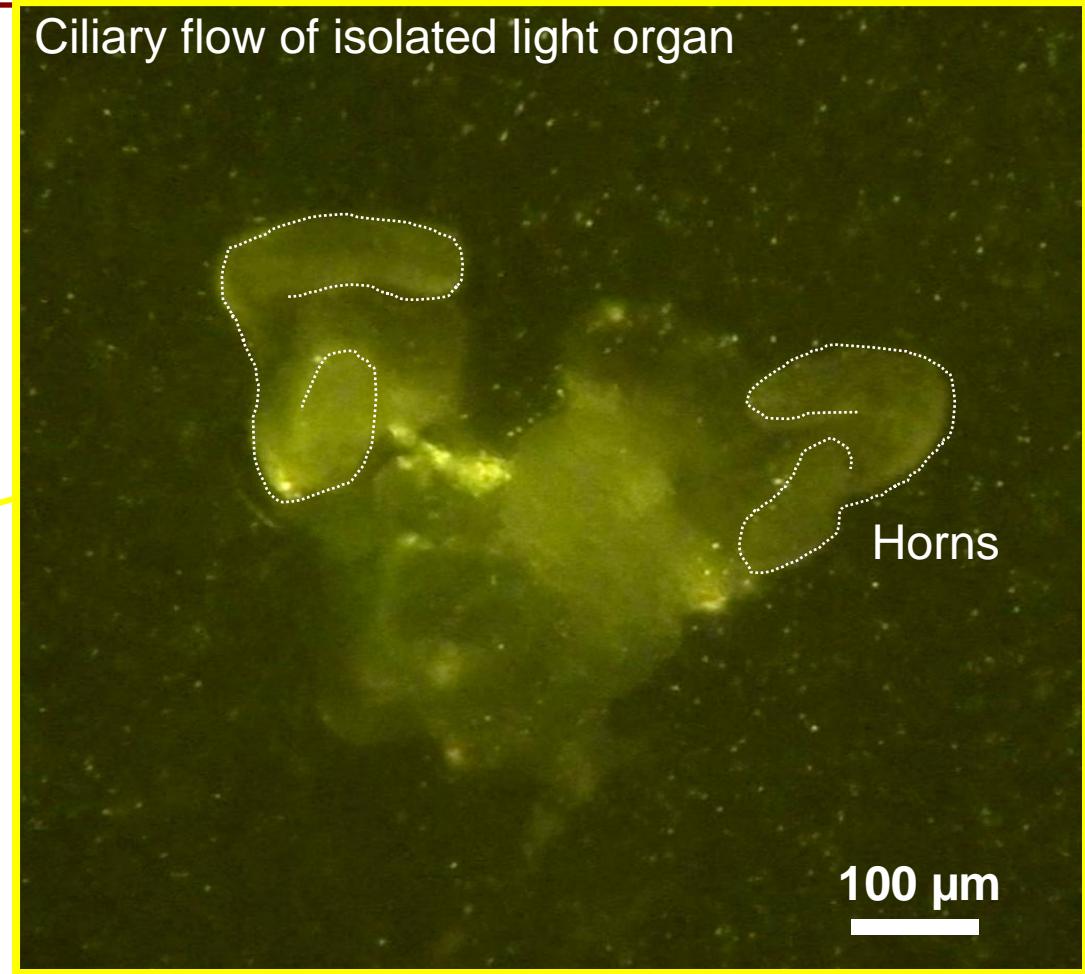


# The light organ is subject low Reynolds number flow

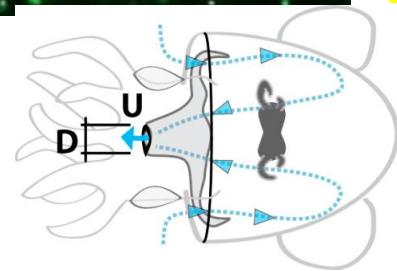
Light organ in mantle flow



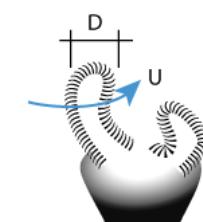
Ciliary flow of isolated light organ

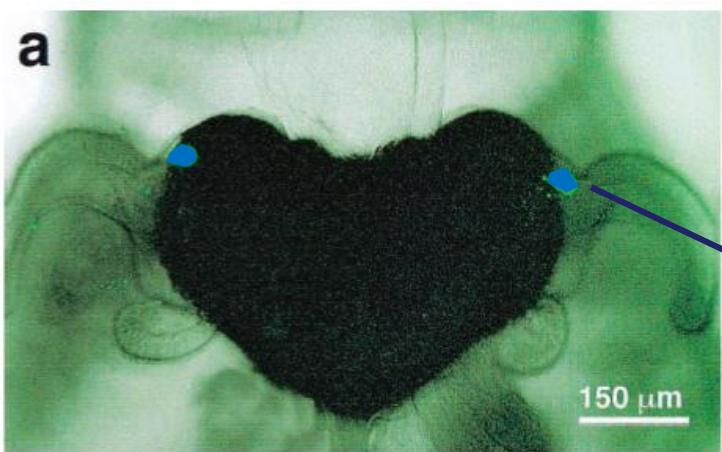


$$Re = \frac{UD}{\nu} = 0.2$$



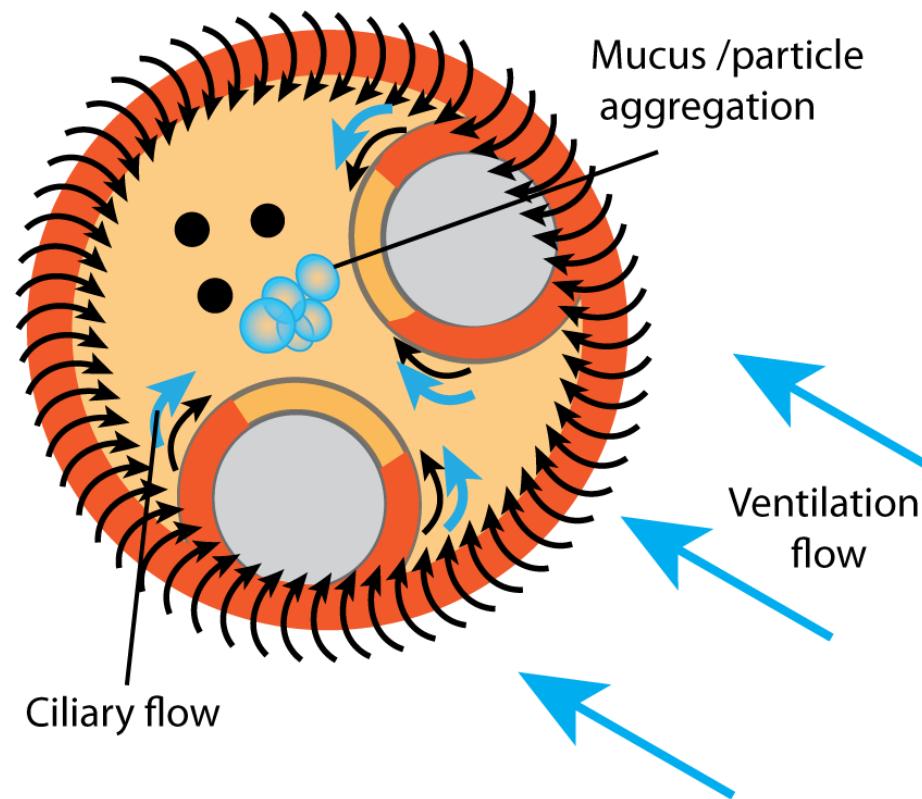
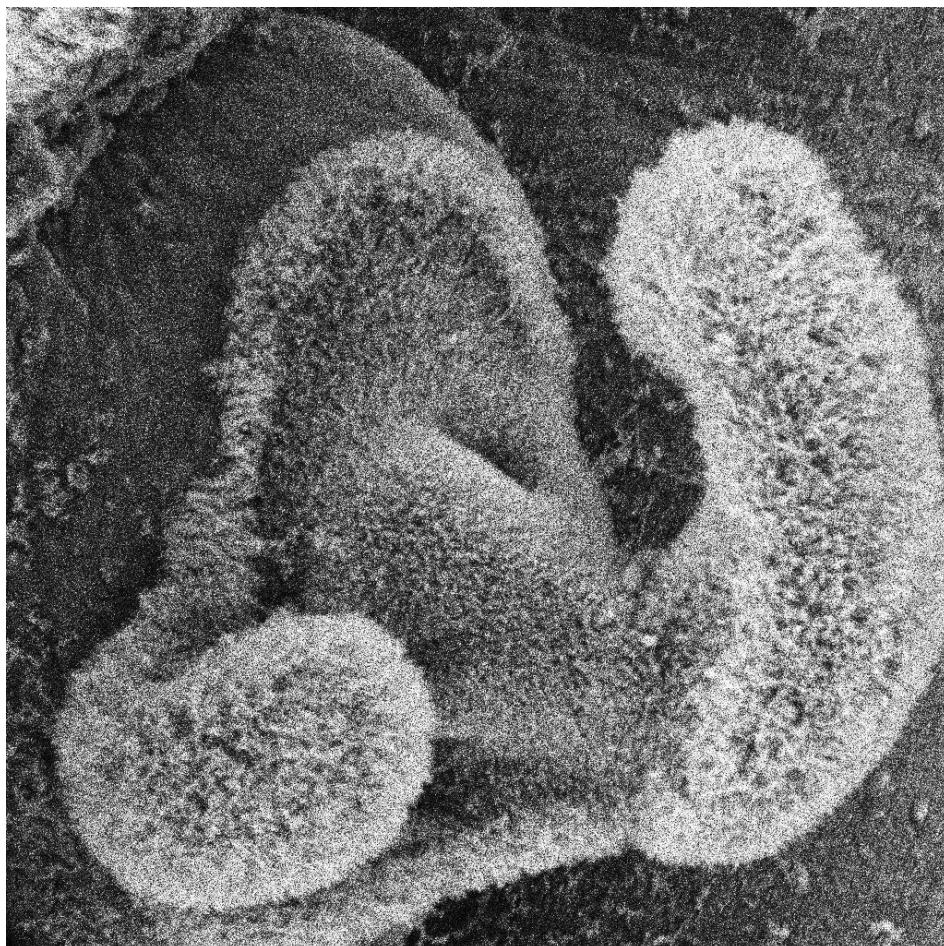
$$Re = \frac{UD}{\nu} = 0.004$$



**a**

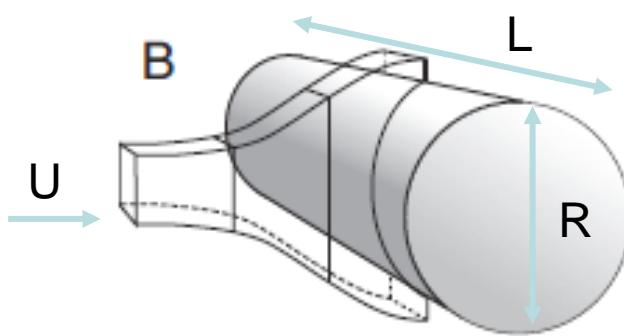
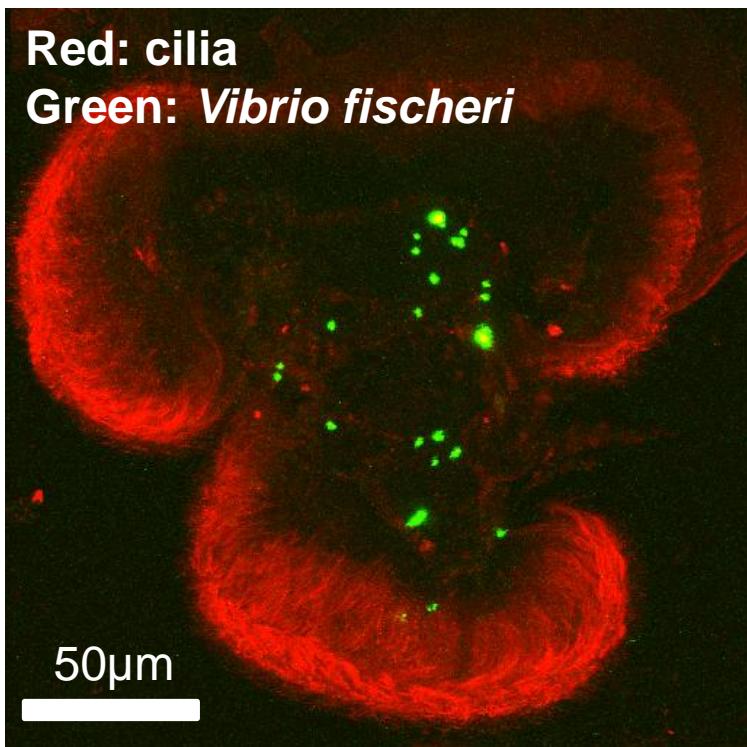
## The mucociliary trap

Mucus aggregation



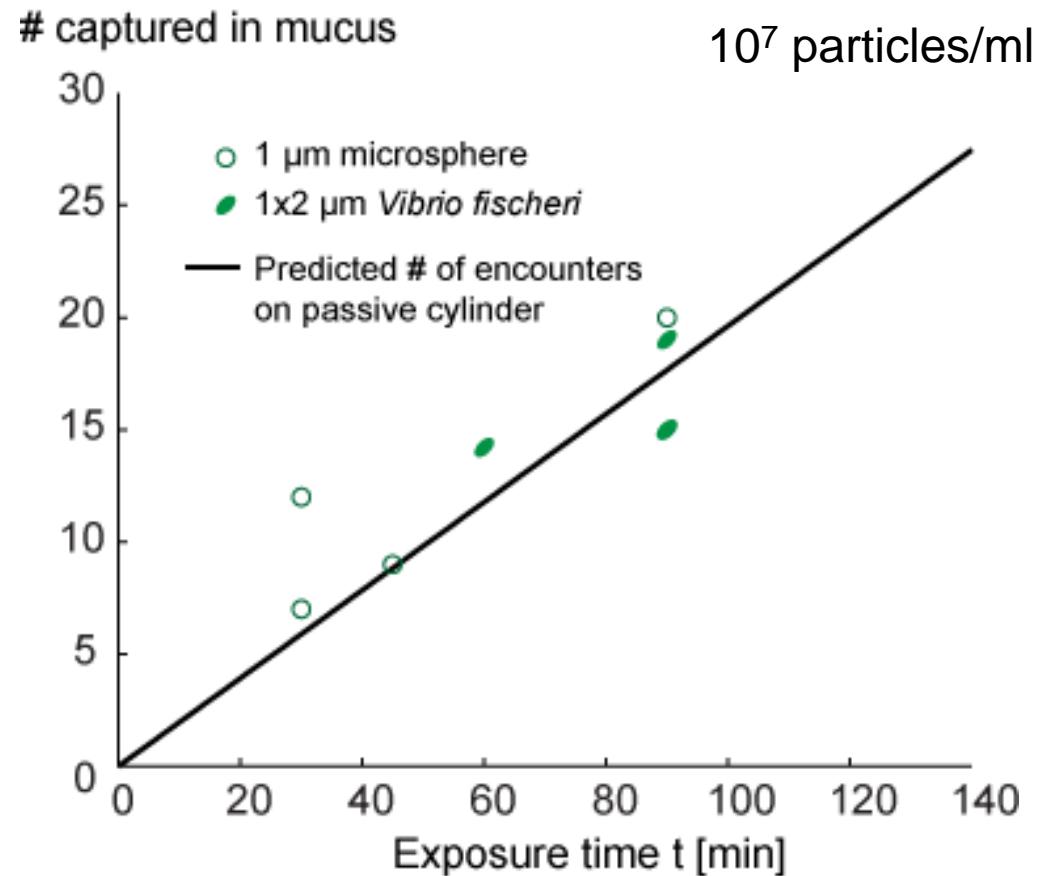
Red: cilia

Green: *Vibrio fischeri*



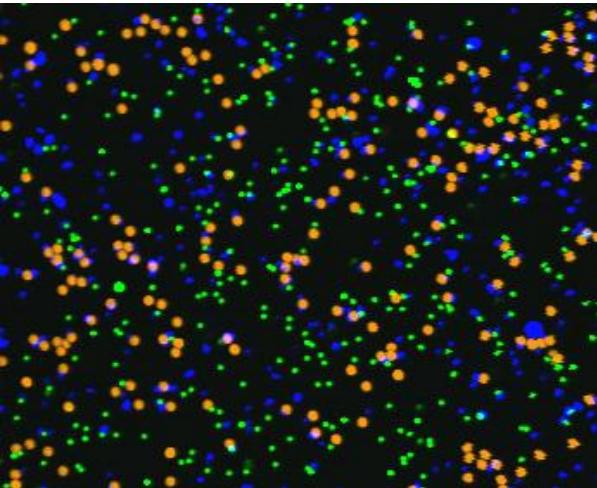
Encounter model for passive cylinder in flow  
(Humphries, PNAS, 2009)

## The mucociliary trap: not such a good trap?

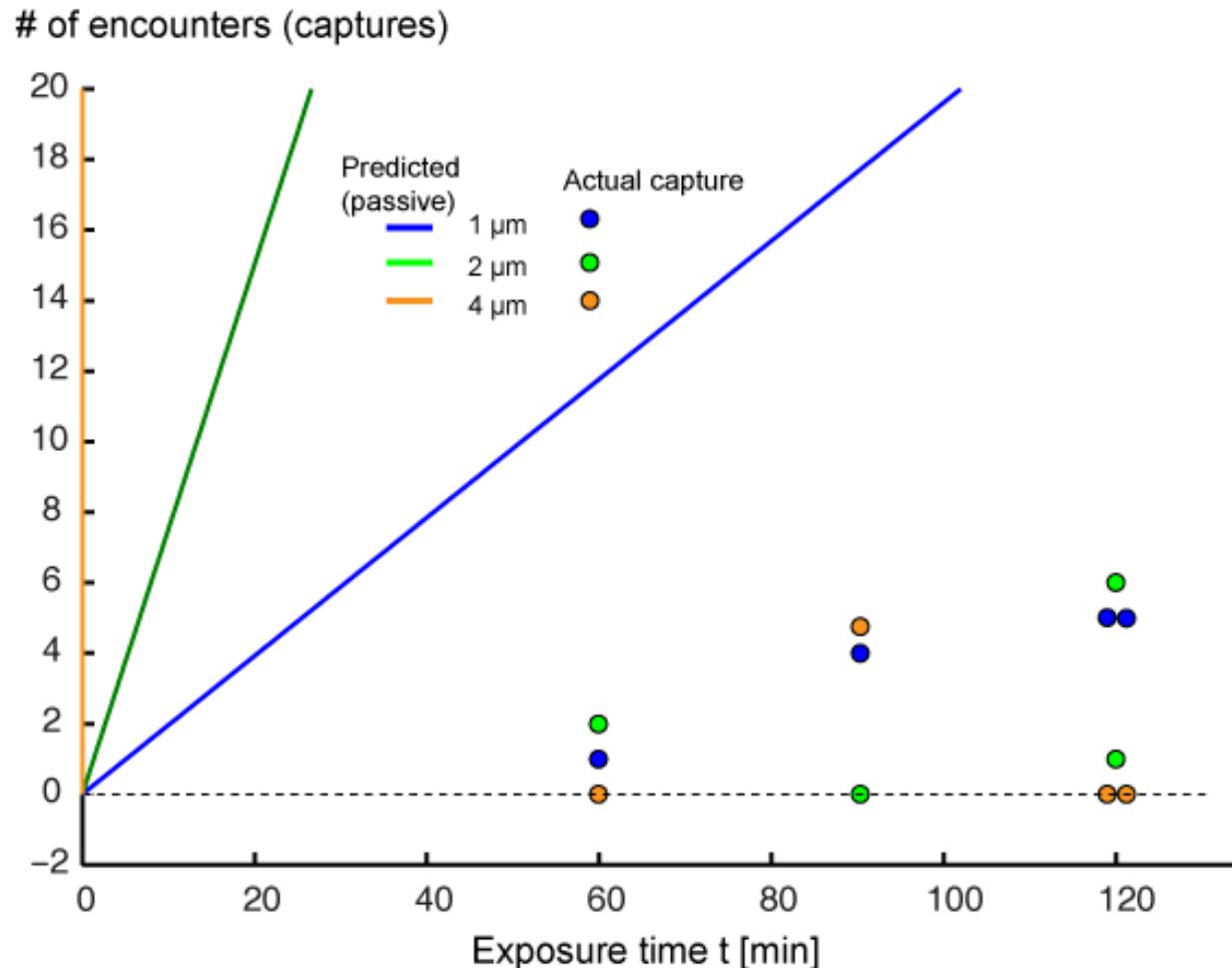
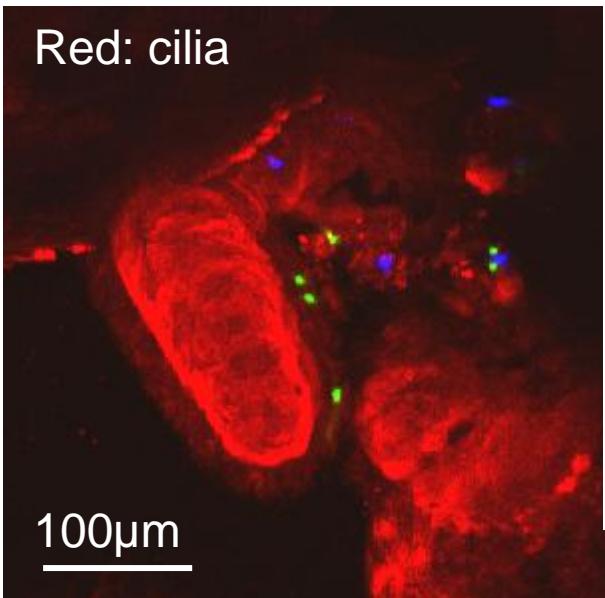


Capture rate similar to predicted encounter rate of passive cylinder in flow  
→ No obvious increase in capture rate compared to chance

# The mucociliary trap - not for everyone: Evidence for size-biased capture

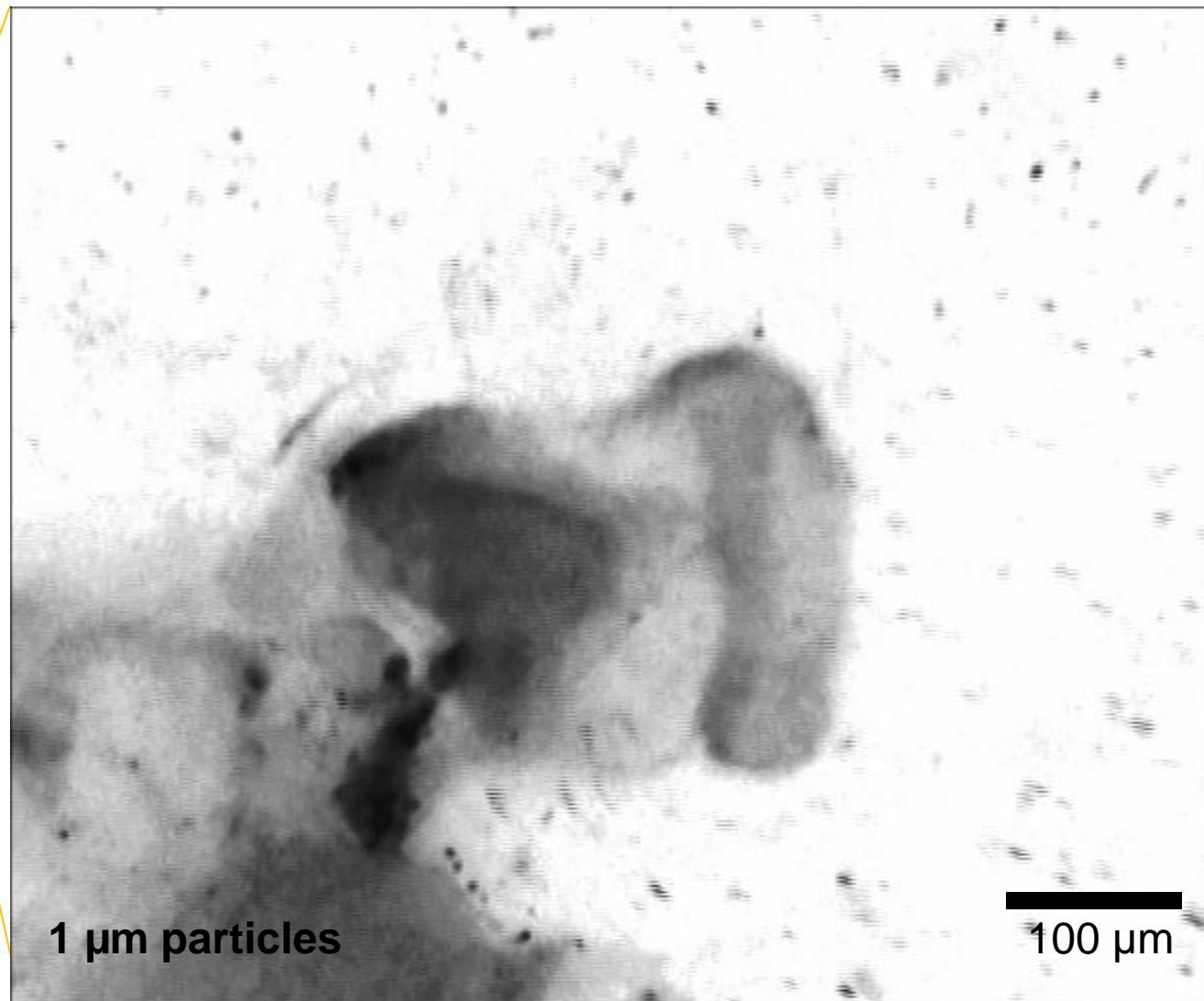
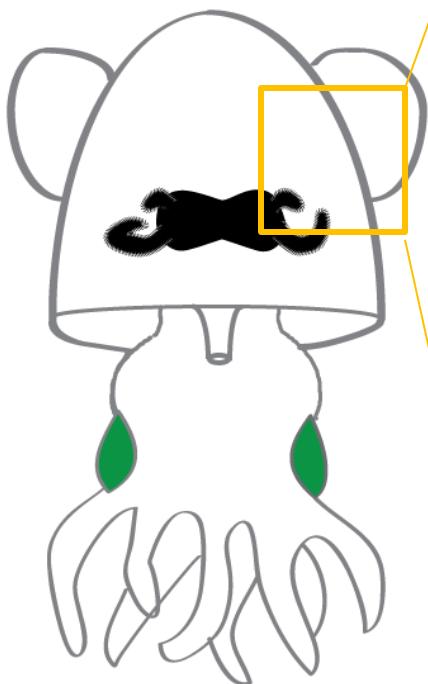


1 $\mu$ m (blue), 2 $\mu$ m (green),  
4 $\mu$ m (orange);  
 $10^7$  particles/ml each



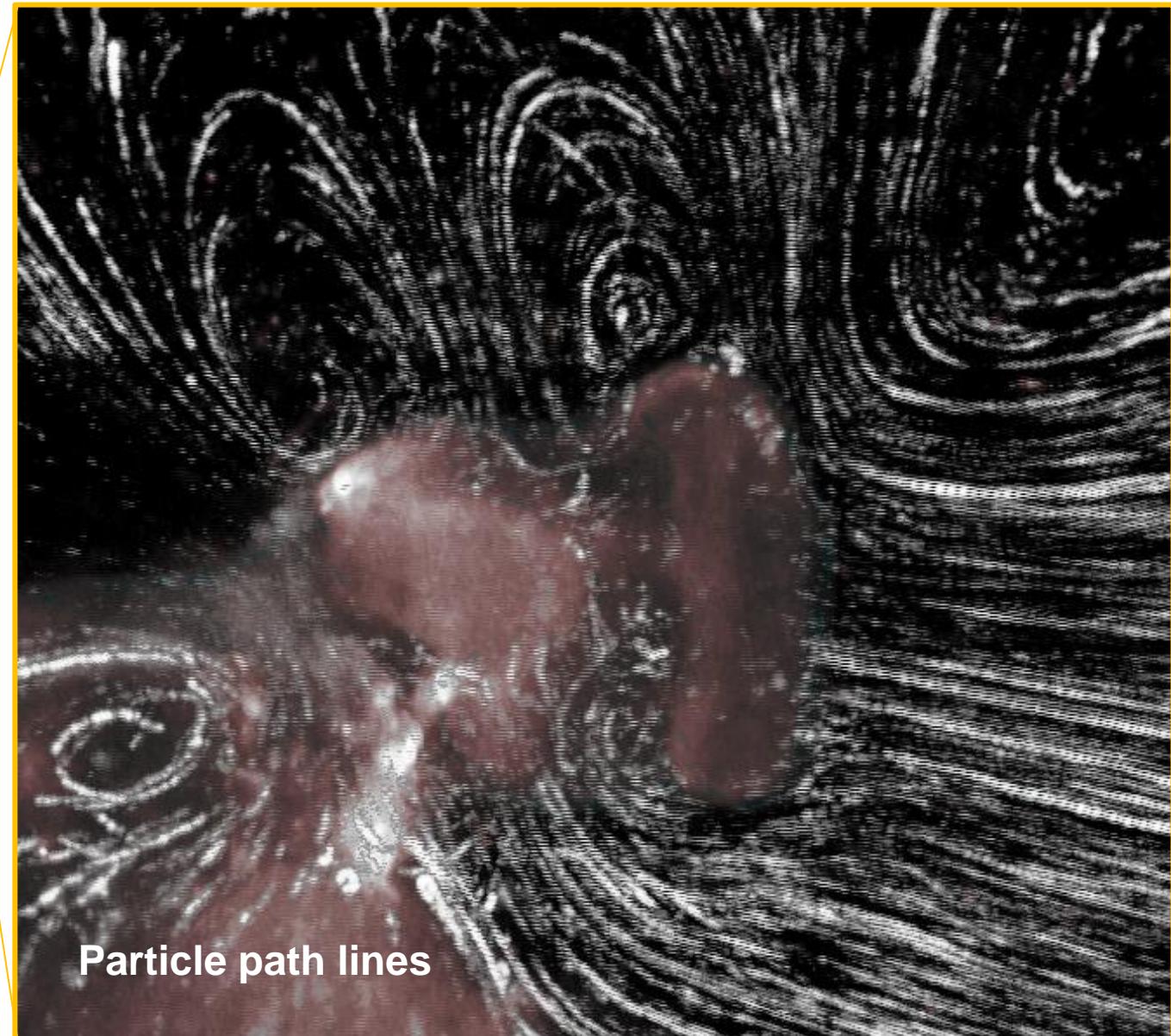
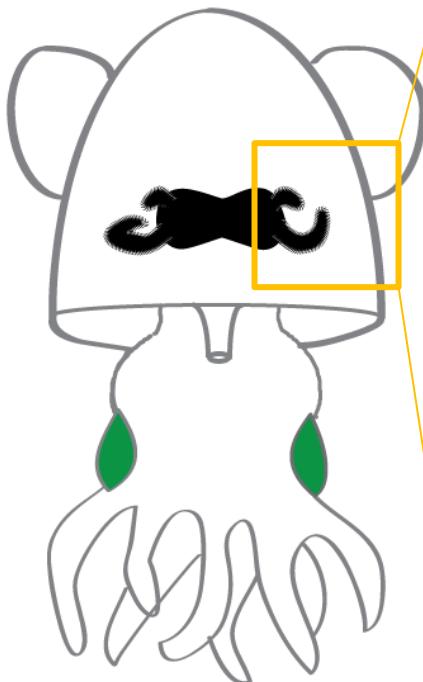


# Cilia-generated flow field





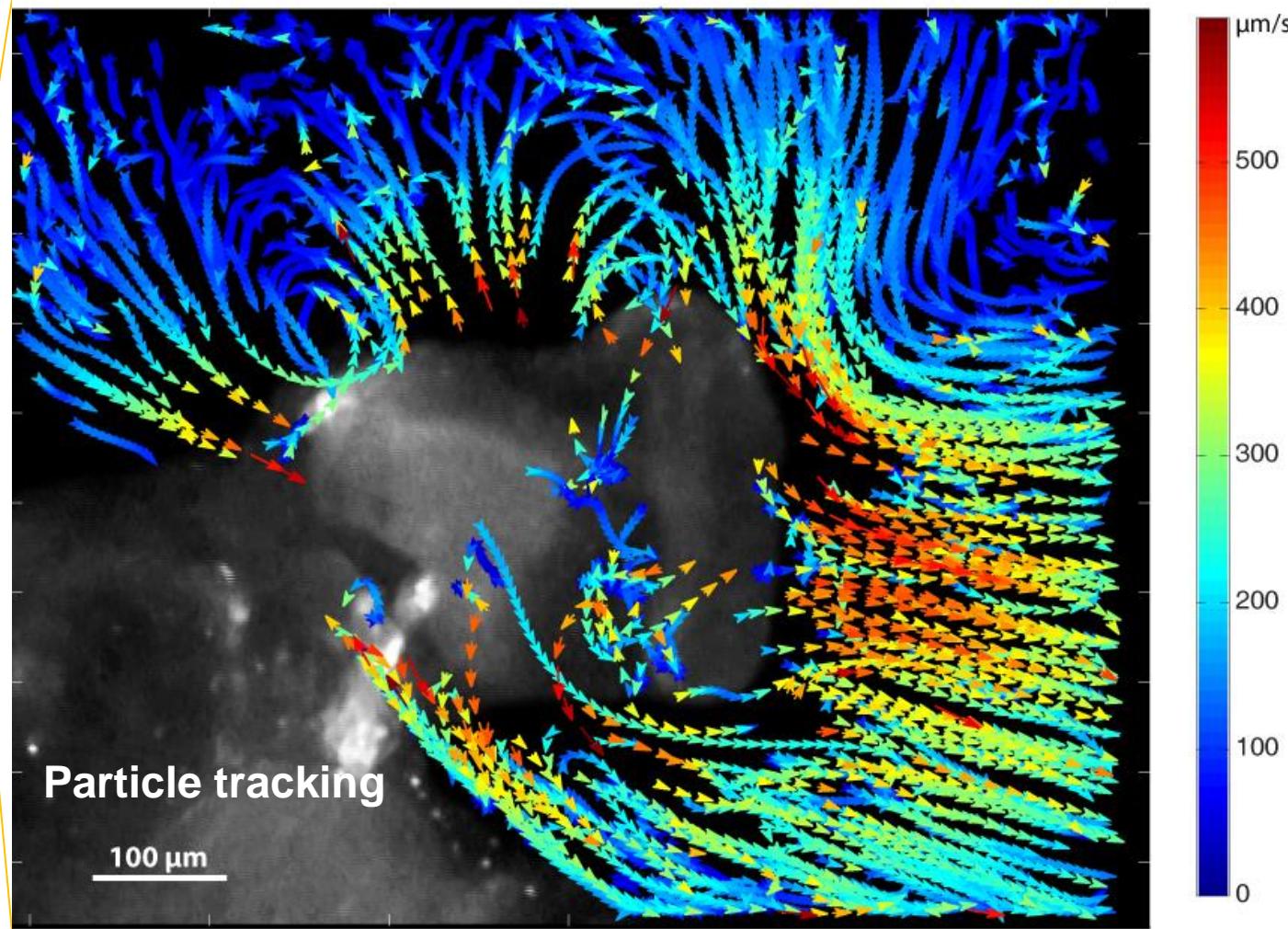
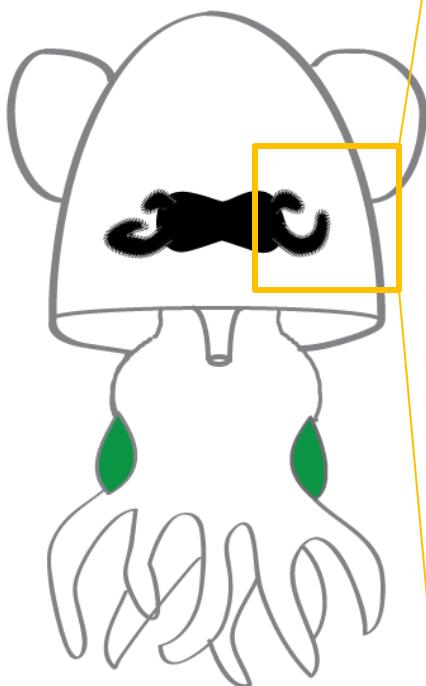
# Cilia-generated flow field



Particle path lines



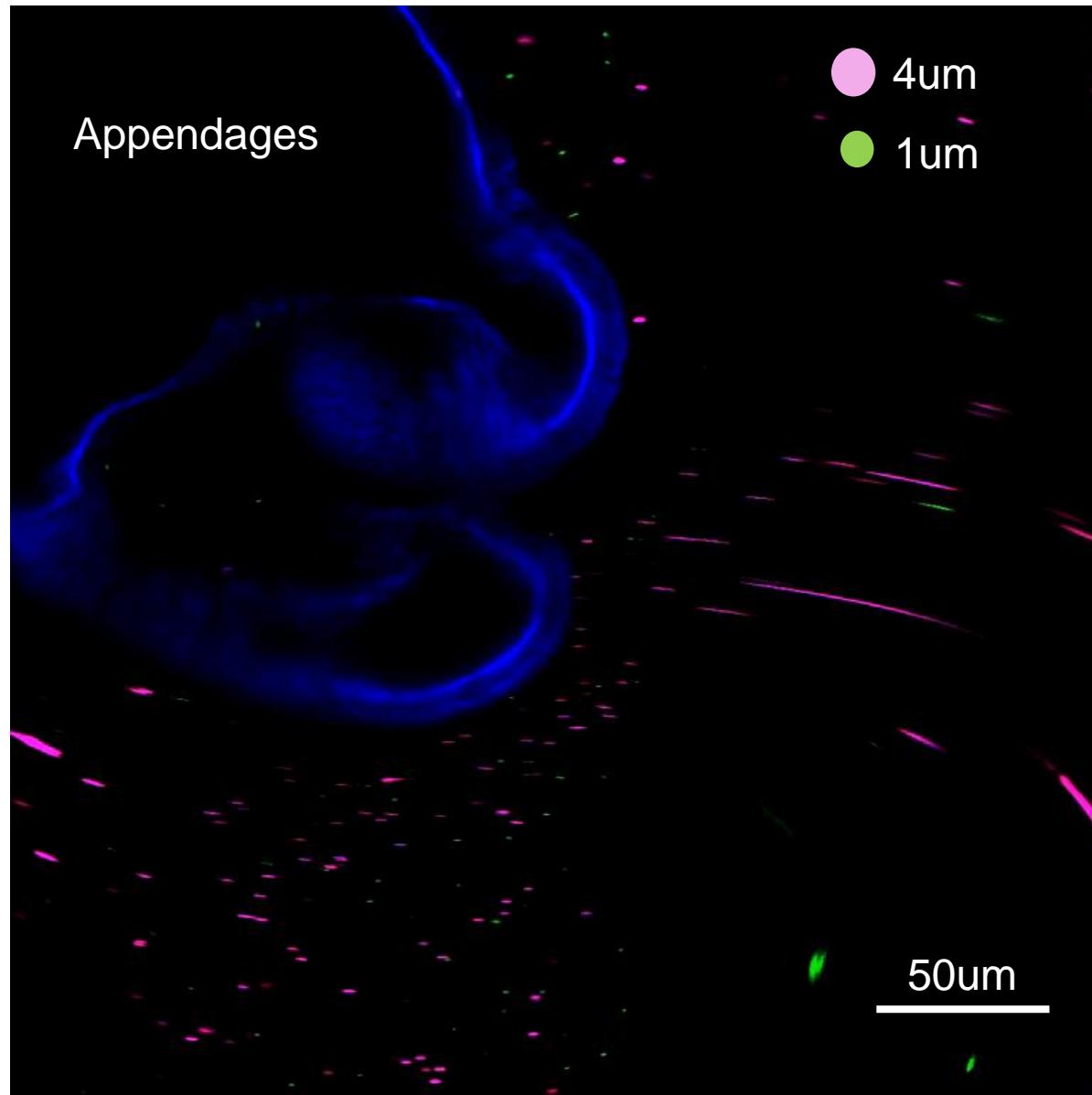
# Cilia-generated hydrodynamic sieve



Ratio advected particles:captured particles  $\approx 50:1$

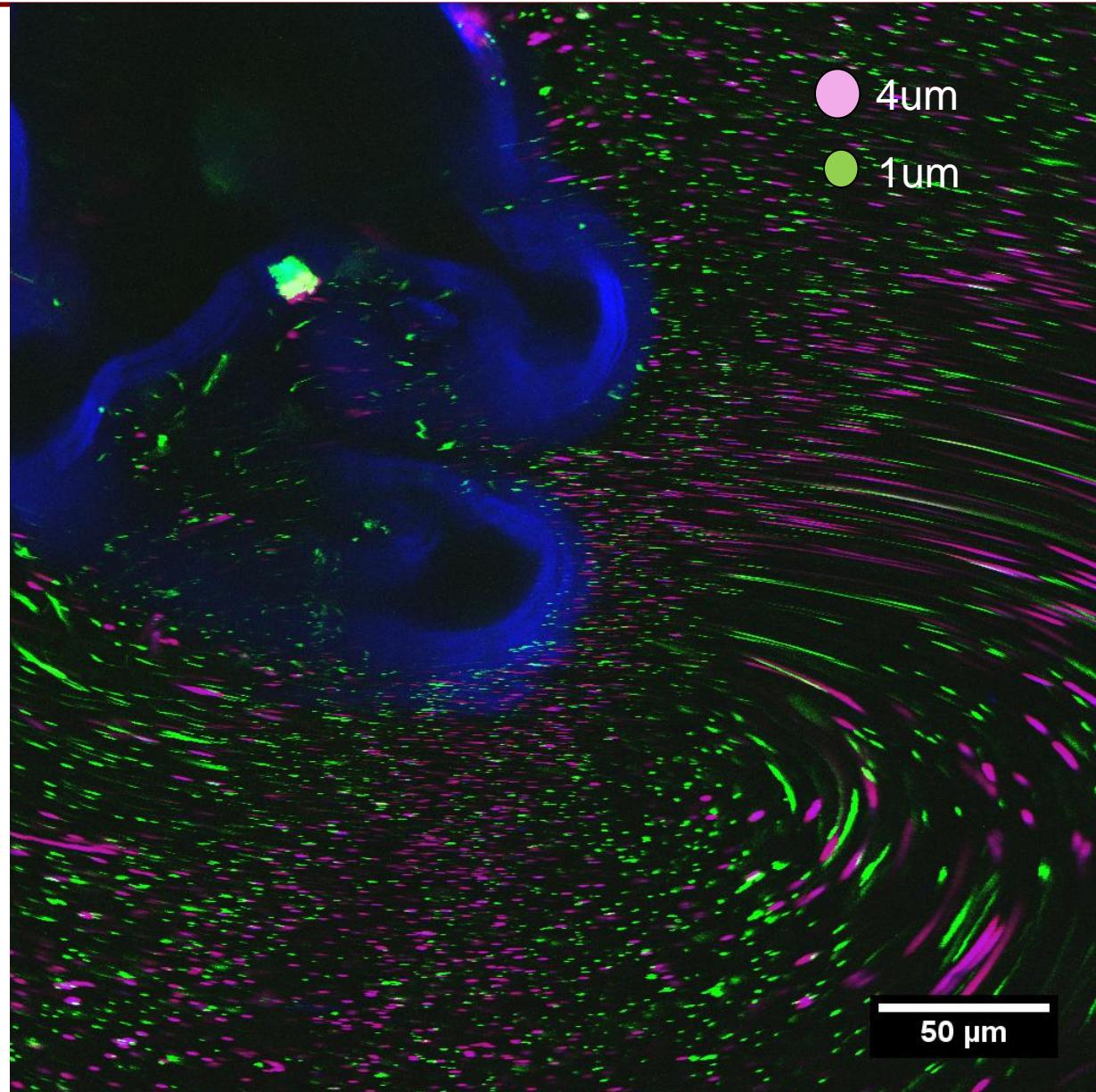


# *In vivo* capture





# *In vivo* capture

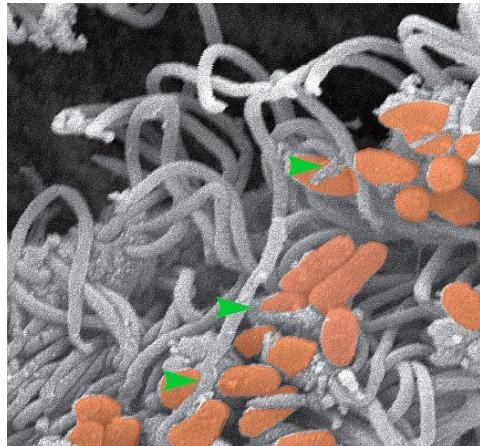




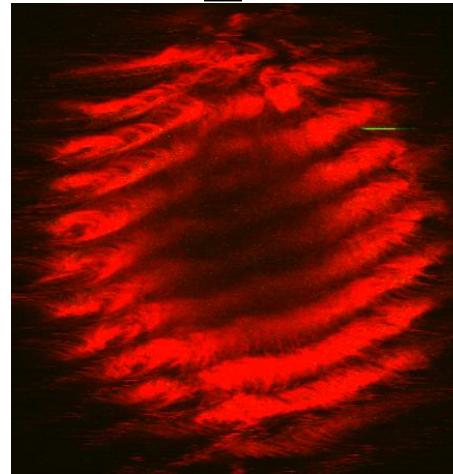
# Current work: Identifying structure-function relationships...

## Cilia fitness

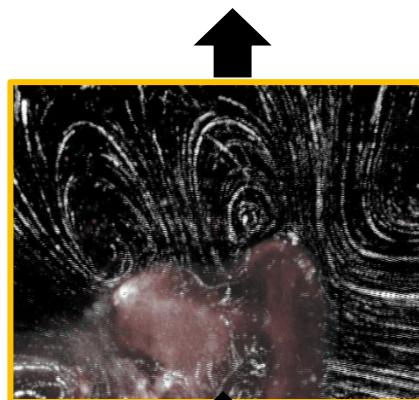
Capture and aggregation of bacteria (-sized particles)



+

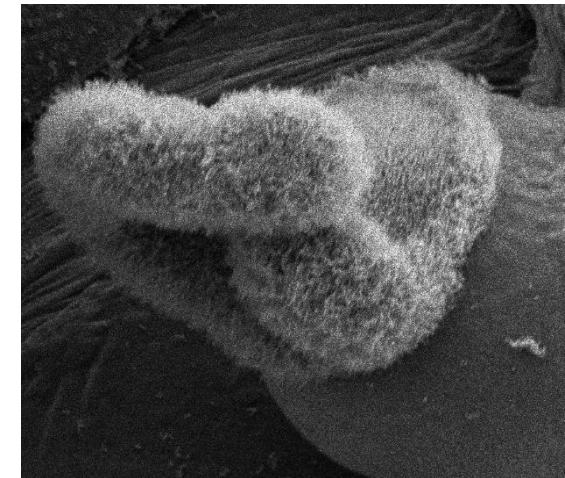


Metachronal wave



Fluid transport  
and mixing

+



Ciliary structure and  
kinematics

Surface geometry



# Current work: Identifying structure-function relationships...

...using a variety of methods and approaches

