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Caracterización morfológica, fisiológica y molecular de entradas de algodón (*Gossypium hirsutum* L.) e identificación de QTL de importancia agronómica

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# **Lista de Abreviaturas**

<b>A1RR</b>	Altura de inserción de la primera rama reproductiva
<b>AP</b>	Altura de planta
<b>D1P</b>	Distancia de la primera posición reproductiva al tallo principal
<b>DCO</b>	Días desde emergencia a cut out
<b>DM</b>	Días desde emergencia a madurez
<b>DPCA</b>	Días desde emergencia a apertura del primer capullo
<b>DPF</b>	Días desde emergencia a primera flor
<b>DPP</b>	Días desde emergencia a primer botón floral
<b>IF</b>	Índice de fibra
<b>IS</b>	Índice de semilla
<b>IU</b>	Uniformidad de fibra
<b>Mic</b>	Micronaire
<b>N1RR</b>	Nudo de inserción de la primera rama reproductiva
<b>NC</b>	Número de capullos por planta
<b>NN</b>	Número de nudos del tallo principal
<b>NRR</b>	Número de ramas reproductivas (simpodiales)
<b>NRV</b>	Número de ramas vegetativas (monopodiales)
<b>NSC</b>	Número de semillas por capullo
<b>PC</b>	Peso de capullo
<b>PN</b>	Fotosíntesis neta
<b>RB</b>	Rendimiento de algodón en bruto
<b>RF</b>	Rendimiento de fibra
<b>RFD</b>	Porcentaje de fibra al desmote
<b>Str</b>	Resistencia de fibra
<b>UHML</b>	Longitud de fibra
<b>gs</b>	Conductancia estomática

# **Resumen**

Primer párrafo del resumen en español.

Segundo párrafo del resumen aquí.

# **Abstract**

Primer párrafo del resumen en inglés.

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# Introducción

El algodón (*Gossypium hirsutum* L.) se cultiva en más de 80 países y desempeña un papel crucial en la producción textil como fuente de fibra. Argentina es el segundo productor de algodón de América Latina después de Brasil y desempeña un papel clave en los sistemas productivos de la región noreste del país. La producción de fibra cubre la demanda interna a la vez que facilita la exportación a países como Vietnam, Pakistán, Turquía, China, Indonesia, Colombia e India (ICAC, 2023; Paytas & Ploschuk, 2013). Se han desarrollado nuevas variedades y sistemas de producción para adaptarse a diversas condiciones medioambientales. En Argentina, el Instituto Nacional de Tecnología Agropecuaria (INTA) cuenta con un programa de mejora genética del algodón que utiliza el germoplasma disponible para desarrollar variedades que mejoren el rendimiento y la calidad de la fibra. El programa ha logrado aumentar tanto el rendimiento como la calidad de la fibra y también ha incorporado la resistencia genética a enfermedades importantes como mancha angular, enfermedad azul y marchitez por fusarium, en numerosas variedades (Royo et al., 2007; Scarpin et al., 2022; Scarpin et al., 2023).

Recientemente, Scarpin et al. (2022) informaron de un progreso genético en el rendimiento de fibra, el porcentaje de desmote, el rendimiento de algodón bruto y el número de cápsulas (NC), mostrando una tasa media de crecimiento anual de  $3,24 \text{ kg ha}^{-1}$ , 0,05 %,  $4,86 \text{ kg ha}^{-1}$  y 0,12 NC, respectivamente. Como resultado, las variedades más modernas tienen un mayor rendimiento de fibra, porcentaje de desmote, rendimiento de algodón bruto y número de cápsulas que las variedades más antiguas. Estos resultados indican

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que los programas de mejoramiento genético del algodón en Argentina han logrado un progreso genético sustancial para el rendimiento de fibra y sus componentes. Además, Scarpin et al. (2023) enfatizaron que este progreso no resultó en una disminución de la calidad del algodón. Sin embargo, se necesita más investigación para mejorar el rendimiento de fibra, la calidad de la fibra y la adaptabilidad a diferentes ambientes. En la provincia de Santa Fe (Argentina), el cultivo de algodón se concentra en los departamentos de 9 de Julio, Vera y General Obligado, cada uno de los cuales presenta disparidades distintivas en cuanto a la composición del suelo y las condiciones ambientales. El tipo de clima en el norte de la provincia es subtropical según la clasificación climática de Köppen, con una estación seca en la región noroeste y sin estación seca en la región noreste de la provincia (ANIDA, 2024). Debido a estas diferencias, es esencial desarrollar genotipos para estas diferentes condiciones para avanzar en la mejora de los cultivos. Además, se necesitan esfuerzos adicionales para identificar genes o QTLs asociados con rasgos agro-nómicos y de calidad de fibra en variedades de algodón en Argentina a través de métodos de mapeo genético.

En un programa de mejora genética del algodón, combinar un alto rendimiento y calidad de la fibra es un reto clave. El primer paso crucial para un programa de mejora eficaz es caracterizar el germoplasma del algodón. Kearsey y Pooni (1996) destacaron la importancia de comprender las diferencias genéticas y la heredabilidad de los rasgos al momento de cruzar plantas. La heredabilidad muestra en qué medida los rasgos están influidos por la genética y puede estimarse comparando las varianzas de las generaciones segregantes y no segregantes. En el algodón, se han notificado estimaciones de heredabilidad para varios rasgos, como el rendimiento de fibra, los componentes del rendimiento, la calidad de la fibra, la altura de la planta, el aceite de semilla, el nudo de la primera rama reproductiva, la cápsulas a prueba de tormentas, entre otros (De Carvalho et al., 2022; Meredith, 1984; Nidagundi et al., 2023; Ribeiro et al., 2017; Tang et al., 1996). Estos estimadores ayudan a los mejoradores a decidir la

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selección individual y a predecir cuánto pueden mejorar ciertos rasgos. Sin embargo, merece la pena tener en cuenta que el nivel de heredabilidad cambia en función del rasgo, la población y el ambiente en el que se cultivan.

# **Capítulo 1**

## **Caracterización de entradas de algodón (*Gossypium hirsutum* L.) del banco de germoplasma de INTA mediante caracteres morfo-fisiológicos**

### **1.1. Introducción**

Upland cotton (*Gossypium hirsutum* L.) is one of the most important natural fiber crops worldwide, playing a crucial role in the global textile industry. Argentina ranks as the second most important cotton fiber crop in Latin America after Brazil, with their production concentrated mainly in the northeastern provinces of Santa Fe, Chaco, and Santiago del Estero. Cotton cultivation not only satisfies domestic demand but also contributes to export markets, supplying countries such as Pakistan, Vietnam, Turkey, China, Indonesia, Bangladesh and others (Paytas et al., 2023; ICAC, 2025). During the 2024/25 growing season, Argentina reached approximately 683,000 hectares of planted cotton, with Santa Fe showing a 26 % increase compared to the previous season and accounting for 34 % of the national planted area, highlighting its growing importance in the national production system (ICAC, 2025; Zorzon, 2025).

With the creation of the Instituto Nacional de Tecnología Agropecuaria (INTA), and the implementation of its cotton breeding program in 1956, efforts were directed toward improving yield potential, fiber quality, and genetic resistance to major diseases such as bacterial blight (*Xanthomonas citri* subsp. *malvacearum*), blue disease (Cotton leafroll dwarf virus), and fusarium wilt (*Fusarium oxysporum* f. sp. *vasinfectum*) (Royo et al., 2007; Scarpin et al., 2022; Scarpin et al., 2023). Significant genetic progress has been achieved in traits such as lint yield, lint percentage, and the number of bolls per plant, without compromising fiber quality (Scarpin et al., 2022; Scarpin et al., 2023). However, sustained progress remains essential to cope with diverse environmental conditions and to satisfy the increasing demand for cultivars adapted to mechanized production systems.

In northern Santa Fe, where most production relies on the stripper-harvesting system, there is an increasing demand for cultivars with compact architecture and reduced vegetative branching to improve mechanized harvest efficiency and minimize fiber contamination. The introduction of the “stripper-type” harvester Javiyú, developed by INTA Reconquista, revolutionized cotton harvesting by replacing manual labor with efficient mechanized techniques (Pilatti et al., 2021). In parallel, technological innovations such as narrow-row and high-density systems have enhanced cotton production. Among producers, the most widely adopted configuration consists of 52-cm row spacing and plant densities of about 200,000 plants per hectare (Paytas et al., 2011). These technological changes underscore the need for cultivars specifically adapted to stripper-based harvesting systems and their associated management practices.

A fundamental step in breeding programs designed to meet these demands is the characterization of available germplasm to identify parents with complementary traits. INTA’s cotton germplasm collection, managed by the National Genetic Resources Network (REDGEN), currently conserves more than 700 accessions, mainly of *G. hirsutum*,

with smaller representations of *G. barbadense* and *G. barbadense* var. *brasiliensis* (Arriel et al., 2023). This genetic diversity constitutes a valuable resource for the development of cultivars adapted to local agroecological and mechanization-oriented systems, and its relevance is further reinforced by regional initiatives such as the +Cotton project coordinated by FAO and the Brazilian Cooperation Agency (ABC), which underscore the strategic importance of conserving and characterizing Latin American germplasm to support breeding efforts focused on climate resilience and fiber quality improvement (FAO and ABC, 2025).

Cotton yield is a complex trait determined by the interaction of multiple components, including boll number, boll weight, lint percentage, and seed index, all of which are strongly influenced by canopy structure, source–sink relationships, and environmental conditions during the growing cycle (Pettigrew, 2004; Constable and Bange, 2015). Physiological traits such as photosynthetic efficiency, biomass accumulation, and assimilate partitioning to reproductive organs have been shown to play key roles in determining lint yield and fiber quality (Constable and Bange, 2015). In this context, parameters such as photosynthetic rate, stomatal conductance, leaf chlorophyll status (commonly estimated through SPAD readings), and vegetative and reproductive biomass provide valuable information to interpret genotypic differences in growth dynamics and yield determination. Therefore, integrating morpho-physiological characterization with traditional agronomic and fiber quality evaluation provides a more comprehensive understanding of genotypic performance. This integrative approach is particularly relevant for identifying parental lines suited to the specific production systems of northern Argentina, where yield stability and mechanical harvest efficiency depend on optimized plant architecture and reproductive balance.

However, despite the relevance of cotton production in Argentina and the genetic progress achieved through breeding, there is still limited availability of cultivars specific-

cally adapted to stripper harvesting and narrow-row configurations. Moreover, although several studies have characterized cotton germplasm for general agronomic and fiber traits (Royo, 1998; Royo et al., 2003; Klein et al., 2011), few of them have focused on morphological and physiological attributes directly related to adaptation under these management conditions (Diaz et al., 2011). This dual limitation, restricted availability of adapted materials and scarce characterization targeted to mechanized systems, hinders the identification and use of parental lines that combine favorable yield, fiber quality, and plant architecture for efficient mechanized production in northern Argentina.

Multivariate statistical approaches, including principal component analysis (PCA) and hierarchical clustering using Gower's distance, have been widely used to explore complex phenotypic datasets and reveal underlying patterns of variation (Spoljaric et al., 2018; Sarwar et al., 2021; Klein et al., 2022; Ullah et al., 2022; Mangi et al., 2024; Anwar et al., 2025). These methods enable the joint assessment of quantitative and qualitative traits, facilitating the identification of relationships among accessions and the delineation of groups sharing similar morphological, agronomic, or fiber quality characteristics. Including these analyses provides a robust framework for identifying genetically divergent parental lines that combine desirable yield, fiber, and architectural traits, which are essential for modern mechanized production systems.

Therefore, this study aimed to conduct a comprehensive evaluation of a diverse set of cotton accessions, with specific aims to (i) characterize morphological, yield, and fiber quality traits; (ii) analyze physiological parameters and assimilate partitioning in selected contrasting genotypes; and (iii) identify promising parental lines for breeding populations targeting improved adaptability to contrasting environments and mechanized production systems.

## 1.2. Objetivos

Caracterizar entradas de algodón del banco de germoplasma de INTA con diferente procedencia mediante caracteres morfológicos relacionados al rendimiento.

Evaluar procesos fisiológicos que intervienen en la determinación del rendimiento de fibra de entradas de algodón del banco de germoplasma de INTA.

## 1.3. Materiales y métodos

### 1.4. Experimental site and plant material

The study was carried out at the Instituto Nacional de Tecnología Agropecuaria (INTA), Reconquista Experimental Station, located in Santa Fe province, Argentina (29°15' S, 59°44' W, 50 m a.s.l.). Experiments were carried out during the 2018–2020 growing seasons under greenhouse conditions.

A total of 26 cotton accessions from the INTA cotton germplasm bank (Sáenz Peña, Chaco, Argentina) were evaluated (Table 1). These accessions were selected based on their diverse geographical origins and contrasting performance for yield and fiber quality traits according to information provided by the germplasm bank. The set included materials from Argentina, Australia, Cameroon, Chad, China, Ivory Coast, Pakistan, Senegal, and the United States.

The plants were grown in 5 L pots filled with 2200 g of undisturbed soil (pH 6.7, available P: 165.7 mg kg<sup>-1</sup>, Na<sup>+</sup>: 0.6 cmol<sup>+</sup> kg<sup>-1</sup>, K<sup>+</sup>: 0.8 cmol<sup>+</sup> kg<sup>-1</sup>, Ca<sup>2+</sup>: 27.0 cmol<sup>+</sup> kg<sup>-1</sup>, Mg<sup>2+</sup>: 2.2 cmol<sup>+</sup> kg<sup>-1</sup>, NH<sub>4</sub><sup>+</sup>: 76.3 mg kg<sup>-1</sup>, NO<sub>3</sub><sup>-</sup>: 86.1 mg kg<sup>-1</sup>) and 400 g of commercial substrate (GrowMix Multipro). Pots were irrigated regularly to maintain moisture close to field capacity.

Experiments were arranged in a randomized complete block design (RCBD), with each

experimental unit consisting of one plant per pot. The design was selected to minimize potential spatial variability within the greenhouse environment, particularly light and temperature gradients, and it was consistently applied all measurements described below. To ensure comparability among accessions under controlled conditions, three independent replications were established, each represented by one plant per accession.

Tabla 1.1: Entradas de *Gossypium hirsutum* L. usadas en el presente estudio y su procedencia

Entradas	Procedencia	Entradas	Procedencia
BGSP-00166*	Argentina	BGSP-00748	China
BGSP-00177	Argentina	BGSP-00752	China
BGSP-00192	Argentina	BGSP-00755	China
BGSP-00193	Argentina	BGSP-00759	China
BGSP-00194	Argentina	BGSP-00779	China
BGSP-00207*	Argentina	BGSP-00067	Costa de Marfil
BGSP-00269*	Argentina	BGSP-00028	EEUU
SP 41255*	Argentina-Línea avanzada	BGSP-00145	EEUU
SP 6565*	Argentina-Línea avanzada	BGSP-00159*	EEUU
BGSP-00514	Australia	BGSP-00425*	EEUU
BGSP-00072	Camerún	BGSP-00428	EEUU
BGSP-00088	Camerún	BGSP-00715	Pakistán
BGSP-00070	Chad	BGSP-00126*	Senegal

Referencias: \* Entradas que fueron seleccionadas para un mayor análisis

## 1.5. Phenotypic diversity of cotton germplasm: morphological, yield and fiber quality traits

In this study, an initial characterization of 26 cotton accessions (Table 1) was conducted. Morphological, phenological, yield, and fiber quality traits were evaluated as summarized in Table 2.

At the cut-out reproductive stage (identified when the number of nodes above the last white flower on the main stem was fewer than four (Bourland et al., 1992)), qualita-

tive morphological traits, such as plant shape, canopy type, flowering type (observed on the lowest fruiting branch; classification based on boll proximity), leaf shape, trichome density, bract type (for the boll at the first fruiting position), and boll shape, were recorded using standardized upland cotton descriptors (UPOV, 2018). At maturity (when all capsules were open), quantitative morphological traits were measured, including plant height (HP), number of nodes on the main stem (STEMNN), number of monopodial (vegetative) branches (MONON), number of sympodial (reproductive) branch nodes (SY-MNN), distance from the first reproductive position to the main stem (DFRP) and first fruiting branch position, recorded both as stem node number (FFBN) and height (FFBH).

Each plant was harvested individually to determine seed cotton yield (SCY), lint yield (LY), lint percentage (LP), boll weight (BW), and boll number per plant (BN). Fiber quality traits, including fiber length (FL), strength (FS), micronaire (MIC), and fiber uniformity index (FU), were measured using a High Volume Instrument (HVI Uster 1000, APPA – Reconquista, Santa Fe) with pooled samples from three replicates.

Phenological traits were determined based on the number of days from seedling emergence to key developmental stages, including first square (DFS), first flower (DFF), cut-out (DCO), first boll opening (DBO), and maturity (DM).

Tabla 1.2: Morphological, phenological, yield, and fiber quality traits measured in 26 cotton accessions

Trait.Category	Trait	Abbreviation	Unit	Description
Fiber Quality	Fiber length	FL	mm	Upper half mean length (UHML), measured with HVI
	Fiber strength	FS	g tex <sup>-1</sup>	Measured with HVI
	Micronaire	MIC	-	Measured with HVI
	Fiber uniformity	FU	%	Uniformity index, measured with HVI
Morphological (Qualitative)	Plant shape, canopy, flowering type, leaf shape, trichome density, bract type, boll shape	-	-	Scored according to Upland cotton descriptors

**Tabla 1.2: Morphological, phenological, yield, and fiber quality traits measured in 26 cotton accessions (continued)**

Trait Category	Trait	Abbreviation	Unit	Description
Morphological (Quantitative)	Plant height	HP	cm	Height of the plant at maturity in cm
	Main stem nodes	STEMNN	count	Number of nodes on main stem
	Monopodial branches	MONON	count	Number of monopodial (vegetative) branches
	Sympodial branch nodes	SYMNN	count	Number of sympodial branch nodes
	First fruiting branch position (node)	FFBN	count	First fruiting branch position measured by number of stem node
	First fruiting branch position (height)	FFBH	cm	First fruiting branch position measured by height in cm
	Distance to first reproductive position	DFRP	cm	Distance from the first reproductive position to the main stem in cm
Phenological	Days to first square	DFS	days	From seedling emergence to first square
	Days to first flower	DFF	days	From seedling emergence to first flower
	Days to cut-out	DCO	days	From seedling emergence to Cut Out (Bourland et al., 1992)
	Days to boll opening	DBO	days	From seedling emergence to first boll opening
	Days to maturity	DM	days	From seedling emergence to physiological maturity
Yield Components	Seed cotton yield	SCY	g plant <sup>-1</sup>	Total seed cotton weight per plant
	Lint yield	LY	g plant <sup>-1</sup>	Total lint weight per plant
	Lint percentage	LP	%	(Lint weight / Seed cotton weight) × 100
	Boll weight	BW	g	Average weight of harvested bolls
	Boll number per plant	BN	bolls plant <sup>-1</sup>	Total number of bolls per plant

## 1.6. Yield, fiber quality, physiological, and biomass partitioning analysis of contrasting cotton accessions

Eight accessions were selected (marked with an asterisk in Table 1) based on their contrasting performance in yield components, especially LP, and fiber quality. Some accessions with compact plant architecture and low MONON were also included. These accessions were subjected to a more detailed evaluation of yield and fiber quality variables and physiological processes influencing yield components. Two independent experiments were conducted at different time points, each including nine replicates per accession, to increase the representativeness of fiber quality parameters and enhance statistical precision. The yield and fiber quality variables described in the section “Phenotypic diversity of cotton germplasm: morphological, yield and fiber quality traits” were also recorded, along with the additional variables listed in Table 3: seed number per boll (SNPB), lint index (LI), and seed index (SI).

These same eight accessions were used to evaluate physiological traits and biomass partitioning as summarized in Table 3. Destructive sampling was performed at four phenological stages (moments): first square (M1), first flower (M2), first open boll (M3), and maturity (M4). Plants were partitioned into vegetative components (stem, vegetative branches and leaves) and reproductive structures (sympodial branches, squares, flowers, green and open bolls, seed, and fiber). Samples were oven-dried at 65 °C until constant weight to determine total and partitioned biomass. The experiment followed a RCBD with split-plot arrangement, with phenological stages as the main plots and accessions as the subplots.

Flowering dynamics and reproductive structure retention were assessed by mapping all reproductive positions following previous studies (Kerby et al., 2010).

Physiological traits, including net photosynthetic rate ( $P_N$ ), stomatal conductance

( $g_s$ ), and chlorophyll content (SPAD), were measured at three moments: at first flower (M2), and 15 and 30 days thereafter, using a LI-COR 6400 photosynthesis system ( $P_N$  and  $g_s$ ) and a Minolta SPAD 502 chlorophyll meter (SPAD). Four replications were included for each accession. Four fully expanded leaves located at the upper canopy (apical leaves) were sampled and labeled at the beginning of the experiment. These same four leaves were measured at each evaluation time. Measurements were made under light intensity ranging from 900 to 1500  $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$  between 11:00 and 13:00 h.

Tabla 1.3: Physiological and biomass partitioning traits measured in eight contrasting cotton accessions

Trait.Category	Trait	Abbreviation	Unit	Method
Additional Yield Traits	Seed number per boll	NSPB	count	Number of seeds per boll
	Lint index	LI	g	Weight of lint from 100 seeds
	Seed index	SI	g	Weight of 100 seeds
Biomass Partitioning	Total biomass	TB	$\text{g plant}^{-1}$	Sum of vegetative and reproductive biomass
	Vegetative biomass	VB	$\text{g plant}^{-1}$	Sum of stem, vegetative branches and leaves
	Reproductive biomass	RB	$\text{g plant}^{-1}$	Sum of sympodial branches, squares, flowers, green and open bolls, seed, and fiber
Physiological	Net photosynthesis rate	PN	$\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$	LICOR 6400 gas exchange system
	Stomatal conductance	gs	$\text{mol H}_2\text{O m}^{-2} \text{ s}^{-1}$	LICOR 6400 gas exchange system
	Chlorophyll content	SPAD	SPAD units	Minolta SPAD 502
Reproductive Dynamics	Global retention	GR	%	Mapping according to Kerby et al. (2010). Global retention as the ratio of the total number of reproductive structures retained by the total number of reproductive positions generated per plant
	First position retention	R1P	%	Mapping according to Kerby et al. (2010). Retention as the ratio of the total number of reproductive structures retained in the first positions to the total number of reproductive positions generated in those same positions per plant

## 1.7. Statistical analysis

All statistical analyses were performed using R software (R Core Team, 2023). The significance level was set at 95 % ( $p \leq 0.05$ ) for all tests.

For the characterization of the 26 accessions, the normality of quantitative traits was verified using the Shapiro–Wilk test. Analysis of variance (ANOVA) was then applied to determine differences among accessions, followed by Fisher's least significant difference (LSD) test for mean comparison (Luo, 2024).

For qualitative morphological traits, the relative frequencies of each descriptor were analyzed. Additionally, hierarchical cluster analysis was performed using Gower's distance metric and the Ward.D method, integrating qualitative and quantitative traits. Homogeneous groups of accessions were visualized in dendograms and heat maps generated with the cluster, dplyr, and ComplexHeatmap packages (Gu, 2022; Wickham et al., 2023; Maechler et al., 2025).

For the eight contrasting and selected accessions, data on yield, yield components, and fiber quality traits were analyzed using linear mixed models (LMM) with the nlme package (Pinheiro et al., 2025), applying the restricted maximum likelihood (REML) method. Accessions were considered as fixed effects, while experiment and replication were treated as random effects. When significant differences were detected, multiple comparisons were performed using Fisher's LSD test. Associations among traits were examined using Spearman's correlation coefficients (Conigrave, 2023).

For biomass partitioning and reproductive retention, LMM adapted to a split-plot design were applied, with moment, accession, and their interaction (Moment:Accession) as fixed effects. Random effects accounted for experiment, blocks, and moments (main plots). Specific variance structures (varIdent) were implemented when heteroscedasticity between measurement times was detected.

Physiological traits were analyzed as repeated measures over time. The models included moment, accession, and their interaction (Moment:Accession) as fixed effects, with block and individual (pot number) as random effects. This approach captured temporal variation and accession-specific dynamics across measurement dates.

Finally, to explore the overall diversity among accessions, a principal component analysis (PCA) was conducted.

## **1.8. Resultados**

### **1.8.1. Phenotypic diversity of cotton germplasm: morphological, yield and fiber quality traits**

#### **1.8.2. Qualitative morphological traits**

The frequency distribution of qualitative morphological descriptors among the evaluated accessions is shown in Fig. 1.1. Boll shapes were predominantly ovate (88.5 %), followed by circular (7.7 %) and broad-elliptic (3.8 %). All accessions had normal bracts. Canopy types were mainly medium (69.2 %) and open (23.1 %), with a few compact forms (7.7 %). Regarding flowering type, semi-clustered plants were the most frequent (69.2 %), followed by non-clustered (23.1 %) and clustered types (7.7 %). Lamina were mostly flat (96.1 %) and leaves as palmate (96.1 %), with a small proportion of cup-shaped (3.9 %) and okra-type (3.9 %) leaves. Pubescence was predominantly medium (76.9 %), with weak and strong levels each accounting for 11.5 %. Finally, plant shape was mainly cylindrical (61.5 %) and conical (38.5 %). Plant shape descriptors reflected qualitative differences observed under greenhouse conditions and should be interpreted as exploratory indicators of architectural diversity. The descriptors of the evaluated cotton germplasm accessions are summarized in Table S1 (Supplementary Material).

### **1.8.3. Quantitative morphological traits, yield and its components and fiber quality**

All mean values of evaluated morphological quantitative traits are shown in Table S2. Quantitative morphological traits varied significantly among accessions, with HP ranging from 89.0 to 126.3 cm and STEMNN from 13.0 to 17.0. The MONON ranged from 0.3 to 2.7, whereas SYMNN varied from 9.7 to 13.3. The FFBN ranged from 3.7 to 5.7, and FFBH varied from 25.0 to 40.7 cm. The DFRP ranged from 3.9 to 14.3 cm. Yield, fiber quality, and phenological traits also showed wide variability among accessions, with SCY ranging from 14.3 to 33.5 g plant<sup>-1</sup>, LY from 4.6 to 12.3 g plant<sup>-1</sup>, BW from 2.6 to 4.7 g, and BN from 4.7 to 8.7. Fiber quality traits varied for FL (25.1–33.9 mm) and FS (21.5–37.6 g tex<sup>-1</sup>). Phenological traits ranged from 19 to 27 DFS, 47 to 55 DFF, 68 to 76 DCO, 97 to 105 DBO, and 118 to 133 DM.

To combine all evaluated traits, a hierarchical cluster analysis was performed using Gower's distance. Fig. 1.2 shows the resulting heatmap together with the dendograms generated by the Ward.D method. Based on the observed patterns, two main clusters were distinguished. Cluster one included accessions with a conical plant shape and non-clustered flowering, some of which exhibited superior fiber quality. Cluster two grouped accessions with a more compact canopy and clustered flowering, generally associated with a higher lint percentage and greater boll number.

Meeting the simultaneous demands for fiber quality, yield and adaptation to intensive management practices requires expanding the genetic bases available to breeders. INTA's germplasm collection, with its documented diversity in morphological attributes (leaf morphology, bract morphology, boll shape, leaf pubescence, presence or absence of leaf nectaries, stormproof traits), yield components and fiber quality (Royo et al., 2003; Klein et al., 2011; Spoljaric et al., 2018; Scarpin et al., 2019; Arriel et al., 2023) offers a unique opportunity to identify parental for future breeding populations. Almost all

traits exhibited phenotypic diversity, with values varying significantly among accessions, highlighting the potential of this germplasm not only for fiber improvement but also for breeding programs aimed at enhancing productivity and developing varieties suited to specific production systems. The variability shown in this work is consistent with reports from U.S. germplasm collections, where extensive allelic diversity has been documented (Hinze et al., 2025), and from Asian collections, including studies in Pakistan that revealed significant genetic variation in yield and fiber quality traits (Farooq et al., 2017). In contrast, recent genomic analyses in China have highlighted the narrow genetic base and relatively low diversity of cultivated cotton, underscoring the importance of broadening germplasm resources for future breeding (Li et al., 2023). Importantly, initiatives in Latin America, such as the +Cotton project (FAO-ABC), have also emphasized the value of regional germplasm diversity for breeding programs, particularly in the context of climate resilience and fiber quality improvement (FAO and ABC, 2025).

## **1.9. Yield, fiber quality, physiological, and biomass partitioning analysis of contrasting cotton accessions**

### **1.9.1. Yield components and fiber quality**

The accessions selected for a more detailed evaluation of yield traits and fiber quality are indicated with an asterisk in Table 1. Selection was based primarily on accessions belonging to different clusters according to the hierarchical cluster analysis (Fig. 1), as well as on the data presented in Table S2. Priority was given to accessions with marked contrasts in yield and their components, particularly lint percentage, as well as fiber quality. In addition, some accessions with lower number of monopodial (vegetative) branch and compact canopy structure were included, such as accession BGSP-00166.

The selected cotton accessions showed significant differences in all traits related to

yield and fiber quality ( $p < 0.05$ , Table 4). Accession SP-41255 had the highest mean values for SCY, LY, LP, and BN, with values of  $34.2 \text{ g plant}^{-1}$ ,  $15.8 \text{ g plant}^{-1}$ ,  $45.8 \%$ , and  $9.2 \text{ bolls plant}^{-1}$ , respectively. However, this accession had the lowest mean value for SI, with  $7.7 \text{ g}$ . BGSP-00159 had the lowest mean SCY with  $26.2 \text{ g plant}^{-1}$ . BGSP-00166 showed the lowest mean values for LY, LP, MIC, and BN ( $8.6 \text{ g plant}^{-1}$ ,  $31.1 \%$ ,  $3.2$ , and  $5.8 \text{ bolls plant}^{-1}$ , respectively), yet it showed the highest mean values for BW, SI, FL, FS, and FU ( $4.6 \text{ g boll}^{-1}$ ,  $11.5 \text{ g}$ ,  $33.2 \text{ mm}$ ,  $37.3 \text{ g tex}^{-1}$ , and  $85.8 \%$ , respectively). BGSP-00269 had the lowest mean values for BW, SNPB, and FL ( $3.4 \text{ g boll}^{-1}$ ,  $22.8 \text{ seeds boll}^{-1}$ , and  $28.3 \text{ mm}$ , respectively), while having the highest mean values for LI and MIC ( $6.5 \text{ g}$  and  $4.3$ , respectively). BGSP-00425 had the highest mean SNPB with  $30.6 \text{ seeds boll}^{-1}$ , while having the lowest mean values for LI and FU, with  $4.0 \text{ g}$  and  $82.2 \%$ , respectively. BGSP-00207 had the lowest mean values for FL and FS, with  $28.3 \text{ mm}$  and  $29.2 \text{ g tex}^{-1}$ , respectively. Notably, accessions BGSP-00166 and SP-41255 were significantly different for nearly all studied traits except SNPB.

These results indicate that there is phenotypic variation among the contrasting cotton accessions evaluated in terms of both yield and fiber quality. These findings are consistent with previous reports documenting wide variability in cotton germplasm (McCarty et al., 2006) and reinforce the well-known trade-off between yield and fiber quality (Li et al., 2023). The greatest variation was observed for LY, BN, LI, SI, LP, and BW traits, while less variation occurred for MIC, SNPB, SCY, FS, and FL. In contrast, the least variation was observed in FU. McCarty et al. (2006) also found diversity with wide ranges for SCY, LY, BW, LP, MIC, FL, and FS.

Table S3 shows the Spearman correlation values for fiber yield traits, their components, and fiber quality. Several phenotypic correlations were statistically significant. Correlations greater than  $0.80$  were considered strong, those between  $0.40$  and  $0.80$  moderate, and those below  $0.40$  weak. LY showed a strong positive correlation with BN and

a moderate positive correlation with BW. LP showed moderate positive correlations with LY and BN but a moderate negative correlation with SI. FL showed moderate negative correlations with LY, indicating that as fiber yield increases, fiber length decreases. When we analyzed the correlations within the cotton germplasm collection, LY exhibited a positive correlation with LP, BW, and BN, while we observed the strongest association with the latter. Scarpin et al. (2022) also identified this association, demonstrating that the rise in LY is primarily influenced by an increase in BN, driven by enhanced retention, and secondarily by an increase in LP and BW. Meredith (1984) reported positive correlations between LP and LY, BN, and LI, while negative correlations were found with SI, BW, FL, FS, and MIC. We obtained similar results, except for MIC, which exhibited a positive correlation. Collectively, these findings suggest that breeding programs aiming to increase LY should focus on BN enhancement. Increased BN may be associated with higher LP, leading to higher LI and MIC, while decreasing SI, BW, FL, and FS. However, achieving a balance with LP is necessary to maintain fiber quality traits. This can be accomplished by increasing LP to a level that does not excessively reduce traits such as FL or FS.

## **1.10. Biomass partitioning, reproductive retention and physiological analysis**

The temporal dynamics of biomass accumulation and its partitioning among organs in eight contrasting cotton accessions are illustrated in Fig. 2. During early development (M1–M2), vegetative biomass (VB), mainly stems and leaves, represented the largest proportion of total dry matter. From flowering onward (M3–M4), reproductive structures became the dominant contributors to total biomass, evidencing a clear shift from vegetative to reproductive allocation (Fig. 2.1). Both moment and accession effects were highly significant ( $p < 0.001$ , Table S4), as well as their interaction, indicating differential growth dynamics among accessions.

At maturity (M4), clear differences among accessions were observed in the distribution of biomass components. BGSP-00207 exhibited the greatest accumulation in reproductive branches, whereas BGSP-00166 showed the lowest biomass accumulation in both vegetative and reproductive branches, consistent with its compact plant architecture compared to the more open structure of BGSP-00207. SP-41255 and BGSP-00126 accumulated the highest vegetative branch biomass (Fig. 2.2). Overall, the figure highlights how cotton accessions differ in their temporal patterns of biomass partitioning and organ-specific allocation throughout development. These differences observed in vegetative and reproductive branch biomass among accessions align with previous studies suggesting that cotton plant architecture, including branching patterns, is influenced by genetic factors (McGarry et al., 2016; Sun et al., 2022; Huang et al., 2022). Genotypic variation may therefore contribute to the contrasting behaviors observed in this study, such as the reduced vegetative and reproductive biomass in accession BGSP-00166. However, plant density also exerts a strong influence on biomass partitioning. Li et al. (2019b) demonstrated that higher plant densities increase intra-specific competition for light and nutrients, leading to reduced vegetative branching and a shift toward reproductive allocation, whereas lower densities allow greater lateral expansion and promote vegetative branch development. These findings indicate that both genetic background and management practices interact to shape the balance between vegetative and reproductive growth, highlighting the relevance of integrating genotypic and agronomic perspectives in cotton breeding programs.

Morpho-physiological characterization highlighted differences in plant architecture and biomass partitioning, traits potentially relevant for mechanized production. Accessions with compact canopy and reduced monopodial branching, such as BGSP-00166, showed allocation patterns consistent with improved harvest efficiency and suitability for high plant density, in line with studies emphasizing the role of architecture in the efficiency of mechanized plant harvest (Huang et al., 2022; Madhu et al., 2023).

The reproductive retention patterns observed in eight contrasting cotton accessions are shown in Fig. 3. The temporal dynamics of global (GR) and first-position (R1P) retention across phenological stages (M2–M4), revealing a consistent decline from first flower (M2) to maturity (M4) as can be seen in Figure 3.1. This reduction reflects the progressive accumulation of dry matter in reproductive organs and the resulting competition for assimilates. Fig. 3.2 compares accessions at maturity (M4), highlighting clear differences in both GR and R1P values. BGSP-00269, SP-6565, BGSP-00425, and SP-41255 maintained higher GR at maturity, while BGSP-00269 and SP-41255 also exhibited the greatest R1P, indicating greater efficiency in early boll set and fruit retention. In contrast, BGSP-00166 and BGSP-00159 displayed the lowest retention values in terms of GR and R1P.

The analysis of reproductive structure retention (Table S5) confirmed the visual trends discussed previously, showing significant effects of moment and accession for GR ( $p = 0.0017$ ) and a significant Moment:Accession interaction ( $p = 0.0119$ ). These results indicate that the genotypes evaluated differed in their capacity to retain fruiting structures across reproductive stages, contributing to the observed variation in yield components.

The spatial distribution of bolls within the plant canopy has been consistently linked to both yield and fiber quality. Bolls set in early fruiting positions or in the mid-section of the plant (nodes 9–14) generally exhibit greater fiber length and strength, while those located in upper internodes (nodes 16–22) tend to produce lower micronaire values (Davidonis et al., 2004; Nie et al., 2019; Zhang et al., 2021). This positional effect reflects differences in assimilate availability and growth conditions, which directly influence boll weight and fiber maturity. Consequently, accessions that combine high global retention with strong first-position retention, such as BGSP-00269 and SP-41255 (Fig. 3), not only secure a larger number of fruits but also retain those bolls most likely to contribute superior fiber properties. Findings from Mao et al. (2015) are consistent with this inter-

pretation, showing that improvements in fiber strength under high plant density and mepiquat chloride application were linked to a greater retention of bolls in lower and middle fruiting positions.

The estimated temporal variation of key physiological traits,  $P_N$ ,  $g_s$ , and SPAD, measured at first flower (M2) and at 15 and 30 days thereafter in eight cotton accessions is shown in Fig. 4. All three variables showed a clear decline over time. At M2, accessions BGSP-00269 and BGSP-00425 exhibited the highest mean  $P_N$ ; however, their rates declined more sharply thereafter compared with the other genotypes, which could explain the lower biomass accumulation observed in BGSP-00425. Significant effects of moments were detected for  $P_N$  and SPAD ( $p < 0.001$ ), whereas accession and the moment  $\times$  accession interaction were significant only for  $P_N$  ( $p = 0.012$  and  $0.005$ , respectively, Table S6). This indicates that genotypes differed mainly in their photosynthetic capacity rather than in stomatal behavior or chlorophyll content.

Physiological measurements revealed distinct accession-specific dynamics in photosynthetic rate. Although  $P_N$  declined over time in all genotypes, as expected, the magnitude of this decline differed among accessions. Genotypes with steeper reductions, such as BGSP-00425, showed lower final biomass, suggesting limitations in carbon supply during boll filling. In contrast, accessions with more stable  $P_N$  profiles tended to exhibit more favorable reproductive outcomes. Notably,  $P_N$  rather than  $g_s$  or SPAD was the main driver of physiological differences, indicating that variation in carbon assimilation capacity, rather than stomatal or chlorophyll traits, played a central role in shaping yield performance. These findings align with previous studies showing that  $P_N$  is positively correlated with biomass accumulation and yield in cotton (Li et al., 2019a)

## 1.11. Principal Component Analysis

Principal component analysis of the significant variables revealed that the first principal component explained 55.89 % of the variance, while the second component explained 14.54 % (Fig. 5). Accessions located in the left quadrants, such as BGSP-00166, BGSP-00159, and BGSP-00425, were associated with higher fiber quality (FL, FS, FU) and larger seed size (SI), but lower values for yield-related traits such as SCY, LY, LI, BN, and LP. In contrast, accessions positioned on the right side of the biplot, including SP-41255, SP-6565, BGSP-00269, BGSP-00207, and BGSP-00126, were related to greater biomass production (stem, total biomass, monopodial and sympodial branches) and higher yield performance, but comparatively lower fiber quality; consistent with their higher biomass and more fruit retention described in the biomass partitioning results (Fig. 2; Fig. 3). Traits such as GR, R1P, and P<sub>N</sub> also contributed positively to the right side, reinforcing their role in yield determination. Both, PCA and correlation coefficients (Table S3) revealed the relationships among the traits within this set of genotypes. For instance, traits such as LY exhibited a strong positive correlation with BN, a moderate positive correlation with BW and LP, and a moderate negative correlation with quality traits such as FL.

Principal Component (PC) analysis routinary is used to identify similarities among the genotypes for traits in cotton (Shakeel et al., 2015; Farooq et al., 2017; Klein et al., 2022). In our study, the principal component analysis found the most contrasting accessions in terms of the measured traits; in particular, BGSP-00166 was associated with higher values of fiber quality (FL, FS, FU) and seed size (SI), whereas SP-41255 was associated with higher values in yield-related traits such as SCY, LY, LI, BN, LP. The cultivar SP-41255 stood out due to its performance, exhibiting the highest values of LP and BN, and the lowest values of BW and SI when Scarpin et al. (2022) performed a study focusing on genetic progress for yield and its components across 20 cotton cultivars. In agreement with our observations, SP-41255 was associated with superior yield-related traits, specifically

higher values of LP and BN (Fig. 5). These findings are similar with the report of McCarty et al. (2006), where accessions that exhibited higher values in yield-related traits also tended to have lower fiber quality values.

Overall, the multivariate analysis supports the identification of complementary parental lines for cotton breeding programs. The contrasting profiles of BGSP-00166 and SP-41255 offer a strategic opportunity to combine fiber quality and yield potential through targeted recombination, contributing to the development of ideotypes suited for mechanized cotton production.

This integrative approach, combining agronomic, physiological, and fiber quality assessments, contributes to the early-stage characterization of germplasm and supports parental candidate identification for breeding programs oriented toward mechanized and high-density production systems.

Tabla 1.4: Descriptores

Entradas	Característica de planta			Característica de hoja			Bráctea	Cápsula
	Forma	Canopia	Tipo Floración	Forma	Lámina	Pubescencia		
BGSP-00028	Cónica	Abierta	Individual	Palmada	Plana	Escasa	Normal	Cónica
BGSP-00067	Cónica	Abierta	Individual	Palmada	Plana	Escasa	Normal	Cónica
BGSP-00070	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00072	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00088	Cónica	Medio	Individual	Palmada	Plana	Fuerte	Normal	Cónica
BGSP-00126	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Escasa	Normal	Cónica
BGSP-00145	Cónica	Medio	Individual	Palmada	Plana	Media	Normal	Cónica
BGSP-00159	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00166	Cilíndrica	Compacta	Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00177	Cónica	Abierta	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00192	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00193	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00194	Cilíndrica	Compacta	Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00207	Cónica	Abierta	Individual	Palmada	Plana	Fuerte	Normal	Cónica
BGSP-00269	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00425	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00428	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Redonda
BGSP-00514	Cónica	Abierta	Individual	Digitada (Okra)	Plana	Media	Normal	Cónica
BGSP-00715	Cónica	Medio	Medio Agrupada	Palmada	Plana	Fuerte	Normal	Cónica
BGSP-00748	Cónica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Elíptica
BGSP-00752	Cónica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
BGSP-00755	Cilíndrica	Medio	Medio Agrupadas	Palmada	Plana	Media	Normal	Cónica
BGSP-00759	Cilíndrica	Abierta	Medio Agrupada	Palmada	En copa	Media	Normal	Redonda
BGSP-00779	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
SP 41255	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica
SP 6565	Cilíndrica	Medio	Medio Agrupada	Palmada	Plana	Media	Normal	Cónica

Tabla 1.5: Datos conjuntos...

Entradas	Características morfológicas							Rendimiento y sus componentes					Calidad de fibra				Fenología				
	AP	NN	NRV	NRR	N1RR	A1RR	D1P	RB	RF	RFD	PC	NC	UHML	Str	Mic	IU	DPP	DPF	DCO	DPCA	DFC
BGSP-00028	106,3 (5,5)	13,3 (0,9)	1,0 (0,6)	10,7 (0,7)	3,7 (0,3)	25,3 (0,7)	9,3 (0,7)	24,1 (2,0)	8,7 (0,6)	36,5 (0,7)	4,3 (0,3)	5,7 (0,3)	30,7	34,0	3,1	84,1	25	51	74	118	163
BGSP-00067	102,0 (4,0)	14,7 (0,7)	1,3 (0,3)	11,0 (0,6)	4,7 (0,3)	27,0 (1,7)	11,0 (1,5)	23,3 (3,3)	8,2 (1,0)	36,0 (1,9)	2,8 (0,2)	8,3 (1,2)	26,8	37,0	2,9	80,8	25	54	74	128	163
BGSP-00070	109,3 (0,3)	14,3 (0,3)	1,3 (0,3)	9,7 (0,3)	5,7 (0,7)	40,7 (4,2)	9,7 (1,9)	19,9 (1,3)	7,7 (0,8)	39,6 (0,8)	3,4 (0,3)	6,0 (0,6)	32,5	31,0	3,8	84,6	27	53	76	128	163
BGSP-00072	113,3 (4,3)	17,0 (0,0)	1,3 (0,3)	13,3 (0,3)	4,7 (0,3)	27,3 (3,2)	7,3 (1,8)	26,3 (2,2)	9,8 (0,9)	38,3 (0,6)	3,5 (0,4)	7,7 (0,3)	28,1	28,2	4,3	82,6	25	54	80	130	162
BGSP-00088	104,3 (5,8)	14,3 (0,7)	1,7 (0,3)	10,7 (0,3)	4,7 (0,3)	28,3 (2,7)	14,3 (1,2)	31,0 (3,2)	11,3 (1,1)	37,1 (0,4)	4,4 (0,2)	7,0 (0,6)	31,6	36,5	3,5	86,2	25	54	78	135	163
BGSP-00126	126,3 (3,8)	14,9 (0,2)	1,5 (0,2)	11,3 (0,3)	4,6 (0,1)	37,4 (1,4)	8,9 (0,9)	16,0 (0,6)	6,6 (0,3)	42,9 (0,3)	2,7 (0,2)	6,0 (0,3)	29,9	32,4	2,8	83,5	25	54	79	132	163
BGSP-00145	89,0 (4,5)	14,3 (0,3)	1,0 (0,6)	11,3 (0,3)	4,0 (0,0)	25,7 (0,9)	9,7 (1,8)	16,1 (3,1)	6,1 (1,1)	38,6 (0,7)	3,6 (0,6)	4,7 (0,9)	26,9	26,6	3,5	80,8	25	52	73	121	163
BGSP-00159	104,3 (3,6)	14,3 (0,3)	1,1 (0,2)	11,0 (0,2)	4,3 (0,2)	31,8 (1,9)	11,4 (0,6)	14,3 (0,6)	5,1 (0,2)	36,8 (0,6)	2,7 (0,1)	5,2 (0,2)	30,0	34,4	2,5	82,7	22	49	74	114	161
BGSP-00166	109,0 (3,0)	15,2 (0,2)	0,8 (0,2)	11,8 (0,3)	4,4 (0,2)	30,6 (1,2)	3,9 (0,6)	18,2 (0,5)	5,5 (0,2)	31,1 (0,4)	3,6 (0,1)	5,1 (0,2)	33,9	37,6	2,5	86,1	23	52	74	124	163
BGSP-00177	103,7 (3,3)	14,3 (0,3)	1,7 (0,3)	10,7 (0,3)	4,7 (0,3)	32,7 (3,4)	13,7 (1,7)	24,5 (2,2)	7,9 (0,8)	33,0 (0,4)	4,2 (0,8)	6,0 (0,6)	29,2	30,7	4,2	82,9	26	53	75	127	164
BGSP-00192	101,3 (5,8)	15,0 (0,0)	1,7 (0,3)	11,0 (0,0)	5,0 (0,0)	29,0 (2,1)	11,3 (0,9)	18,7 (3,3)	7,1 (1,1)	37,9 (2,0)	3,3 (0,4)	5,7 (0,3)	28,2	31,9	3,5	83,3	25	54	76	124	162
BGSP-00193	106,7 (5,5)	15,3 (0,3)	2,7 (0,3)	11,3 (0,3)	5,0 (0,0)	30,7 (0,9)	8,0 (1,7)	29,4 (3,9)	10,9 (1,4)	38,1 (1,8)	4,2 (0,6)	7,0 (0,0)	30,2	33,8	3,5	85,5	25	55	78	137	162
BGSP-00194	92,7 (3,2)	14,7 (0,3)	2,3 (0,3)	10,7 (0,3)	5,0 (0,0)	31,3 (0,7)	5,7 (3,2)	24,0 (3,1)	9,6 (1,4)	40,9 (0,8)	3,7 (0,4)	6,7 (0,9)	29,9	32,6	3,7	85,0	25	55	75	118	163
BGSP-00207	105,0 (8,5)	15,3 (0,2)	0,7 (0,2)	12,2 (0,2)	4,2 (0,1)	29,4 (0,8)	10,1 (1,0)	16,2 (0,6)	6,7 (0,2)	43,6 (0,5)	2,8 (0,1)	5,8 (0,2)	28,6	28,4	2,8	83,2	23	52	79	135	164
BGSP-00269	113,0 (3,7)	14,7 (0,2)	1,2 (0,2)	11,2 (0,2)	4,4 (0,1)	34,2 (1,6)	8,3 (0,8)	16,9 (0,5)	7,1 (0,3)	43,7 (0,5)	2,9 (0,2)	6,1 (0,3)	29,2	33,4	3,9	84,1	23	52	75	133	163
BGSP-00425	105,2 (3,3)	13,8 (0,1)	0,9 (0,3)	10,8 (0,1)	4,0 (0,0)	25,5 (0,7)	8,4 (0,9)	15,1 (0,5)	4,6 (0,2)	31,5 (0,3)	2,6 (0,1)	5,8 (0,3)	32,1	32,6	2,3	82,2	25	53	73	130	163
BGSP-00428	105,3 (7,3)	15,3 (0,3)	0,7 (0,3)	12,0 (0,6)	4,3 (0,3)	28,0 (2,9)	8,7 (2,4)	21,4 (3,6)	8,7 (1,4)	40,1 (0,4)	4,3 (0,7)	5,0 (0,6)	25,1	23,1	4,0	80,0	23	49	74	113	163
BGSP-00514	99,0 (8,0)	16,0 (0,0)	2,7 (0,3)	12,3 (0,3)	4,7 (0,3)	25,0 (1,5)	11,7 (1,7)	28,0 (2,8)	11,7 (1,2)	42,3 (0,4)	4,2 (0,2)	6,7 (0,9)	29,0	32,0	2,7	83,6	23	52	76	117	161
BGSP-00715	102,0 (8,5)	14,7 (0,9)	0,3 (0,3)	11,7 (0,9)	4,0 (0,0)	27,3 (2,3)	13,0 (1,2)	24,0 (3,6)	9,8 (1,2)	41,6 (1,2)	3,4 (0,4)	7,0 (0,6)	27,2	31,7	4,0	83,5	24	55	78	134	163
BGSP-00748	105,7 (0,9)	15,0 (0,0)	1,7 (0,3)	12,0 (0,0)	4,0 (0,0)	25,7 (1,2)	12,0 (0,6)	33,5 (1,8)	12,3 (1,0)	39,0 (2,4)	4,4 (0,1)	7,7 (0,3)	26,3	21,5	3,9	80,9	23	52	74	135	164
BGSP-00752	103,3 (5,2)	15,3 (0,3)	2,7 (0,3)	11,3 (0,3)	5,0 (0,0)	29,3 (0,7)	10,3 (0,9)	27,2 (2,1)	11,3 (1,2)	42,1 (1,9)	4,1 (0,1)	6,7 (0,3)	31,8	26,7	3,6	84,1	25	55	79	135	162
BGSP-00755	97,3 (3,3)	13,0 (0,0)	2,7 (0,3)	10,0 (0,0)	4,0 (0,0)	28,7 (0,3)	10,3 (0,7)	26,3 (2,0)	10,0 (0,7)	38,5 (0,8)	4,7 (0,2)	5,7 (0,7)	29,8	31,4	3,3	84,2	25	53	75	124	163
BGSP-00759	108,3 (5,0)	14,0 (0,6)	1,0 (0,6)	10,0 (1,0)	5,0 (0,6)	38,3 (5,5)	9,7 (2,4)	24,6 (3,6)	8,7 (1,5)	38,3 (1,1)	4,7 (0,6)	5,7 (1,5)	27,8	23,9	4,1	82,3	25	54	77	134	163
BGSP-00779	112,3 (1,9)	16,0 (0,0)	1,3 (0,3)	12,3 (0,3)	4,7 (0,3)	30,3 (1,8)	11,7 (0,7)	26,3 (1,5)	10,4 (0,5)	40,5 (0,7)	3,0 (0,1)	8,7 (0,3)	30,2	31,5	2,7	84,4	25	55	76	126	161
SP 41255	122,3 (2,9)	15,2 (0,1)	1,2 (0,2)	11,8 (0,2)	4,4 (0,1)	32,7 (1,1)	11,2 (0,8)	20,7 (0,7)	8,8 (0,4)	44,7 (0,5)	2,8 (0,1)	7,3 (0,3)	30,9	32,3	3,0	84,1	23	52	76	124	162
SP 6565	120,8 (1,9)	15,7 (0,2)	1,3 (0,2)	12,2 (0,2)	4,5 (0,2)	35,8 (1,6)	8,4 (0,6)	17,3 (0,5)	6,9 (0,2)	41,2 (0,3)	2,6 (0,1)	6,8 (0,3)	29,9	33,3	3,0	84,6	25	55	79	131	162
p-valor	<0,001	<0,001	<0,001	<0,001	<0,001	<0,001	<0,001	<0,001	<0,001	<0,001	<0,001	<0,001	-	-	-	-	-	-	-	-	
L.S.D Fisher	17,96	1	0,99	1,05	0,74	6,29	3,89	4,3	1,76	2,46	0,76	1,42	-	-	-	-	-	-	-	-	

Referencias: AP: altura de planta a madurez en cm; NN: número de nudos en el tallo principal; NRV: número de ramas monopodiales (vegetativas); NRR: número de nudos de ramas simpodiales; N1RR: posición de la primera rama fructífera medida como número de nudo del tallo; A1RR: posición de la primera rama fructífera medida como altura en cm; D1P: distancia desde la primera posición reproductiva al tallo principal en cm. RB: rendimiento de algodón en bruto en g; RF: rendimiento de fibra en g; RFD: porcentaje de fibra en %; PC: peso de capullo en g; NC: número de capullos por planta. UHML: longitud de fibra en mm; Str: resistencia de fibra en g tex-1; Mic: micronaire; IU: uniformidad de fibra en %. DPP: días desde emergencia de plántula hasta primer botón floral; DPF: días desde emergencia hasta primera flor; DCO: días desde emergencia hasta cut out; DPCA: días desde emergencia hasta apertura del primer capullo; DM: días desde emergencia hasta madurez.

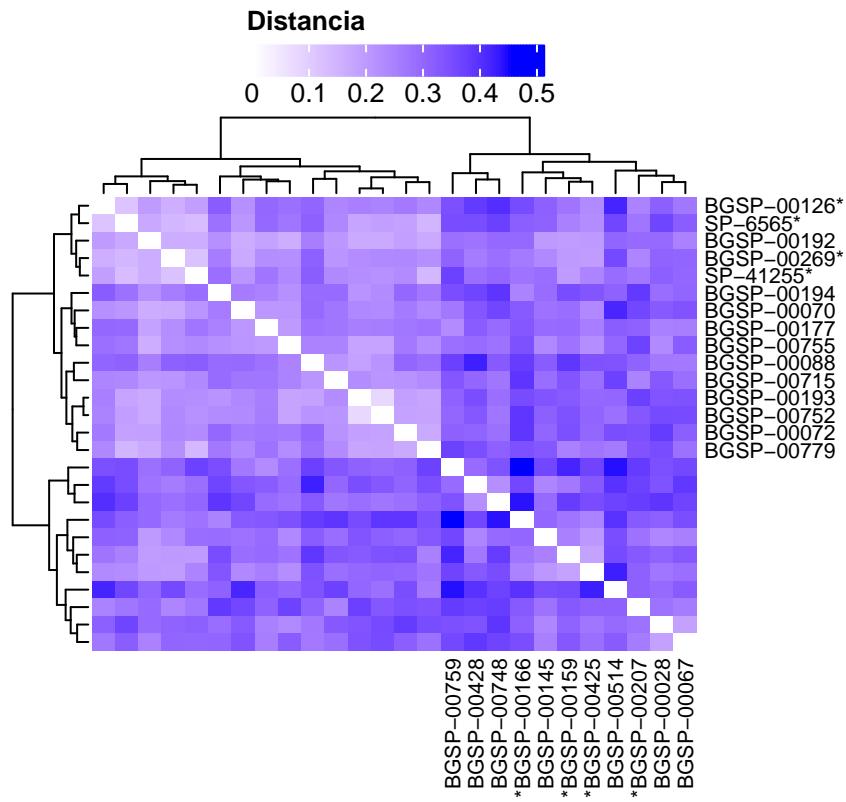
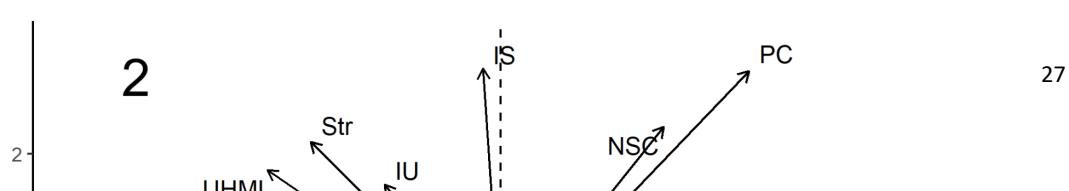
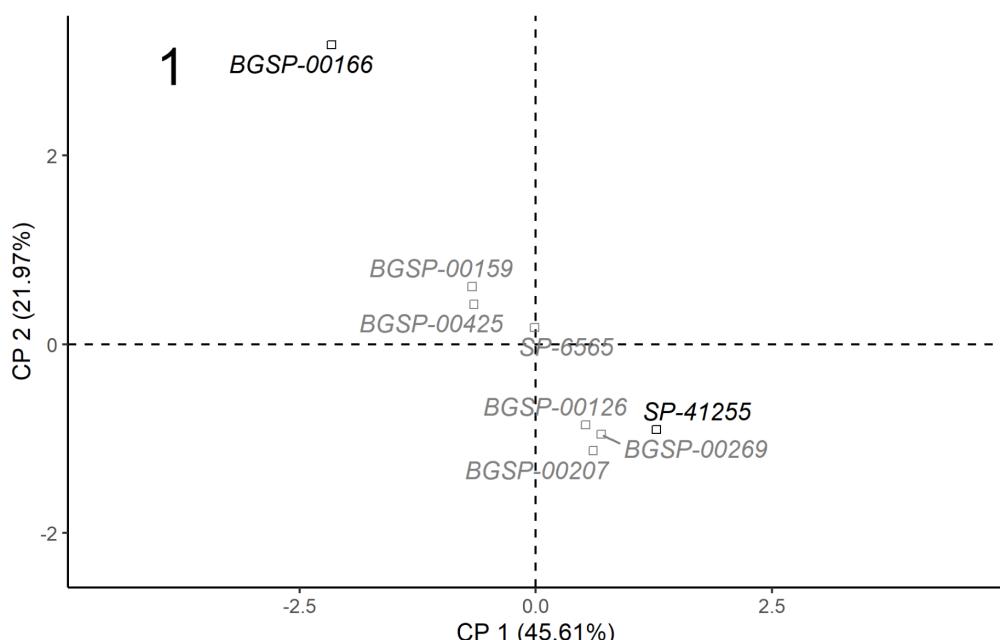


Figura 1.1: Heatmap que muestra las distancias calculadas entre los datos utilizando la métrica de Gower.

Tabla 1.6: Valores medios, error estándar (entre paréntesis) y prueba L.S.D. (diferencia mínima significativa) de Fisher para rendimiento de fibra, sus componentes y calidad.

Entradas	Rendimiento y sus componentes								Calidad de fibra			
	RB	RF	RFD	PC	NC	IS	IF	NSC	UHML	Str	Mic	IU
BGSP-00126	30,5 (6,6)	13,2 (2,9)	43,1 (0,4)	3,6 (0,4)	8,0 (1,0)	8,7 (0,3)	6,2 (0,3)	25,0 (1,6)	29,8 (0,3)	30,4 (1,3)	3,5 (0,4)	83,8 (1,0)
BGSP-00159	26,2 (5,4)	9,5 (1,9)	36,3 (0,8)	4,1 (0,5)	6,2 (0,6)	10,3 (1,0)	5,6 (0,7)	26,8 (0,7)	28,4 (0,9)	33,2 (2,1)	3,7 (0,5)	82,7 (0,5)
BGSP-00166	27,5 (4,3)	8,6 (1,4)	31,2 (0,3)	4,6 (0,4)	5,8 (0,5)	11,5 (0,5)	5,0 (0,4)	28,9 (0,8)	33,2 (0,5)	37,3 (2,0)	3,2 (0,3)	85,8 (0,6)
BGSP-00207	30,2 (6,3)	13,1 (2,7)	43,5 (0,4)	3,6 (0,4)	8,0 (1,0)	8,3 (0,2)	6,2 (0,3)	25,2 (2,0)	28,3 (0,4)	29,2 (0,7)	3,4 (0,3)	83,5 (0,4)
BGSP-00269	29,8 (5,8)	12,9 (2,5)	43,4 (0,4)	3,4 (0,2)	8,6 (1,1)	8,8 (0,4)	6,5 (0,2)	22,8 (1,9)	28,3 (0,5)	31,6 (1,2)	4,3 (0,2)	83,7 (0,3)
BGSP-00425	31,5 (7,4)	11,3 (2,9)	34,2 (1,2)	3,6 (0,4)	8,3 (1,1)	7,9 (0,4)	4,0 (0,6)	30,6 (0,9)	30,4 (1,0)	31,3 (1,6)	3,4 (0,5)	82,2 (0,2)
SP-41255	34,2 (6,1)	15,8 (3,0)	45,8 (0,6)	3,7 (0,5)	9,2 (1,0)	7,7 (0,1)	6,2 (0,3)	26,8 (2,8)	29,8 (0,7)	32,2 (1,2)	3,8 (0,3)	83,6 (0,7)
SP-6565	30,1 (5,8)	12,5 (2,4)	41,4 (1,0)	3,7 (0,6)	8,2 (0,7)	8,9 (0,4)	5,9 (0,2)	25,5 (3,0)	30,1 (0,5)	32,5 (0,9)	3,7 (0,3)	84,9 (0,6)
p-value	0,005	<0,001	<0,001	0,002	<0,001	<0,001	<0,001	0,014	<0,001	0,017	<0,001	0,003
Fisher's L.S.D	3,68	1,91	1,99	0,57	1,19	1,35	0,75	4,04	1,38	4,03	0,43	1,66

Referencias: RB: rendimiento de algodón en bruto en g; RF: rendimiento de fibra en g; RFD: porcentaje de fibra en %; PC: peso de capullo en g; NC: número de capullos por planta; IS: índice de semilla en g; IF: índice de fibra en g; NSC: número de semillas por capullo. UHML: longitud de fibra en mm; Str: resistencia de fibra en g tex-1; Mic: micronaire; IU: uniformidad de fibra en %.

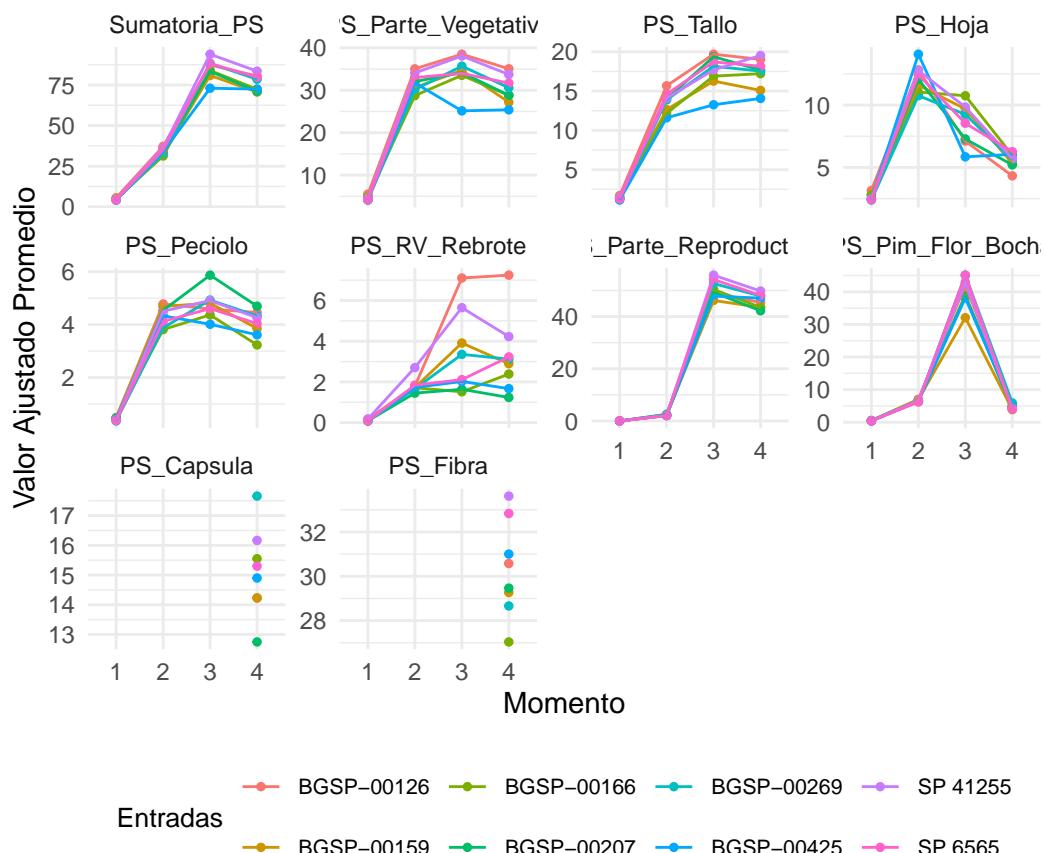


**Tabla 1.7: Correlación de Spearman entre los rasgos evaluados en el germosplasma de colección**

	RB	RF	RFD	PC	NC	IS	IF	NSC	UHML	Str	Mic
RF	.94***	-									
RFD	.24	.48***	-								
PC	.75***	.67***	-.15	-							
NC	.86***	.89***	.42**	.43**	-						
IS	.01	-.09	-.51***	.41**	-.27	-					
IF	.52***	.70***	.54***	.49***	.49***	.20	-				
NSC	.54***	.38**	-.34*	.69***	.30*	-.03	-.06	-			
UHML	-.35*	-.46**	-.36*	-.15	-.42**	.14	-.46**	.06	-		
Str	-.14	-.27	-.42**	.07	-.30*	.51***	-.17	-.08	.38**	-	
Mic	.79***	.83***	.35*	.64***	.77***	.06	.71***	.28	-.53***	-.23	-
IU	-.05	-.06	-.01	.09	-.10	.40**	.09	-.14	.34*	.46***	-.10

*Referencias:* \*p <0.05, \*\*p <0.01 and \*\*\* p <0.001. RB: rendimiento de algodón en bruto en g; RF: rendimiento de fibra en g; RFD: porcentaje de fibra en %; PC: peso de capullo en g; NC: número de capullos por planta; IS: índice de semilla en g; IF: índice de fibra en g; NSC: número de semillas por capullo. UHML: longitud de fibra en mm; Str: resistencia de fibra en g tex-1; Mic: micronaire; IU: uniformidad de fibra en %.

### Valores Ajustados Promedios por Momento y Entradas



**Tabla 1.8: Acumulacion de biomasa en estructuras vegetativas, tallo, hojas, peciolos, ramas vegetativas, rebrote, biomasa reproductiva, pimpollo, flor, bocha, cápsula, semilla**

ANOVA						
Rasgos	Fuente	GLNum	GLDen	F	p-valor	
Biomasa Total	(Intercept)	1	193	62.54	<0.0001	
	Momento	3	21	92.71	<0.0001	
	Entradas	7	193	6.34	<0.0001	
	Momento:Entradas	21	193	2.37	0.0011	
1 Biomasa Vegetativa	(Intercept)	1	193	210.27	<0.0001	
	Momento	3	21	196.04	<0.0001	
	Entradas	7	193	4.55	0.0001	
	Momento:Entradas	21	193	1.65	0.0416	
1.1 Tallo	(Intercept)	1	193	332.03	<0.0001	
	Momento	3	21	249.72	<0.0001	
1.2 Hoja	Entradas	7	193	6.30	<0.0001	
	Momento:Entradas	21	193	2.41	0.0009	
	(Intercept)	1	193	211.00	<0.0001	
	Momento	3	21	39.10	<0.0001	
1.3 Peciolo+Ramas	Entradas	7	193	0.70	0.6741	
	Momento:Entradas	21	193	1.81	0.0195	
	(Intercept)	1	193	148.48	<0.0001	
	Momento	3	21	123.02	<0.0001	
1.4 RV+Rebrote	Entradas	7	193	2.73	0.0102	
	Momento:Entradas	21	193	1.46	0.0956	
	(Intercept)	1	193	63.81	<0.0001	
	Momento	3	21	21.61	<0.0001	
2 Biomasa Reproductiva	Entradas	7	193	4.02	0.0004	
	Momento:Entradas	21	193	1.49	0.0829	
	(Intercept)	1	193	10.52	0.0014	
	Momento	3	21	62.70	<0.0001	
2.1 Prim+Flor+Bocha	Entradas	7	193	3.15	0.0035	
	Momento:Entradas	21	193	1.45	0.0992	
	(Intercept)	1	193	132.53	<0.0001	
	Momento	3	21	94.71	<0.0001	
2.2 Cápsula (M4)	Entradas	7	193	0.94	0.4797	
	Momento:Entradas	21	193	0.63	0.8961	
2.3 Fibra+Semilla (M4)	(Intercept)	1	47	10.82	0.0019	
	Entradas	7	47	3.85	0.0022	
	(Intercept)	1	47	13.01	0.0007	
	Entradas	7	47	2.36	0.0373	

Tabla 1.9: Retención global, y en posiciones 1, 2 y 3, en los momentos M2, M3, y M4

ANOVA						
Rasgos	Fuente	GLNum	GLDen	F	p-valor	
Ret_Global	(Intercept)	1	144	90.21	<0.0001	
	Momento	2	14	52.91	<0.0001	
	Entradas	7	144	3.48	0.0018	
	Momento:Entradas	14	144	2.21	0.0098	
Ret_P1	(Intercept)	1	144	338.14	<0.0001	
	Momento	2	14	89.26	<0.0001	
	Entradas	7	144	1.47	0.1810	
	Momento:Entradas	14	144	3.05	0.0004	
Ret_P2	(Intercept)	1	144	22.78	<0.0001	
	Momento	2	14	8.89	0.0032	
Ret_P3	Entradas	7	144	2.65	0.0131	
	Momento:Entradas	14	144	1.29	0.2206	
	(Intercept)	1	144	13.92	0.0003	
	Momento	2	14	6.59	0.0096	
Ret_P3	Entradas	7	144	1.06	0.3914	
	Momento:Entradas	14	144	0.92	0.5359	

### Valores Ajustados Promedios por Momento y Entradas

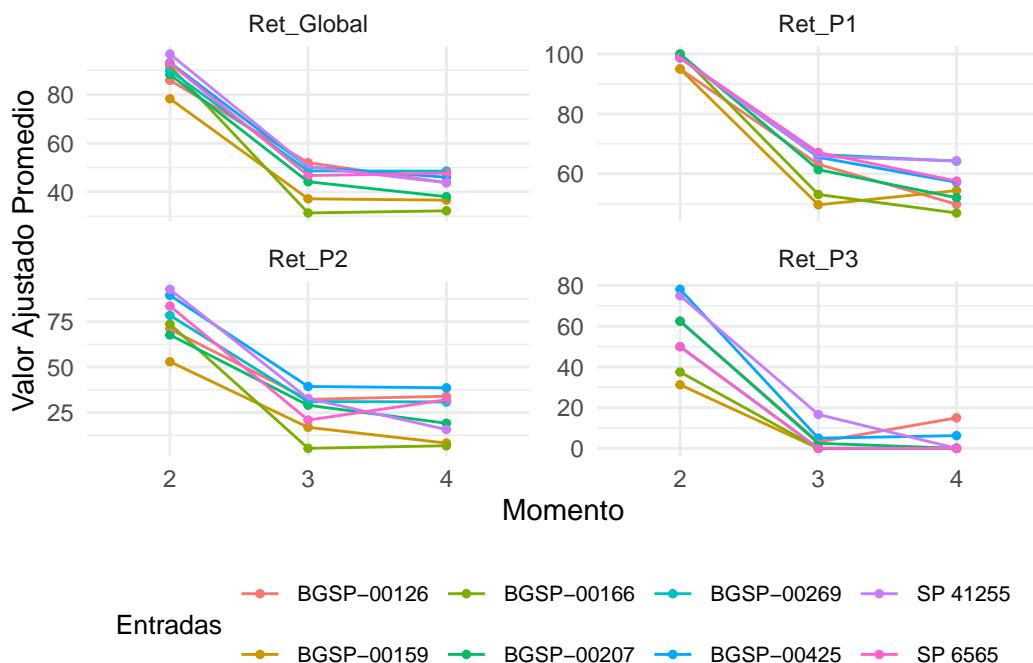
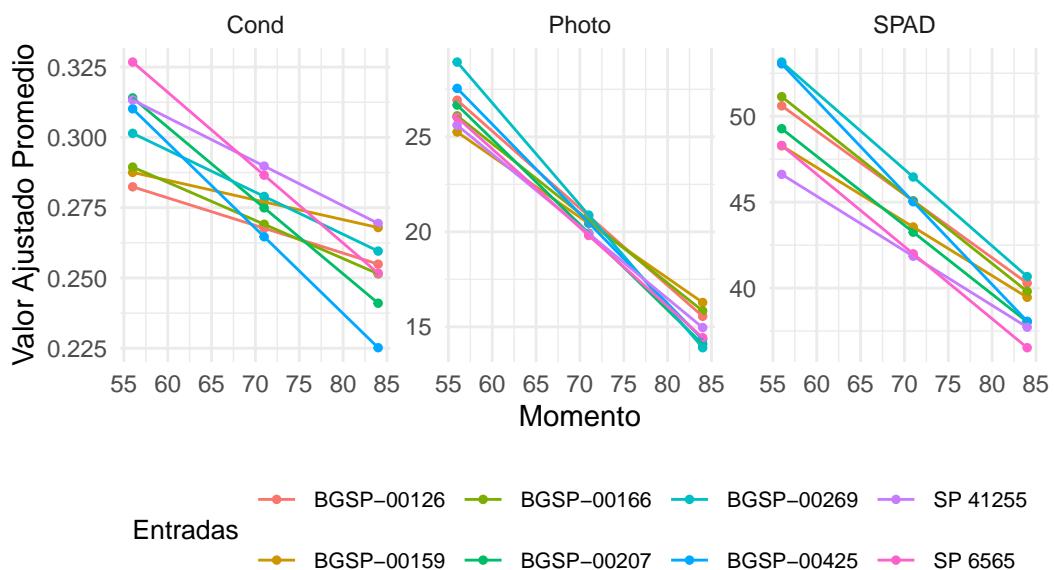


Tabla 1.10: Fotosintesis, conductancia estomática y SPAD

ANOVA					
Rasgos	Fuente	GLNum	GLDen	F	p-valor
SPAD	(Intercept)	1	56	139.88	<0.0001
	DDE	1	56	19.76	<0.0001
	Entradas	7	21	1.04	0.4363
	DDE:Entradas	7	56	0.74	0.6355
Cond	(Intercept)	1	56	26.32	<0.0001
	DDE	1	56	1.18	0.2814
	Entradas	7	21	0.90	0.5224
	DDE:Entradas	7	56	0.91	0.5085
Photo	(Intercept)	1	56	339.17	<0.0001
	DDE	1	56	122.17	<0.0001
	Entradas	7	21	3.49	0.0122
	DDE:Entradas	7	56	3.36	0.0046

### Valores Ajustados Promedios por Momento y Entradas



## 1.12. Discusión

Aquí la discusión del capítulo 1

plant mapping specifically refers to the recording and evaluating of plant structure and the distribution and retention of fruit on plants at a specific time (Kerby et al., 2010)

## 1.13. Conclusión

This study revealed substantial phenotypic, physiological, and agronomic variation among the evaluated cotton germplasm, confirming the diversity preserved in the INTA collection and its potential for cotton crop breeding. Morphological descriptors, yield components, and fiber traits showed broad ranges, enabling the identification of accessions with contrasting and complementary characteristics.

Among the eight selected accessions, SP-41255 consistently exhibited superior performance for LY, LP, and BN, whereas BGSP-00166 combined a compact plant architecture with the highest FL, FS, and FU. Differences in biomass partitioning and reproductive retention reflected distinct growth strategies, with high-yielding accessions allocating more assimilates to reproductive structures and maintaining higher fruit retention. Physiological measurements indicated that variation in net photosynthesis, rather than stomatal or chlorophyll traits, contributed most to differences in biomass accumulation and yield formation.

Multivariate analyses supported these patterns by separating accessions primarily along a yield–fiber quality gradient. The contrasting profiles of BGSP-00166 and SP-41255 highlight a promising parental combination for simultaneously improving fiber quality and yield potential. Overall, the combined assessment of agronomic, physiological, and fiber traits provides a framework for the early identification of promising parental candidates, supporting subsequent field-based evaluation and breeding decisions aimed at mechanized and high-density production systems. Future multi-environment field studies will be required to confirm trait stability and harvest adaptability.

# **Capítulo 2**

## **Identificación de QTL de importancia agronómica**

### **2.1. Introducción**

Aquí una breve introducción del capítulo

### **2.2. Objetivo**

Mapear QTL (Quantitative Trait Loci) asociados a caracteres de rendimiento de fibra mediante el análisis de una población segregante F<sub>2</sub> obtenida del cruzamiento de progenitores contrastantes.

### **2.3. Materiales y métodos**

Se caracterizaron marcadores moleculares del tipo SSR (Single Sequence Repeats o microsatélites) asociados a QTLs de importancia agronómica. Se utilizaron 55 SSR chequear n° de marcadores que fueron seleccionados acorde tanto a trabajos científicos (Ademe et al., 2017; An et al., 2010; Baytar et al., 2018; Iqbal & Rahman, 2017; Li et al., 2017; R. Liu et al., 2012; R. Liu et al., 2018; X. Liu et al., 2017; Qin et al.,

2015; Shen et al., 2007; Shi et al., 2015; Su et al., 2016; B. Wang et al., 2007; H. Wang et al., 2015; M. Wang et al., 2014; Xia et al., 2014; S. Zhang et al., 2016; Z.-S. Zhang et al., 2005) como la base de datos de CottonGen (<http://www.cottongen.org>). Dicha actividad se realizó en el laboratorio de biotecnología de INTA Reconquista. A partir de las entradas contrastantes para las características de interés, se realizaron cruzamientos y por autofecundación de las F<sub>1</sub> se obtuvo la población F<sub>2</sub>. A continuación, se detallan las etapas:

I. Obtención de la población segregante para los loci de los marcadores (M) y los QTLs. Luego de la caracterización morfológica y fisiológica, las entradas fueron seleccionadas para ser utilizadas como parentales contrastantes en los respectivos cruzamientos de este estudio. Para los mismos se utilizaron las entradas de alto porcentaje de desmote como parentales masculinos. Considerando que por cada bocha se obtiene, en promedio, 20 semillas (Naeem et al., 2017) se tomó una flor de cada parental para dicho cruzamiento, para la generación de la población F<sub>1</sub> (20 plantas). La población segregante (F<sub>2</sub>) resultó de la autofecundación de las F<sub>1</sub>, para llegar a una población F<sub>2</sub> aproximada de 200 plantas (Bardak et al., 2018; T. Zhang et al., 2003).

BGSP-00166 y SP-41255 fueron los genotipos parentales seleccionados. Ambos parentales son de origen argentino. El primer parental es una línea endogámica derivada del cruce entre Chaco 510 x SP 6269, mientras que el segundo parental es otra línea endogámica seleccionada del cruce entre SP-81270-7 x Mij. 3. Morfológicamente, difieren en su estructura de planta (Figure 2.2). BGSP-00166 es más compacta, con una distancia más corta entre la primera posición de fructificación y el tallo, mientras que SP-41255 tiene una estructura de planta más abierta. El cruzamiento se realizó en ambas direcciones para evaluar la existencia de efecto recíproco. La emasculación se realizó en las plantas utilizadas como parentales fe-

meninos mediante la eliminación de los estambres con pinzas antes de la antesis floral (estadio en forma de vela), durante las horas de la tarde. Al día siguiente, se realizó el cruce mediante la transferencia de polen de los parentales masculinos a los femeninos (flores emasculadas), tal y como se describe en Acquaah (2012). A continuación, las semillas  $F_1$  de los cruces de algodón se sembraron en condiciones de invernadero y se autopolinizaron para producir  $F_2$ . En la temporada de cultivo 2019-2020, la población  $F_2$  se sembró en hiladas no replicadas en condiciones de campo. Cada hilada tenía 10 m de longitud y una separación de 1 m, con 50 plantas por hilada (Figura 2.1). Las generaciones no segregantes (tanto las  $F_1$ s, SP-41255 x BGSP-00166 y BGSP-00166 x SP-41255, como los genotipos parentales) también se sembraron en las mismas condiciones. Todas estas generaciones se sometieron a evaluación fenotípica. En la dirección de cruzamiento de SP-41255 x BGSP-00166, se consiguió una población segregante  $F_2$  de 205 plantas, mientras que para la dirección de cruzamiento inversa, se obtuvieron 234 plantas. En el mismo ensayo, como se muestra en la Figura 2.1, se sembraron y consiguieron uniformemente 17 plantas del progenitor BGSP-00166, 19 plantas del progenitor SP-41255 y 11 plantas de la  $F_1$  (mitad y mitad generadas a partir de una dirección de cruzamiento y su recíproca).

- II. Medición de la característica fenotípica controlada por los QTL. Se realizarán las mediciones de rendimiento y calidad de fibra detalladas en el apartado "Caracterización morfológica". Para el experimento del cruce biparental, cada planta (de poblaciones segregantes y no segregantes) también se cosechó individualmente en la fase de madurez para evaluar el rendimiento y las características relacionadas con la calidad: SCY, LY, LP, BW, BN, FL, FS, FU y MIC se midieron como se ha descrito anteriormente.
- III. Caracterización molecular por SSR de los de los parentales y cada planta de la po-

blación F<sub>2</sub>. El ADN genómico se aisló utilizando hojas jóvenes procedentes de los progenitores y la población segregante, mediante método CTAB modificado (Paterson et al., 1993; J. Zhang & Stewart, 2000). Se evaluó la cantidad y calidad del ADN mediante espectrofotometría para comparación. La amplificación por PCR y la tinción con plata se realizó según Lin et al. (2005) . Los productos de PCR de los SSR se separaron en geles de poliacrilamida desnaturizantes al 2 % y revelados con carbonato de sodio(Lin et al., 2005).

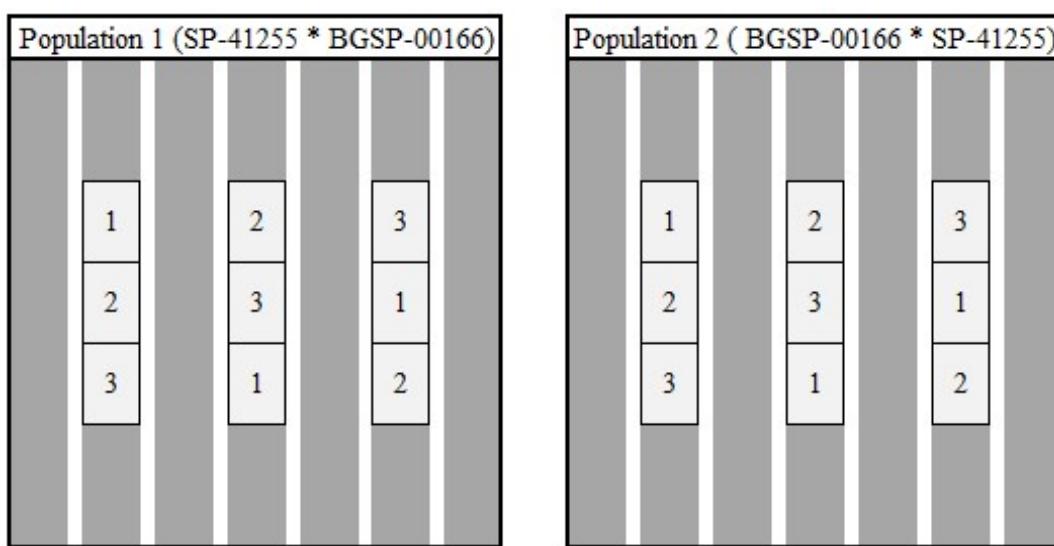


Figura 2.1: Descripción del ensayo de campo, mostrando las dos poblaciones generadas a partir del cruce en la dirección SP-41255 x BGSP-00166 y su recíproco. En el ensayo se distribuyeron al azar plantas del parental SP-41255 (1), Filial 1 (2) y del parental BGSP-00166 (3).

### 2.3.1. Análisis estadístico

Para el experimento de cruzamiento biparental, se realizó una prueba de Levene para confirmar la varianza homogénea en las generaciones no segregantes, mientras que la distribución normal se confirmó mediante la prueba de Shapiro-Wilk. Se realizó la prueba L.S.D. de Fisher para comparar las parentales P<sub>1</sub>, P<sub>2</sub> y la generación F<sub>1</sub>. Además, también se realizó una comparación de las medias de las generaciones F<sub>1</sub> y F<sub>2</sub> recíprocas para detectar la existencia de un efecto recíproco en las características objeto de estudio. Se

realizó un test de bondad de ajuste Chi-Cuadrado para detectar segregación transgresiva siguiendo el enfoque propuesto por Rodriguez et al. (2005) . Se consideró que la generación parental tenía un tamaño igual al de la generación F<sub>2</sub>. Las frecuencias esperadas se calcularon asumiendo igual tamaño entre la generación parental y la F<sub>2</sub>, y se consideraron fenotipos extremos aquellos individuos de la F<sub>2</sub> que superaban la media del rasgo en la generación parental y dos desvíos estándar (test de una cola, el 2,5 % de los individuos en una distribución normal). Bajo estos supuestos, cuando la proporción observada de individuos extremos en la generación F<sub>2</sub> superaba esta proporción esperada (un χ<sup>2</sup> significativo), se consideraba que el rasgo mostraba un patrón de herencia transgresivo.

La heredabilidad en sentido amplio ( $h_{bs}^2$ ) se estimó a partir de los componentes de varianza de las generaciones no segregantes (P<sub>1</sub>, P<sub>2</sub> and F<sub>1</sub>) y de la generación F2 de la siguiente manera (Kearsey & Pooni, 1996):

$$h_{bs}^2 = \frac{\sigma^2 G}{\sigma^2 Ph} \times 100$$

Donde  $h_{bs}^2$  = heredabilidad en sentido amplio,  $\sigma^2 G$  = varianza genotípica,  $\sigma^2 Ph$  = varianza fenotípica.

La varianza entre plantas dentro de las F<sub>1</sub>'s y los parentales P<sub>1</sub> y P<sub>2</sub> se consideró como varianza ambiental y se restó de la varianza entre plantas F<sub>2</sub> para obtener estimaciones de la varianza genotípica.

$$\sigma^2 E = \frac{\sigma^2 P_1 + \sigma^2 P_2 + \sigma^2 F_1}{3}$$

Donde  $\sigma^2 E$  = Varianza ambiental,  $\sigma^2 P_1$  = Varianza del parental 1,  $\sigma^2 P_2$  = Varianza del parental 2,  $\sigma^2 F_1$  = varianza de la generación F<sub>1</sub> .

$$\sigma^2 G = \sigma^2 Ph - \sigma^2 E$$

Para estimar la correlación genética entre rasgos se utilizó la metodología propuesta por Kearsey y Pooni (1996) .

## 2.4. Resultados



Figura 2.2: Planta de los genotipos parentales SP-41255 y BGSP-00166

### 2.4.1. Generación de variabilidad a partir del cruce biparental BGSP-00166 x SP-41255

Los genotipos parentales BGSP-00166 y SP-41255 fueron significativamente diferentes para todos los caracteres evaluados, a excepción de RB (Tabla 2.1). Por otra parte, los valores medios de todos los caracteres no mostraron diferencias significativas entre la F<sub>1</sub> y la F<sub>2</sub> recíproca ( $p > 0,05$ ), como se muestra en la Tabla S1. Esto significa que no existe el efecto recíproco en este cruce para los caracteres analizados. Por lo tanto, decidimos fusionar las poblaciones recíprocas F<sub>1</sub> y F<sub>2</sub>.

Tabla 2.1: Mean and variance (in parentheses) of the traits in the non-segregating generations (BGSP-00166, SP-41255 and F1) and the segregating generation F2

Generations	Caracteres								
	SCY	LY	LP	BW	BN	FL	FS	MIC	FU
BGSP-00166	89.5a (1266.4)	25.8a (110.3)	30.9a (0.9)	6.2b (0.4)	14.5a (29.8)	32.5b (0.3)	37.1b (2.8)	3.9a (0.1)	85.5b (1.8)
SP-41255	86.9a (761.8)	36.6b (126.1)	43.4c (0.8)	4.6a (0.3)	19.1b (29.6)	26.9a (0.6)	30.7a (2.9)	4.8c (0.1)	82.0a (2.1)
F~1~	123.5b (1013.1)	44.1b (126.6)	37.4b (0.4)	6.4b (0.6)	19.4b (26.7)	32.0b (0.3)	37.4b (0.9)	4.2b (0.1)	86.1b (0.9)
F~2~	114.1 (1957.2)	37.5 (227.3)	37.2 (4.4)	5.9 (0.9)	19.7 (52.5)	31.3 (1.5)	36.1 (5.2)	4.3 (0.2)	84.9 (1.7)

RB: Seed cotton yield in g, LY: Lint yield in g,

LP: Lint percentage in %, BW: Boll weight in g, BN: Boll number

per plant, SI: Seed index in g, LI: Lint index in g, SNPB: Seed

number per boll, FL: Fibre length in mm, FS: Fibre strength

g tex<sup>-1</sup>, Mic: Micronaire, FU: Fibre uniformity in %.

## 2.5. Discusión

Aquí la discusión del capítulo 3

## 2.6. Conclusión

Aquí la conclusión del capítulo 3

# **Capítulo 3**

## **Si es necesario un tercer capítulo**

### **3.1. Introducción**

Aquí una breve introducción del capítulo

### **3.2. Objetivo**

Aquí objetivo del capítulo 3

### **3.3. Materiales y métodos**

Aquí M y M del capítulo 3

### **3.4. Discusión**

Aquí la discusión del capítulo 3

### **3.5. Conclusión**

Aquí la conclusión del capítulo 3



# **Conclusión**

Las conclusiones de la tesis aquí..

# **Apéndice A**

## **Primer Apéndice**

Sí es necesario incluir un apéndice, iría aquí..

**En capítulo 1:**

Descripción aquí..

## **Apéndice B**

### **Segundo Apéndice**

Este sería el segundo apéndice..

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