## National Health and Nutrition Examination Survey

2015-2016 Data Documentation, Codebook, and Frequencies

Lead, Cadmium, Total Mercury, Selenium & Manganese - Blood (PBCD\_I)

Data File: PBCD\_I.xpt

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### Component Description

#### Lead

Lead is a known environmental toxin that has been shown to affect deleteriously the nervous, hematopoietic, endocrine, renal, and reproductive systems. In young children, lead exposure is a particular hazard because children more readily absorb lead than do adults, and children's developing nervous systems also make them more susceptible to the effects of lead. The primary sources of exposure for children are lead-laden paint chips and dust as a result of deteriorating lead-based paint. The risk for lead exposure is disproportionately higher for children who are poor, non-Hispanic black, living in large metropolitan areas, or living in older housing. Among adults, the most common high exposure sources are occupational. Blood lead levels measured in previous NHANES programs have been the cornerstone of lead exposure surveillance in the U.S. The data have been used to document the burden and dramatic decline of elevated blood lead levels, to promote the reduction of lead use, and to help to redefine national lead poisoning prevention guidelines, standards, and abatement activities.

#### Cadmium

A cadmium assay is performed to identify cases of cadmium toxicity. Occupational exposure is the most common cause of elevated cadmium levels.

#### Manganese

The greatest demand for manganese is for the production of iron and steel. In addition, it is a key component of low-cost stainless steel and certain aluminum alloys. At low concentrations, it is used to decolorize glass, while at higher concentrations; it is used to make violet-colored glass. Manganese dioxide, besides being a useful pigment, is a catalyst and a component of certain dry cell batteries. Potassium permanganate is a potent oxidizer and disinfectant. Manganese (in the form of manganese ions) is an essential trace nutrient in all known forms of life. On the other hand, excess manganese is toxic.

#### **Total Mercury**

Uncertainties exist regarding levels of exposure to methyl mercury from fish consumption and potential health effects resulting from this exposure. Past estimates of exposure to methyl mercury have been obtained from results of food consumption surveys and measures of methyl mercury in fish. Measures of a biomarker of exposure are needed for improved exposure assessments. Blood mercury levels will be assessed in two subpopulations particularly vulnerable to the health effects from mercury exposure: children 1-5 years old and women of childbearing age. Blood measures of total and inorganic mercury will be important for evaluation of exposure from exposure to mercury in interior latex paints.

#### Selenium

Selenium salts are toxic in large amounts, but trace amounts are necessary for cellular

function in many organisms, including all animals. Selenium is a component of the antioxidant enzymes glutathione peroxidase and thioredoxin reductase (which indirectly reduce certain oxidized molecules in animals and some plants). It is also found in three deiodinase enzymes, which convert one thyroid hormone to another. Selenium requirements in plants differ by species, with some plants requiring relatively large amounts, and others apparently requiring none.

### Eligible Sample

All examined participants aged 1-11 years old, and a one-half sample from participants aged 12 years and older were eligible.

### **Description of Laboratory Methodology**

This method directly measures lead (Pb), cadmium (Cd), total mercury (Hg), manganese (Mn) and selenium (Se) content of whole blood specimens using mass spectrometry after a simple dilution sample preparation step.

During the sample dilution step, a small volume of whole blood is extracted from a larger whole blood patient specimen after the entire specimen is mixed (vortexed) to create a uniform distribution of cellular components. This mixing step is important because some metals (e.g., Pb) are known to be associated mostly with the red blood cells in the specimen and a uniform distribution of this cellular material must be produced before a small volume extracted from the larger specimen will accurately reflect the average metal concentration of all fractions of the larger specimen. Coagulation is the process in which blood forms solid clots from its cellular components. If steps are not taken to prevent this process from occurring, i.e., addition of anti-coagulant reagents such as EDTA in the blood collection tube prior to blood collection, blood will immediately begin to form clots once leaving the body and entering the tube. These clots prevent the uniform distribution of cellular material in the blood specimen even after rigorous mixing, making a representative sub-sample of the larger specimen unattainable. It is important that prior to or during sample preparation the analyst identify any sample having clots or micro-clots (small clots). Clotted samples are not analyzed by this method due to the inhomogeneity concerns (i.e., all results for the sample are processed as "not reportable").

Dilution of the blood in the sample preparation step prior to analysis is a simple dilution of 1 part sample + 1 part water + 48 parts diluent. The effects of the chemicals in the diluent are to release metals bound to red blood cells making them available for ionization, reduce ionization suppression by the biological matrix, prevent clogging of the sample introduction system pathways by undissolved biological solids, and allow introduction of internal standards to be utilized in the analysis step. Tetramethylammonium hydroxide (TMAH, 0.4% v/v) and Triton X-100TM (0.05%) in the sample diluent solubilizes blood components. Triton X-100TM also helps prevent biological deposits on internal surfaces of the instrument's sample introduction system and reduce collection of air bubbles in sample transport tubing. Ammonium pyrrolidine dithiocarbamate (APDC) in the sample diluent (0.01%) aids in solubilizing metals released from the biological matrix. Ethyl alcohol in the sample diluent (1%) aids solubility of blood components and aids in aerosol generation by reduction of the surface tension of the solution. The internal standards, rhodium, iridium, and tellurium, are at a constant concentration in all blanks, calibrators, QC, and samples. Monitoring the instrument signal ratio of a metal to its internal standard allows correction for instrument noise and drift, and sample-to-sample matrix differences.

Liquid samples are introduced into the mass spectrometer through the inductively coupled plasma (ICP) ionization source. The liquid diluted blood sample is forced through a nebulizer, which converts the bulk liquid into small droplets in an argon aerosol. The smaller droplets from the aerosol are selectively passed through the spray chamber by a flowing argon stream into the ICP. By coupling radio-frequency power into flowing argon, plasma is created in which the predominant species are positive argon ions and electrons and has a temperature of 6000-8000 K. The small aerosol droplets pass through a region of the plasma and the thermal energy vaporizes the liquid droplets, atomizes the molecules of the sample and then ionizes

the atoms. The ions, along with the argon, enter the mass spectrometer through an interface that separates the ICP (at atmospheric pressure, ~760 torr) from the mass spectrometer (operating at a pressure of 10-5 torr). The ions first pass through a focusing region, then the dynamic reaction cell (DRC), the quadrupole mass filter, and finally are selectively counted in rapid sequence at the detector allowing individual isotopes of an element to be determined.

Generally, the DRC operates in one of two modes. In 'vented' (or 'standard') mode the cell is not pressurized and ions pass through the cell to the quadrupole mass filter unaffected. In 'DRC' mode, the cell is pressurized with a gas for the purpose of causing collisions and/or reactions between the fill gas and the incoming ions. In general, collisions or reactions with the incoming ions selectively occur to either eliminate an interfering ion, change the ion of interest to a new mass, which is free from interference, or collisions between ions in the beam and the DRC gas can focus the ion beam to the middle of the cell and increase the ion signal. In this method, the instrument is operated in DRC mode when analyzing for manganese, mercury, and selenium. For selenium, the DRC is pressurized with methane gas (CH4, 99.999%) which reduces the signal from 40Ar2+ while allowing the 80Se+ ions to pass relatively unaffected through the DRC on toward the analytical quadrupole and detector. Manganese and mercury are both measured when the DRC is pressurized with oxygen gas (O2, 99.999%). They are analyzed at the same flow rate of oxygen to the DRC cell to avoid lengthening analysis time due to pause delays that would be necessary if different gas flows were used for the two analytes. The oxygen reduces the ion signal from several interfering ions (37Cl18O+, 40Ar15N+, 38Ar16O1H+, 54Fe1H+) while allowing the Mn+ ion stream to pass relatively unaffected through the DRC on toward the analytical quadrupole and detector. In the case of mercury, collisional focusing of the mercury ions occurs, increasing the observed mercury signal at the detector by approximately a factor of two (2x).

Once ions pass through the DRC cell and electrically selected for passage through the analytical quadrupole, electrical signals resulting from the ions striking the discrete dynode detector are processed into digital information that is used to indicate the intensity of the ions. The intensity of ions detected while aspirating an unknown sample is correlated to an elemental concentration through comparison of the analyte: internal standard signal ratio with that obtained when aspirating calibration standards. This method was originally based on the method by Lutz (Lutz et al., 1991). The DRC portions of the method are based on work published by Tanner (Tanner, et al. 1999; 2002).

Refer to the Laboratory Method Files section for a detailed description of the laboratory methods used.

There were no changes to lab methods, lab equipment or lab site for this component for the 2015-2016 cycle.

## **Laboratory Method Files**

Cadmium, Lead, Manganese, Mercury, and Selenium Lab Procedure Manual (January 2018)

### Laboratory Quality Assurance and Monitoring

Whole blood samples are processed, stored, and shipped to the National Center for Environmental Health, and Centers for Disease Control and Prevention, Atlanta, GA for analysis.

Detailed instructions on specimen collection and processing are discussed in the NHANES Laboratory Procedures Manual (LPM). Vials are stored under appropriate frozen (–30°C) conditions until they are shipped to National Center for Environmental Health for testing.

The NHANES quality assurance and quality control (QA/QC) protocols meet the 1988 Clinical Laboratory Improvement Act mandates. Detailed QA/QC instructions are discussed in the NHANES LPM.

Mobile Examination Centers (MECs)

Laboratory team performance is monitored using several techniques. NCHS and contract consultants use a structured competency assessment evaluation during visits to evaluate both the quality of the laboratory work and the quality-control procedures. Each laboratory staff member is observed for equipment operation, specimen collection and preparation; testing procedures and constructive feedback are given to each staff member. Formal retraining sessions are conducted annually to ensure that required skill levels were maintained.

#### **Analytical Laboratories**

NHANES uses several methods to monitor the quality of the analyses performed by the contract laboratories. In the MEC, these methods include performing blind split samples collected on "dry run" sessions. In addition, contract laboratories randomly perform repeat testing on 2% of all specimens.

NCHS developed and distributed a quality control protocol for all the contract laboratories, which outlined the use of Westgard rules (Westgard et al., 1981) when running NHANES specimens. Progress reports containing any problems encountered during shipping or receipt of specimens, summary statistics for each control pool, QC graphs, instrument calibration, reagents, and any special considerations are submitted to NCHS quarterly. The reports are reviewed for trends or shifts in the data. The laboratories are required to explain any identified areas of concern.

All QC procedures recommended by the manufacturers were followed. Reported results for all assays meet the Division of Environmental Health Laboratory Sciences quality control and quality assurance performance criteria for accuracy and precision, similar to the Westgard rules (Caudill, et al., 2008).

#### Data Processing and Editing

The data were reviewed. Incomplete data or improbable values were sent to the performing laboratory for confirmation.

Five additional variables were created for this data file. The formula for their creation is as follows:

- The cadmium in µg/L was converted to nmol/L by multiplying by 8.897.
- The lead in µg/dL was converted to µmol/L by multiplying by 0.0483.
- The manganese in µg/L was converted to nmol/L by multiplying by 18.202.
- The selenium in  $\mu$ g/L was converted to  $\mu$ mol/L by multiplying by 0.0127.
- The mercury in µg/L was converted to nmol/L by multiplying by 4.99.

### **Analytic Notes**

Refer to the 2015-2016 Laboratory Data Overview for general information on NHANES laboratory data.

#### **Subsample Weights**

The appropriate sample weights are provided in the variable WTSH2YR in this data file for all participants and should be used when analyzing these data.

The analytes included in this dataset were measured for all examined participants aged 1-11 years, and in a one-half subsample of participants 12 years and older. For participants aged 1-11 years their WTSH2YR are equivalent to their MEC exam sample weights. These 1-11 years old participants have completed at least one physical exam component in the MEC; therefore, they all have an exam sample weight larger than "0," regardless of their lab test results. For participants 12 years and older, special sample weights were created for the subsample. These special weights accounted for the additional probability of selection into the subsample, as well as the additional nonresponse to these lab tests. Therefore, if a participant 12 years and older was selected as part of the one-half subsample, but did not provide a blood specimen, he/she would have the sample weight value assigned as "0" in his/her record.

#### **Demographic and Other Related Variables**

The analysis of NHANES laboratory data must be conducted using the appropriate survey design and demographic variables. The NHANES 2015-2016 Demographics File contains demographic data, health indicators, and other related information collected during household interviews as well as the sample design variables. The recommended procedure for variance estimation requires use of stratum and PSU variables (SDMVSTRA and SDMVPSU, respectively) in the demographic data file.

The Fasting Questionnaire File includes auxiliary information, such as fasting status, length of fast and the time of venipuncture.

This laboratory data file can be linked to the other NHANES data files using the unique survey participant identifier (i.e., SEQN).

#### **Detection Limits**

The detection limits were constant for all of the analytes in the data set. Two variables are provided for each of these analytes. The variable name ending in "LC" (ex., LBDBCDLC) indicates whether the result was below the limit of detection: the value "0" means that the result was at or above the limit of detection, "1" indicates that the result was below the limit of detection. For analytes with analytic results below the lower limit of detection (ex. LBDBCDLC=1), an imputed fill value was placed in the analyte results field. This value is the lower limit of detection divided by the square root of 2 (LLOD/sqrt[2]). The other variable prefixed URX (ex., LBXBCD) provides the analytic result for the analyte.

The lower limit of detection (LLOD, in  $\mu g/L$ ) for cadmium, manganese, total mercury and selenium, and (LLOD, in  $\mu g/L$ ) for lead:

Variable Name	SAS Label	LLOD
LBXBCD	Cadmium, blood	0.1
LBXBPB	Lead, blood	0.07
LBXMN	Manganese, blood	0.99
LBXTHG	Mercury, total, blood	0.28
LBXBSE	Selenium, blood	24.48

Please refer to the NHANES Analytic Guidelines and the on-line NHANES Tutorial for further details on the use of sample weights and other analytic issues.

#### References

- Caudill, S.P., Schleicher, R.L., Pirkle, J.L. Multi-rule quality control for the age-related eye disease study. Statist. Med. (2008) 27(20):4094-40106.
- Lutz, T.M., P.M.V. Nirel, and B. Schmidt, Whole-blood analysis by ICP-MS. Applications of Plasma Source Mass Spectrometry, ed. G. Holland and A.N. Eaton. 1991, Cambridge: Royal Soc Chemistry. 96-100.
- Tanner, S.D., Baranov, Vladimir I, Theory, Design, and Operation of a Dynamic Reaction Cell for ICP-MS. Atomic Spectroscopy, 1999. 20(2): p. 45-52.
- Tanner, S.D., V.I. Baranov, and D.R. Bandura, Reaction cells and collision cells for ICP-MS: a tutorial review. Spectrochimica Acta Part B-Atomic Spectroscopy, 2002. 57(9): p. 1361-1452.
- Westgard J.O., Barry P.L., Hunt M.R., Groth T. A multi-rule Shewhart chart for quality control in clinical chemistry. Clin Chem. 1981 Mar; 27(3):493-501.

## Codebook and Frequencies

### SEQN - Respondent sequence number

Variable Name: SEQN

SAS Label: Respondent sequence number

**English Text:** Respondent sequence number

Target: Both males and females 1 YEARS - 150 YEARS

## WTSH2YR - Blood metal weights

Variable Name: WTSH2YR

SAS Label: Blood metal weights

**English Text:** Blood metal weights

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to Item
5099.649848 to 499733.23816	Range of Values	5597	5597	
0	Participants 12+ years with no lab specimen	218	5815	
	Missing	0	5815	

## LBXBPB - Blood lead (ug/dL)

Variable Name: LBXBPB

SAS Label: Blood lead (ug/dL)

English Text: Blood lead (ug/dL)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0.05 to 23.51	Range of Values	4988	4988	
	Missing	827	5815	

# LBDBPBSI - Blood lead (umol/L)

Variable Name: LBDBPBSI

SAS Label: Blood lead (umol/L)

English Text: Blood lead (umol/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to Item
0.002 to 1.136	Range of Values	4988	4988	
	Missing	827	5815	

## LBDBPBLC - Blood lead comment code

Variable Name: LBDBPBLC

**SAS Label:** Blood lead comment code

**English Text:** Blood lead comment code

**Target:** Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0	At or above the detection limit	4983	4983	
1	Below lower detection limit	5	4988	
	Missing	827	58/15	

# LBXBCD - Blood cadmium (ug/L)

Variable Name: LBXBCD

SAS Label: Blood cadmium (ug/L)

English Text: Blood cadmium (ug/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0.07 to 9.17	Range of Values	4988	4988	
	Missing	827	5815	

## LBDBCDSI - Blood cadmium (umol/L)

Variable Name: LBDBCDSI

SAS Label: Blood cadmium (umol/L)

English Text: Blood cadmium (umol/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0.62 to 81.59	Range of Values	4988	4988	
	Missing	827	5815	

### LBDBCDLC - Blood cadmium comment code

Variable Name: LBDBCDLC

SAS Label: Blood cadmium comment code

English Text: Blood cadmium comment ode

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Descripti	ion	Count	Cumulative	Skip to I tem
0	At or above the detection	n/limit	3711	3711	
1	Below lower detection	mit	1277	4988	
	Missing		827	5815	

## LBXTHG - Blood mercury, total (ug/L)

Variable Name: LBXTHG

SAS Label: Blood mercury, total (ug/L)

English Text: Blood mercury, total (ug/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0.2 to 36.26	Range of Values	4988	4988	
	Missing	827	5815	

## LBDTHGSI - Blood mercury, total (umol/L)

Variable Name: LBDTHGSI

SAS Label: Blood mercury, total (umol/L)

English Text: Blood mercury, total (umol/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
1 to 180.9	Range of Values	4988	4988	
	Missing	827	5815	

## LBDTHGLC - Blood mercury, total comment code

Variable Name: LBDTHGLC

**SAS Label:** Blood mercury, total comment code

**English Text:** Blood mercury, total comment code

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0	At or above the detection limit	3718	3718	
1	Below lower detection limit	1270	4988	
	Missing	827	5815	

## LBXBSE - Blood selenium (ug/L)

Variable Name: LBXBSE

SAS Label: Blood selenium (ug/L)

**English Text:** Blood selenium(ug/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
59.35 to 419.15	Range of Values	4987	4987	
	Missing	828	5815	

## LBDBSESI - Blood selenium (umol/L)

Variable Name: LBDBSESI

SAS Label: Blood selenium (umol/L)

English Text: Blood selenium (umol/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0.75 to 5.32	Range of Values	4987	4987	
	Missing	828	5815	

### LBDBSELC - Blood selenium comment code

Variable Name: LBDBSELC

SAS Label: Blood selenium comment code

**English Text:** Blood selenium comment code

**Target:** Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0	At or above the detection limit	4987	4987	
1	Below lower detection limit	0	4987	
	Missing	828	5815	

# LBXBMN - Blood manganese (ug/L)

Variable Name: LBXBMN

SAS Label: Blood manganese (ug/L)

English Text: Blood manganese (ug/L)

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
2.21 to 100.41	Range of Values	4987	4987	
	Missing	828	5815	

## LBDBMNSI - Blood manganese (umol/L)

Variable Name: LBDBMNSI

SAS Label: Blood manganese (umol/L)

**English Text:** Blood manganese (umol/L)

**Target:** Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative /	Skip to Item
40.23 to 1827.66	Range of Values	4987	4987	
	Missing	828	5815	

## LBDBMNLC - Blood manganese comment code

Variable Name: LBDBMNLC

**SAS Label:** Blood manganese comment code

**English Text:** Blood manganese comment code

Target: Both males and females 1 YEARS - 150 YEARS

Code or Value	Value Description	Count	Cumulative	Skip to I tem
0	At or above the detection limit	4987	4987	
1	Below lower detection limit	0	4987	
	Missing	828	5815	