

Assignments for Next Monday:

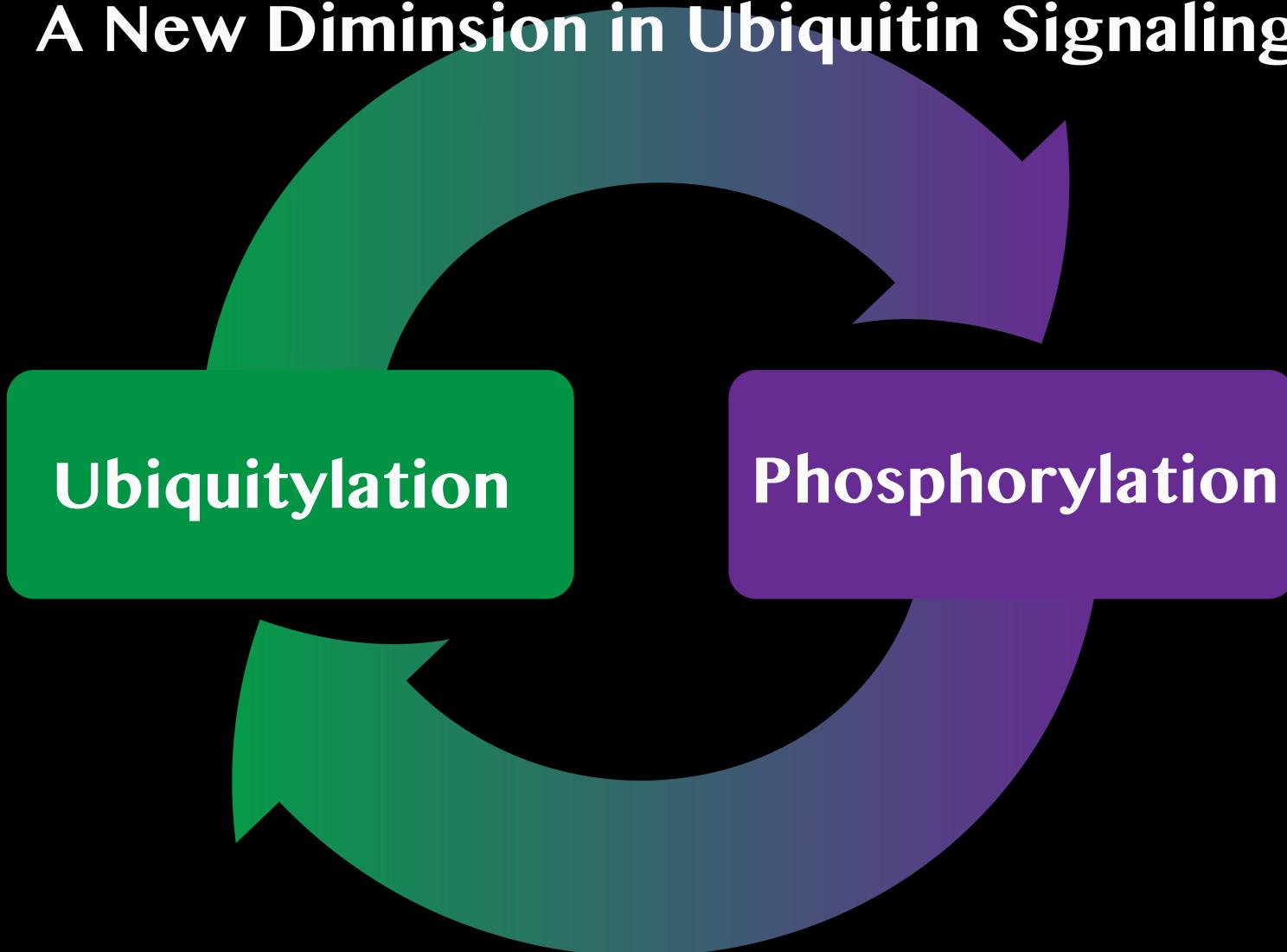


Nathan (Lab work Day 1, 9/28)



Ryan (Journal Club on [Wauer et al, 2015], 9/28)

Phosphorylation of Ubiquitin: A New Dimension in Ubiquitin Signaling

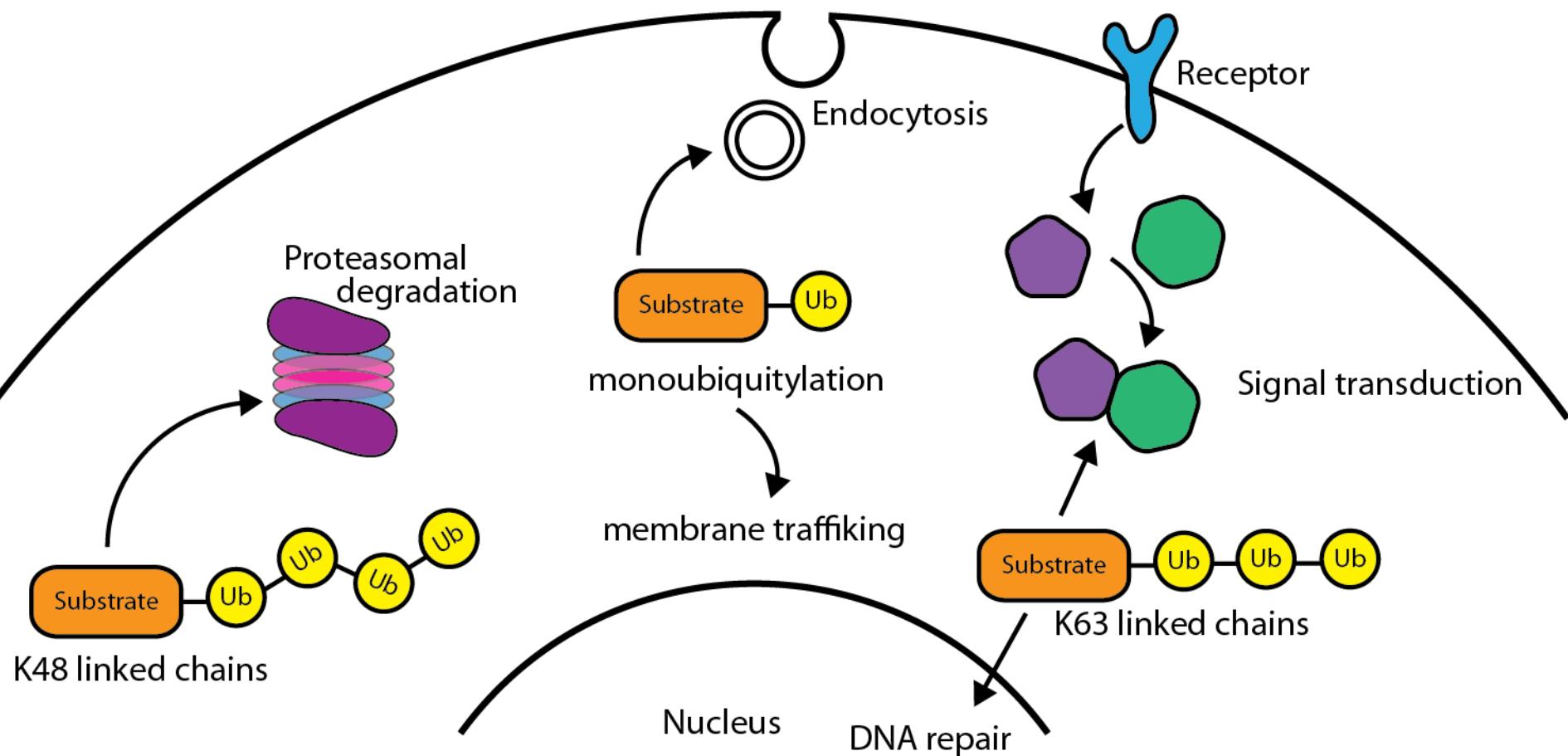


Danielle Swaney
Assistant Adjunct Professor, QB3

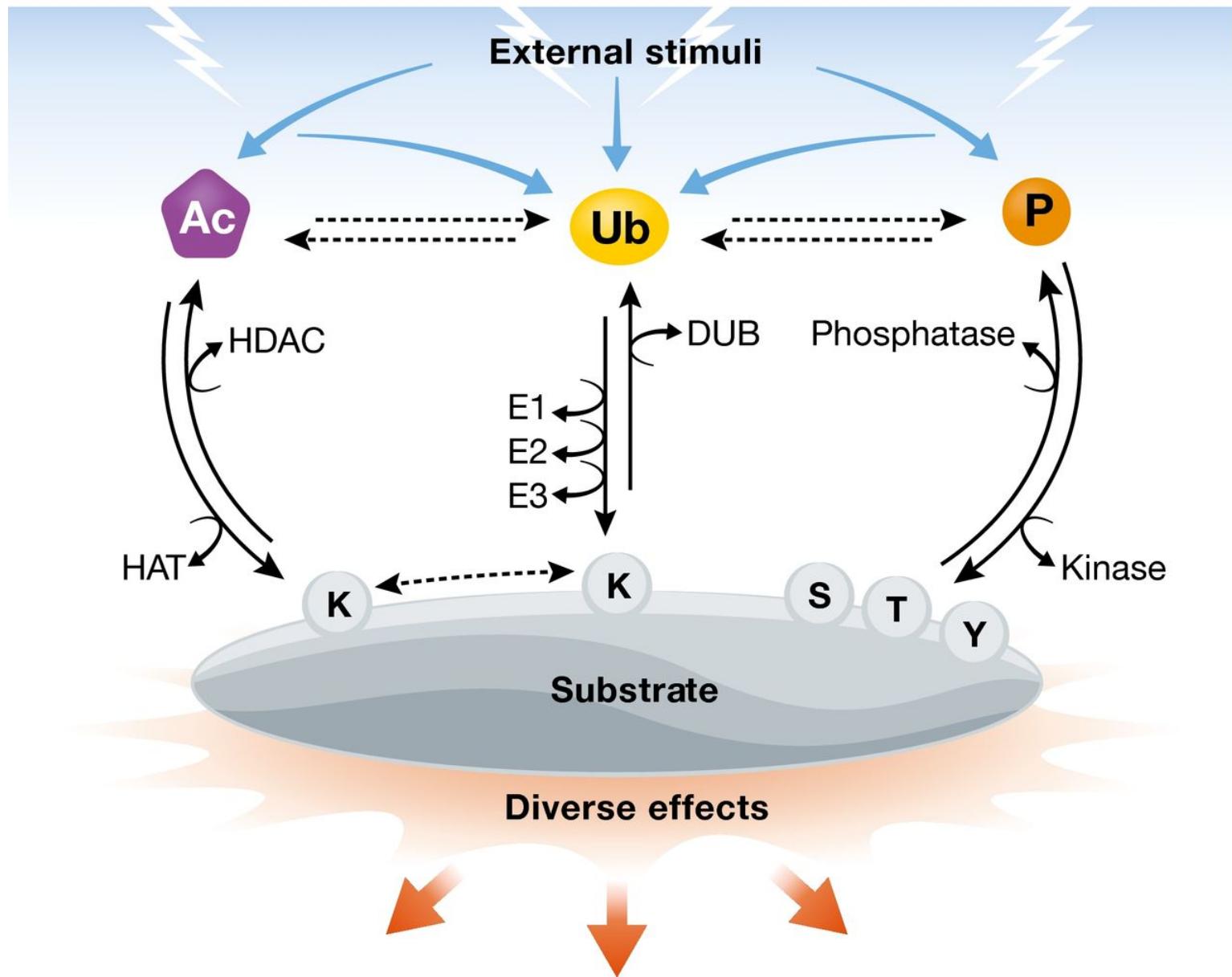
Outline

- Rationale for mass spectrometry project
- Approaches to study kinase-substrate interactions
- Introduction to mass spectrometry

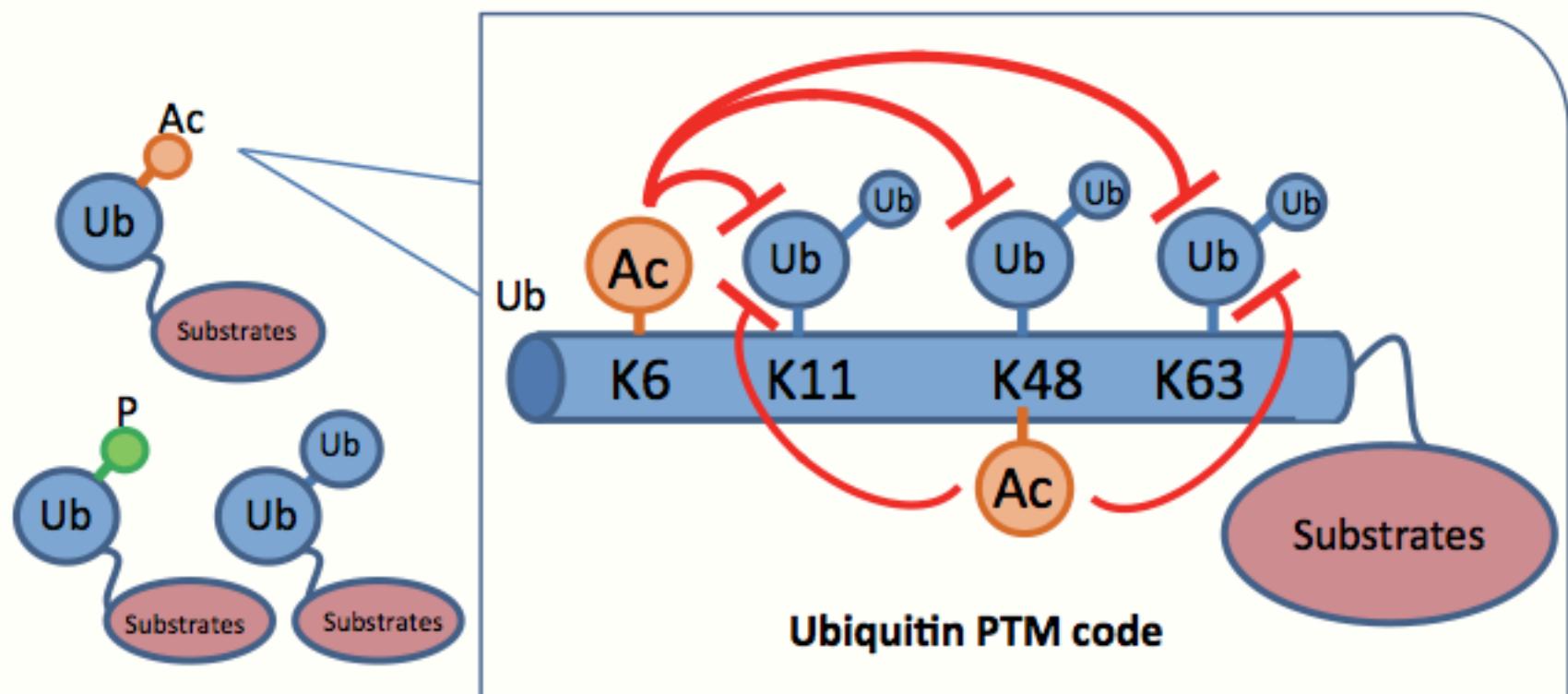
Ubiquitin is a protein post-translational modification with a wide variety of roles



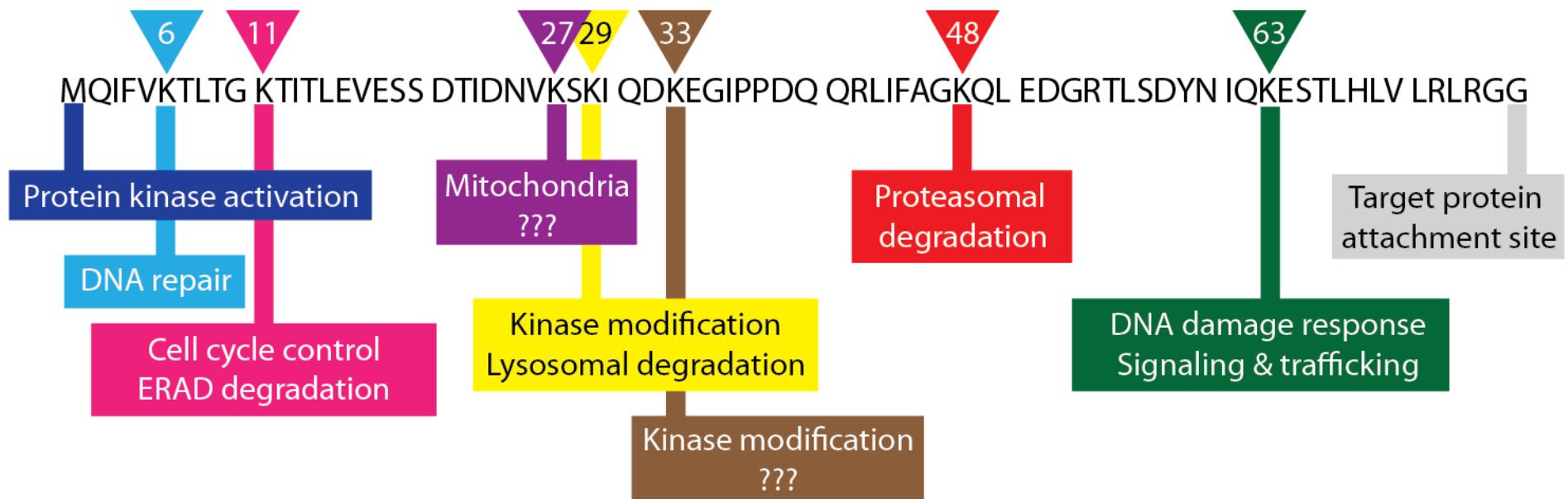
Ubiquitin itself can be modified by other post-translational modifications



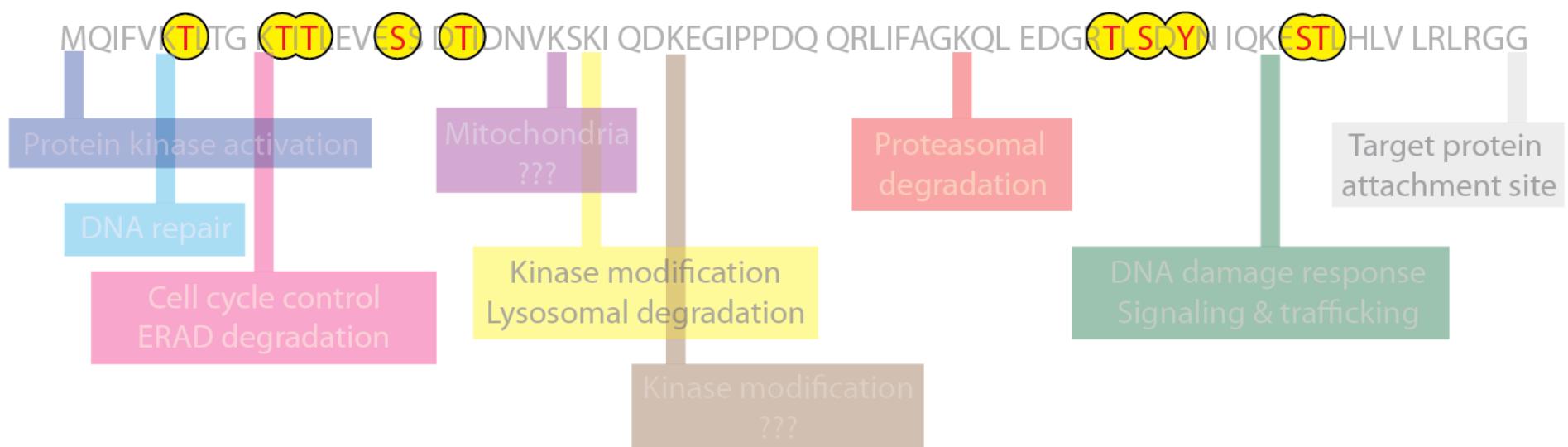
Lysine acetylation of ubiquitin – blocks ubiquitin chain elongation



Functions of ubiquitin

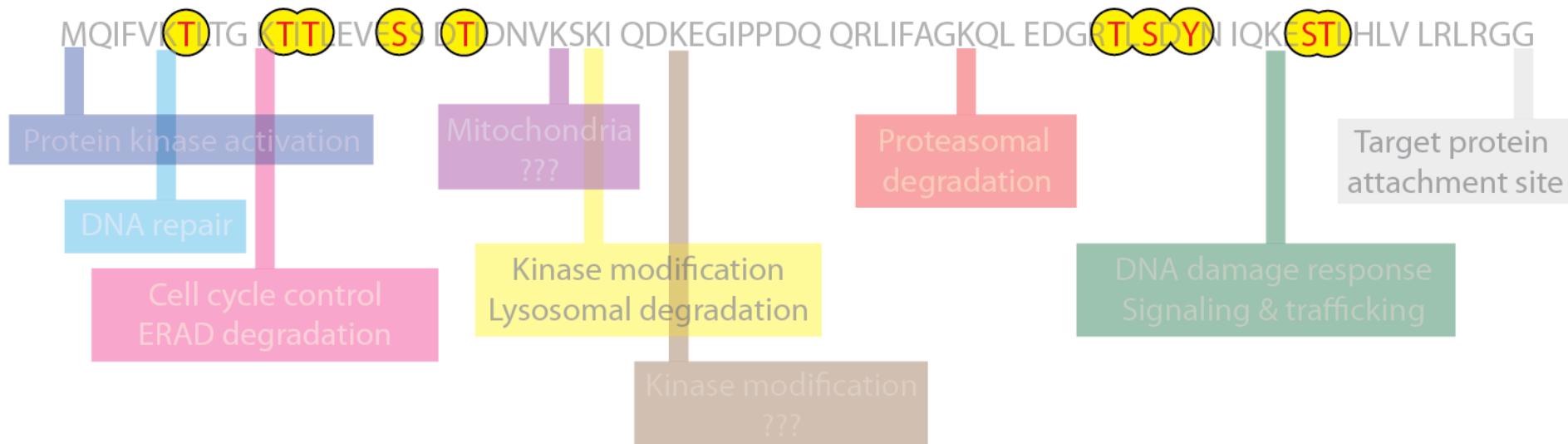


Nearly every S/T/Y on ubiquitin (the most conserved protein) is phosphorylated and conserved from human to yeast



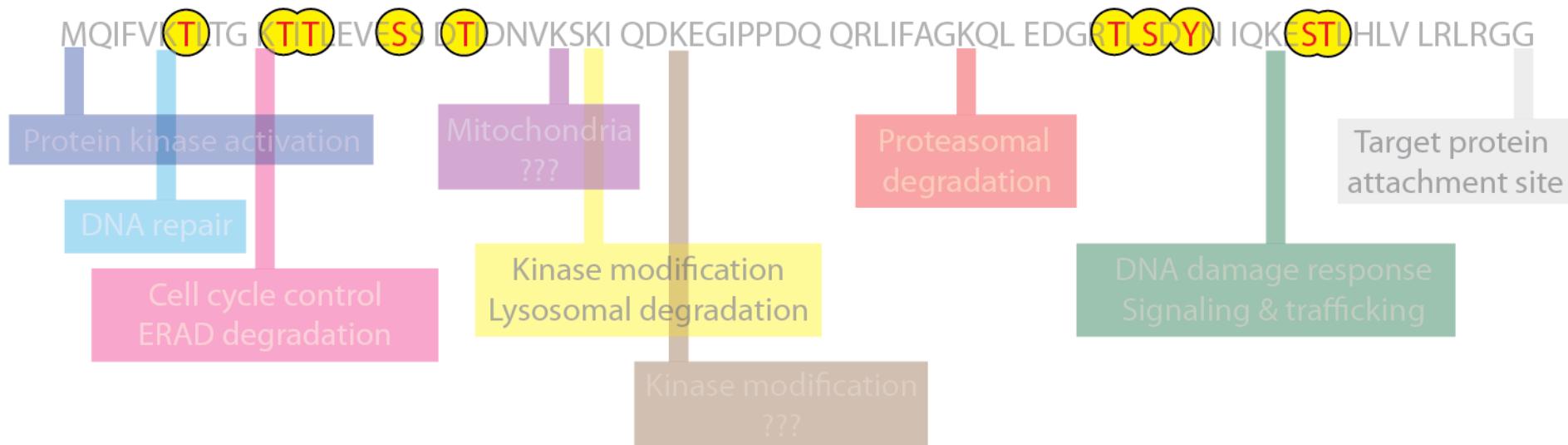
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What kinases phosphorylate ubiquitin?



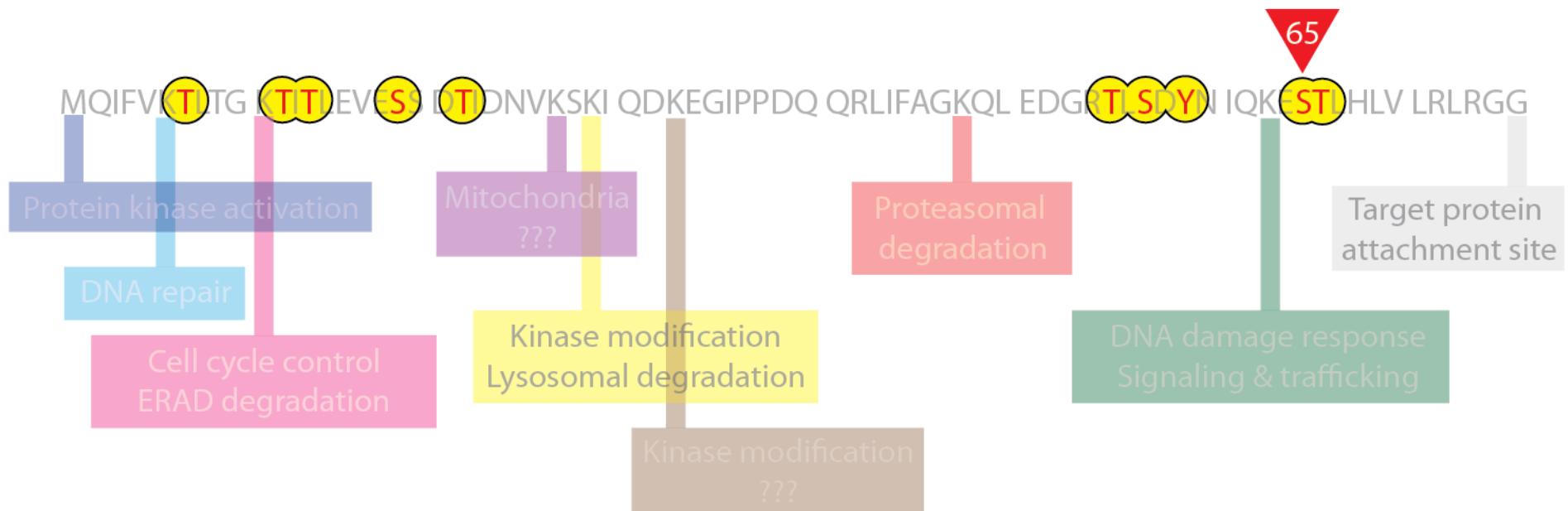
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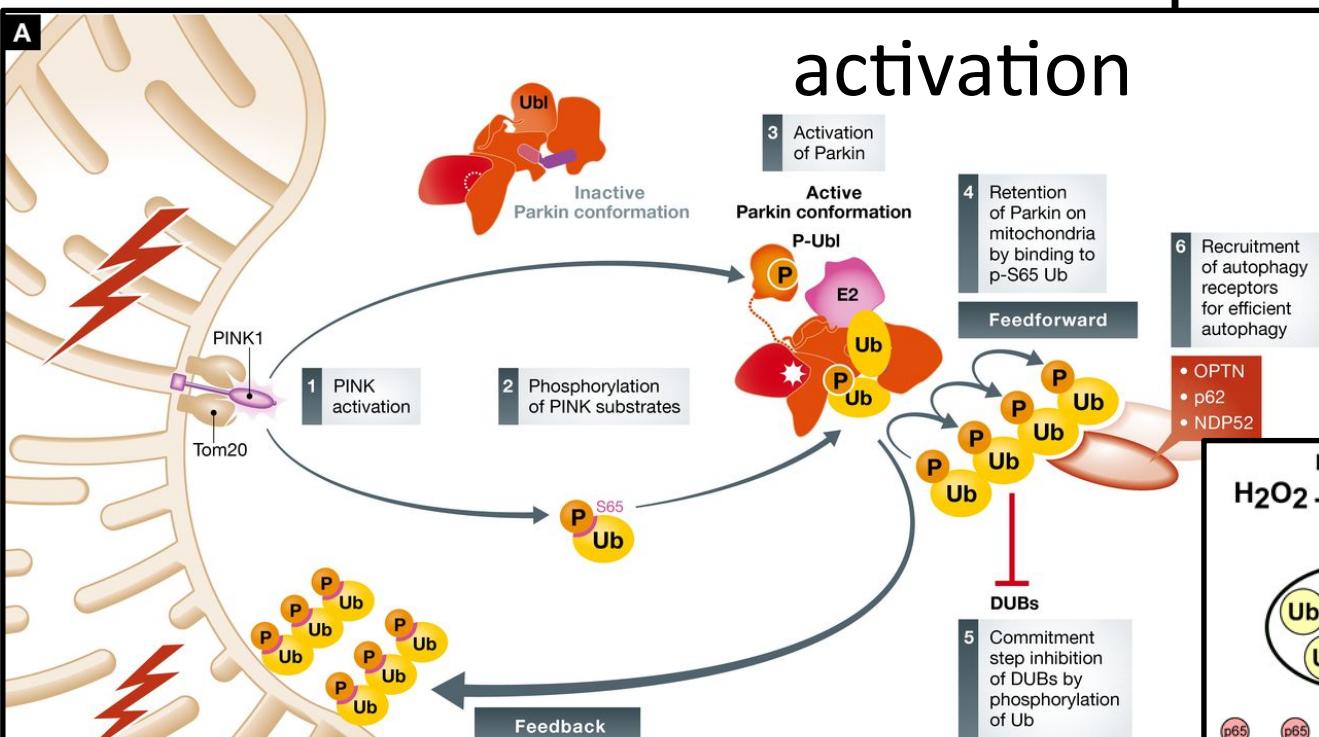


How does phosphorylation regulate ubiquitin function?

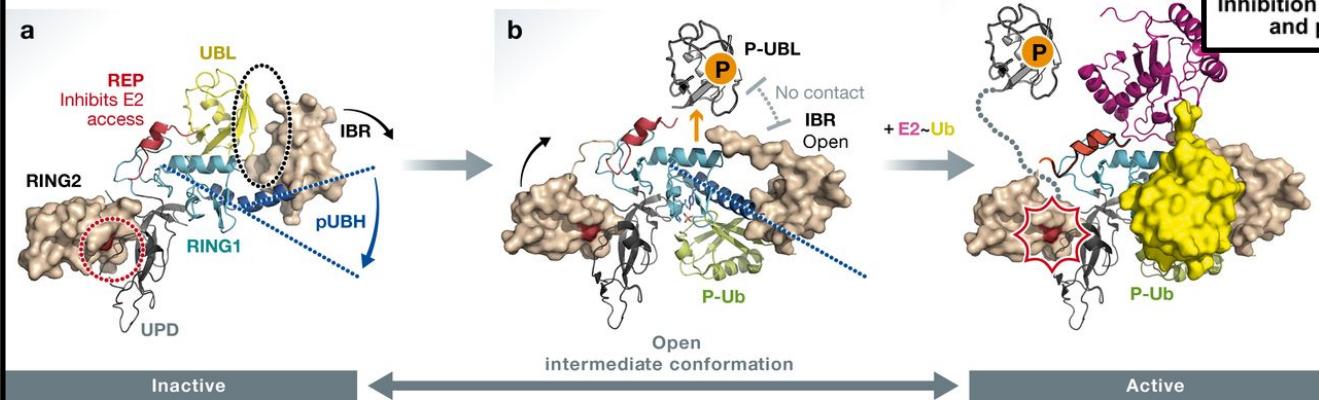
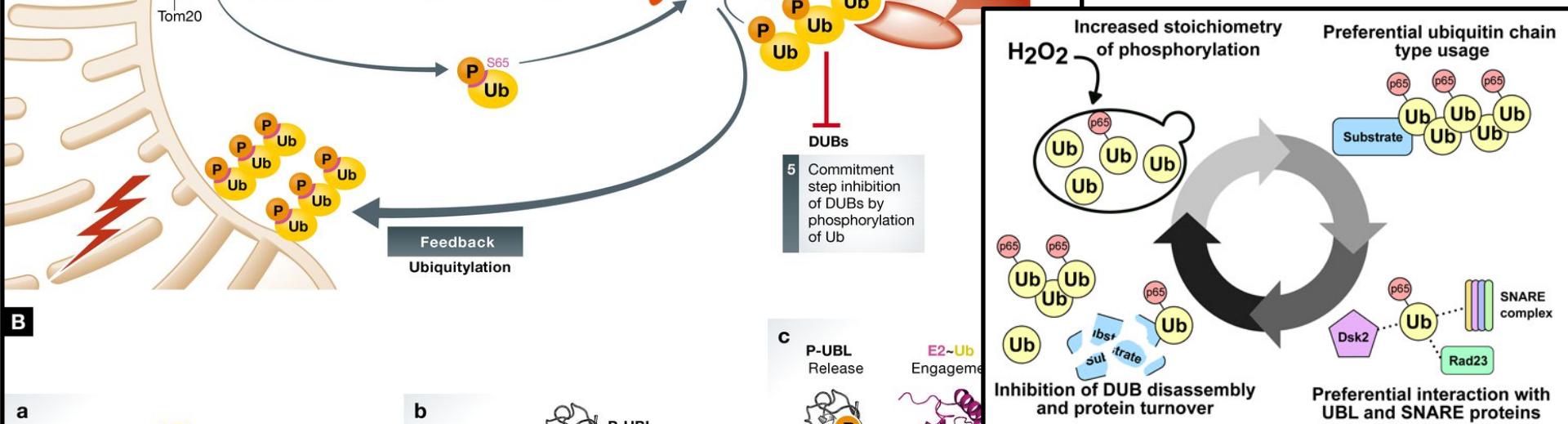
How does phosphorylation regulate ubiquitin function?



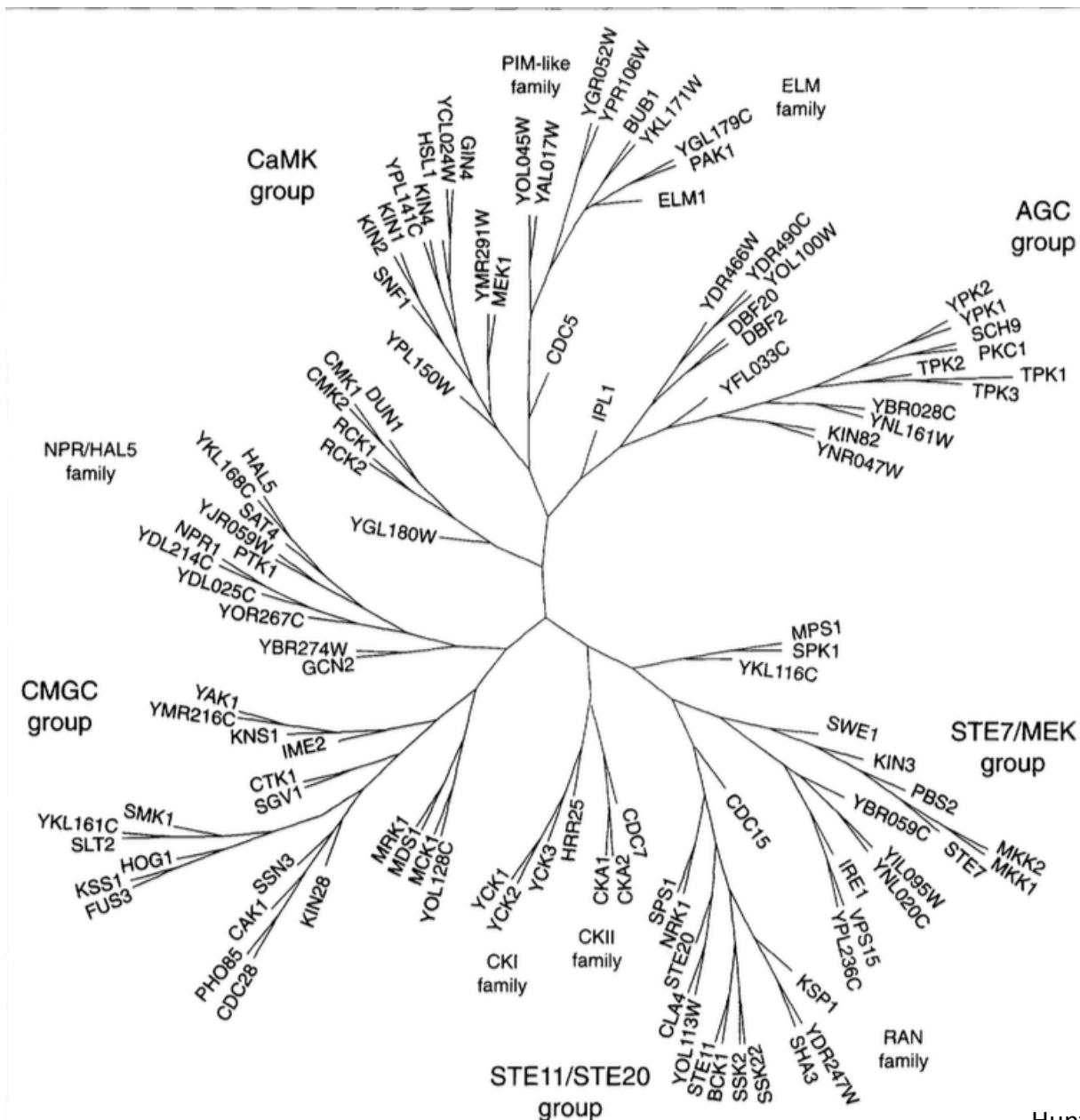
Human: Pink1 kinase → Ub S65p → Parkin E3 ligase activation



Ub S65p also exists in yeast, despite no Pink1 ortholog. What is the kinase?

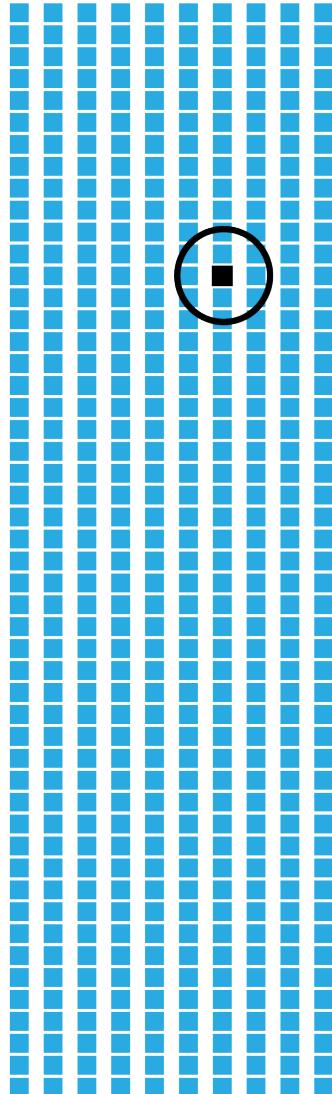


Approaches to study kinase-substrate interactions

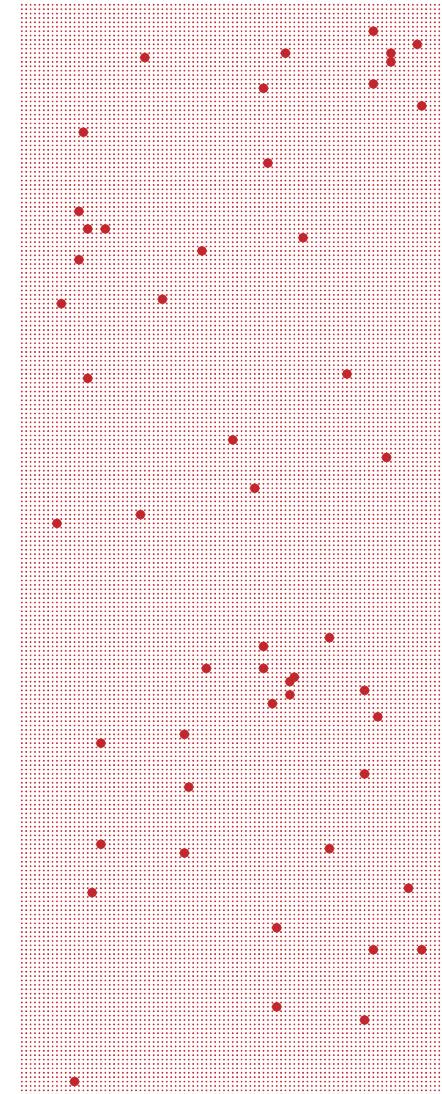


Connecting enzymes and substrates is challenging

> 500 Protein Kinases



> 25,000 Phosphosites



How can we connect these?



Two lines of evidence
are typically required:

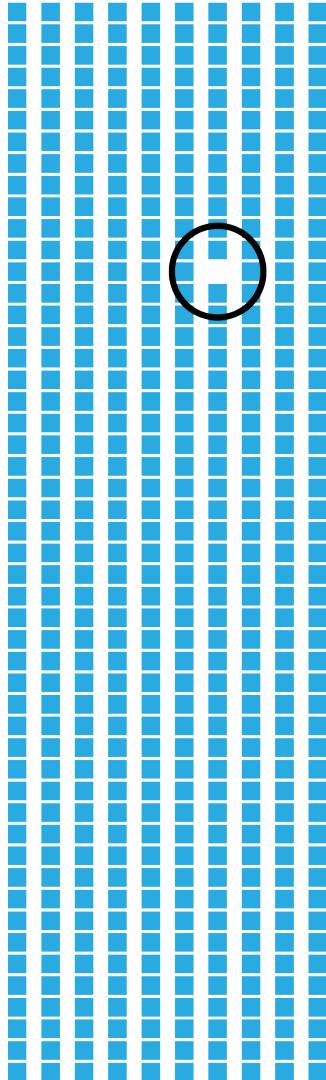
1. *in vivo*

- Overexpression

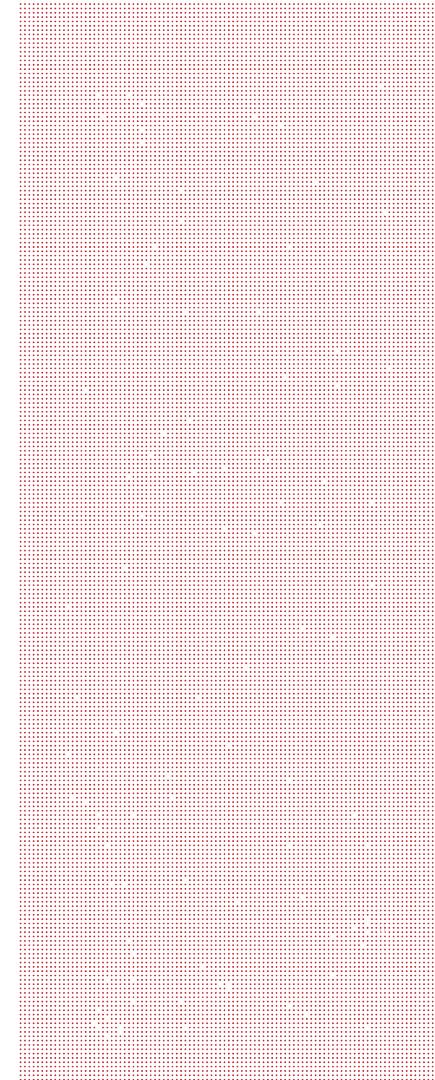


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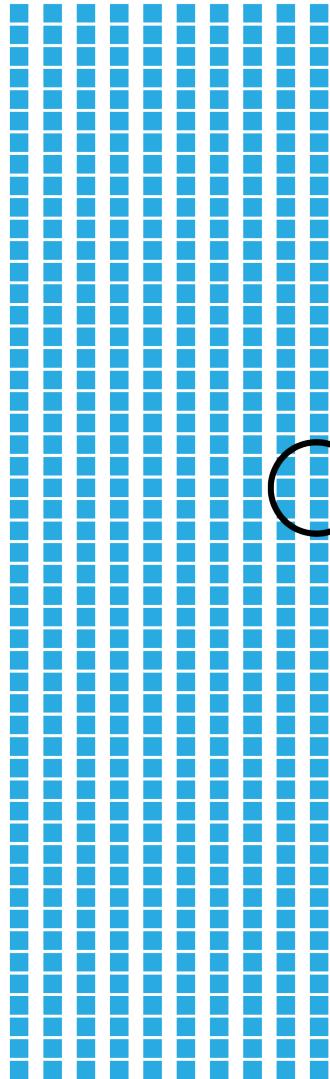
1. *in vivo*

- Overexpression
- Knockdown

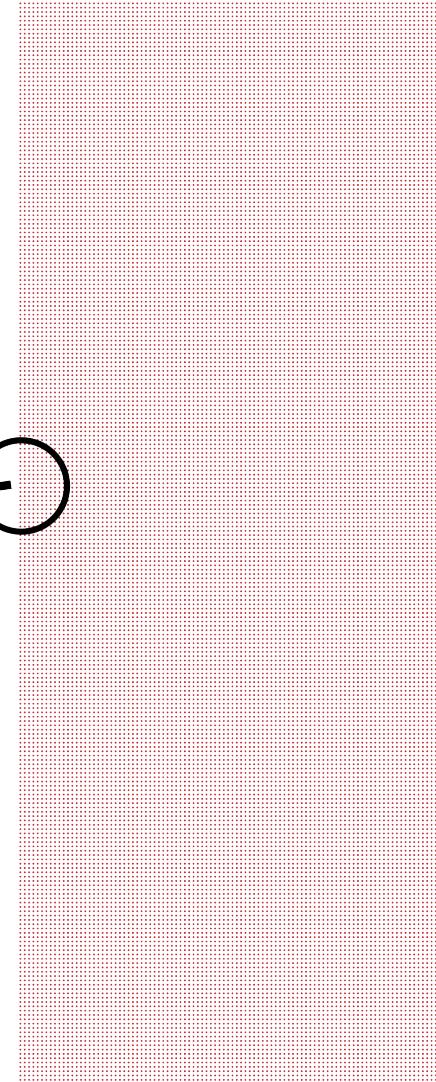


Connecting enzymes and substrates is challenging

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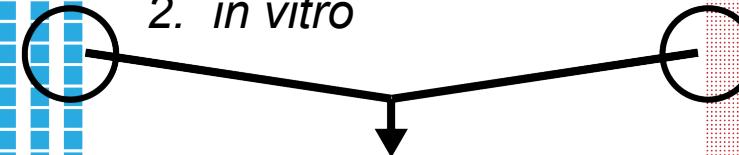


How can we connect these?



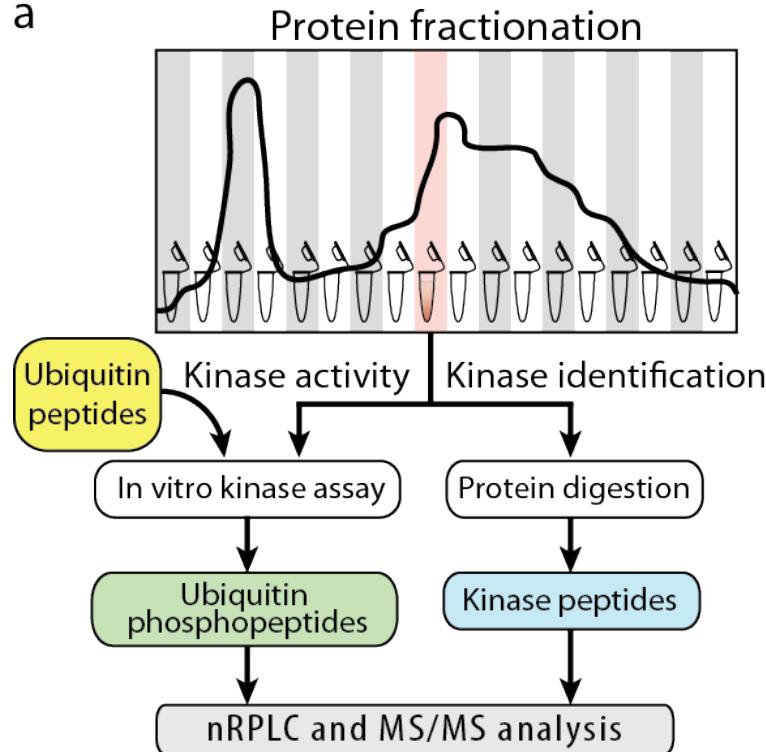
Two lines of evidence
are typically required:

2. *in vitro*

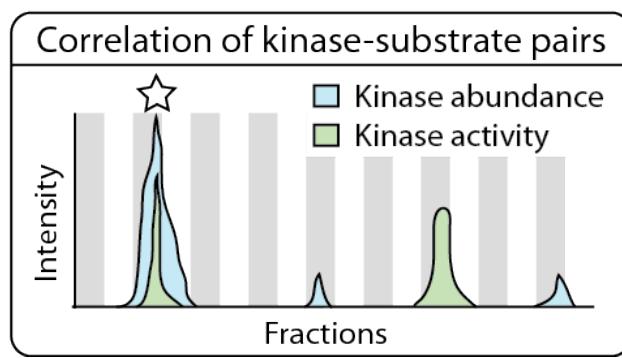


Kinase activity profiling approach

a

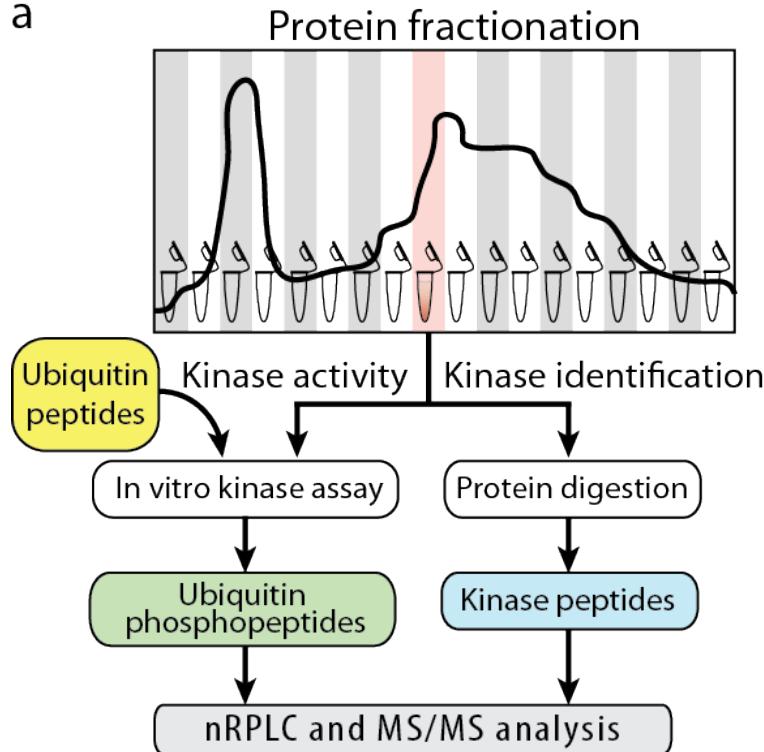


b



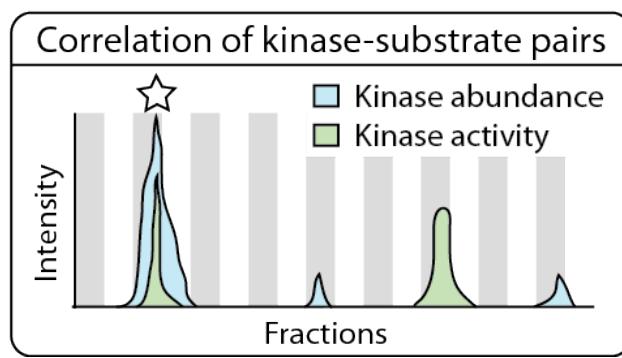
Kinase activity profiling approach

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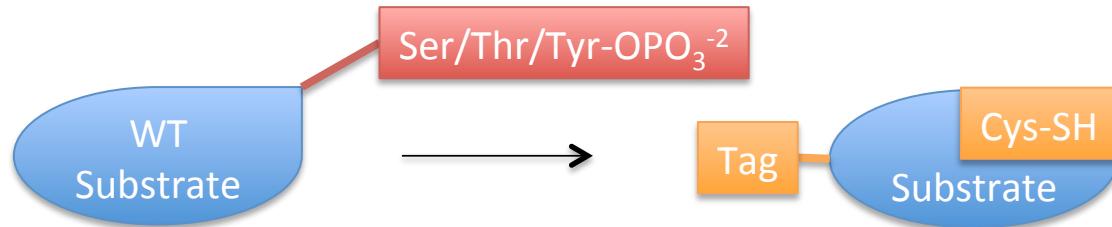


PROS: Unbiased
CONS: labor intensive, high-false positive

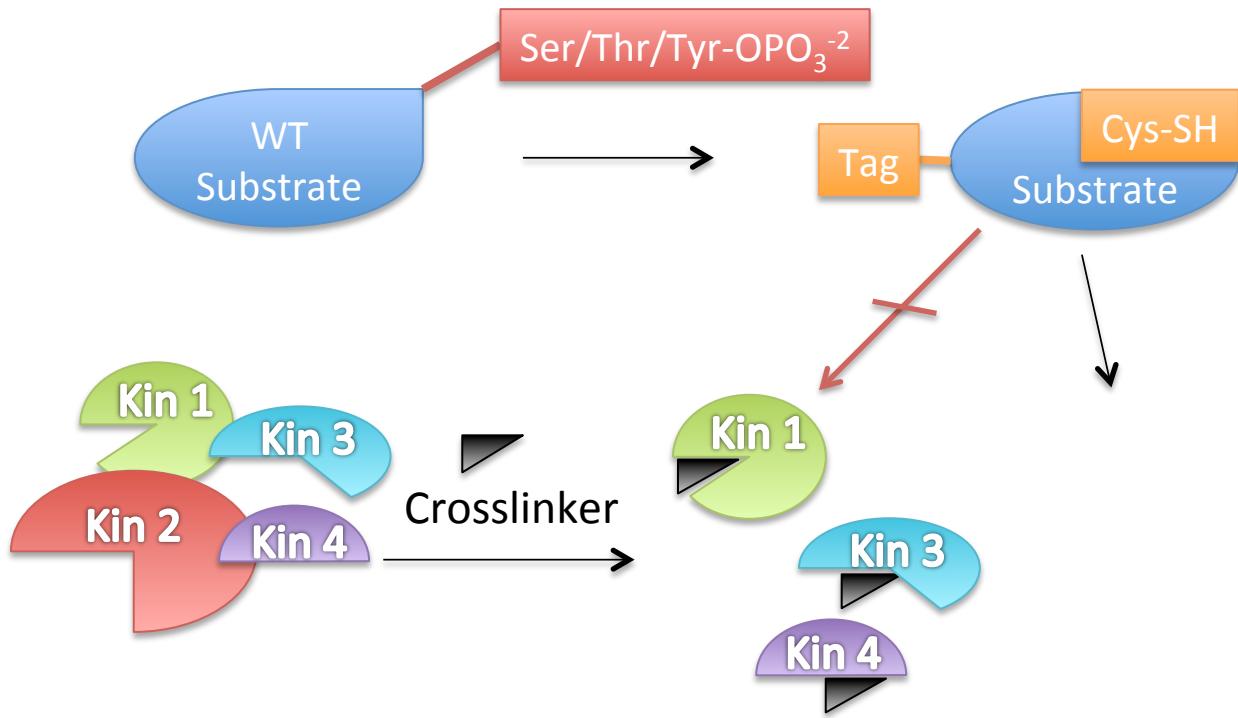
b



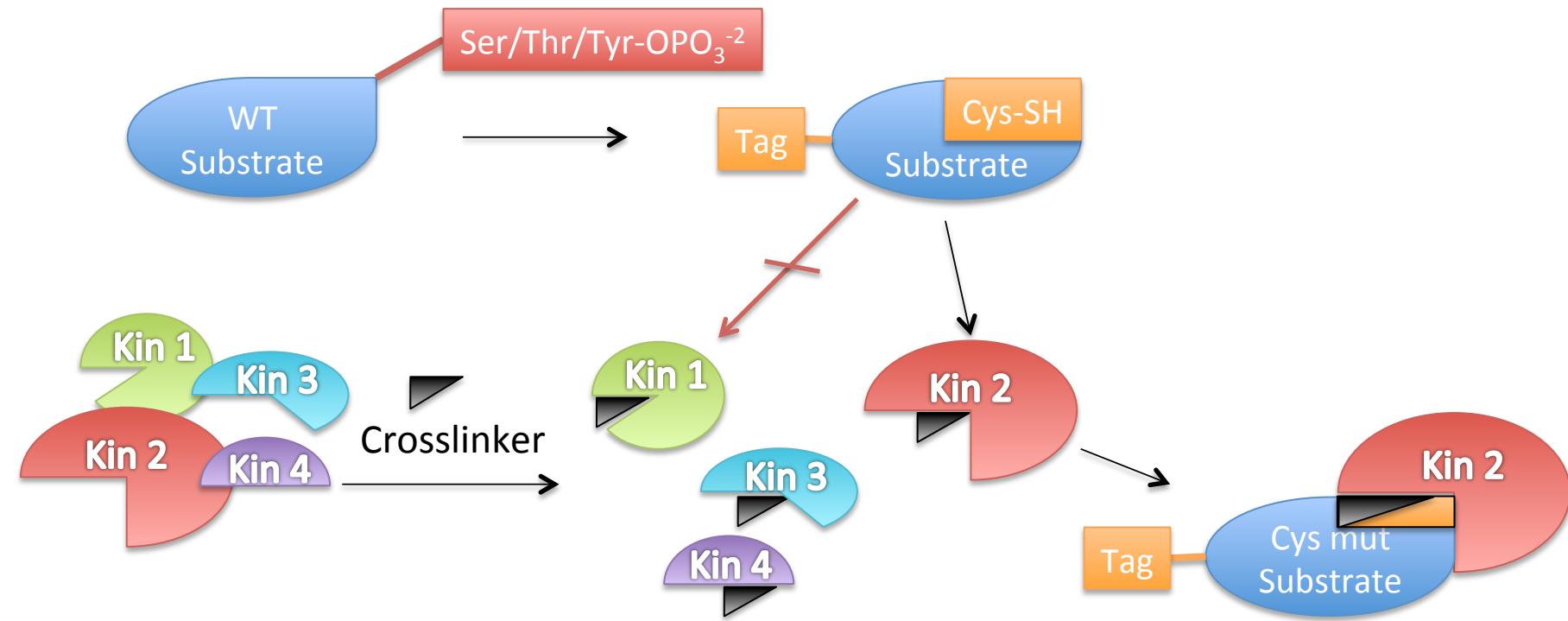
Chemical Biology approach (Shokat method)



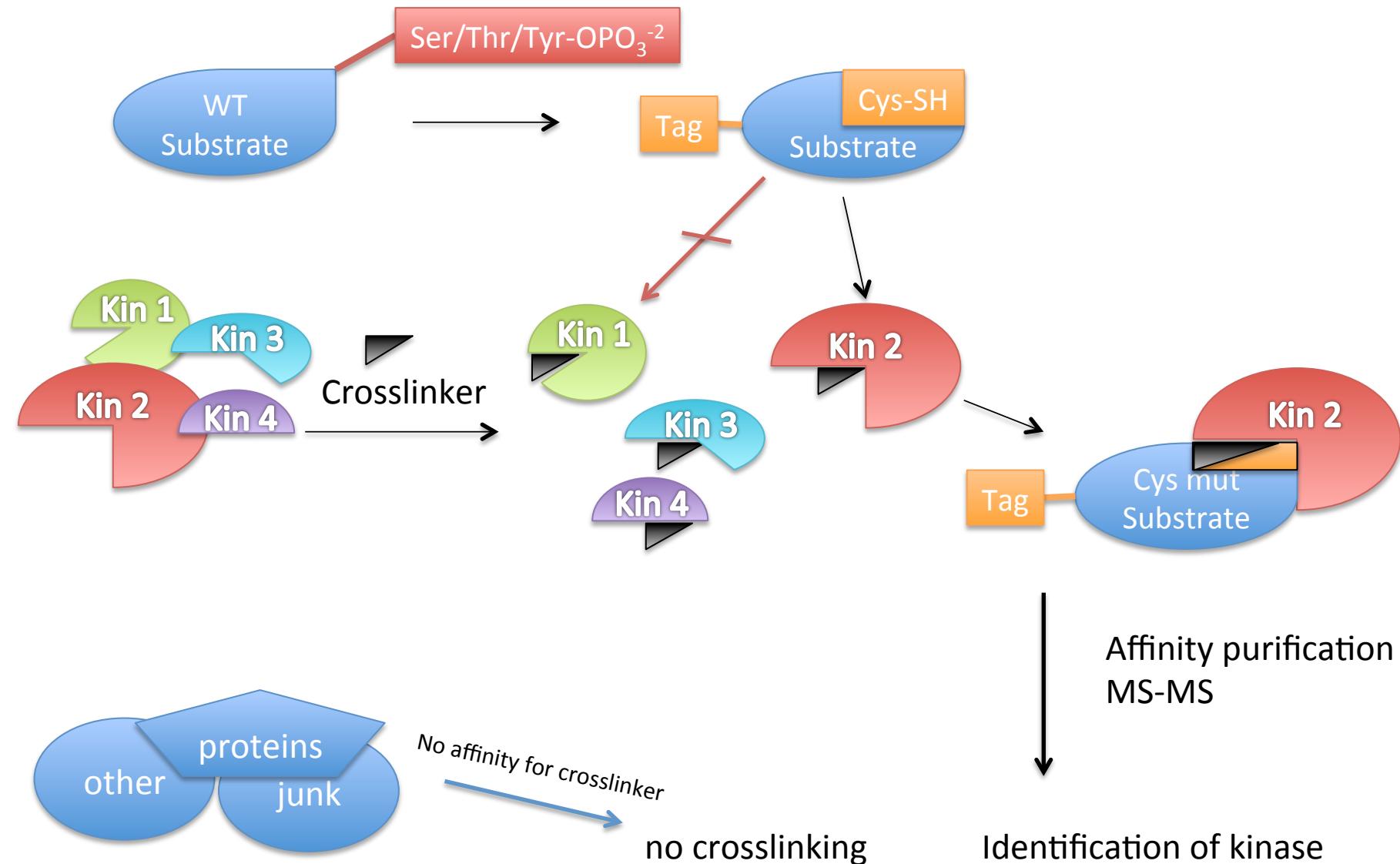
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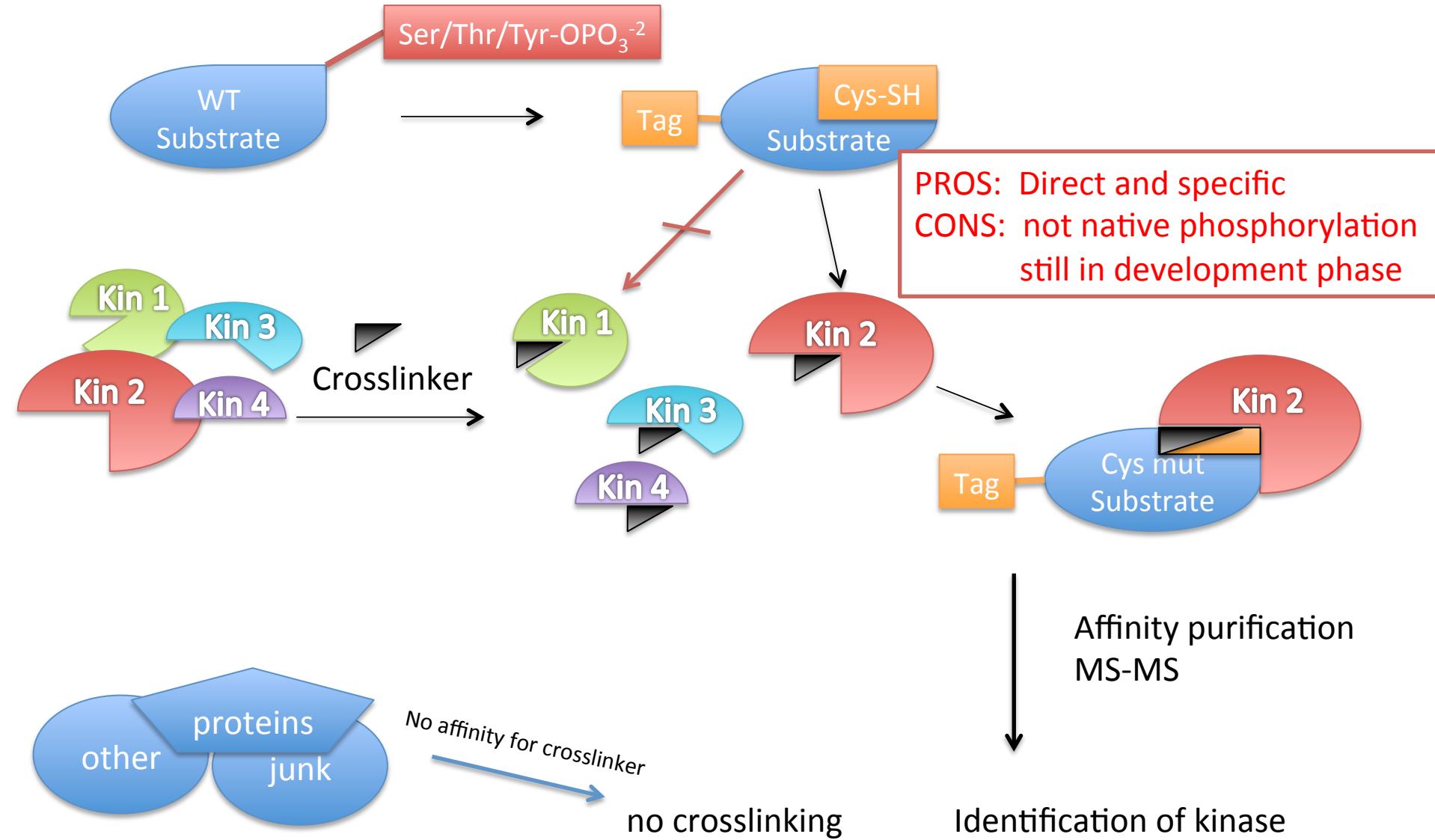
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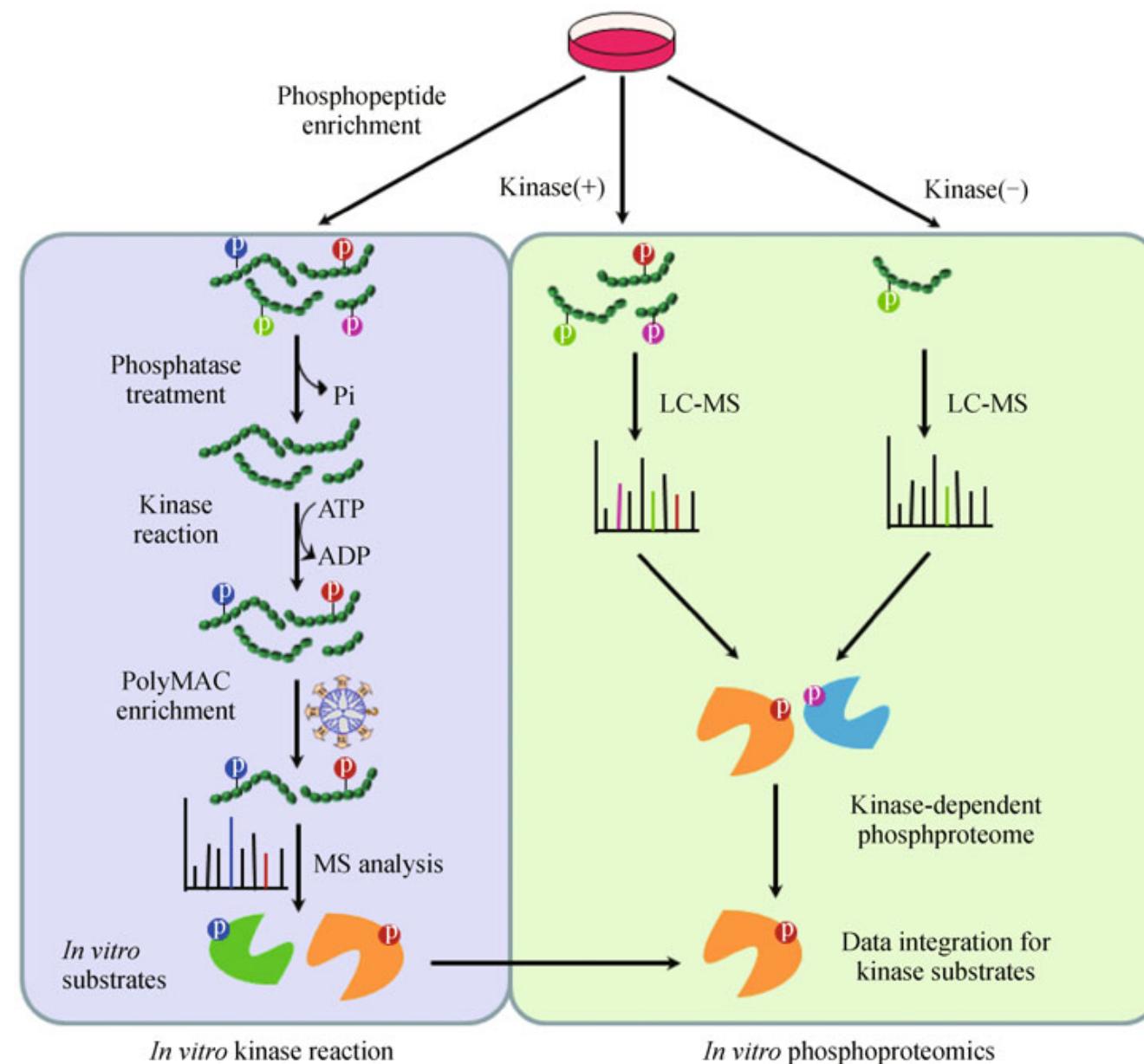
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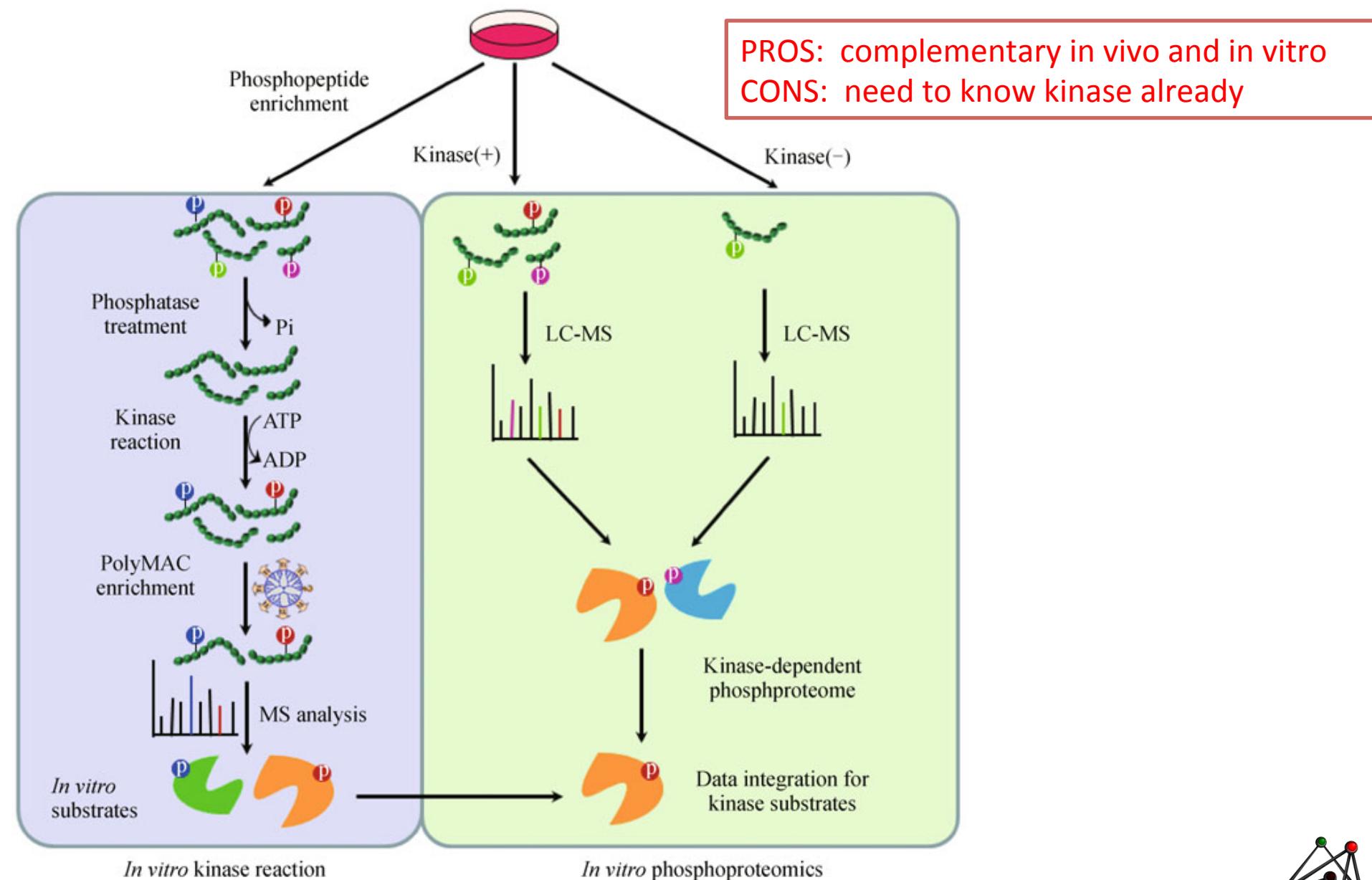
Chemical Biology approach (Shokat method)



Kinase directed approaches



Kinase directed approaches



Selection of kinases for this course: protein array approach

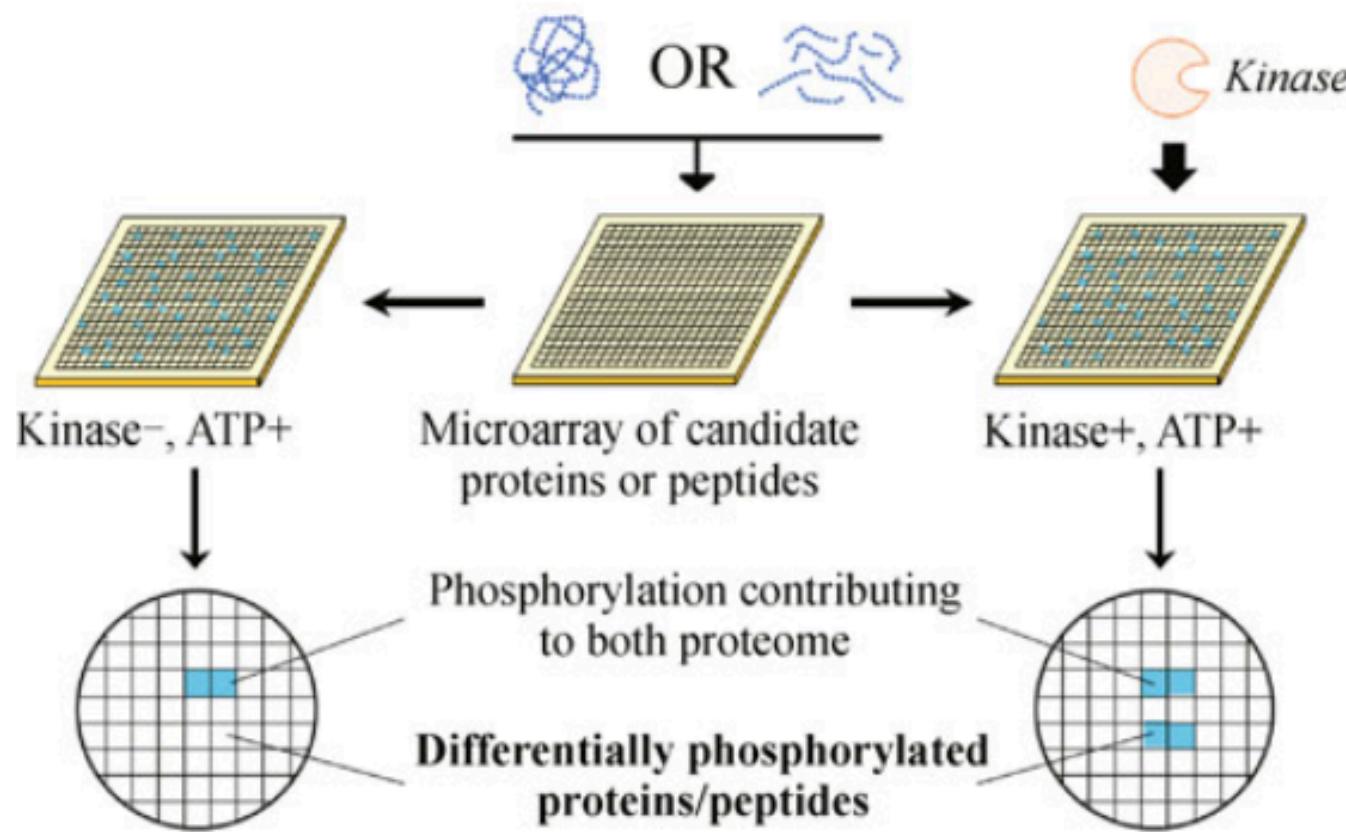


Figure 2 Kinase assay based on protein array or peptide array. Protein/peptide collections are spotted on the microarray, followed by the incubation with a purified active kinase under the reaction condition. Phosphorylation is detected by various methods.

Xue, L. & Tao, W. A. Current technologies to identify protein kinase substrates in high throughput. *Front. Biol.* **8**, 216–227 (2013).

Ptacek, J. et al. Global analysis of protein phosphorylation in yeast. *Nat Cell Biol* **438**, 679–684 (2005).

Newman, R. H. et al. Construction of human activity-based phosphorylation networks. *Molecular Systems Biology* **9**, 655–655 (2013).



Selection of kinases for this course: protein array approach

PROS: High-throughput

CONS: In vitro, prone to high
false positive rates

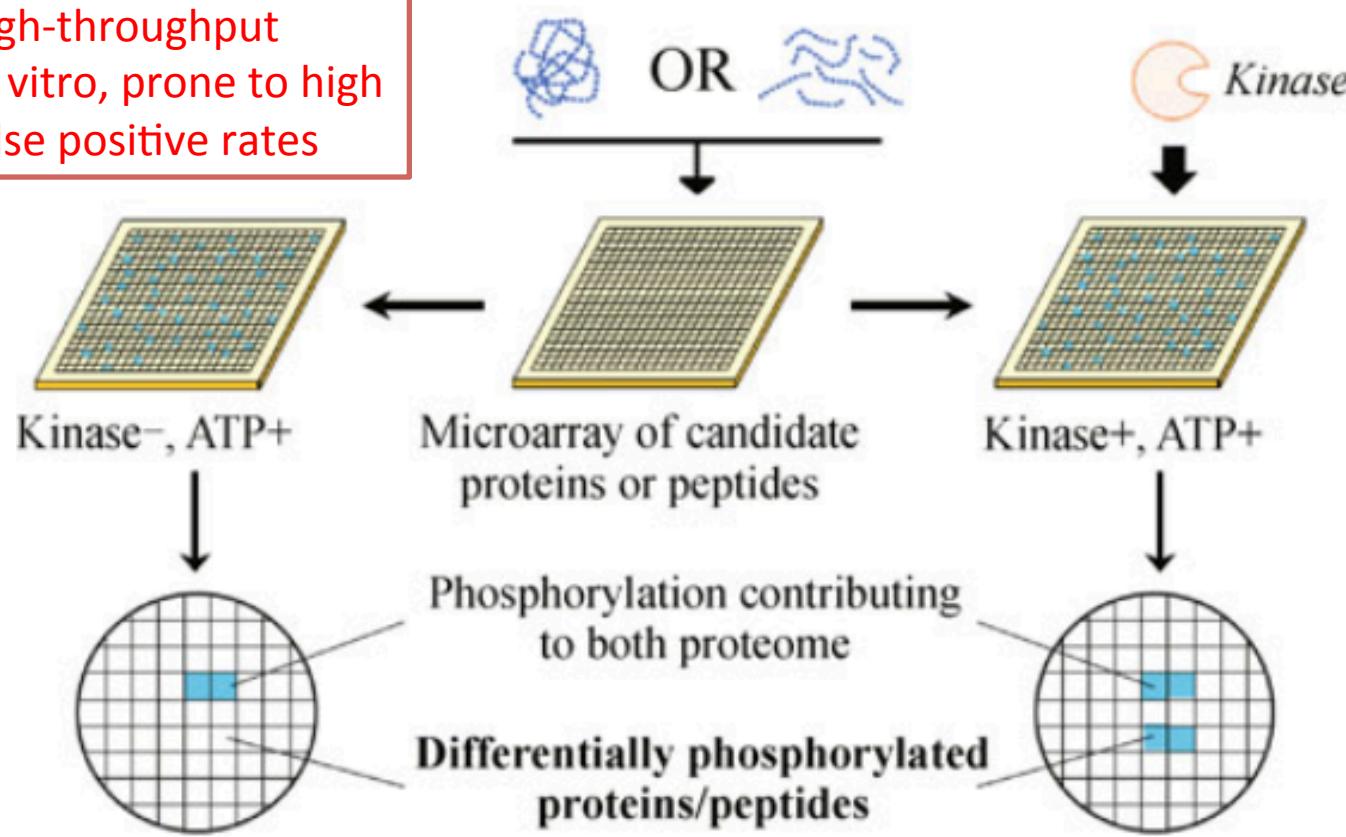


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In short, well-established methods to map kinase-substrate relationship require one of the following:

(1) Prior knowledge of kinase
→ substrate hunting

(2) A labor intensive brute force approach
→ kinase hunting

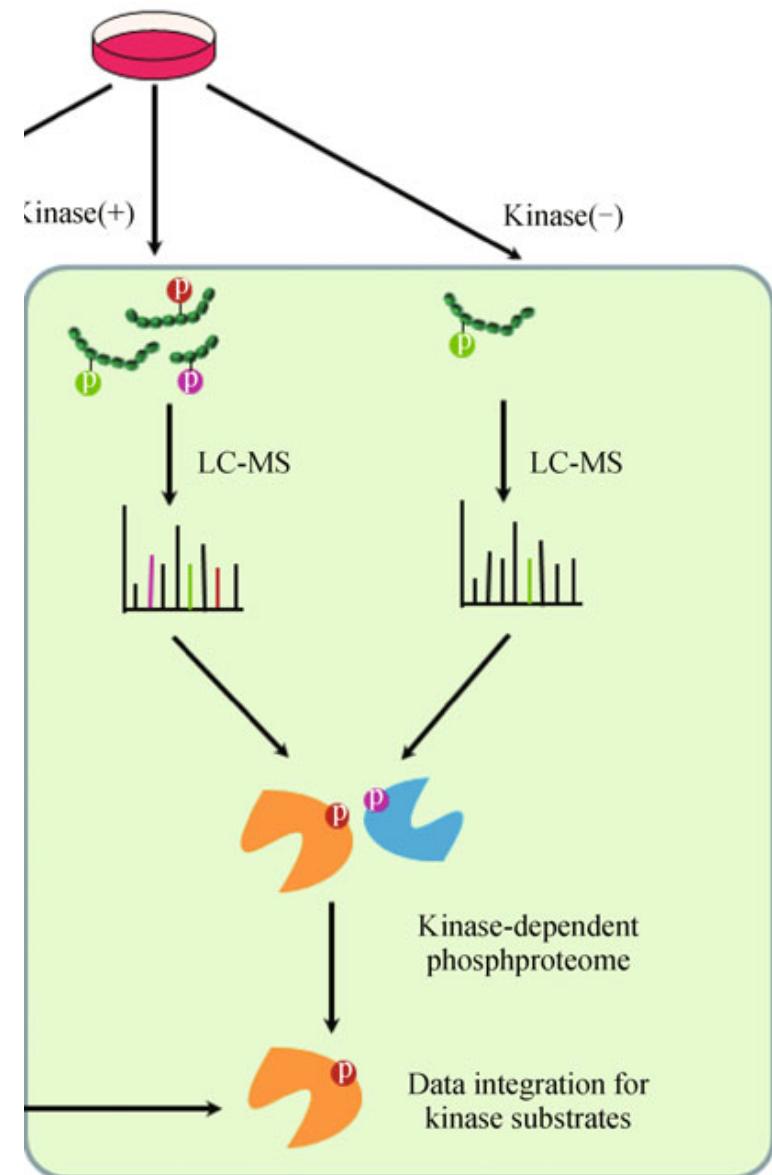
(3) Serendipitous luck

***note other approaches do exist: phage display, yeast 2-hybrid, genetic interaction. But they all suffer from one of the primary CONS listed for methods described here.

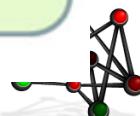


Selection of kinases for this course: protein array results directing a kinase-directed approach

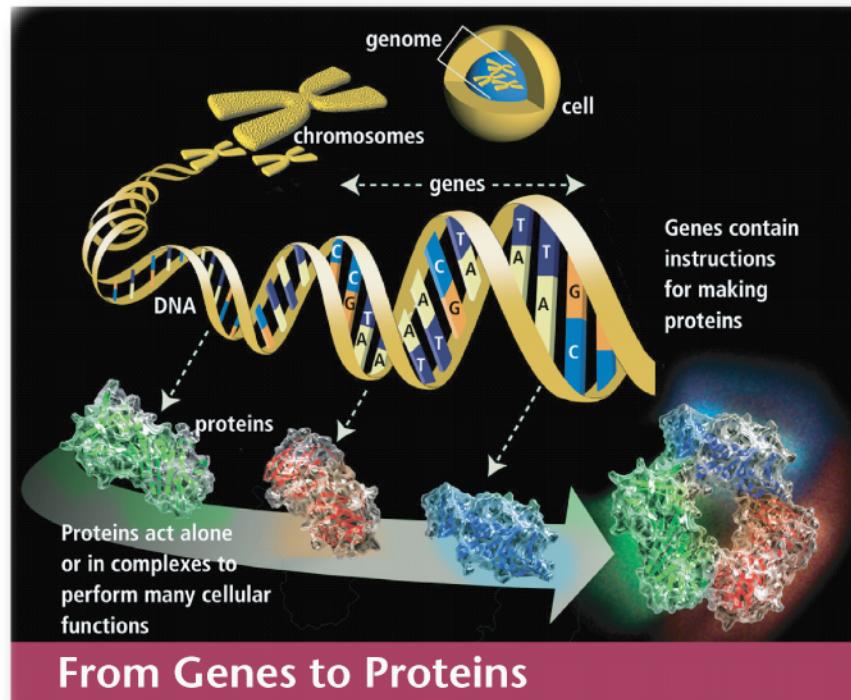
Paper	Human Kinase	Yeast kinase?
Human kinase array	NEK2	KIN3
Human kinase array and YEAST kinase array	WEE1	SWE1
Yeast kinase array		CMK1
Yeast kinase array		TPK1
Yeast kinase array		ALK1
Yeast kinase array (Youle autophagy paper connected Pink1, phosphoubiquitin, and autophagy signaling)		ATG1



In vitro phosphoproteomics



From genes to proteins



www.doegenomes.org

- The human genome codes ~ 25,000 proteins after modification > 500,000
 - the human body consists of 10^{14} cells
 - each cell makes ~ 15,000 different proteins

- what are they?
- where are they?
- how many copies are present?
- what is their function?
- when are they made?
- what proteins do they interact with?
- how are they modified?

Mass spectrometry based proteomics



Velos Orbitrap



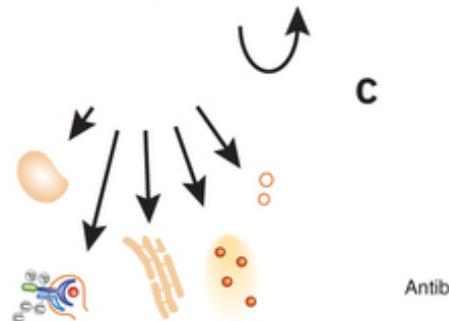
Orbitrap Fusion

Proteomics Workflow

a Cells or tissue



b

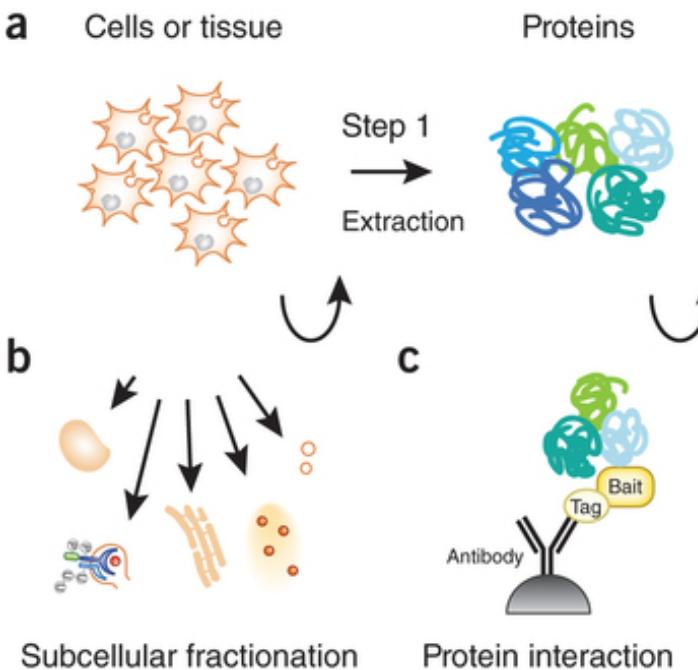


Prote

Considerations:

- Qualitative: what proteins are there?
- Quantitative: What differences in proteins or PTMs between conditions?
 - Different cell types
 - Kinase KO
 - Chemical perturbation
 - Etc.

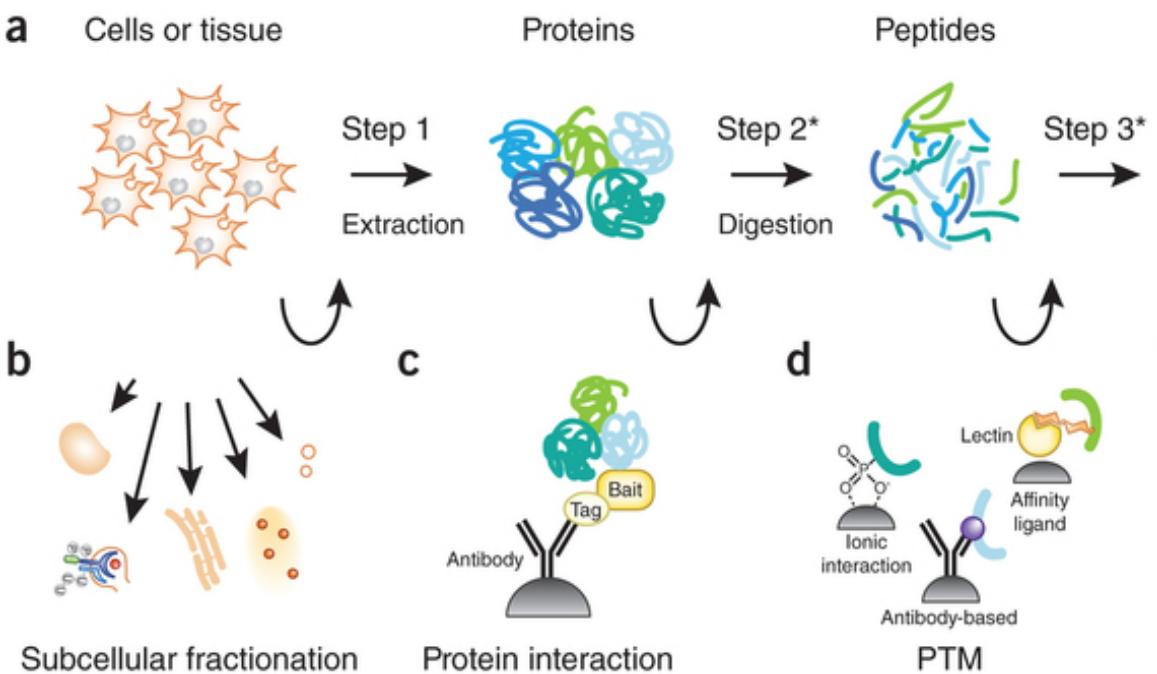
Proteomics Workflow



Considerations:

- Native or denaturing?
- Protein purification
- PTM stability

Proteomics Workflow



Considerations:

- PTM purification
- Fractionation to reduce complexity

Proteomics Workflow

a Cells or tissue



Step 1
→
Extraction

Proteins



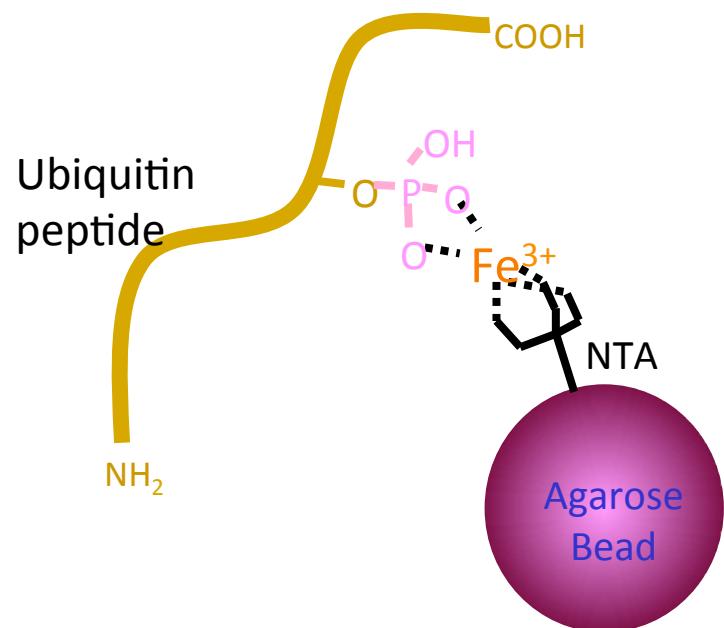
Peptides

Step 2*
→
Digestion

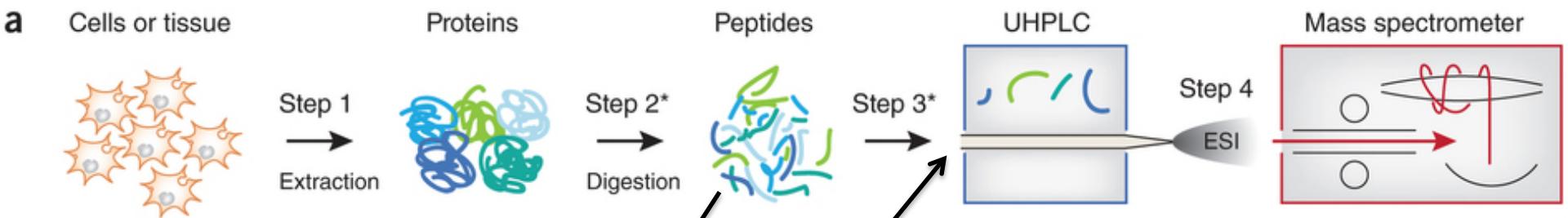


Step 3*
→

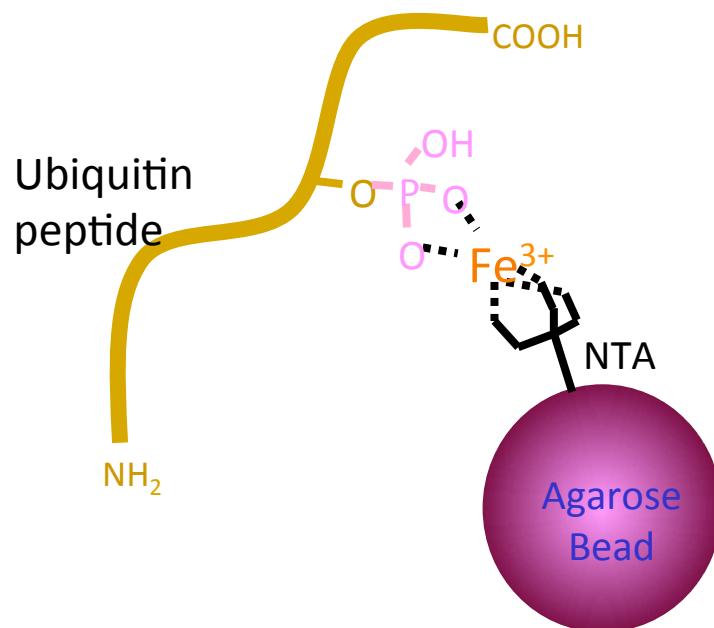
Phosphopeptide enrichment
Immobilized Metal Affinity Chromatography (IMAC)



Proteomics Workflow



Phosphopeptide enrichment
Immobilized Metal Affinity Chromatography (IMAC)



Proteomics Workflow

a Cells or tissue



Step 1
→
Extraction

Proteins



Step 2*
→
Digestion

Peptides



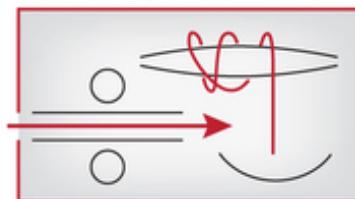
Step 3*
→

UHPLC

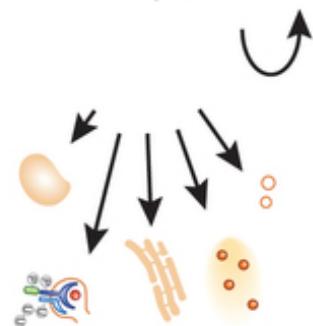


Step 4
→
ESI

Mass spectrometer



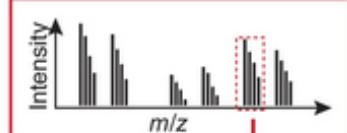
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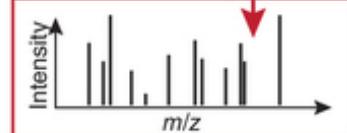
Subcellular fractionation



MS1



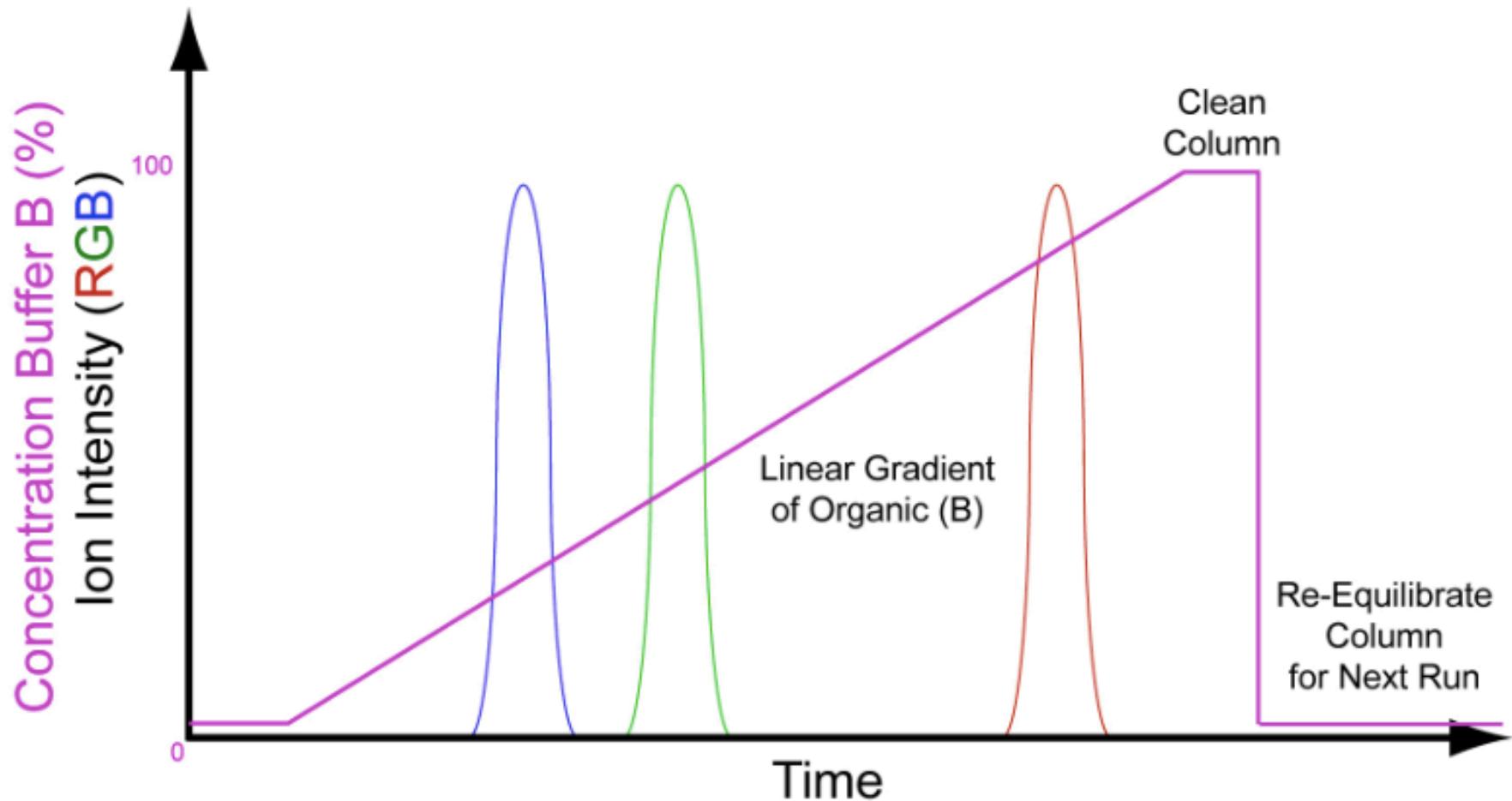
MS2



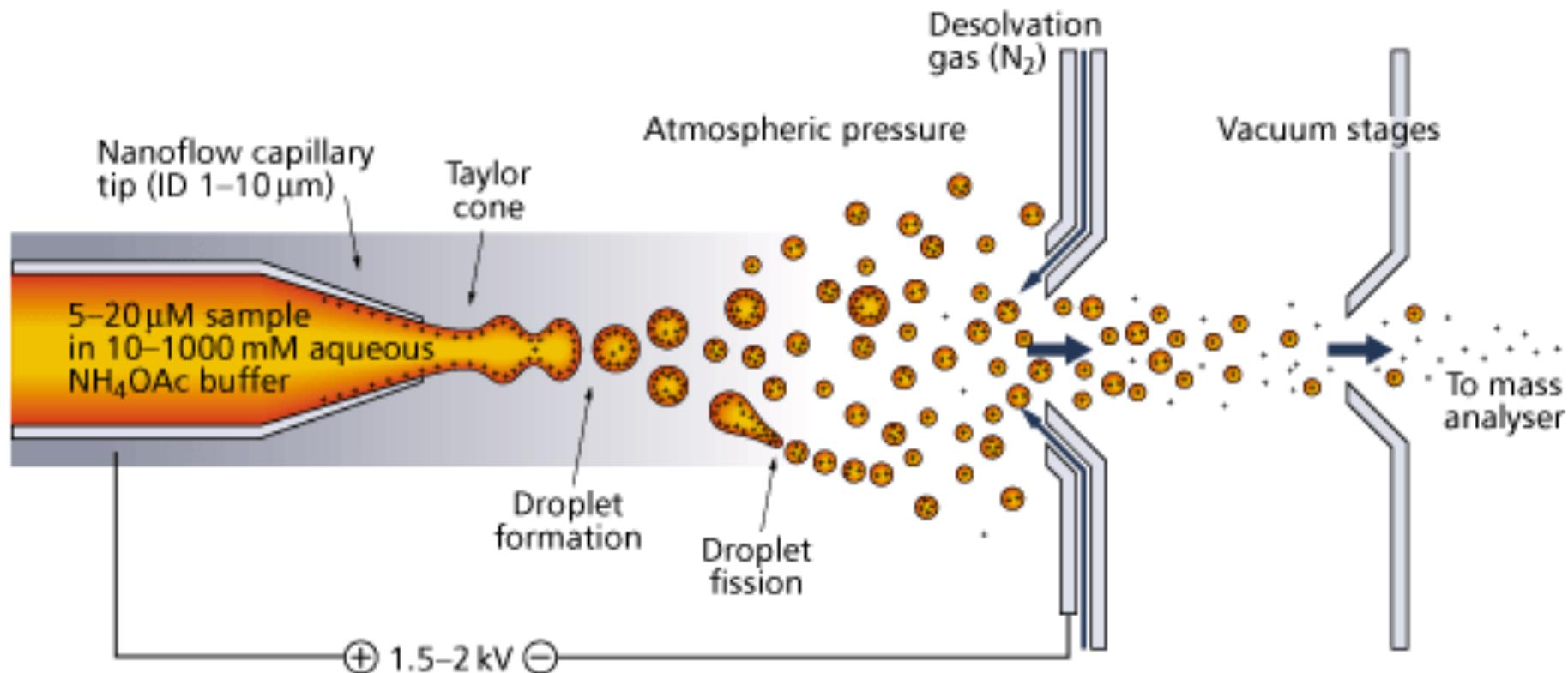
Reversed-Phase HPLC

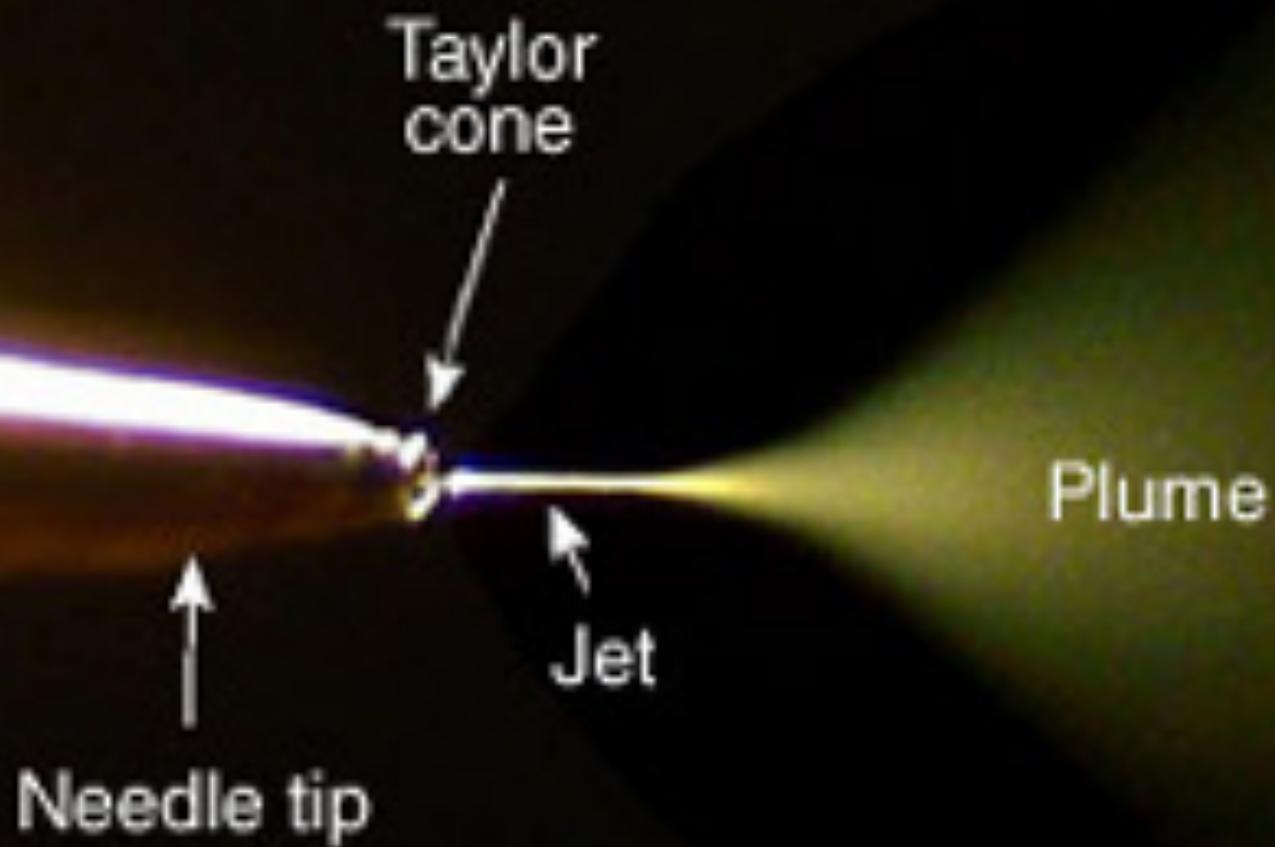
Load peptides: 100% water, 0.1% formic acid

Elute with increasing gradient of acetonitrile, 0.1% formic acid



Getting your sample into mass spec – electrospray ionization



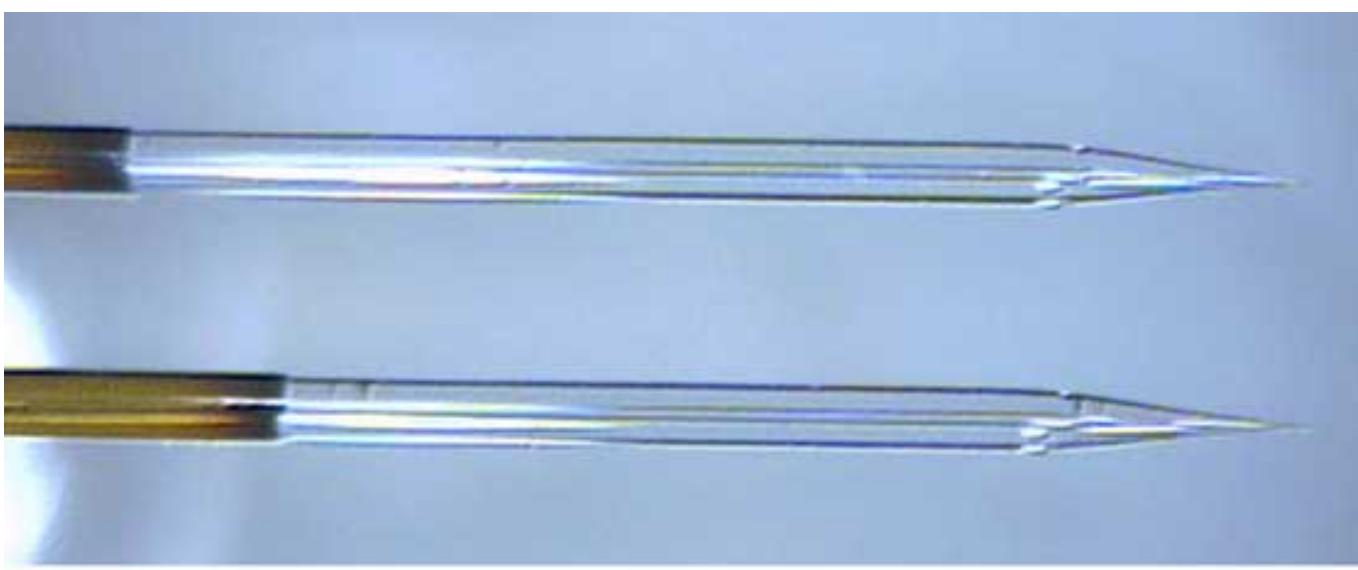


Taylor
cone

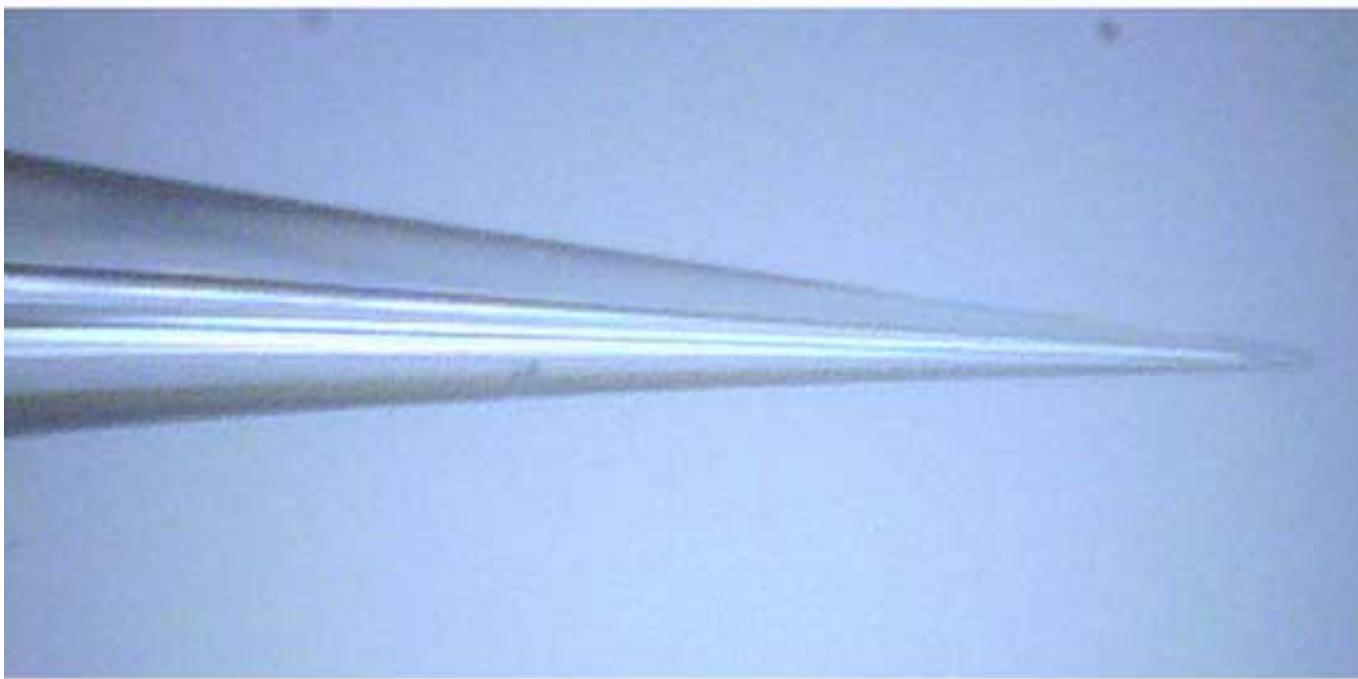
Jet

Plume

Needle tip

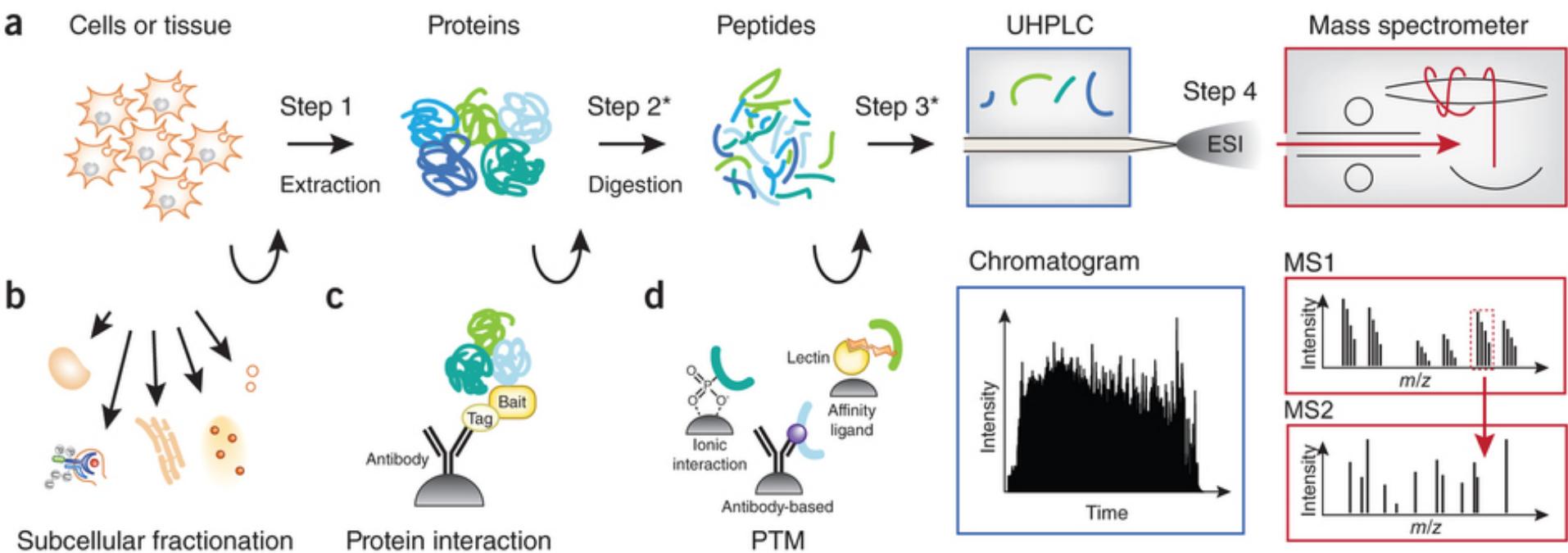


Electrospray Tip - 20x



Electrospray Tip - 400x

Proteomics Workflow



Considerations:

- Chemical nature of peptides of interest (PTM or un-modified)
- Complexity and dynamic range of mixture

RADIO FREQUENCY TWO DIMENSIONAL QUADUPOLE LINEAR ION TRAP

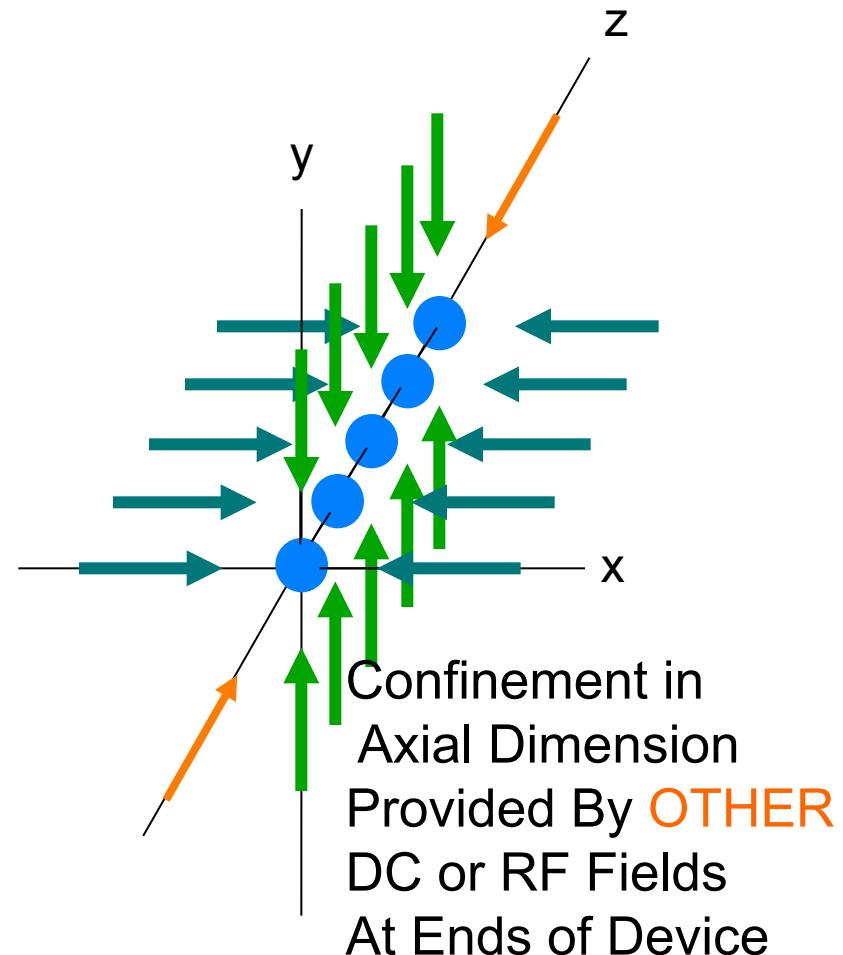
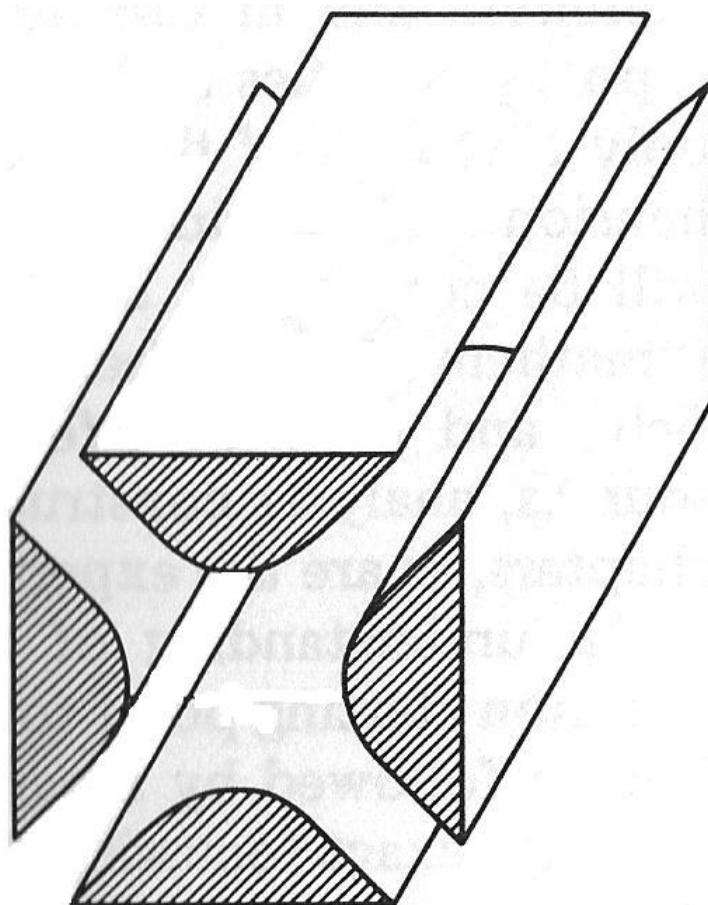
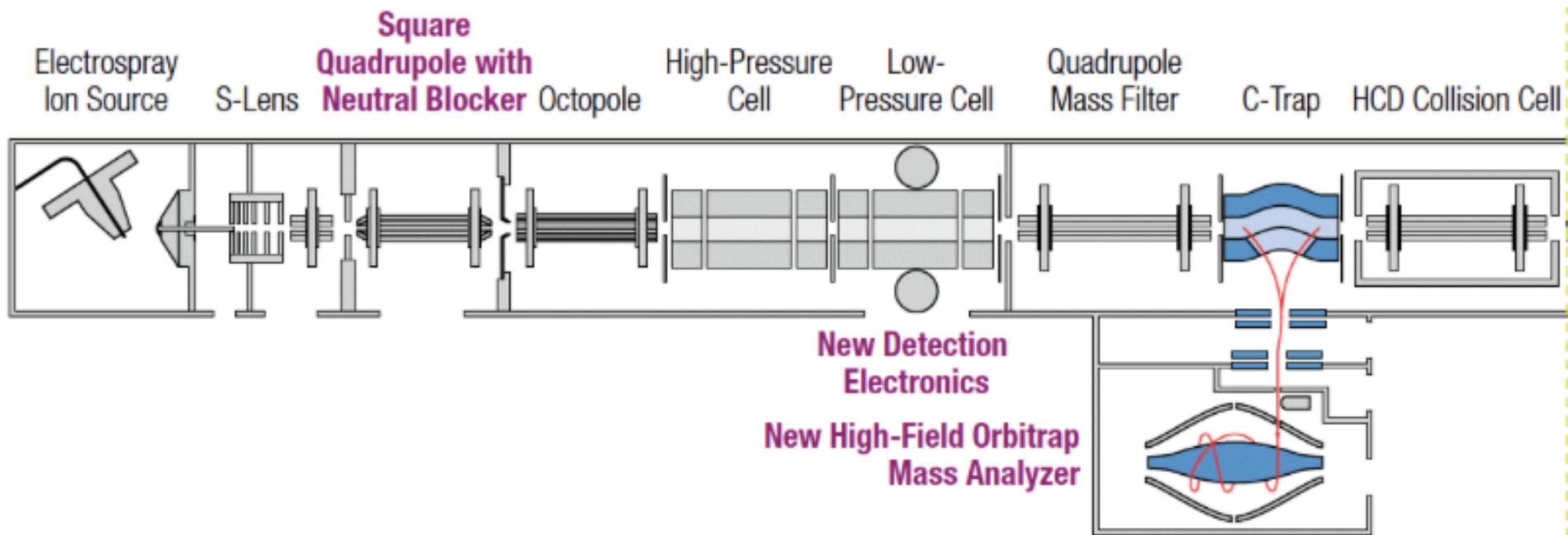


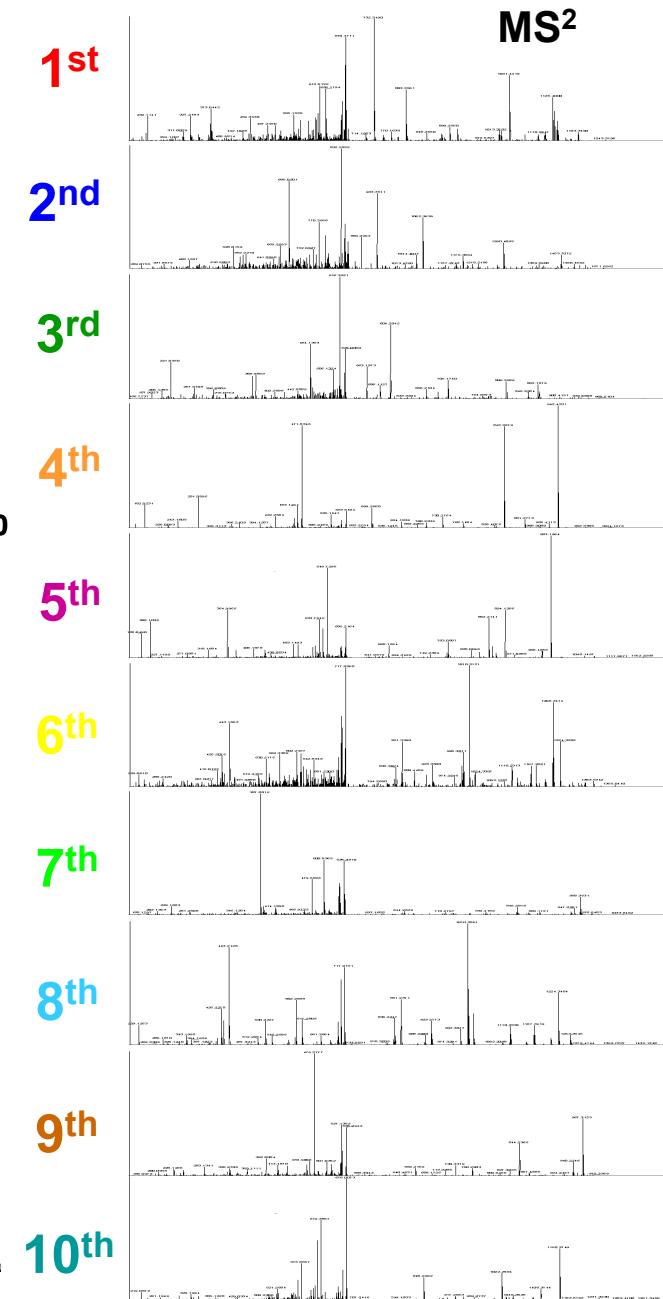
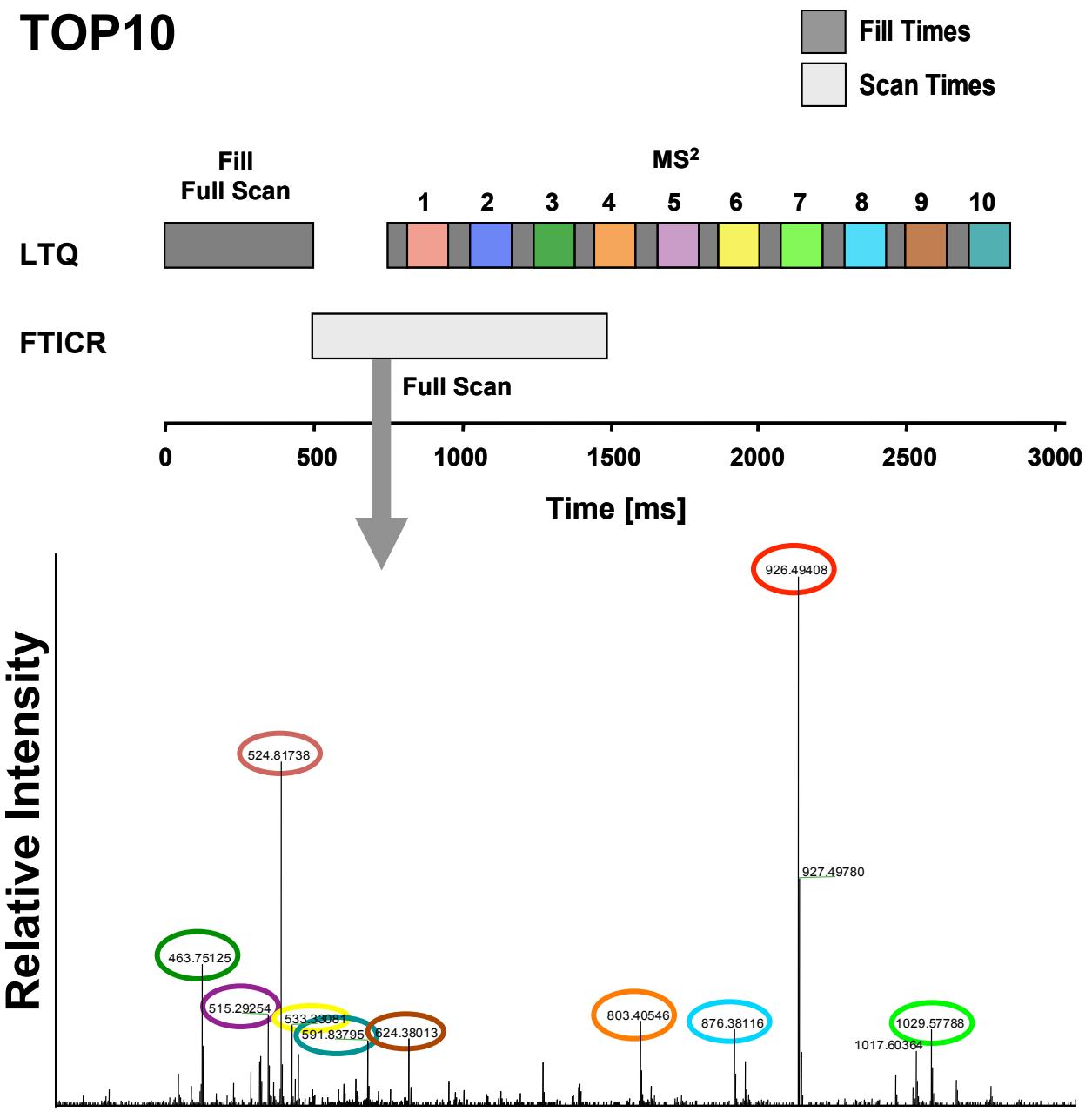
Figure From
Quadrupole Mass Spectrometry and Its Applications
P.H. Dawson Ed., Reprinted AIP Press 1995

Mass spec schematic and duty cycle

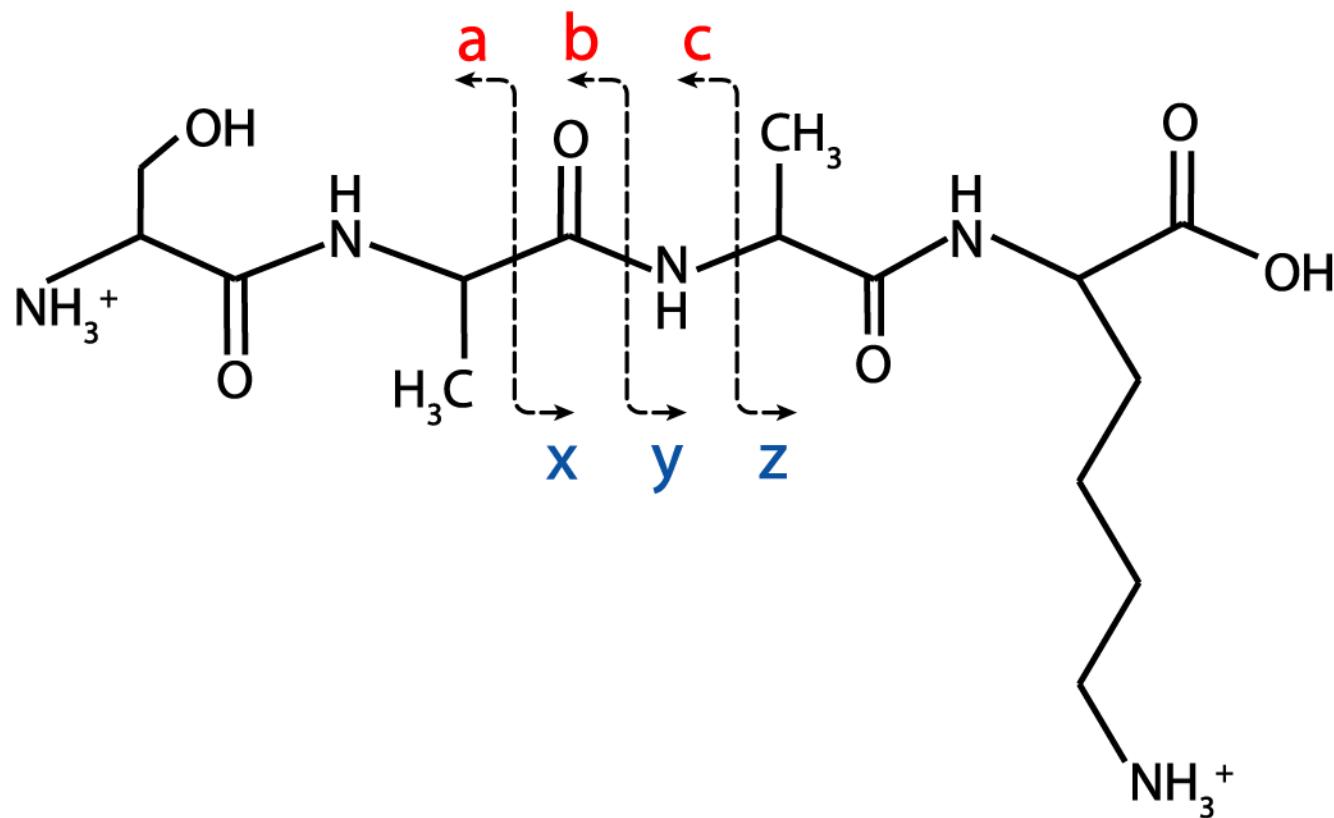


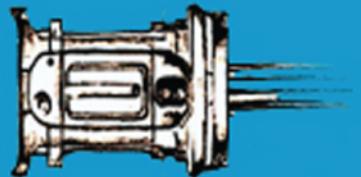
“shotgun sequencing”

TOP10



Fragmentation nomenclature

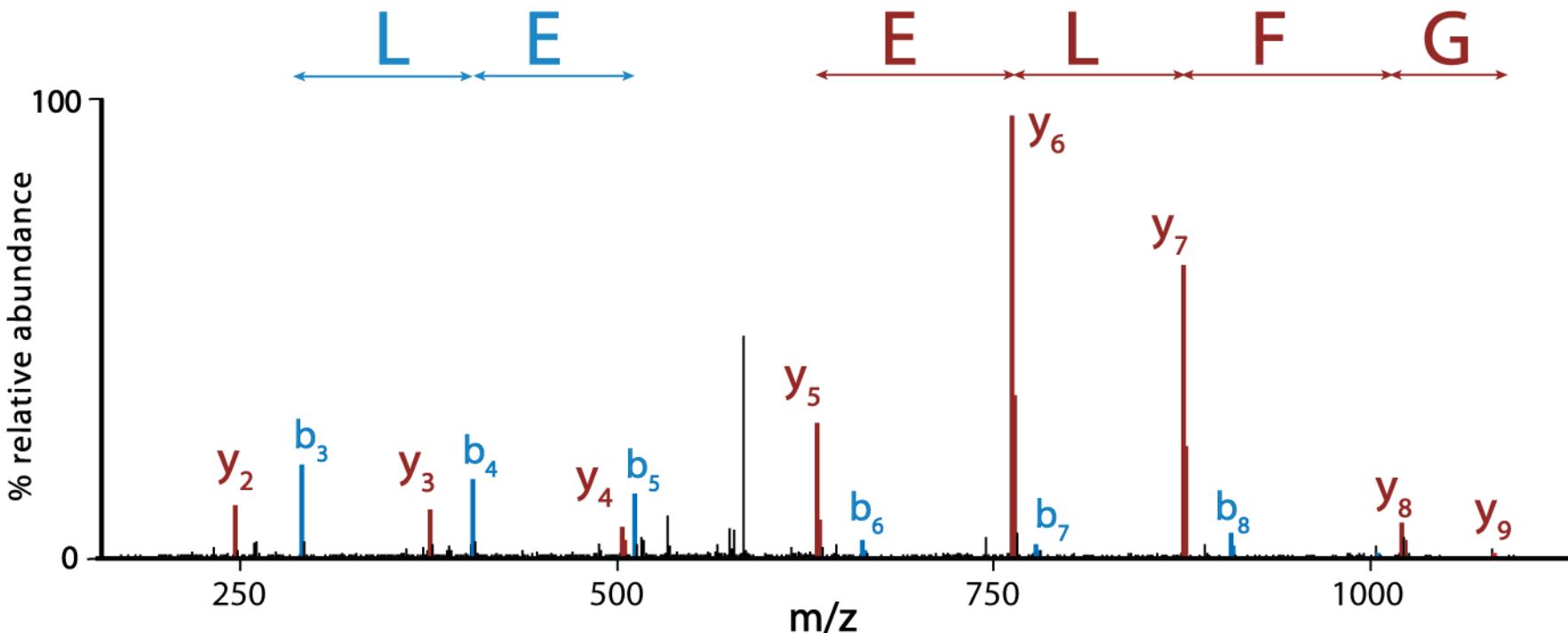




Peptide Sequencing (MS/MS)

collision-activated dissociation (CAD)

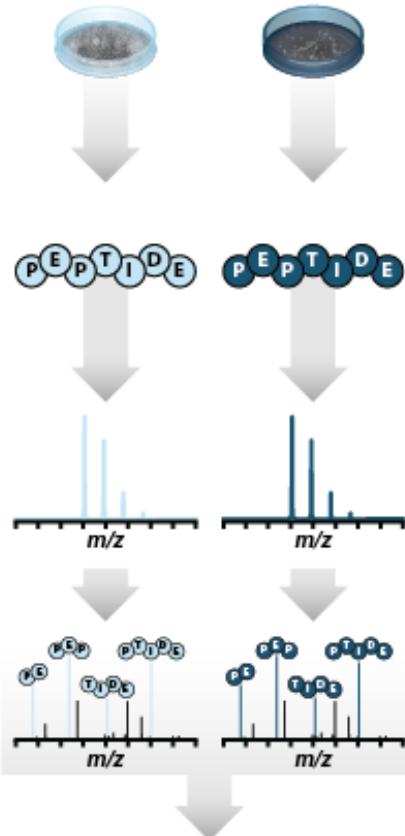
b ⁺	88	145	292	405	534	663	778	907	1020	1166
y ⁺	<u>1166</u>	<u>1080</u>	<u>1022</u>	<u>875</u>	<u>762</u>	<u>633</u>	<u>504</u>	<u>389</u>	<u>260</u>	147



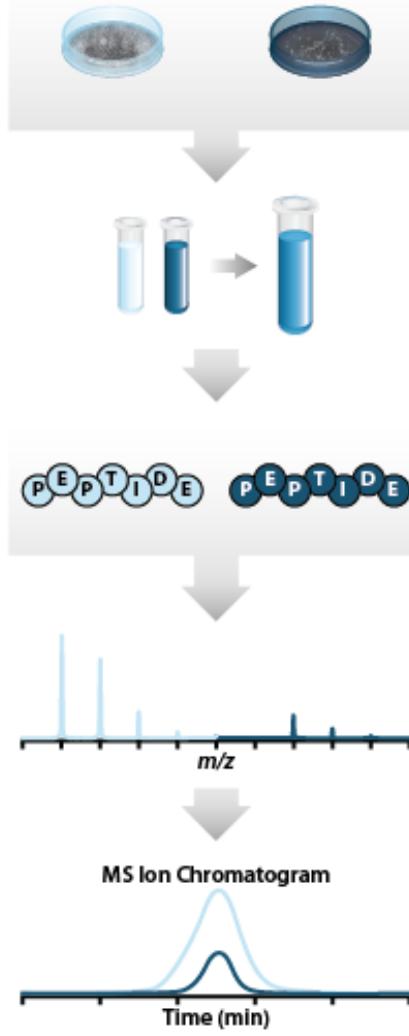
Mass spec operation animation:

Label-Based Quantitation

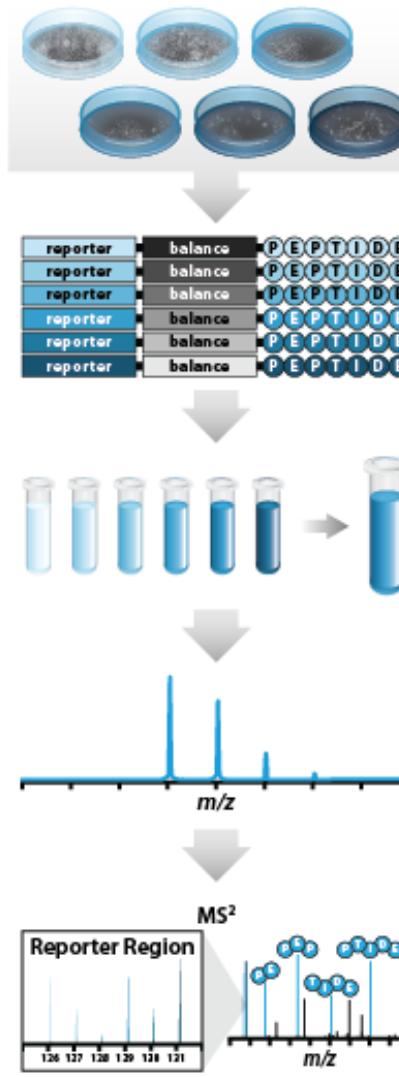
A) Label Free



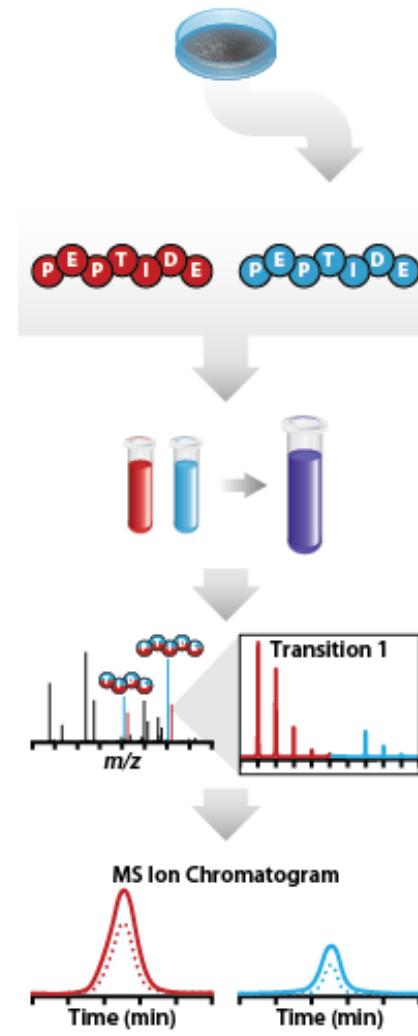
B) Metabolic labeling



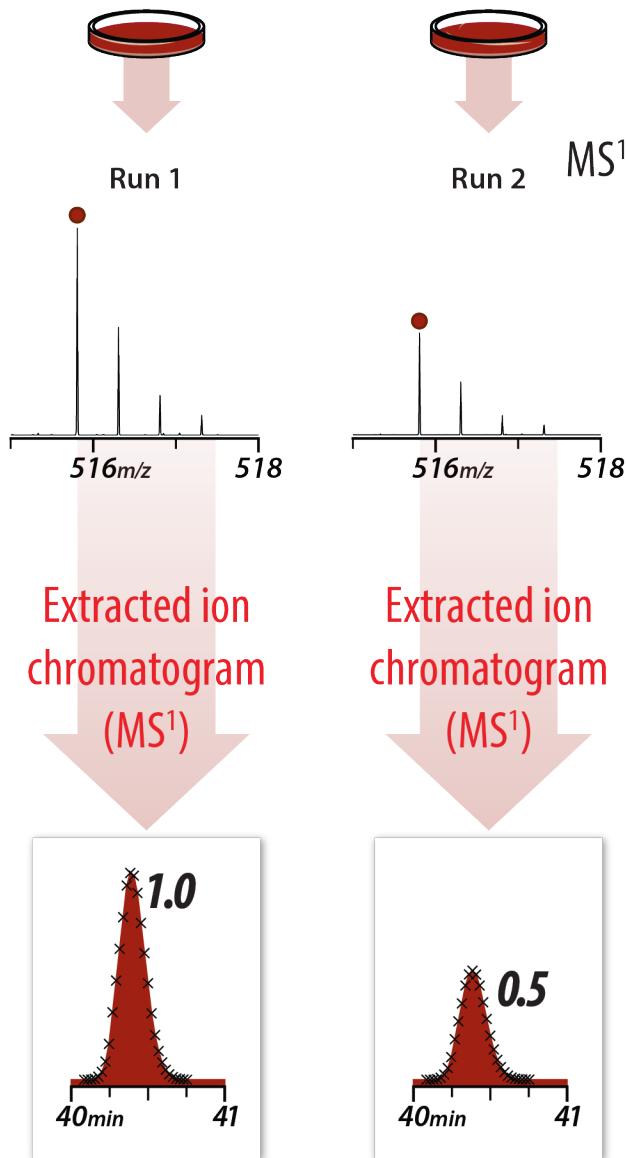
C) Isobaric tagging



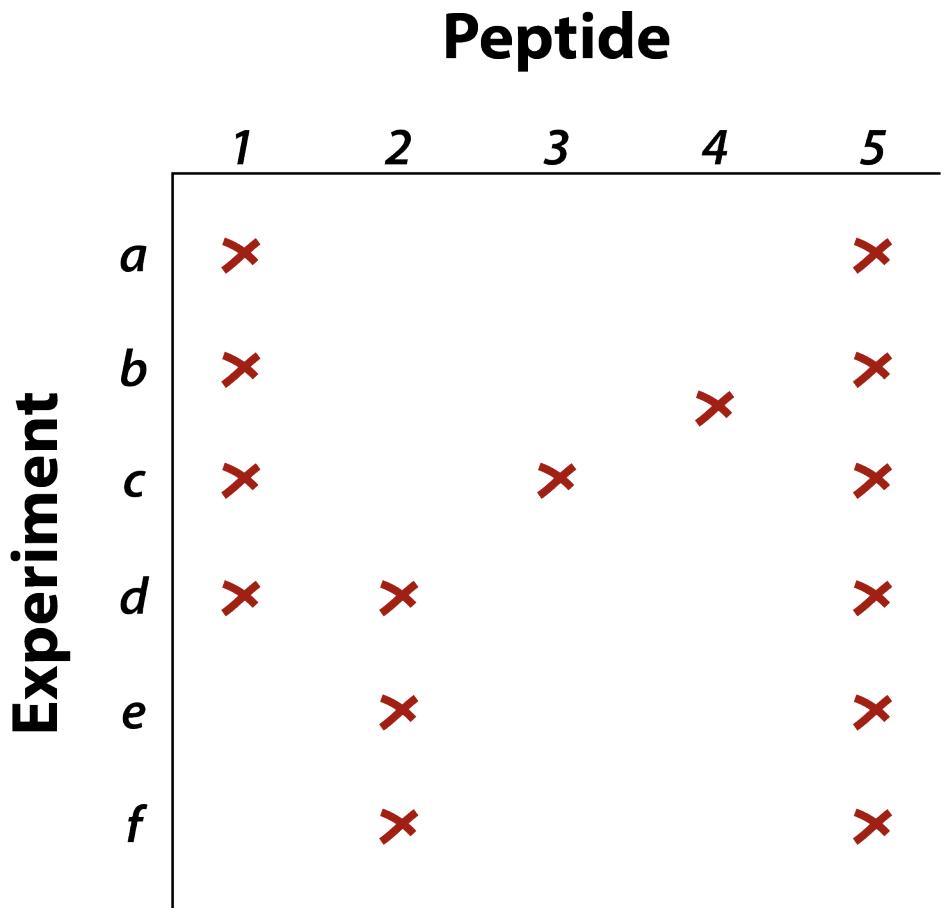
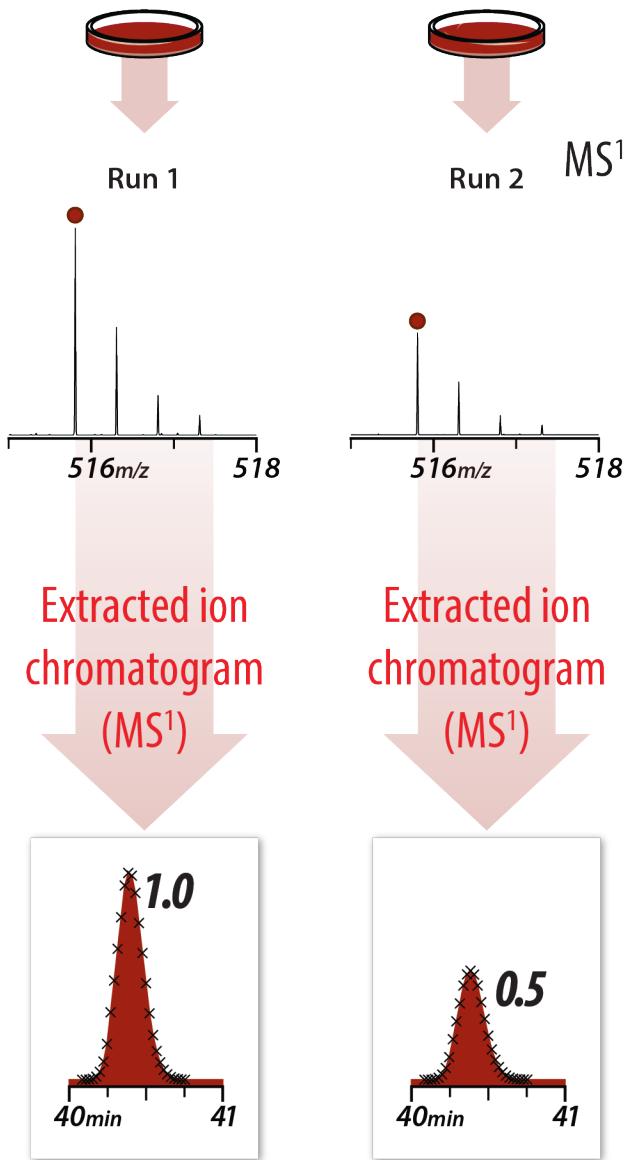
D) SRM



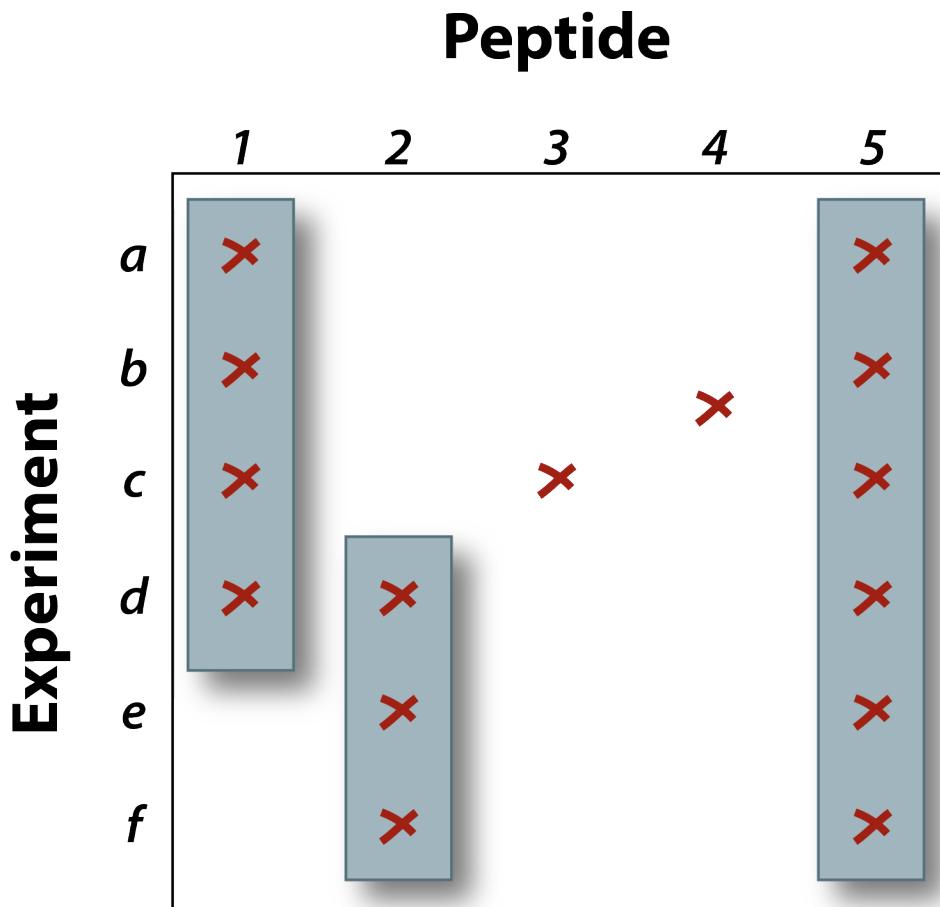
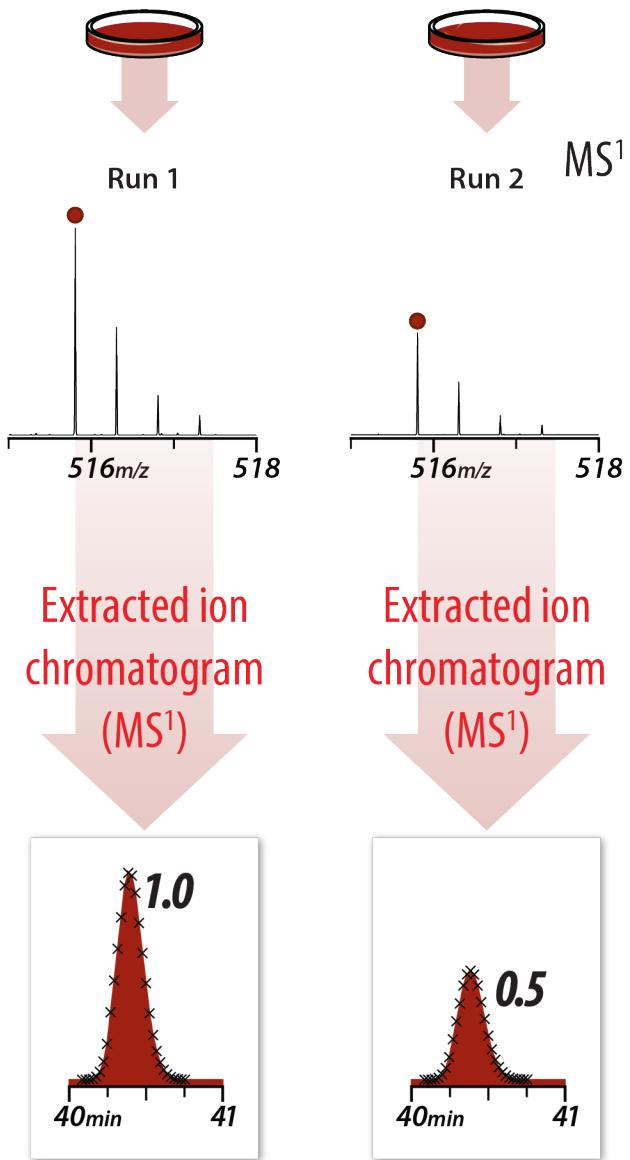
Label free quantitation AUC



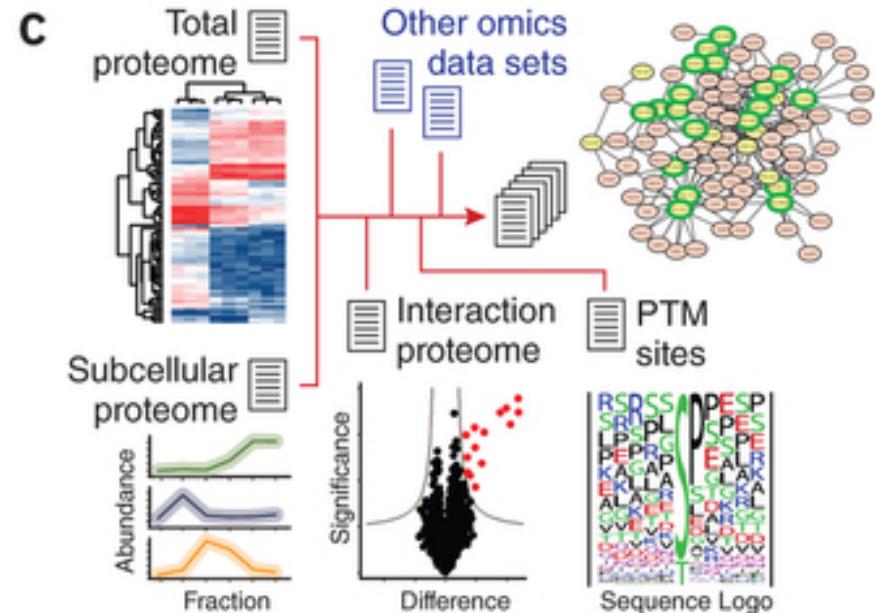
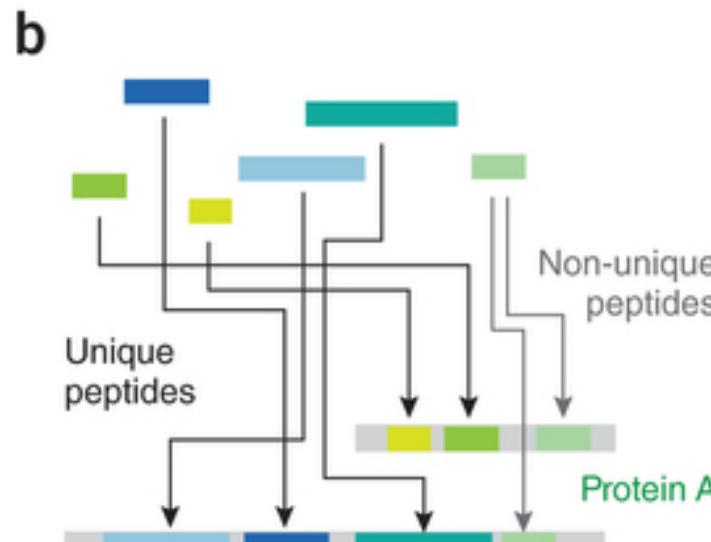
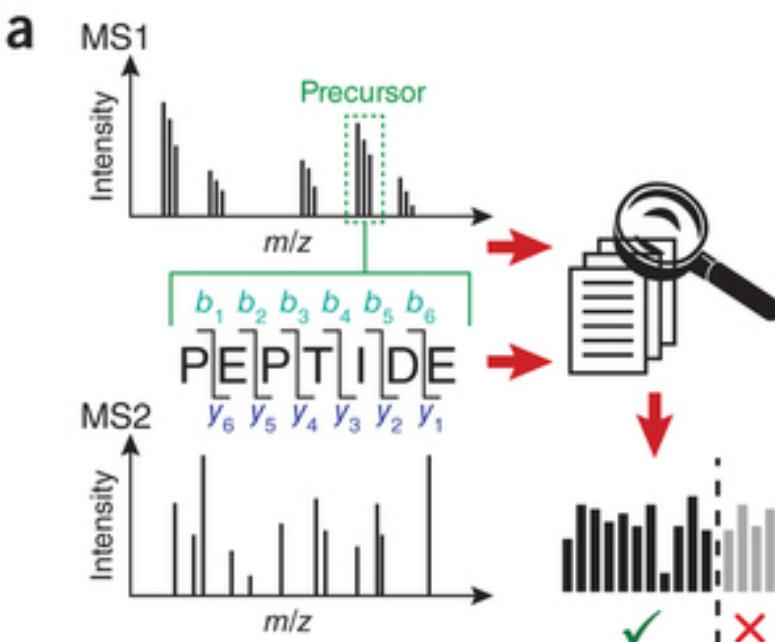
Label free quantitation AUC



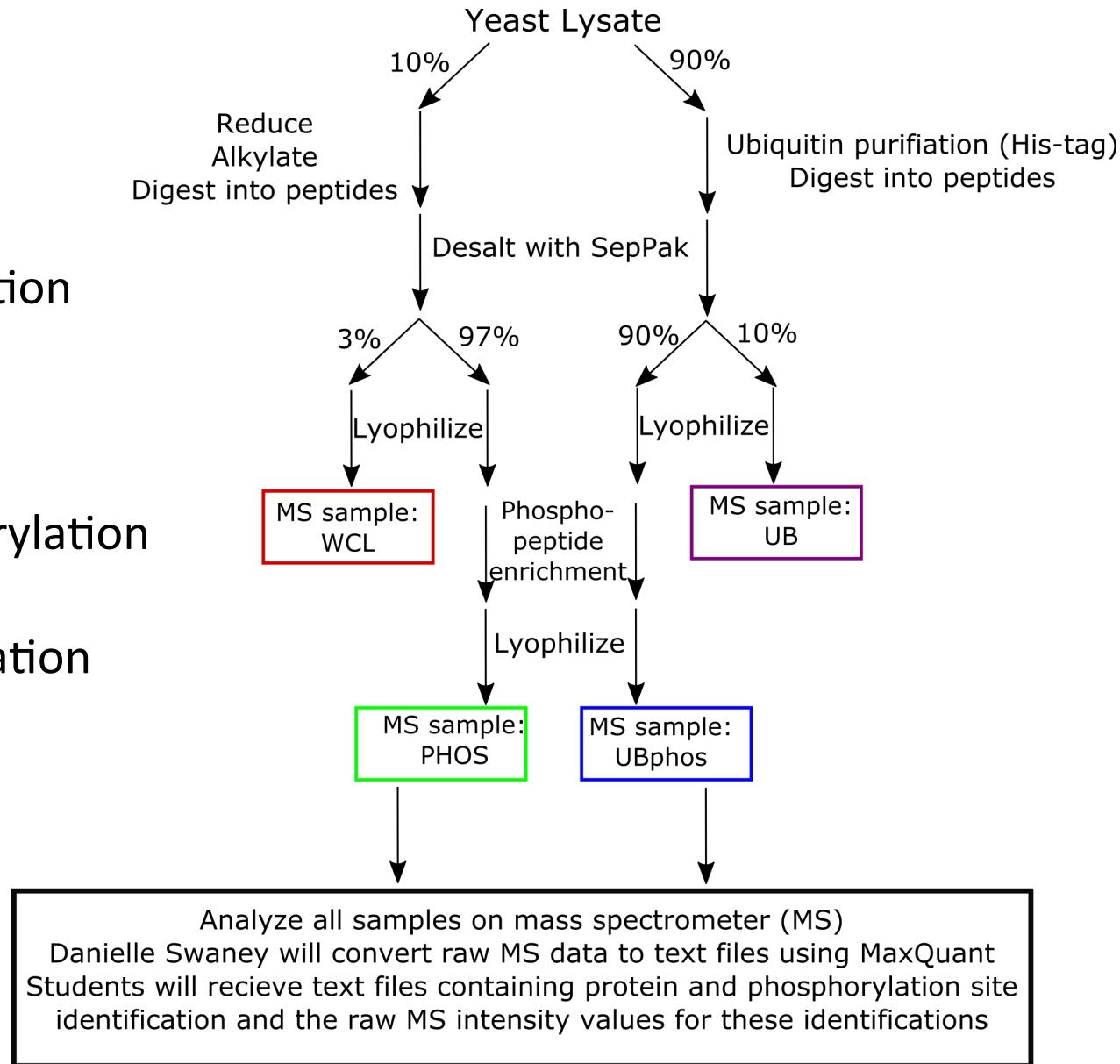
Label free quantitation AUC



What proteins did we identify? MaxQuant software

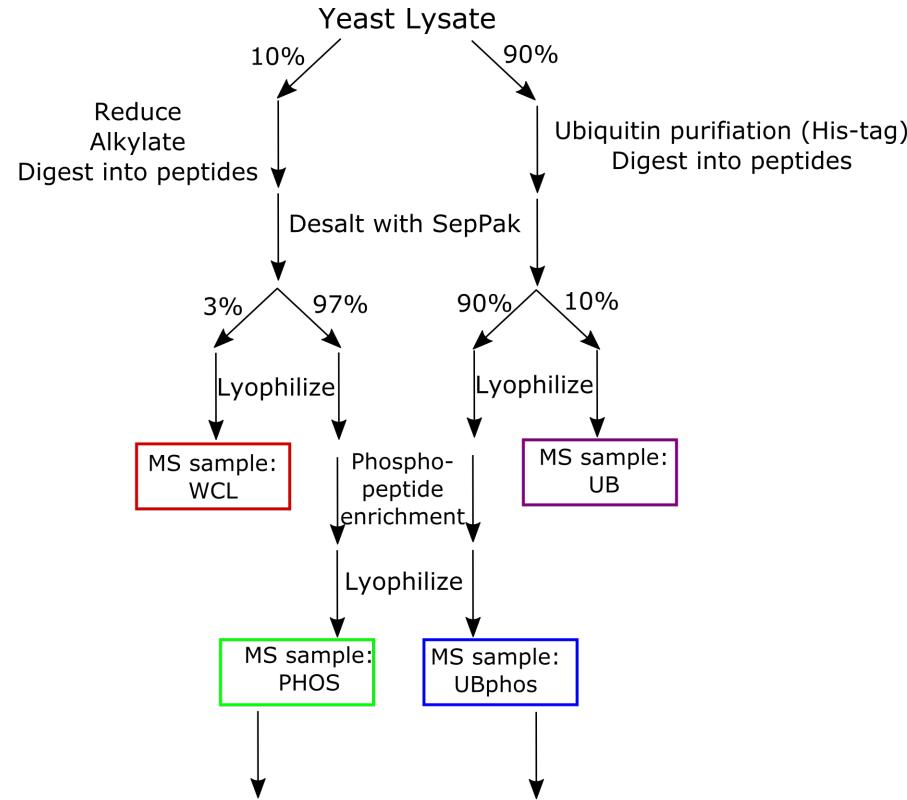


- Conditions:
 - Control
 - Kinase KO
 - Chemical perturbation
- Measurements:
 - Ubiquitin
 - Ubiquitin phosphorylation
 - Global proteome
 - Global phosphorylation



*****PRO TIPS*****

- You are purifying a **MINORITY** population from a complex mixture.
- Focus on **REMOVING** as much of what you don't want from the sample as possible
- Worry less about maintaining 100% of your analyte of interest.
- Understand where your sample is.



Analyze all samples on mass spectrometer (MS)
Danielle Swaney will convert raw MS data to text files using MaxQuant
Students will receive text files containing protein and phosphorylation site identification and the raw MS intensity values for these identifications